NIH-PA Author Manuscript

NIH-PA Author Manuscript

NIH-PA Author Manuscript

NIH Public Access
Author Manuscript
DNA Cell Biol. Author manuscript; available in PMC 2013 August 29.

Published in final edited form as:
DNA Cell Biol. 2006 June ; 25(6): 359–364. doi:10.1089/dna.2006.25.359.

Increased Recombination Between Active tRNA Genes

MATTHEW J. PRATT-HYATT[1,2], **KEVIN M. KAPADIA**[1], **THOMAS E. WILSON**[2,3], and **DAVID R. ENGELKE**[1,2]

[1]Department of Biological Chemistry, University of Michigan Medical School, Ann Arbor, Michigan

[2]Graduate Program in Cellular and Molecular Biology, University of Michigan Medical School, Ann Arbor, Michigan

[3]Department of Pathology, University of Michigan Medical School, Ann Arbor, Michigan

Abstract

Transfer RNA genes are distributed throughout eukaryotic genomes, and are frequently found as multicopy families. In *Saccharomyces cerevisiae*, tRNA gene transcription by RNA polymerase III suppresses nearby transcription by RNA polymerase II, partially because the tRNA genes are clustered near the nucleolus. We have tested whether active transcription of tRNA genes might also suppress recombination, since recombination between identical copies of the repetitive tRNA genes could delete intervening genes and be detrimental to survival. The opposite proved to be the case. Recombination between active tRNA genes was elevated, but only when both genes are transcribed. We also tested the effects of tRNA genes on recombination between the direct terminal repeats of a neighboring retrotransposon, since most Ty retrotransposons reside next to tRNA genes, and the selective advantage of this arrangement is not known.

INTRODUCTION

Most tRNA genes exist in multiple copies distributed throughout the genome, and appear to have duplicated in dispersed locations through RNA-mediated transposition (Hani and Feldmann, 1998; Dujon *et al*., 2004). Little is known about which predicted tRNA genes in multicellular eukaryotes are transcribed, but there are indications from a limited number of studies that there might be considerable developmental regulation in the transcription of tRNA gene subclasses (Koski and Clarkson, 1982; Wilson *et al*., 1985). In addition to tRNA genes, vertebrates have highly repetitive DNA elements, termed SINEs (short interspersed repetitive elements) that are derived from small RNA genes with tRNA-class promoters for RNA polymerase III (pol III) (Jurka, 2004). The major SINE in humans, the Alu elements, are originally derived from the 7SL RNA found in signal recognition particles, and are found in more than 500,000 copies per haploid genome (Gilbert and Labuda, 1999). Although few of these elements appear transcriptionally active under normal conditions, extensive Alu expression can be observed during viral infection or in response to cellular stress (Fornace and Mitchell, 1986; Liu *et al*., 1995), and most cloned Alu repeats can be transcribed by pol III *in vitro* (Elder *et al*., 1981; Liu and Schmid, 1993).

The existence of such frequent, highly similar DNA sequences in the genome raises the question of whether there is some mechanism to protect against frequent deletion of chromosome segments between repeats by homologous recombination, since such deletions

© Mary Ann Liebert, Inc.

Address reprint requests to: David R. Engelke, Ph.D., University of Michigan Medical School, 1150 W. Medical Center Drive, 3200 MSRB III, Ann Arbor, MI 48109-0606, engelke@umich.edu.

NIH-PA Author Manuscript
NIH-PA Author Manuscript
NIH-PA Author Manuscript

could lead to the death of single cells or developmental abnormalities in complex organisms. In adults, such deletions can lead to unregulated growth, such as human mammary tumors with deletions between Alu elements at the BRCA1 loci (Rohlfs *et al.*, 2000; Pavlicek *et al.*, 2004; Tournier *et al.*, 2004).

Transcription of genes by RNA polymerase II (pol II) increases their susceptibility to recombination (Smith *et al.*, 1996), but this has not been addressed for pol III transcription units, which are very short and almost entirely covered by the active transcription complex when present. Although transfer of genetic information between identical pol III genes is possible (e.g., tRNA) (Munz *et al.*, 1982), recombination that deletes intervening material might be predicted to be suppressed as being counter to survival. Here we use a yeast model system to address the effects of pol III transcriptional activity on recombination between and near tRNA genes in the yeast chromosome. We examined the rate of recombination between two nearly identical tRNA genes in the yeast *Saccharomyces cerevisiae* that were either transcriptionally active or inactive. The results were unexpected, in that recombination increased when both tRNA genes were active.

