

Different Factors Associated with CTX-M-Producing ST131 and Non-ST131 *Escherichia coli* Clinical Isolates

Marie-Hélène Nicolas-Chanoine^{1,2,3*}, Jérôme Robert^{4,5}, Marie Vigan⁶, Cédric Laouénan^{6,7}, Sylvain Brisse⁸, France Mentré^{6,7}, Vincent Jarlier^{4,5}, the Coli β study group[†]

1 Service de Microbiologie, Hôpital Beaujon AP-HP, Clichy, France, **2** Faculté de Médecine D. Diderot, Paris, France, **3** Institut National de la Santé et de la Recherche Médicale, U773, Centre de Recherche Biomédicale Bichat-Beaujon (CRB3), Université Paris Diderot, Paris, France, **4** Bactériologie-Hygiène, EA 1541/ER 5, Université Pierre et Marie Curie-Paris 6, Site Pitié, Paris, France, **5** Hôpitaux universitaires Pitié Salpêtrière-Charles Foix, AP-HP, Paris, France, **6** UMR-S 738 INSERM - Université Paris Diderot, Sorbonne Paris Cité, Paris, France, **7** Service de Biostatistique, Hôpital Bichat AP-HP, Paris, France, **8** Institut Pasteur, Plateforme Génomique des Pathogènes et Santé Publique, Paris, France

Abstract

Objectives: To determine factors associated with CTX-M-producing ST131 *Escherichia coli* which is the worldwide predominant lineage among CTX-M-producing *E. coli* isolates.

Methods: Consecutive inpatients with a clinical sample positive for CTX-M-producing *E. coli* and considered as cases in a previous 8-month (2008–2009) case-control study performed in ten university hospitals in the Paris area were included in the present sub-population study. Patients with a CTX-M-producing ST131 *E. coli* clinical isolate were compared with those with a CTX-M-producing non-ST131 *E. coli* clinical isolate with regard to 66 variables. Variables were first compared using univariate logistic regression, then a multivariate analysis using a backward selection with variables with p-value <0.1 in univariate analysis was carried out.

Results: Fifty-five patients with a CTX-M-producing ST131 *E. coli* clinical isolate were compared to 97 patients with a CTX-producing non-ST131 *E. coli* clinical isolate. Multivariate analysis showed that only previous residence in long term care facilities (OR = 4.4; 95% CI = 1.3–14.7) was positively associated with a CTX-M-producing ST131 *E. coli* isolate. However, it also showed that regular consumption of poultry products (OR = 0.2; 95% CI = 0.1–0.6), having had at least one device in the preceding 6 months (OR = 0.3; 95% CI = 0.1–0.7) and stay in ICU (OR = 0.2; 95% CI = 0.05–0.8) were negatively associated with isolation of CTX-M-producing ST131 *E. coli* from clinical samples.

Conclusions: This study provides more insight into the epidemiological features of ST131 and non-ST131 *E. coli* producing CTX-M enzymes. It shows, for the first time, that isolation of CTX-M-producing ST131 *E. coli* from clinical samples is not linked to consumption of various foods and confirms that residence in long term care facilities is a predictor of these isolates.

Citation: Nicolas-Chanoine M-H, Robert J, Vigan M, Laouénan C, Brisse S, et al. (2013) Different Factors Associated with CTX-M-Producing ST131 and Non-ST131 *Escherichia coli* Clinical Isolates. PLoS ONE 8(9): e72191. doi:10.1371/journal.pone.0072191

Editor: Axel Cloeckert, Institut National de la Recherche Agronomique, France

Received: May 7, 2013; **Accepted:** July 5, 2013; **Published:** September 4, 2013

Copyright: © 2013 Nicolas-Chanoine et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This study was supported by a grant (PAS⁷010) from the Programme Régional de Recherche Clinique AP-HP/Institut Pasteur, Direction de la Recherche Clinique AP-HP, Paris, France (a publicly funded non-profit organization). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: mhn.chanoine@bjn.aphp.fr

† Membership of the Coli β study group is provided in the Acknowledgments.

Introduction

The polyclonal structure of *Escherichia coli* from clinical and commensal human isolates, and from environmental isolates has clearly been shown by studies recently carried out in the Netherlands (clinical and commensal human isolates and chicken meat isolates), England (clinical isolates) and France (clinical and commensal isolates) on the basis of sequence types (ST) [1–5]. However, some *E. coli* lineages were identified as predominant in the five above cited studies independent of the source of the isolate or the production of extended-spectrum β -lactamase (ESBL). In particular, *E. coli* ST131 was predominant among the clinical and commensal human isolates, producers of

