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## Circulating folate, vitamin B<sub>12</sub>, homocysteine, vitamin B<sub>12</sub> transport proteins and risk of prostate cancer: a case-control study, systematic review and meta-analysis

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## Abstract

**Background**—Disturbed folate metabolism is associated with an increased risk of some cancers. Our objective was to determine whether blood levels of folate, vitamin B<sub>12</sub> and related metabolites were associated with prostate cancer risk.

**Methods**—Matched case-control study nested within the UK population-based ProtecT study of PSA-detected prostate cancer in men aged 50–69 years. Plasma concentrations of folate, B<sub>12</sub> (cobalamin), holo-haptocorrin, holo- and total-transcobalamin, and total homocysteine (tHcy) were measured in 1,461 cases and 1,507 controls. ProtecT study estimates for associations of folate, B<sub>12</sub>, and tHcy with prostate cancer risk were included in a meta-analysis, based on a systematic review.

**Results**—In the ProtecT study, increased B<sub>12</sub> and holo-haptocorrin concentrations showed positive associations with prostate cancer risk (highest vs lowest quartile of B<sub>12</sub> odds ratio (OR)=1.17 (95% CI 0.95–1.43), P-for-trend=0.06; highest vs lowest quartile of holo-haptocorrin OR=1.27 (1.04–1.56), P-for-trend=0.01); folate, holo-transcobalamin and tHcy were not associated with prostate cancer risk. In the meta-analysis, circulating B<sub>12</sub> levels were associated with an increased prostate cancer risk (pooled OR=1.10 (1.01–1.19) per 100 pmol/L increase in B<sub>12</sub>, P=0.002); the pooled OR for the association of folate with prostate cancer was positive (OR=1.11 (0.96–1.28) per 10 nmol/L, P=0.2) and conventionally statistically significant if ProtecT (the only case-control study) was excluded (OR=1.18 (1.00–1.40) per 10 nmol/L, P=0.02).

**Conclusion**—Vitamin B<sub>12</sub> and (in cohort studies) folate were associated with increased prostate cancer risk.

**Impact**—Given current controversies over mandatory fortification, further research is needed to determine whether these are causal associations.

## Keywords

folate; vitamin B<sub>12</sub>; cobalamin; transcobalamin; haptocorrin; homocysteine; folate-mediated one-carbon metabolism; prostate cancer

## Introduction

The folate-mediated one-carbon metabolic pathway is fundamental to DNA synthesis, repair and methylation (1). The role of folate antagonists in treating haematological (2) and trophoblastic (3) malignancies is well-known, and genetic studies have suggested that folate-pathway gene polymorphisms may be associated with colorectal and gastric cancers (4). Several epigenetic mechanisms related to folate metabolism, including CpG island and histone methylation, DNA uracil mis-incorporation, and chromosomal re-arrangements, have been observed in prostate tumour cells (5, 6).

Studies of dietary intake and blood levels of folate, vitamin B<sub>6</sub>, methionine, and homocysteine have generally found no associations with risk of prostate cancer (7–17), although there is some evidence that high dietary intake and blood levels of vitamin B<sub>12</sub> are associated with increased risk (12–15). One recent study reported a positive association between folic acid supplementation and prostate cancer risk (18). However, results from the same study suggested inverse associations with baseline dietary and plasma folate, as did three other studies (8, 19, 20), and the main trial finding was not replicated in a larger trial (21). Although differences in study design may partly explain these contradictory findings

(22), any role of folate metabolism is likely to be complex, possibly involving a dual effect in which low folate concentrations are associated with increased risk of cancer initiation, while high concentrations, or folic acid supplementation, are associated with more rapid progression following disease onset (23). Answers to these research questions are urgently needed to inform the debate over mandatory fortification of food with folic acid and vitamin B<sub>12</sub>.

We used data from a cross-sectional case-control study nested within the UK population-based Prostate testing for cancer and Treatment ( ProtecT) study to investigate whether plasma concentrations of folate, vitamin B<sub>12</sub>, and total homocysteine (tHcy) were associated with risk of localized and/or advanced prostate cancer detected by means of prostate-specific antigen (PSA) testing. We included our results in a meta-analysis of data from studies identified by systematic review of the literature.

In the ProtecT case-control study, we also measured concentrations of total- and holo-transcobalamin, and calculated the concentration of holo-haptocorrin. Haptocorrin and transcobalamin are B<sub>12</sub> transport proteins to which total circulating B<sub>12</sub> is bound, as holo-haptocorrin and holo-transcobalamin, in an approximate 80:20 ratio (24). Holo-transcobalamin is an alternative marker of impaired B<sub>12</sub> absorption (25), and decreased levels have been associated more strongly than total B<sub>12</sub> with conditions related to impaired folate and B<sub>12</sub> metabolism (26, 27). Raised levels of holo-haptocorrin have been reported in some cancers (28, 29), possibly as a result of up-regulated haptocorrin production by tumour cells (30).