In addition to testing the effect of pol III transcription on recombination between tRNA genes, there was reason to suspect that homologous recombination within a few hundred base pairs of active tRNA genes might also be suppressed. Most of the Ty retrotransposons in yeast (Ty1–Ty4) have evolved mechanisms for preferential insertion and retention near tRNA genes (Chalker and Sandmeyer, 1992; Hull *et al.*, 1994; Kendall *et al.*, 2000; Bolton and Boeke, 2003). A rare fifth class, Ty5, prefers transcriptionally silent regions such as telomeres and silent mating type loci (Zou *et al.*, 1995). Proximity to a tRNA gene might have the selective advantage of conferring conditional pol II transcriptional silencing (Hull *et al.*, 1994), but reduced recombination between the direct, long terminal repeats (LTRs) of the Ty retrotransposons might also have provided the selective advantage for the Ty elements to have developed this insertion preference.

MATERIALS AND METHODS

Yeast strains

Strains were derived from *Saccharomyces cerevisiae* W3031A (MAT a *leu*2–3, 112 *his*3–11, 15 *ade*2–1 *trp*1–1 *ura*3–1 *can*1–100) and BY4741 (MATa his3Δ1 leu2Δ0 met15Δ0 ura3Δ0). Growth was performed in YPD (Rose *et al.*, 1990) except where noted. For testing recombination between identical $tRNA^{Leu}$ genes, constructs were made in W3031A, the coding sequence of the *LEU2* gene, on chromosome III, adjacent to the *SUP53* $tRNA^{Leu}$ gene was precisely replaced with the coding region of *URA3*. A second copy of the *SUP53* gene was introduced 200 bp downstream from the 3′ end of the *URA3* coding region, 1.3 kb downstream from the first tRNA gene and in the same orientation. The insertion contained *SUP53* sequence from 59 bp upstream to 29 bp downstream of the tRNA coding sequence. The strains contained either a wild-type $tRNA^{SUP53}$ promoter at each site or a transcriptionally inactive promoter containing previously characterized point mutations in both the A box and B box (Newman *et al.*, 1983; Hull *et al.*, 1994). The mutated gene is designated *sup53*ΔAB (Fig. 1A).

To test recombination rates near a tRNA gene, *URA3* was inserted at the Ty3–1 locus on chromosome VII in strain BY4741, (YGRWTy3–1) replacing all but 100 bp of each end of the central epsilon region. A second strain was created by then mutating the neighboring $tRNA^{Cys}$ gene promoter (triple B box mutation G52A, T53A, C55G) by homologous recombination of a PCR product with integration selected by G418 resistance and confirmed by sequencing genomic PCR fragments from the strains. The Kan gene was inserted 152 bp

NIH-PA Author Manuscript

NIH-PA Author Manuscript

NIH-PA Author Manuscript

downstream of the tRNA gene in the opposite orientation in the strain with and without mutations to the neighboring $tRNA^{Cys}$.

PCR fragments for deletions of *RAD51* and *RAD52* were produced by replacement of their coding regions with the *KAN* cassette from the plasmid pFA6a–KanMX6 (Longtine *et al.*, 1998).

Recombination assay

For each experiment three cultures were inoculated from individual colonies and were grown in SDC-Ura (Rose *et al.*, 1990) medium to ensure retention of the *URA3* genes. These cultures were used to inoculate 200 ml of YPD cultures to a starting OD600 of 0.2 and grown for 7 h to allow loss of *URA3*. Cells were collected and resuspended in 10 ml of 10 mM Tris-HCl pH 7.5, 1 mM EDTA. Cells are plated on SDC + 5-fluorootic acid (5-FOA) medium, to select against *URA3*. The number of 5-FOA-resistant colonies is expressed relative to the number of cells plated.

To test whether 5-FOA resistance arose from homologous recombination to delete *URA3*, 24 to 32 5-FOA-resistant colonies were tested in two or more separate experiments by PCR of genomic DNA from flanking primers.