ESBL or not. In contrast, it was not identified in ESBL-producing *E. coli* isolated from chicken meat in the Netherlands. The absence of clone ST131 has also been confirmed recently in Spain among the *E. coli* isolates contaminating raw chicken meat [6] although another previous Spanish study had found that 7% of retail chicken samples were contaminated by *E. coli* ST131 [7]. Vincent *et al.* also had identified *E. coli* ST131 from retail chicken samples in Canada but at a significant lower prevalence (0.4%) than in Spain [8]. In contrast, isolates of ST10, comprising ESBL and non-ESBL producers, were frequent both among the clinical and commensal human isolates as well as among the meat isolates [1,2,4,5,8]. As shown by the Dutch, Canadian and French studies, the

recognized avian pathogenic *E. coli* ST117 was another predominant lineage among the clinical and meat isolates [1,5,9]. On the other hand, although CTX-M-15 was shown to be the predominant CTX-M enzyme (46%) among the French clinical isolates, it should be stressed that CTX-M-1 was the only ESBL found in the ST117 clinical isolates in France and was the predominant ESBL found among the ST117 meat isolates in the Netherlands [1,5].

These reports suggest that epidemiological differences exist between CTX-M-producing strains of ST131 and non-ST131 clones. Therefore, we sought to analyse characteristics associated with CTX-M-producing *E. coli* ST131 isolated from clinical samples by performing a sub-population analysis of data collected during a case-control study carried out from November 2008 to June 2009 to determine factors independently associated with a clinical sample positive for a CTX-M-producing *E. coli* isolate in ten hospitals of the Paris area [10]. The analysis of the population structure of CTX-M-producing *E. coli* and non-ESBL-producing *E. coli* isolates which was performed in addition to the case-control study, was also used as a basis for the present study [1].

Materials and Methods

Ethics Statement

Written informed consent was obtained from all adult participants and from parents for child participants. The study and the consent procedure were approved by the Ethics Committee of the Groupe Hospitalier Universitaire Nord (Institutional review board N°IRB00006477).

Study Design and Participants

All consecutive inpatients with a clinical sample positive for CTX-M-producing *E. coli* and considered as cases in a previous 8-month (2008–2009) case-control study performed in ten university hospitals in the Paris area were included in the present sub-population study [10]. Patients with a clinical sample positive for CTX-M-producing ST131 *E. coli* (n = 55) were compared with those with a clinical sample positive for CTX-M-producing non-ST131 *E. coli* (n = 97) with regard to 66 characteristics collected during the case-control study, including basic demographic data, patient's lifestyle (housing, travel abroad, diet, pet, sport practice...), medical history (hospitalisation and invasive devices in the preceding six months, antibiotic in the preceding month, comorbidity...) and data on the current hospitalisation (hospitalisation wards, invasive devices, antibiotic regimens ...). The 97 non-

Table 1. Univariate and multivariate analyses of demographic and lifestyle factors associated with a CTX-M-producing ST131 or non-ST131 *E. coli* clinical isolate.

Characteristic	Univariate analysis				Multivariate analysis	
	ST131 (n = 55)	Non-ST131 (n = 97)	Odds ratio	P value	Odds ratio	P value
	No. (%)	No. (%)	(95% CI)		(95% CI)	
<i>Demographic data</i>						
Age (mean ± SD) in years	70.2±25.8	60.5±24.0	1.0 (1.0–1.0)	0.02		
Age <15 years	3 (5.4)	6 (6.2)	0.9 (0.2–3.6)	0.8		
Age ≥ 65 years	37 (67.3)	47 (48.5)	2.2 (1.1–4.3)	0.03		
Age ≥80 years	28 (50.9)	21 (21.7)	3.8 (1.8–7.7)	0.0003		
Female	41 (74.6)	58 (59.8)	2.0 (1.0–4.1)	0.07		
Country of birth outside of Europe	15 (27.3)	36 (37.1)	0.6 (0.3–1.3)	0.2		
Living in a country outside of Europe	1 (1.8)	10 (10.3)	0.2 (0.02–1.3)	0.09		
<i>Lifestyle</i>						
Collective housing	17 (30.9)	10 (10.3)	3.9 (1.6–9.3)	0.002		
Individual housing (>2 household members)	11 (20.0)	33 (34.0)	0.5 (0.2–1.1)	0.07		
Live alone	13 (23.6)	21 (21.7)	1.1 (0.5–2.5)	0.8		
Functionally dependent before hospitalisation	29 (52.7)	20 (20.6)	4.3 (2.1–8.8)	<0.0001		
Patients not working	43 (78.2)	64 (66.0)	1.8 (0.9–4.0)	0.12		
Retired patients	37 (67.3)	51 (52.6)	1.9 (0.9–3.7)	0.08		
<i>Consumption of:</i>						
- ≥7 raw vegetables/week	26 (68.4)	59 (69.4)	1.0 (0.4–2.2)	0.9		
- poultry ≥ twice a week	15 (39.5)	56 (66.7)	0.3 (0.1–0.7)	0.006	0.2 (0.1–0.6)	0.002
- beef ≥ twice a week	21 (55.3)	57 (67.9)	0.6 (0.3–1.3)	0.2		
Consumption of raw meat	9 (16.4)	28 (28.9)	0.5 (0.2–1.1)	0.09		
Community meal	29 (52.7)	52 (53.6)	1.0 (0.5–1.9)	0.9		
Practice of a sport	3 (5.5)	8 (8.3)	0.6 (0.2–2.5)	0.5		
Pets or livestock	5 (9.1)	14 (14.4)	0.6 (0.2–1.7)	0.3		
Travel abroad in the preceding 6 months	3 (5.5)	14 (14.4)	0.3 (0.09–1.2)	0.1		