## Methods

### Study population

The ProtecT study is a randomized controlled trial of treatments for localized prostate cancer. Between 2001 and 2009, all (approximately 227,300) men aged 50–69 years in 300 general practices located around nine UK cities (centres) were invited to have a PSA test at a prostate check clinic appointment. Participants with a PSA level between 3.0 and 19.9 ng/mL (approximately 10% of men tested) were invited to attend the centre's urology department for digital rectal examination and 10-core trans-rectal ultrasound-guided biopsy. Men with a PSA level  $\geq$  20 ng/mL were referred as a matter of urgency to a urologist, and were eligible to participate in the treatment trial only if localized cancer was confirmed. A diagnosis of localized prostate cancer was defined as a positive biopsy, clinical stage T1–T2, NX, M0; advanced prostate cancer was defined as positive biopsy, clinical stage T3–T4 or N1 or M1. All men provided written informed consent. Trent Multicentre Research Ethics Committee approved the ProtecT study and allied prostate cancer research under the auspices of ProMPT (Prostate Mechanisms of Prostate cancer and Treatment).

### Selection of cases and controls

The study size (1,500 cases and 1,500 controls) was determined *a priori* to detect an effect estimate (odds ratios) of 1.26 (exposure odds in cases = 0.42) comparing the highest vs lowest three quartiles of vitamin and metabolite concentration at 5% significance, 80% power. Cases were selected at random from among all men diagnosed (by July 2008) with localized or advanced cancer who had consented to a blood sample for research. Eligible controls were men who had a PSA level < 3 ng/mL, or who had a PSA level  $\geq$  3 ng/mL and a negative biopsy result, and who had consented to provide a research blood sample. Controls were stratum-matched to cases by five-year age group and by the primary care practice from which they were recruited, thereby matching for calendar time as prostate check clinics were completed sequentially. For the assays investigated in the current

analysis, one control per case was selected at random from the pool of eligible controls in each stratum.

### Blood sample handling

A standardized blood collection and storage protocol was in place across all collecting centres. Blood was drawn from non-fasting participants at the time of their initial PSA test. Plasma samples were collected using the BD Vacutainer® PPT™ 8.5 mL polymer gel and spray-dried K2EDTA separator tube (Becton, Dickinson & Co, Oxfordshire, UK; catalogue number 362799), centrifuged at 2,200 relative centrifugal force within 10 minutes of blood draw and transported upright at 4 °C to the local processing laboratory. The plasma was transferred to intermediate cryo-vials for medium-term storage and frozen at –80 °C within 36 hours of draw. Samples were transferred to the central bio-repository hub on dry ice. Plasma samples were thawed at 4 °C in a shaking water bath, mixed thoroughly using the Stuart SB3 Blood Rotator Mixer (Bibby Scientific Ltd, Staffordshire, UK) for 10 minutes, centrifuged for 10 minutes at 4 °C at 4,500 revolutions per minute in the Beckman 25R Allegra centrifuge (Beckman Coulter Ltd, Buckinghamshire, UK), and aliquotted into Starlab 1.5 mL cryo-tubes (STARLAB Ltd, Buckinghamshire, UK). The plasma was then stored at –80 °C and transferred on dry ice to the Department of Physiology, Anatomy and Genetics, University of Oxford for assay.

### Biochemical analyses

Plasma concentrations of folate, B<sub>12</sub>, holo- and total-transcobalamin were measured by automated (Perkin Elmer MultiProbe 11 liquid handling system, Perkin Elmer Life and Analytical Sciences (LAS), Buckinghamshire, UK) microbiological assay using *Lactobacillus casei* for folate (31) and *Lactobacillus leichmannii* for B<sub>12</sub> (32), holo- and total-transcobalamin (33). Plasma tHcy was measured by automated (Abbott IMx system, Abbott Laboratories, Chicago, IL, USA) fluorescence polarization immunoassay (34). Between-batch coefficients of variation were, respectively: folate 7.4%; B<sub>12</sub> 7.1%; holo- and total-transcobalamin 8.2%; holo-haptocorrin 10%; tHcy 3.3%. Each batch contained an approximate 1:1 mix of case and control samples and laboratory staff were blind to case-control status. Assays that gave out-of-range results were repeated with diluted (if too high) or larger (if too low) samples. Full results were obtained for all but one sample (insufficient volume for tHcy assay). Folate was measured as <2 nmol/L in 4 participants and holo-transcobalamin as <9 pmol/L in 13 participants; values of 1.8 nmol/L and 8 pmol/L, respectively, were substituted for these results. Holo-haptocorrin concentration was calculated by subtracting holo-transcobalamin concentration from B<sub>12</sub> concentration, hence does not include cobalamin analogues.

### Other Covariates

Self-reported data on ethnicity, smoking, alcohol, medications and dietary supplements, family history of prostate cancer (father and brother), height and weight were collected from Diet, Health and Lifestyle (DHL) questionnaires which are completed before receipt of the initial PSA test result. Self-reported height was used to calculate body mass index (BMI, kg/m<sup>2</sup>), along with nurse-measured weight where available (92.5% of cases and 92.8% of controls) and self-reported weight otherwise.