RESULTS

Recombination between identical tRNA genes

Since many tRNA genes in yeast are duplicated up to 15 times throughout the genome, suppression of recombination between duplicates could have survival value. The question addressed here is whether transcription of tRNA genes by RNA polymerase III affects the recombination rate between identical tRNA genes. We modified the *LEU2* locus on chromosome III as shown in Figure 1A. The $tRNA^{SUP53}$ gene is normally located upstream of the *LEU2* gene, and its transcription has been extensively characterized (Newman *et al.*, 1983; Huibregtse and Engelke, 1989). The coding sequence of *LEU2* was precisely replaced with that of *URA3*, along with inserting a second copy of the $tRNA^{SUP53}$ gene (*SUP53*) 200 base pairs downstream from *URA3*. Strains were produced containing either active tRNA genes or tRNA genes inactivated by point mutations in the internal promoters ($G_{19}C$, $C_{56}G$; *sup53*ΔAB) that prevent formation of any part of the RNA polymerase III complex (Newman *et al.*, 1983; Huibregtse *et al.*, 1989). The four tested constructs had either both tRNA genes active (*SUP53/SUP53*), both inactive (*sup53*ΔAB/*sup53*ΔAB), or only one at a time active.

Loss of *URA3* function was selected on media containing the 5-FOA, and the normalized rate of number of 5-FOA-resistant colonies is shown in Figure 1B. These data are averages from five different experiments, each done in duplicate.

To determine if 5-FOA resistance arose through deletion between the tRNA genes, PCR was done on genomic DNA from independent isolates using primers flanking the tRNA genes. The starting strains produce a 1.8-kb product spanning both tRNA genes and the *URA3* gene, whereas precise homologous recombination between the tRNA repeats produces a 175-bp product. Sixteen 5-FOA resistant colonies from two separate experiments on each strain all showed a 175-bp product, consistent with a precise deletion between sequences in the tRNA genes.

In contrast to our original hypothesis that recombination would be repressed between transcribed tRNA genes, the highest homologous recombination rate was found to be the one where both $tRNA^{SUP53}$ genes were active. This strain had a recombination rate

NIH-PA Author Manuscript

NIH-PA Author Manuscript

NIH-PA Author Manuscript

averaging five times greater than the other three strains in four separate experiments. It is particularly interesting that inactivating either tRNA gene gave the same recombination as inactivating both. The failure of only a single active tRNA transcription unit to stimulate recombination implies that the increase in recombination between active tRNA genes is the result of some direct or indirect communication between components of active pol III complexes on both genes.

Recombination path

Single-strand annealing (SSA) is one of the common mechanisms leading to deletions between direct repeats. When a break occurs between the two repeats the 5′ ends are resected. This allows the 3′ strands to anneal within the repeated elements, followed by removal of the nonhomologous 3′ tails corresponding to the sequence between the repeats, gap filling, and ligation (Liu and Schmid, 1993; Prado *et al.*, 2003; Tournier *et al.*, 2004;). Deletions caused by recombination between direct repeats of untranscribed regions have been shown to be reduced 10- to 100-fold in *rad52* mutants consistent with the role of Rad52p in SSA (Jackson and Fink, 1981; Prado and Aguilera, 1995). In contrast, the *rad52* mutation did not affect deletion rates of direct repeats in rDNA and high-copy *CUP1* tandem arrays (Ozenberger and Roeder, 1991). Another participant in homologous recombination, Rad51p, acts by promoting pairing and strand exchange with an intact homologous duplex. Rad51p is required for most recombination but dispensable for SSA (Ozenberger and Roeder, 1991). To examine whether recombination between the tRNA genes was proceeding through a SSA path, *RAD52* and *RAD51* were individually deleted from our strains with either two active or two inactive tRNA genes flanking the *URA3* gene.

The *rad52*Δ strains consistently showed reduced 5-FOA resistant colonies. In the *SUP53/SUP53* strain there was a 20-fold decrease, and in the *sup53*ΔAB/*sup53*ΔAB strain there was a 33-fold decrease in homologous recombination (Table 1). This sensitivity to *rad52* deletion distinguishes recombination between these direct repeats transcribed by pol III from the rDNA and *CUP1* repeats transcribed by RNA polymerases I and II. In contrast, *rad51*Δ caused little change in the number of 5-FOA resistant colonies in the *SUP53/SUP53* strain, and caused a slight increase in 5-FOA resistance in *sup53*ΔAB/*sup53*ΔAB strains (Fig. 1B). This is consistent with previous demonstrations that recombination between direct repeats can occur in the absence of strand exchange (McDonald and Rothstein, 1994; Rattray and Symington, 1995; Ivanov *et al.*, 1996). These results suggest that recombination between tRNA genes is occurring in the absence of strand exchange regardless of their transcriptional activity, with Rad52p facilitating SSA.