doi:10.1371/journal.pone.0072191.t001

Table 2. Univariate and multivariate analyses of medical history-related factors associated with a CTX-M-producing ST131 or non-ST131 *E. coli* clinical isolate.

Characteristic	Univariate analysis		Multivariate analysis			
	ST131 (n = 55)	Non-ST131 (n = 97)	Odds ratio	P value	Odds ratio	P value
	No. (%)	No. (%)	(95% CI)		(95% CI)	
In the preceding 6 months						
- hospitalised	34 (61.8)	63 (65.0)	0.9 (0.4–1.7)	0.7		
- hospitalised ≥ 10 days	19 (34.6)	42 (43.3)	0.7 (0.3–1.4)	0.3		
- hospitalised <10 days	15 (27.3)	21 (21.7)	1.4 (0.6–2.9)	0.4		
- hospitalised outside of France	1 (1.8)	7 (7.2)	0.2 (0.03–2.0)	0.2		
- at least one invasive device	30 (54.6)	66 (68.0)	0.6 (0.3–1.1)	0.1	0.3 (0.1–0.7)	0.01
• urine drainage	16 (29.6)	31 (32.3)	0.9 (0.4–1.8)	0.7		
• mechanical ventilation	3 (5.6)	10 (10.4)	0.5 (0.1–1.9)	0.3		
• intravascular devices	29 (53.7)	62 (64.6)	0.6 (0.3–1.3)	0.2		
• colonoscopy, endoscopy	9 (17.3)	28 (30.4)	0.5 (0.2–1.1)	0.09		
Surgery during the last month	10 (18.2)	34 (35.4)	0.4 (0.2–0.9)	0.03		
Prosthesis within the last year	2 (3.7)	8 (8.3)	0.4 (0.09–2.1)	0.3		
Antibiotic in the month preceding hospitalisation	16 (29.1)	37 (38.1)	0.7 (0.3–1.4)	0.3		
- cotrimoxazole	2 (3.6)	8 (8.3)	0.4 (0.09–2.1)	0.3		
- fluoroquinolones	4 (7.3)	7 (7.2)	1.0 (0.3–3.6)	1.0		
- extended spectrum cephalosporins	4 (7.3)	7 (7.2)	1.0 (0.3–3.6)	1.0		
- penicillins	6 (10.9)	11 (11.3)	1.0 (0.3–2.7)	0.9		
- ≥ 5 days	9 (16.4)	22 (22.7)	0.7 (0.3–1.6)	0.4		
Nursing or physiotherapy before hospitalisation	9 (16.4)	17 (17.5)	0.9 (0.4–2.2)	0.9		
At least one co-morbidity	33 (60.0)	57 (58.8)	1.1 (0.5–2.1)	0.9		
- recurrent urinary tract or chronic skin infections	18 (32.7)	21 (21.7)	1.8 (0.8–3.7)	0.1		
- obstructive bronchial pulmonary disease	2 (3.6)	5 (5.2)	0.7 (0.1–3.7)	0.7		
- cancer	10 (18.2)	27 (27.8)	0.6 (0.3–1.3)	0.2		
- diabetes	12 (21.8)	22 (22.7)	0.9 (0.4–2.1)	0.9		

doi:10.1371/journal.pone.0072191.t002

ST131 *E. coli* isolates displayed 51 ST types of which 38 were displayed by a single isolate and 13 by several isolates: 14 isolates for ST10, 7 for ST167 and ST648, 4 for ST88 and ST410, 3 for ST38, ST93, ST117, ST354, ST405, ST617 and ST1284 and 2 for ST44 [1].

Statistical Analysis

Variables were first compared using univariate logistic regression and odds ratio (OR) and 95% confidence interval (CI) were estimated. We next used a multivariate analysis using a backward selection with variables with p-value <0.1 in univariate analysis. P-values were assessed at the 0.05 level. All statistical analyses were performed with SAS software, version 9.3 (SAS Institute, Cary, North Carolina).