### Systematic review and data extraction

Eligible studies of the association of prostate cancer risk with serum or plasma folate, B<sub>12</sub> or tHcy levels were identified by searching the Medline and Embase online databases using text search terms for “folate/folic”, “B12/cobalamin”, and “tHcy”; each in conjunction with the MeSH term “prostatic neoplasm” and text terms “prostate cancer” and “prostatic

carcinoma". No language or publication date restrictions were imposed. All databases were last searched on 26/09/2009. References of retrieved articles were screened. Case-control and cohort studies that reported associations of blood (serum or plasma) levels of folate, B<sub>12</sub> and tHcy with prostate cancer risk were included. We also included data from the placebo arms of randomized controlled trials of folic acid and B<sub>12</sub> supplementation. Studies reported their results in a number of different ways and presented various models with different adjustments. We selected the age-adjusted estimate, or a more fully adjusted estimate where available, except where the model was deemed by us to be over-adjusted (eg. adjusted for vegetable intake). Data were extracted independently by two investigators (SMC and RH).

## Statistical methods

**Vitamins and metabolites**—Circulating vitamin and metabolite concentrations were categorized into quartiles (with cut-points based on their distributions among controls), and odds ratios as a measure of relative risk of prostate cancer per quartile of vitamin and metabolite were estimated by conditional logistic regression to account for the matching variables (5-year age group and recruiting general practice), further adjusted for exact age as a continuous variable. Linear trends across quartiles were tested in these models using the mean value for each quartile. Odds ratios for associations with advanced and localized cancer versus controls were compared using a multinomial logistic regression model. This model provides a statistical test for heterogeneity in odds ratios comparing associations of the vitamins and metabolites of interest with localized *vs* advanced prostate cancers. It is an unconditional model, hence it was adjusted for exact age and the study centre where the recruiting general practice was based (9-level variable). Pairwise correlations between circulating vitamin and metabolite concentrations (and with PSA level) in controls were measured by Spearman's rank correlation coefficient. Given previous suggestions of possible U-shaped relationships (23), we used fractional polynomials to investigate possible departures from linearity in the relationships between vitamin and metabolite levels (as continuous measures) and prostate cancer risk (35). Circulating vitamin and metabolite concentrations were natural log-transformed (all had non-normal distributions) for inclusion in multivariable linear regression models. These models were used to assess potential confounders (see "other covariates"), to assess the effects of mutually adjusting vitamin and metabolite levels for each other, and to test for interaction between folate and B<sub>12</sub> and between folate and alcohol on prostate cancer risk. We investigated by linear regression whether log-transformed vitamin and metabolite concentrations were associated with PSA level among controls, because any such association could bias the PSA-based detection of prostate cancer.

**Meta-analysis**—To compare across studies, we calculated the log odds ratio (OR) or hazard ratio (HR) per unit increase in vitamin and metabolite concentration. For studies presenting their results within categories of exposure (*e.g.* quantiles), we used the mean or median exposure in each category when they were reported, and calculated the log OR per unit increase in exposure using the method of Greenland and Longnecker (36). When the mean or median in each group was not reported, and a range of exposure in each group was given, we estimated the mean exposure in each group using the method of Chêne and Thompson (37). Having fitted means to each group, the data were analyzed using the Greenland and Longnecker method (36). We used Stata's *metainf* command to investigate whether exclusion of any one study would significantly change the pooled estimate, *i.e.* whether the pooled point estimate with one study excluded would lie outside the 95% confidence interval of the pooled estimate with all studies included (38).

**Software**—All statistical analyses were performed using Stata Release 11 (StataCorp. 2009, College Station, TX).

## Results

### Baseline characteristics

Of the 3,019 cases and controls for whom plasma concentrations were measured, 51 were in unmatched strata. The remaining 2,968 men were in 587 strata. There was a small surplus of controls in 66 strata and a small deficit in 29 strata, hence the final analysis was based on 1,461 cases (1,298 (89%) localized, 163 (11%) advanced) and 1,507 controls. There were no differences in baseline characteristics of cases and controls (Table 1), but anthropometric (height and weight) and lifestyle data (smoking, alcohol consumption, vitamin supplement) were missing (mainly due to non-return of questionnaires) for a higher proportion of controls (26–27%) than cases (18–19%).

The majority of pairwise combinations of folate, B<sub>12</sub>, holo-haptocorrin, holo- and total-transcobalamin, and tHcy were correlated (Supplementary Table 1). Folate was most strongly correlated with tHcy (correlation coefficient = -0.51), but was also correlated with holo-transcobalamin (correlation coefficient = 0.31) and with B<sub>12</sub> (correlation coefficient = 0.22). None of the vitamin or metabolite concentrations were associated with PSA levels among controls.

### Plasma vitamin and metabolite levels and prostate cancer risk

Of the six vitamins and metabolites, circulating concentrations of B<sub>12</sub>, holo-haptocorrin, and total-transcobalamin were associated with prostate cancer risk in the basic conditional logistic regression models (Table 2). Higher quartiles of B<sub>12</sub> showed a trend (P-for-trend = 0.06) towards increased risk, although this positive association was evident only weakly for the highest *vs* lowest quartile (OR=1.17, 95% CI 0.95–1.43, P=0.1). Holo-haptocorrin concentration was positively associated (P-for-trend = 0.006) with prostate cancer risk (OR=1.27, 95% CI 1.04 – 1.56, P=0.02 comparing highest *vs* lowest quartiles). Total-transcobalamin had an inverse association (P-for-trend = 0.04) with risk of localized prostate cancer, evident only weakly for the highest *vs* lowest quartile (OR=0.80, 95% CI 0.64–1.00, P=0.1).