To test whether the residual 5-FOA resistant colonies arose through recombination between the tRNA genes or some other defect, we again tested genomic numerous 5-FOA isolates by PCR. This analysis showed that only 39% of the *SUP53/SUP53* and 5% of the *sup53*ΔAB/*sup53*ΔAB 5FOA-resistant colonies underwent deletion by homologous recombination in the Δ*rad52* strain, compared to 100% in *RAD52* strains. In the *SUP53/SUP53–*Δ*rad52* strain 22% of the 5FOA resistant colonies came through a probable *URA3* mutation, since deletions were not detected, and 39% appear to have a large chromosomal deletion in the region. In the *sup53*ΔAB/*sup53*ΔAB-Δ*rad52* strain most (95%) of the 5-FOA-resistant colonies represented a probable small mutation in *URA3* (Table 1).

Recombination between retrotransposon LTR elements

We also tested the effect of a nearby tRNA gene on the recombination between LTR elements of a well-characterized yeast retrotransposon locus Ty3–1, in its native chromosomal position on chromosome VII (Bilanchone *et al.*, 1993). We replaced the interior epsilon region of the retrotransposon Ty3–1 with *URA3*. We then either left the

NIH-PA Author Manuscript NIH-PA Author Manuscript NIH-PA Author Manuscript

neighboring tRNA^{Cys} gene intact or mutated three base pairs in the B box internal promoter (Fig. 2), which dramatically decreases pol III transcription (Allison *et al.*, 1983; Kinsey and Sandmeyer, 1991; Aguilera, 2002).

The strains with active versus inactive tRNA genes were tested for their relative frequency of *URA3* loss. The averages from four separate experiments demonstrated that the frequencies for the active tRNA gene and inactive gene were essentially identical (9.5×10^{-6} and 9.0×10^{-6}, respectively) within error for these experiments. Recombination between the LTR direct repeats therefore appears to be unaffected by the presence of a nearby active tRNA gene.

DISCUSSION

Contrary to our original hypothesis, the work presented here showed that the occurrences of recombination between two identical tRNA genes is higher when both tRNA genes are being actively transcribed. In this respect, pol III transcription appears to behave similarly to pol II transcription in that transcription stimulates recombination (Aguilera, 2002). However, there are also distinct differences. It is unclear why both tRNA genes needed to be actively transcribed in order for the increase in homologous recombination to occur. One possibility is having two active tRNA genes causes an increase in breakage, consistent with tRNA genes causing replication fork pause (Deshpande and Newlon, 1996), which have been shown to cause increased homologous recombination (Bilanchone *et al.*, 1993; Aguilera, 2002). If this is the case, it is not clear why a single active tRNA gene might not give an intermediate level of recombination increase. A second possibility is that having two active tRNA genes changes the type of repair that is used. If this is the case, perhaps the spatial organization of these tRNA genes is playing a part in their repair. However, the *rad52*Δ data suggest that the path of this increased recombination is still through SSA.

Recent findings showing spatial clustering of the linearly dispersed tRNA genes suggest a possible explanation for our results. Yeast tRNA genes are largely localized to the nucleolus when actively transcribed, but not when inactivated by promoter point mutations (Thompson *et al.*, 2003). The mechanism of this colocalization is not currently known, but it is conceivable that tRNA transcription complexes associate with some sort of framework that brings them into proximity, increasing the likelihood of physical interactions. This would be consistent with evidence that the frequency of disease-specific chromosomal translocations are nonrandom, and have been correlated to the spatial proximity of the sites involved (Lukasova *et al.*, 1997; Neves *et al.*, 1999; Roix *et al.*, 2003).

The increased recombination between identical tRNA genes when they are transcriptionally active could help explain why families of tRNA genes in yeast are not found tandemly repeated, or even in close proximity. The closest pair of identical tRNA genes occurs on chromosome IX, where two tRNA genes Asp are about 12 kb apart from each other. Studies using plasmid constructs and non-tRNA repeats have shown that increasing the distance between repeats decreases the efficiency of SSA in competition with gene conversion (Fishman-Lobell *et al.*, 1992; Haber and Leung, 1996; Puget *et al.*, 1999).

The results presented here might be pertinent in considering the occurrence of deletions, inversions, and duplications at Alu elements throughout the human genome. Alu elements contain tRNA-class internal promoters from the 7SL RNA genes (Gilbert and Labuda, 1999). While this pol III promoter in cloned Alu repeats is generally found to be transcriptionally competent in vitro, most of the hundreds of thousands of Alu repeats in human cells are inactive in cells, and little is known as to which of the elements are activated in response to cellular insults (Fornace and Mitchell, 1986; Liu *et al.*, 1995; Gilbert

NIH-PA Author Manuscript NIH-PA Author Manuscript NIH-PA Author Manuscript

and Labuda, 1999). An example of recombination between Alu repeats has been studied is the human tumor suppressor gene, BRCA1. BRCA1 genomic sequence is composed of 41.5% Alu sequence, corresponding to an Alu element every 650-bp average (Smith *et al.*, 1996). Alu recombination seems to be the main source of genomic rearrangements in patients with a hereditary predisposition to breast and ovarian cancers (Puget *et al.*, 1999; Rohlfs *et al.*, 2000; Pavlicek *et al.*, 2004). The data in this report suggests that the transcriptional activation of these Alu repeats, possibly through stress response or viral infection, might increase their ability to recombine.

Acknowledgments

This work was supported by the National Institutes of Health (GM063142 to D.R.E. and CA102563 to T.E.W.). M.J.P.-H. is a predoctoral fellow supported by the Cellular and Molecular Biology Training Grant at the University of Michigan (T32 GM007315).

References

AGUILERA A. The connection between transcription and genomic instability. EMBO J. 2002; 21:195–201. [PubMed: 11823412]

ALLISON DS, GOH SH, HALL BD. The promoter sequence of a yeast tRNA tyr gene. Cell. 1983; 34:655–664. [PubMed: 6352054]

BILANCHONE VW, CLAYPOOL JA, KINSEY PT, SANDMEYER SB. Positive and negative regulatory elements control expression of the yeast retrotransposon Ty3. Genetics. 1993; 136:685–700. [PubMed: 8394262]

BOLTON EC, BOEKE JD. Transcriptional interactions between yeast tRNA genes, flanking genes and Ty elements: A genomic point of view. Genome Res. 2003; 13:254–263. [PubMed: 12566403]

CHALKER DL, SANDMEYER SB. Ty3 integrates within the region of RNA polymerase III transcription initiation. Genes Dev. 1992; 6:117–128. [PubMed: 1309715]

DESHPANDE AM, NEWLON CS. DNA replication fork pause sites dependent on transcription. Science. 1996; 272:1030–1033. [PubMed: 8638128]

DUJON B, SHERMAN D, FISCHER G, DURRENS P, CASAREGOLA S, LAFONTAINE I, DE MONTIGNY J, MARCK C, NEUVEGLISE C, TALLA E, et al. Genome evolution in yeast. Nature. 2004; 430:35–44. [PubMed: 15229592]

ELDER JT, PAN J, DUNCAN CH, WEISSMAN SM. Transcriptional analysis of interspersed repetitive polymerase III transcription units in human DNA. Nucleic Acids Res. 1981; 9:1171–1189. [PubMed: 7232214]

FISHMAN-LOBELL J, RUDIN N, HABER JE. Two alternative pathways of double-strand break repair that are kinetically separable and independently modulated. Mol Cell Biol. 1992; 12:1291–1303.

FORNACE AJ JR, MITCHELL JB. Induction of B2 RNA polymerase III transcription by heat shock: Enrichment for heat shock induced sequences in rodent cells by hybridization subtraction. Nucleic Acids Res. 1986; 14:5793–5811. [PubMed: 2426659]

GILBERT N, LABUDA D. Evolutionary inventions and continuity of CORE-SINEs in mammals. J Mol Biol. 2000; 5:365–377. [PubMed: 10772856]

HABER JE, LEUNG WY. Lack of chromosome territoriality in yeast: Promiscuous rejoining of broken chromosome ends. Proc Natl Acad Sci USA. 1996; 93:13949–13954. [PubMed: 8943041]

HANI J, FELDMANN H. tRNA genes and retroelements in the yeast genome. Nucleic Acids Res. 1998; 26:689–696. [PubMed: 9443958]

HUIBREGTSE JM, ENGELKE DR. Genomic footprinting of a yeast tRNA gene reveals stable complexes over the 5′-flanking region. Mol Cell Biol. 1989; 9:3244–3252. [PubMed: 2677668]

HULL M, ERICKSON J, JOHNSTON M, ENGELKE DR. tRNA genes as transcriptional repressor elements. Mol Cell Biol. 1994; 14:1266–1277. [PubMed: 8289806]