For B<sub>12</sub>, total-transcobalamin, and holo-haptocorrin, the lowest (or highest) odds ratios appeared in the second quartile, but models incorporating fractional polynomial terms for each vitamin or metabolite as continuous variables gave no indication of departure from linearity (all P > 0.1). There were no differences in the associations of vitamin or metabolite concentrations with prostate cancer according to whether the cancer was localized or advanced with the exception of total transcobalamin concentration, which was inversely associated with risk of localized cancer but not associated with advanced cancer (P-for-heterogeneity = 0.06).

Table 2 also shows age-adjusted estimates for each vitamin and metabolite as continuous log-transformed variables (OR per log<sub>e</sub> approximates to a doubling in concentration) that were included in multivariable conditional logistic regression models. We found little or no change in the estimates for each vitamin and metabolite when mutually adjusted for each other, and there was no interaction between levels of folate and B<sub>12</sub>. In multivariable conditional logistic regression based on men for whom anthropometric (BMI) and lifestyle (smoking, alcohol consumption, vitamin supplementation) data were available (811 cases, 779 controls), these covariates did not confound the associations of vitamin and metabolite concentrations with prostate cancer risk, and there was no interaction between levels of folate and alcohol intake.

## Systematic review and meta-analysis

Our literature search identified 414 studies, of which 20 were eligible. Of these, 6, 5, and 3 studies provided data on blood concentrations of folate (7, 13, 15, 18, 19, 21), B<sub>12</sub> (7, 13, 15, 18, 21) and tHcy (7, 13, 21), respectively (Supplementary Table 2). The excluded studies reported data on dietary folate or B<sub>12</sub> intake only, or concentrations of other folate-pathway vitamins and metabolites, or outcomes other than prostate cancer risk. The results of the Hultdin *et al* study (13) were replicated in a subgroup analysis by Johansson *et al* (15) using the same samples but a different assay (*Lactobacillus leichmannii* microbiological assay instead of Quantaphase II radioassay) and adjusted for BMI, smoking and concentrations of folate and tHcy. We used the Hultdin *et al* data, although the Johansson *et al* data gave very similar results. We used unpublished data from the placebo arms of two randomized controlled trials: one of folic acid and aspirin supplementation for the chemoprevention of colorectal adenomas (18); one of folic acid, vitamin B<sub>12</sub> and vitamin B<sub>6</sub> supplementation for the lowering of tHcy among patients with ischaemic heart disease (21).

We conducted dose-response meta-analyses, combining our results for serum concentrations of folate, B<sub>12</sub> and tHcy with those extracted from the literature and those obtained directly from authors (Figure 1). The pooled (random effects) estimates were: OR=1.11 (95% CI 0.96–1.28) per 10nmol/L folate (P=0.2); OR=1.10 (95% CI 1.01–1.19) per 100pmol/L B<sub>12</sub> (P=0.002); and OR=0.91 (95% CI 0.69–1.19) per 10µmol/L tHcy (P=0.5). Hence, there was no strong statistical evidence for an association of folate or tHcy with prostate cancer risk, but there was evidence to support a 10% higher odds per 100pmol/L increase in circulating B<sub>12</sub>. There was some heterogeneity in the associations of folate (I<sup>2</sup>=40%) and B<sub>12</sub> (I<sup>2</sup>=47%) with prostate cancer risk. There were too few published studies to assess small-study bias. Influence analysis using Stata's *metainf* command showed that excluding any one study from the meta-analysis of associations of B<sub>12</sub> and tHcy with prostate cancer risk did not significantly change either the fixed or random effects pooled estimates (Figure 2). However, exclusion of the ProtecT case-control study from the meta-analysis of the associations of folate with prostate cancer risk changed the fixed-effects pooled estimate to OR=1.19 (95% CI 1.03–1.37) per 10nmol/L folate (P=0.02), and left little heterogeneity between the remaining studies (I<sup>2</sup>=13%), all of which were prospective cohort studies (Figure 2). The median baseline concentrations of B<sub>12</sub> and tHcy were similar in all studies, but baseline concentrations of folate were higher in the Figueiredo (USA) and ProtecT studies (Figure 2), and among non-Swedish subjects in the Johansson (pan-European) study. The number of studies was too small to support meta-regression analysis to test whether the measures of effect in each study were related to these baseline values. However, examination of effect estimates against the median baseline values of folate, vitamin B<sub>12</sub> and tHcy gave no indication that heterogeneity between studies was attributable to differences in baseline concentrations.

## Discussion

Data from the ProtecT study, when combined with results from all other studies, suggest that high circulating concentrations of vitamin B<sub>12</sub> may be associated with increased risk of prostate cancer (Figure 1). The ProtecT data also showed that high circulating concentrations of holo-haptocorrin were associated with increased risk, and high circulating concentrations of total-transcobalamin with decreased risk of prostate cancer. We found no association of folate with prostate cancer risk in the ProtecT data. This result strongly influenced the meta-analysis, which would otherwise have shown a clear positive association of circulating folate with increased prostate cancer risk (Figure 2). It is possible that folate is positively associated with the rate of progression of localized prostate cancer, hence the ProtecT study (based on PSA-detected prevalent cases) would not detect an effect observed in European cohort studies based mainly on clinically-detected cases. We found no

associations of tHcy with prostate cancer risk, either in our own data or in the meta-analysis. Although there was considerable variation in baseline folate levels between studies, this did not appear to explain between-study differences in the measures of association of folate with prostate cancer risk. This is consistent with Johansson *et al* (15), who found no difference of effect between Swedish vs non-Swedish subjects in their pan-European study, even though baseline levels of folate were much lower in Sweden.