NIH-PA Author Manuscript

NIH-PA Author Manuscript

NIH-PA Author Manuscript

IVANOV EL, SUGAWARA N, FISHMAN-LOBELL J, HABER JE. Genetic requirements for the single-strand annealing pathway of double-strand break repair in *Saccharomyces cerevisiae*. Genetics. 1996; 142:693–704. [PubMed: 8849880]

JACKSON JA, FINK GR. Gene conversion between duplicated genetic elements in yeast. Nature. 1981; 292:306–311. [PubMed: 6265790]

JURKA J. Evolutionary impact of human Alu repetitive elements. Curr Opin Genet Dev. 2004; 14:603–608. [PubMed: 15531153]

KENDALL A, HULL MW, BERTRAND E, SINGER PD, ENGELKE DR. A CBF5 mutation that disrupts nucleolar localization of early tRNA biosynthesis in yeast also suppresses tRNA gene-mediated transcriptional silencing. Proc Natl Acad Sci USA. 2000; 97:13108–13113. [PubMed: 11069303]

KINSEY PT, SANDMEYER SB. Adjacent pol II and pol III promoters: Transcription of the yeast retrotransposon Ty3 and a target tRNA gene. Nucleic Acids Res. 1991; 19:1317–1324. [PubMed: 1851556]

KOSKI RA, CLARKSON SG. Synthesis and maturation of *Xenopus laevis* methionine tRNA transcripts in homologous cell-free extracts. J Biol Chem. 1982; 257:4514–4521. [PubMed: 6802833]

LIU WM, SCHMID CW. Proposed roles for DNA methylation in Alu transcriptional repression and mutational inactivation. Nucleic Acids Res. 1993; 21:1351–1359. [PubMed: 8464725]

LIU WM, CHU WM, CHOUDARY PV, SCHMID CW. Cell stress and translational inhibitors transiently increase the abundance of mammalian SINE transcripts. Nucleic Acids Res. 1995; 23:1758–1765. [PubMed: 7784180]

LONGTINE MS, MCKENZIE A III, DEMARINI DJ, SHAH NG, WACH A, BRACHAT P, PHILIPPSEN A, PRINGLE JR. Additional modules for versatile and economical PCR-based gene deletion and modification in *Saccharomyces cerevisiae*. Yeast. 1998; 14:953–961. [PubMed: 9717241]

LUKASOVA E, KOZUBEK S, KOZUBEK M, KJERONSAK J, RYZNAR L, HORAKOVA J, KRAHULCOVA E, HORNECK G. Localization and distance between ABL and BCR genes in interphase nuclei of bone marrow cells of control donors and patients with chronic myeloid leukemia. Hum Genet. 1997; 100:525–535. [PubMed: 9341866]

MCDONALD JP, ROTHSTEIN R. Unrepaired heteroduplex DNA in *Saccharomyces cerevisiae* is decreased in RAD1, RAD52-independent recombination. Genetics. 1994; 137:393–405. [PubMed: 8070653]

MUNZ P, AMSTUTZ H, KOHLI J, LEUPOLD U. Recombination between dispersed serine tRNA genes in *Schizosac-charomyces pombe*. Nature. 1982; 300:225–231. [PubMed: 6924071]

NEVES H, RAMOS C, DA SIVA MG, PARREIRA A, PARREIRA L. The nuclear topography of AMB, BCR, PML, and RAR genes: Evidence for gene proximity in specific phases of the cell cycle and stages of hematopoietic differentiation. Blood. 1999; 93:1197–1207. [PubMed: 9949162]

NEWMAN AJ, OGDEN RC, ABELSON J. tRNA gene transcription in yeast: effects of specified base substitutions in the intragenic promoter. Cell. 1983; 35:117–125. [PubMed: 6226365]

OZENBERGER BA, ROEDER GS. A unique pathway of double-strand break repair operates in tandemly repeated genes, Mol. Cell Biol. 1991; 11:1222–1231.