The ProtecT study is by far the largest study to date of associations of circulating folate-pathway vitamin and metabolite concentrations with risk of prostate cancer, contributing 81%, 48% and 62% by (inverse variance) weight to the fixed effects meta-analytical results for folate, B<sub>12</sub> and tHcy, respectively (Figure 1). The coefficients of variation for our assay results were low, and any measurement error would attenuate effect estimates to the null rather than generate the observed associations. The study's main limitation is that blood samples were drawn after occurrence of disease, hence causality cannot be directly inferred from our results. However, men were unaware of their disease status so were unlikely to have changed their behaviour or diet in response to disease. Also, our results were similar for advanced and localised disease, which is contrary to what would be expected if effects were secondary to disease status. As with all prostate cancer case-control studies based on PSA testing followed by biopsies, some measurement error would be present due to the imperfect nature of the diagnostic process (39). Non-Caucasian men were not represented in the ProtecT study, or in any of the other studies in our meta-analysis (except for a small proportion (7%) in the Aspirin/Folic Acid Polyp Prevention trial), therefore our findings may not be generalizable to all populations. The small number of advanced cases in ProtecT means we may have been underpowered to detect true differences in associations of vitamin or metabolite concentrations with prostate cancer according to whether the cancer was localized or advanced. Fractional polynomial analysis of the ProtecT data did not indicate departure from linearity for the associations of B<sub>12</sub>, holo-haptocorrin and total-transcobalamin with prostate cancer risk, but these associations were evident only in the top quartiles when analysed as categorical variables. Whether there really is only an effect at extreme elevations could not be discerned from our review of the literature, although two studies showed a similar pattern, with associations evident only in the top quartile of circulating folate (13, 15).

That we found no confounding or effect modification by BMI, smoking or alcohol consumption of associations between vitamin and metabolite concentrations and prostate cancer is consistent with other studies (7, 13, 14, 40). Among men with prostate cancer in the ProtecT study, those who reported little or no sexual activity had higher plasma B<sub>12</sub> concentrations [data not shown]. However, among men with raised PSA levels in the ProtecT study, there was no association between sexual dysfunction and prostate cancer, hence confounding by sexual activity is unlikely to explain our findings (41).

The positive association of B<sub>12</sub> with prostate cancer risk could be causal, or due to reverse-causation or coincidental. A causal association would be consistent with epigenetic mechanisms of prostatic carcinogenesis if these were triggered by elevated levels of B<sub>12</sub> independently of folate and tHcy. Although some associations of B<sub>12</sub> with DNA methylation have been observed in rats (42, 43) and humans (44, 45), such a process as a cause of prostate cancer remains speculative. Vitamin B<sub>12</sub> is an essential co-factor of methionine synthase (MTR), and we reported in a previous meta-analysis that the *A2756G* polymorphism of this enzyme is associated with increased risk of prostate cancer (46). We proposed a causal 'activating polymorphism' mechanism that might be consistent with our finding for B<sub>12</sub>, but this is speculative.



One possible mechanism of reverse causation could be prostate tumour cells having an increased demand for B<sub>12</sub> due to increased biosynthesis of polyamines (47), which in turn up-regulate the activity of MTR (48). Another mechanism could be that elevated levels of plasma B<sub>12</sub> are due to increased production of haptocorrin by prostate tumour cells (49), an effect which may explain high levels of B<sub>12</sub> observed in myelogenous leukemias and metastatic cancers (30). Whether prostate carcinomas can raise plasma concentrations of haptocorrin and B<sub>12</sub> by such processes remains hypothetical, and we did not observe stronger positive associations of plasma B<sub>12</sub> and holo-haptocorrin with advanced *vs* localized prostate cancer. However, the plausibility of reverse-causation as an explanation for our findings is perhaps reinforced by the absence of an association of holo-transcobalamin, representing the bioavailable fraction of B<sub>12</sub>, with prostate cancer risk. We cannot suggest a clear biological explanation for the inverse association of total-transcobalamin with localized prostate cancer. Finally, it is conceivable that folate-pathway genes could be affected coincidentally by epigenetic alterations which play a causal role in prostate cancer. For example, the multidrug resistance protein gene (*MRP1*), which is over-expressed in several cancers including prostate cancer (50), has recently been shown to play a role in cellular efflux of B<sub>12</sub> (51).

That we found similar associations of folate, B<sub>12</sub> and tHcy with advanced and localized prostate cancer is consistent with previous studies (7, 13–15), although Johansson *et al* reported (from a cohort study) borderline heterogeneity (P=0.05) between localized and advanced cases for B<sub>12</sub>: a doubling in vitamin B<sub>12</sub> concentration was associated with an OR=1.69 (95% CI 1.05–2.72, P = 0.03) of advanced cancer, whereas B<sub>12</sub> concentrations were not associated with risk for localized disease (OR=0.96; 95% CI 0.71–1.29, P = 0.8) (15). We found marginal evidence of similar heterogeneity (P=0.09) for total-transcobalamin (advanced disease OR=1.46 (95% CI 0.65–3.26); localized disease OR=0.70 (95% CI 0.50 – 0.99)). Levels of B<sub>12</sub> and total-transcobalamin in our data were uncorrelated, and our finding may be due to chance rather than a corroboration of Johansson *et al*'s finding for B<sub>12</sub>.

The only studies to date of dietary B<sub>12</sub> intake and prostate cancer risk both reported positive associations: Vlainjac *et al* found two-fold higher odds (OR=2.07, 95% CI 1.08–3.97; P-for-trend=0.02) for the highest *vs* lowest tertile (12); Weinstein *et al* (among smokers) found 36% higher odds in the highest *vs* lowest quintile (OR=1.36, 95% CI 1.14–1.62; P-for-trend=0.01) (14). Both studies reported that these results withstood adjustment for dietary covariates; Vlainjac *et al* for intake of total energy, protein, total fat, saturated fatty acids, carbohydrate, total sugar, fibre, retinol equivalent,  $\alpha$ -tocopherol, folate, sodium, potassium, calcium, phosphorus, magnesium and iron (12); Weinstein *et al* for total energy, total protein, animal protein, total fat, animal fat, folate, B<sub>6</sub>, methionine, iron, and specific foods which are correlates of B<sub>12</sub> intake (fish, organ meats, sausages, cholesterol, fatty acids, vitamins and minerals) (14). Hence, there was no confounding by other nutrients which co-occur in foods high in B<sub>12</sub>, and which may be associated with prostate cancer risk. Both studies also adjusted for non-dietary covariates. Although these results may suggest a possible causal relationship, the limitations of studies based on food-frequency questionnaires are well-known, particularly with regard to diet-cancer associations (52). Indeed, the B<sub>12</sub> dietary intake finding by Weinstein *et al* was not found in their earlier, albeit smaller, study of circulating concentrations of B<sub>12</sub> (7).

We conclude that our finding of a positive association of circulating B<sub>12</sub> with increased prostate cancer risk could be explained by reverse-causality. However, given current controversies over mandatory B<sub>12</sub> fortification (53), further research to eliminate a causal role of vitamin B<sub>12</sub> in prostate cancer initiation and/or progression is required, including Mendelian randomisation analyses (54) and repeat measurements of B<sub>12</sub> and holo-

haptocorrin levels during prostate cancer development and/or before and after treatment. Our meta-analysis did not entirely rule out a positive association of circulating folate with increased prostate cancer risk. As with B<sub>12</sub>, even a weak positive association would be a significant public health issue, given the high prevalence of prostate cancer, and legitimate concern about the potential harms *vs* benefits of mandatory folic acid fortification (23).

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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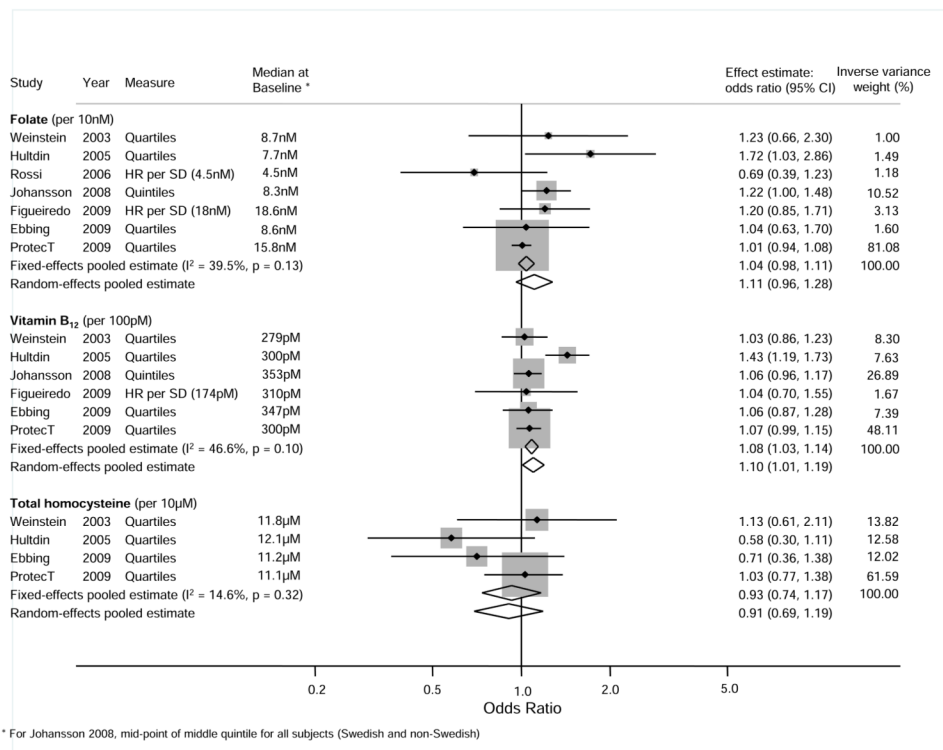
**Role of the Funders:** The funders were nonprofit organizations with no participating role in the study.