PAVLICEK A, NOSKOV VN, KOUPRINA N, BARRETT JC, JURKA J, LARIONOV V. Evolution of the tumor suppressor BRCA1 locus in primates: implication for cancer predisposition. Hum Mol Genet. 2004; 13:2727–2751. [PubMed: 15459176]

PRADO F, AGUILERA A. Role of reciprocal exchange, one ended invasion crossover and single-strand annealing on inverted and direct repeat recombination in yeast: Different requirements for the RAD1, RAD10. Genetics. 1995; 139:109–123. [PubMed: 7705617]

PRADO F, CORTES-LEDESMA F, HUERTAS P, AGUILERA A. Mitotic recombination in *Saccharomyces cerevisiae*. Curr Genet. 2003; 42:185–198. [PubMed: 12589470]

PUGET N, STOPPA-LYONNET D, SINILNKOVA OM, PAGE S, LYNCH HT, LENOIR S, MAZOYER GM. Screening for germ-line rearrangements and regulatory mutations in BRCA1 led to the identification of four new deletions. Cancer Res. 1999; 59:455–461. [PubMed: 9927062]

NIH-PA Author Manuscript NIH-PA Author Manuscript NIH-PA Author Manuscript

RATTRAY AJ, SYMINGTON LS. Multiple pathways for homologous recombination in *Saccharomyces cerevisiae*. Genetics. 1995; 139:45–56. [PubMed: 7705645]

ROHLFS EM, PUGET N, GRAHAM ML, WEBER BL, GARBER JE, SKRZYNIA C, HALPERIN JL, LENOIR GM, SILVERMAN LM, MAZOYER S. An Alu-mediated 7.1 kb deletion of BRCA 1 exon 8 and 9 in breast and overian cancer families that results in alternative splicing of exon 10. Genes Chromosomes Cancer. 2000; 28:300–307. [PubMed: 10862036]

ROIX JJ, MCQUEEN PG, MUNSON PJ, PARADA LA, MISTELI T. Spatial proximity of translocation-prone gene loci in human lymphomas. Nat Genet. 2003; 34:287–291. [PubMed: 12808455]

ROSE, MD.; WINSTON, F.; HIETER, P. Methods in Yeast Genetics: A Laboratory Course Manual. Cold Spring Harbor Press; Cold Spring Harbor, NY: 1990.

SMITH TM, LEE MK, SZABO CI, JEROME N, McEUEN M, TAYLOR M, HOOD L, KING MC. Complete genomic sequence and analysis of 117 kb of human DNA containing the gene BRCA1. Genome Res. 1996; 6:1029–1049. [PubMed: 8938427]

THOMPSON M, HAEUSLER RA, GOOD PD, ENGELKE DR. Nucleolar clustering of dispersed tRNA genes. Science. 2003; 302:1399–1401. [PubMed: 14631041]

TOURNIER I, PAILLERETS BB, SOBOL H, STOPPA-LYON-NET D, LIDEREAU R, BARROIS S, MAZOYER M, COULET F, HARDOUIN A, CHOMPRET A, et al. Significant contribution of germline BRCA2 rearrangements in male breast cancer families. Cancer Res. 2004; 64:8143–8147. [PubMed: 15548676]

WILSON ET, LARSON D, YOUNG LS, SPRAGUE KU. A large region controls tRNA transcription. J Mol Biol. 1985; 183:153–163. [PubMed: 3847510]

ZOU S, WRIGHT DA, VOYTAS DF. The *Saccharomyces* retrotransposon Ty5 integrates preferentially into regions of silent chromatin at the telomeres and mating loci. Genes Dev. 1995; 10:634–645. [PubMed: 8598292]

NIH-PA Author Manuscript
NIH-PA Author Manuscript
NIH-PA Author Manuscript

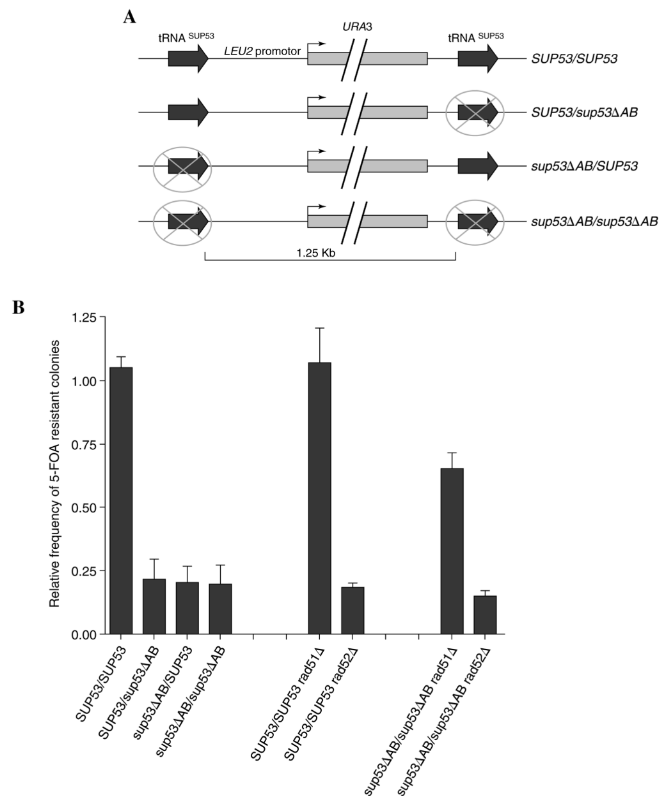

FIG. 1.

(**A**) Recombination between tRNA genes. Four strains were modified by replacing *LEU2* coding sequence with that of *URA3* in strains containing either an active or transcriptionally inactive tRNA gene. An additional active or inactive $tRNA^{SUP53}$ gene was then placed 200 bp downstream of *URA3* in each strain. Deletion events between the two tRNA genes was selected by growth on plates containing 5-FOA media and deletions were confirmed by PCR. (**B**) The first four bars are average values of five independent experiments. The following four values are an average from three experiments. The number of viable colonies on the 5-FOA plates was divided by the number of cells plated and normalized to recombination in the *SUP53/SUP53* strain. The frequency of 5-FOA resistance in the *SUP53/SUP53* strain was ($9.47 \pm 0.71 \times 10^{-10}$). *RAD51* and *RAD52* were deleted in the strain constructs (Fig. 1) containing either two active tRNA genes (*SUP53/ SUP53)* or two inactive tRNA genes (*sup53*ΔAB/*sup53*ΔAB). Deletion of *RAD51* causes a significant increase in 5-FOA resistant colonies between inactive tRNA genes, but only a modest, if any, increase between active tRNA genes. Deletion of *RAD52* caused about a 10-fold decrease in the amount of 5-FOA resistant colonies in the *SUP53*/*SUP53* strain, but only a modest decrease in the *sup53*ΔAB/*sup53*ΔAB strain.

NIH-PA Author Manuscript

NIH-PA Author Manuscript

NIH-PA Author Manuscript

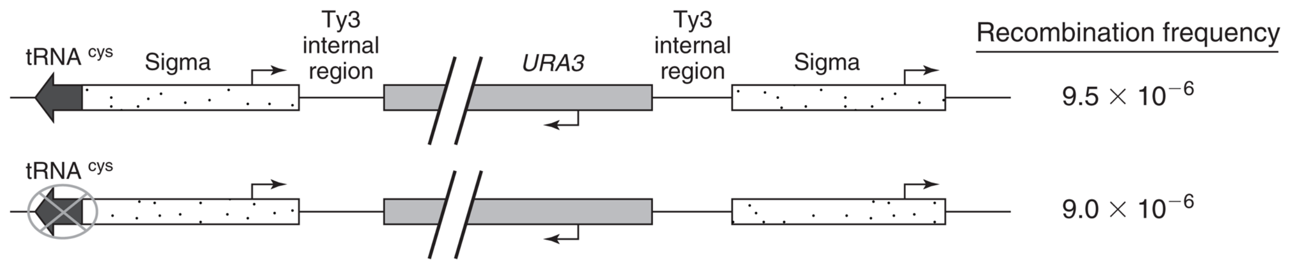

FIG. 2.

Recombination between Ty3 sigma direct repeats. The *URA3* gene was inserted at the Ty3–1 locus, replacing all but 100 bp of either end of the epsilon region. Multiple B box promoter mutations were then created in the neighboring tRNA^{Cys} gene in a second strain, inactivating the transcription promoter. Recombination between the two sigma elements was identified by section against the *URA3* gene through growth on 5-FOA media and screening for precise recombination between sigmas by PCR from genomic DNA. Recombination frequency is shown as number of recombinants divided by the total number of cells plated.

NIH-PA Author Manuscript

NIH-PA Author Manuscript

NIH-PA Author Manuscript

Table 1

Rates of Homologous Recombination

Strain	Rate of 5-FOA Resistance ($\times 10^{-10}$)	% Undergone homologous recombination
SUP53/SUP53	9.47 ± 0.71	100
sup53ΔAB/sup53ΔAB	1.76 ± 0.75	100
SUP53/SUP53 rad52Δ	1.23 ± 0.28	39
sup53ΔAB/sup53ΔAB rad52Δ	1.04 ± 0.19	5
SUP53/SUP53 rad51Δ	10.17 ± 2.61	ND[a]
sup53ΔAB/sup53ΔAB rad51Δ	5.68 ± 1.23	ND

[a]ND = not determined.