## Reference List

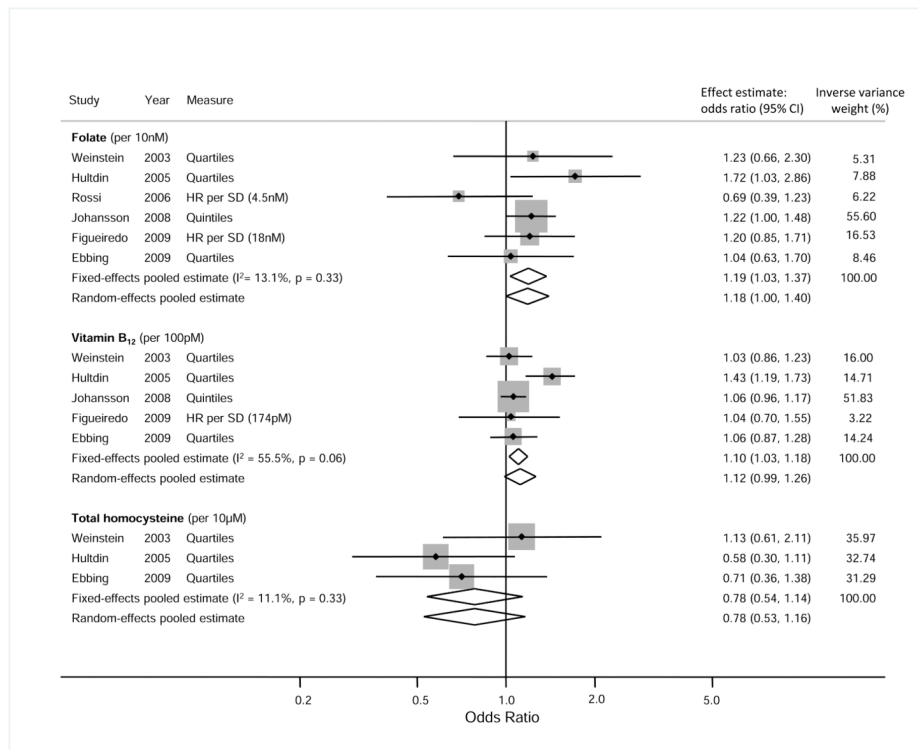
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**Figure 1.** Meta-analyses of associations of circulating folate, vitamin B<sub>12</sub> and tHcy concentrations with prostate cancer risk



**Figure 2.**  
Meta-analyses of associations of circulating folate, vitamin B<sub>12</sub> and tHcy concentrations with prostate cancer risk in prospective cohort studies

**Table 1**

Baseline characteristics of participants stratified by case-control status

Characteristic	Measure	Cases (N=1,461)	Controls (N=1,507)	p*
Age	mean (SD)	62.5 (5.1)	62.3 (5.1)	0.2
Ethnicity (white)		98.5%	98.7%	0.3
BMI (kg/m <sup>2</sup> )**	mean (SD)	27.9 (3.7)	28.2 (4.1)	0.1
Tobacco use**	Current smoker	20.2%	17.3%	0.1
	Ever smoked	63.3%	65.5%	0.3
Alcohol consumption in the past 12 months**	Almost daily or more often	38.5%	37.8%	0.5
	Once or twice per week	41.2%	42.4%	
	Once or twice per month	8.5%	6.9%	
	Special occasions or never	11.9%	12.9%	
Vitamin supplement use in past 12 months (yes/no)**		7.8%	7.9%	0.9
<b>Circulating vitamin and metabolite concentrations</b>				
	<b>median (5<sup>th</sup> – 95<sup>th</sup> percentile)</b>	<b>median (5<sup>th</sup> – 95<sup>th</sup> percentile)</b>		
Folate (nmol/L)	15.9 (6.1 – 51.4)	15.8 (6.2 – 52.3)		0.6
Vitamin B <sub>12</sub> (pmol/L)	307 (156 – 542)	299 (158 – 521)		0.1
Holo-haptocorrin (pmol/L)	247 (119 – 439)	240 (120 – 425)		0.04
Holo-transcobalamin (pmol/L)	57 (23 – 117)	56 (24 – 119)		0.9
Total-transcobalamin (pmol/L)	860 (612 – 1269)	874 (602 – 1270)		0.1
Total homocysteine (μmol/L)	11.1 (7.8 – 17.6)	11.1 (7.7 – 17.7)		0.9

\* Chi-squared test (proportions), Student's t-test (means) or two-sample Wilcoxon rank-sum (Mann-Whitney) test (medians)

\*\* Numbers of cases and controls for whom these data were available: BMI (1,183 cases, 1,105 controls); current smoker (846 cases, 808 controls); ever smoked (1,208 cases, 1,117 controls); alcohol consumption (1,204 cases, 1,116 controls); vitamin supplementation (875 cases, 831 controls)

**Table 2**

Odds ratios for prostate cancer by categorical (quartiles) and continuous ( $\log_e$ -transformed) measures of plasma vitamin and metabolite concentrations

Metabolite	Quartiles in controls (n=1,507)	All cases (n=1,461) vs controls	Localized cases (n=1,298) vs controls	Advanced cases (n=163) vs controls
		Odds Ratio* (95% CI)	Odds Ratio* (95% CI)	Odds Ratio* (95% CI)
Folate (nmol/L)	<10.8	1.00 (reference)	1.00 (reference)	1.00 (reference)
	10.8 – 15.8	1.12 (0.91 – 1.37)	1.18 (0.96 – 1.46)	0.67 (0.39 – 1.18)
	15.9 – 26.2	1.18 (0.96 – 1.44)	1.17 (0.94 – 1.44)	1.47 (0.89 – 2.45)
	>26.2	1.01 (0.82 – 1.24)	1.07 (0.86 – 1.34)	0.71 (0.42 – 1.22)
	P for trend**	0.7	1.0	0.4
	per $\log_e$	0.94 (0.84 – 1.06)	0.96 (0.86 – 1.08)	0.90 (0.67 – 1.21)
P for heterogeneity*** 0.5				
Vitamin B <sub>12</sub> (pmol/L)	<239	1.00 (reference)	1.00 (reference)	1.00 (reference)
	239 – 299	0.91 (0.74 – 1.12)	0.95 (0.77 – 1.18)	0.70 (0.40 – 1.22)
	300 – 376	0.99 (0.80 – 1.22)	0.99 (0.80 – 1.24)	0.93 (0.55 – 1.57)
	>376	1.17 (0.95 – 1.43)	1.20 (0.97 – 1.48)	1.00 (0.61 – 1.65)
	P for trend**	0.06	0.05	0.7
	per $\log_e$	1.19 (0.98 – 1.43)	1.21 (0.99 – 1.48)	1.04 (0.64 – 1.68)
P for heterogeneity*** 0.4				
Holo-haptocorrin (pmol/L)	<187	1.00 (reference)	1.00 (reference)	1.00 (reference)
	187 – 240	0.94 (0.76 – 1.16)	0.95 (0.76 – 1.19)	0.85 (0.50 – 1.44)
	241 – 304	0.99 (0.80 – 1.22)	1.04 (0.84 – 1.29)	0.68 (0.39 – 1.19)
	>304	1.27 (1.04 – 1.56)	1.27 (1.02 – 1.57)	1.24 (0.75 – 2.04)
	P for trend**	0.006	0.01	0.3
	per $\log_e$	1.21 (1.01 – 1.44)	1.23 (1.02 – 1.48)	1.05 (0.67 – 1.64)
P for heterogeneity*** 0.3				
Holo-transcobalamin (pmol/L)	<42	1.00 (reference)	1.00 (reference)	1.00 (reference)
	42 – 56	1.08 (0.88 – 1.33)	1.11 (0.89 – 1.37)	1.02 (0.60 – 1.75)
	57 – 78	1.09 (0.89 – 1.34)	1.12 (0.90 – 1.38)	1.00 (0.61 – 1.65)
	>78	1.04 (0.84 – 1.28)	1.09 (0.87 – 1.35)	0.85 (0.50 – 1.44)
	P for trend**	0.9	0.6	0.5
	per $\log_e$	0.99 (0.86 – 1.14)	1.01 (0.87 – 1.18)	0.93 (0.66 – 1.32)
P for heterogeneity*** 0.7				
Total-transcobalamin (pmol/L)	<749	1.00 (reference)	1.00 (reference)	1.00 (reference)
	749 – 874	1.05 (0.86 – 1.29)	1.00 (0.81 – 1.23)	1.63 (0.94 – 2.83)
	875 – 1024	1.02 (0.83 – 1.25)	1.00 (0.80 – 1.24)	1.36 (0.78 – 2.37)
	>1024	0.84 (0.68 – 1.05)	0.80 (0.64 – 1.00)	1.36 (0.77 – 2.40)
	P for trend**	0.09	0.04	0.6



Metabolite	Quartiles in controls (n=1,507)	All cases (n=1,461) vs controls	Localized cases (n=1,298) vs controls	Advanced cases (n=163) vs controls
		Odds Ratio* (95% CI)	Odds Ratio* (95% CI)	Odds Ratio* (95% CI)
	per log <sub>e</sub>	0.76 (0.55 – 1.05)	0.71 (0.50 – 0.99)	1.45 (0.65 – 3.25)
P for heterogeneity***0.06				
Total homocysteine (3mol/L)	<9.52	1.00 (reference)	1.00 (reference)	1.00 (reference)
	9.52 – 11.13	1.10 (0.90 – 1.35)	1.11 (0.89 – 1.37)	0.93 (0.54 – 1.59)
	11.14 – 13.21	1.03 (0.84 – 1.27)	1.01 (0.81 – 1.26)	1.16 (0.68 – 1.95)
	>13.21	1.04 (0.84 – 1.28)	1.01 (0.81 – 1.26)	1.05 (0.61 – 1.79)
	P for trend**	1.0	0.8	0.8
	per log <sub>e</sub>	0.90 (0.69 – 1.19)	0.88 (0.66 – 1.17)	0.94 (0.47 – 1.91)
P for heterogeneity***0.3				

\* from conditional logistic regression matching on 5-year age group and recruiting centre, further adjusted for exact age (continuous)

\*\* from conditional logistic regression matching on 5-year age group and recruiting centre, further adjusted for exact age (continuous)

\*\*\* from multinomial logistic regression, adjusted for age (continuous) and study centre, using log-transformed concentration