Results

A total of 55 patients with a CTX-M-producing ST131 *E. coli* clinical isolate were compared to 97 patients with a CTX-M-producing non-ST131 *E. coli* clinical isolate with regard to the 66 variables studied (Tables 1, 2 and 3). In univariate analysis, patients harbouring *E. coli* ST131 were more likely than those harbouring non-ST131 *E. coli* to be aged ≥ 65 years (OR = 2.2; 95% CI = 1.1–4.3) and ≥ 80 years (OR = 3.8; 95% CI = 1.8–7.7)

(Table 1). Among factors focusing on patient's lifestyle (Table 1), living in collective housing (OR = 3.9; 95% CI = 1.6–9.3) and being functionally dependent before hospitalisation (OR = 4.3; 95% CI = 2.1–8.8) were significantly associated with a ST131 *E. coli* clinical isolate. On the opposite, consumption of poultry at least twice a week was inversely associated with a ST131 *E. coli* clinical isolate (OR = 0.3; 95% CI = 0.1–0.7) (Table 1). Patients with a ST131 *E. coli* clinical isolate were more likely than others to have been in long term care facilities (LTCF) between admission and study inclusion (OR = 2.8; 95% CI = 1.2–6.3), and to have a urinary tract infection during the current hospitalisation (OR = 2.2; 95% CI = 1.0–4.6) (Table 3). On the opposite, patients with a ST131 *E. coli* clinical isolate were less likely to have surgery in the last month (OR = 0.4; 95% CI = 0.2–0.9) (Table 2), to have been in intensive care unit (ICU) (OR = 0.3; 95% IC = 0.1–0.9), and to have invasive devices within the week prior inclusion (OR = 0.2; 95% CI = 0.1–0.5), notably a urinary catheter (OR = 0.3; 95% CI = 0.1–0.6), and intravascular devices (OR = 0.2; 95% CI = 0.1–0.5) (Table 3).

In multivariate analysis, only previous residence in LTCF (OR = 4.4; 95% CI = 1.3–14.7) remained positively associated with *E. coli* ST131 (Table 3). However, consumption of poultry at least twice a week (OR = 0.2; 95% CI = 0.1–0.6) (Table 1), having had at least one device in the preceding 6 months (OR = 0.3; 95%

Table 3. Univariate and multivariate analyses of current hospitalisation-related factors associated with a CTX-M-producing ST131 or non-ST131 *E. coli* clinical isolate.

Characteristic	Univariate analysis		Multivariate analysis			
	ST131 (n = 55)	Non-ST131 (n = 97)	Odds ratio	P value	Odds ratio	P value
	No. (%)	No. (%)	(95% CI)		(95% CI)	
Transferred from another hospital	32 (25.5)	18 (18.6)	1.5 (0.7–3.3)	0.3		
Mc Cabe score 2	11 (21.6)	23 (27.1)	0.7 (0.3–1.7)	0.5		
Immunocompromised	15 (27.3)	36 (37.1)	0.6 (0.3–1.3)	0.2		
Between admission and inclusion						
- Ward						
• ICU	4 (7.3)	27 (27.8)	0.3 (0.1–0.9)		0.2 (0.05–0.8)	0.02
• LTCF	21 (38.2)	14 (14.4)	2.8 (1.2–6.3)	0.0009*	4.4 (1.3–14.7)	0.02
• Others	30 (54.5)	56 (57.8)	1		1	
- Invasive device during the last week	33 (60.0)	84 (86.6)	0.2 (0.1–0.5)	0.0003		
• urine drainage	11 (20.0)	45 (46.4)	0.3 (0.1–0.6)	0.002		
• mechanical ventilation	5 (10.6)	19 (20.0)	0.5 (0.2–1.4)	0.2		
• intravascular devices	31 (56.4)	81 (84.4)	0.2 (0.1–0.5)	0.0003		
- Antibiotic receipt	24 (43.6)	57 (58.8)	0.5 (0.3–1.1)	0.07		
• cotrimoxazole	3 (5.5)	6 (6.2)	0.9 (0.2–3.6)	0.9		
• fluoroquinolones	7 (12.7)	10 (10.3)	1.3 (0.5–3.5)	0.7		
• penicillins	10 (18.2)	26 (26.8)	0.6 (0.3–1.4)	0.2		
• extended spectrum cephalosporins	6 (10.9)	12 (12.4)	0.9 (0.3–2.5)	0.8		
• aminoglycosides	2 (3.6)	11 (11.3)	0.3 (0.06–1.4)	0.1		
• carbapenems	1 (1.8)	8 (8.3)	0.2 (0.03–1.7)	0.14		
• ≥5 days	13 (23.6)	35 (36.1)	0.5 (0.3–1.2)	0.1		
Specimen and infection data						
- specimen sampled after 48 h of hospitalisation	35 (63.6)	51 (52.6)	1.6 (0.8–3.1)	0.2		
- specimen sampled after >10 days of hospitalisation	21 (38.2)	35 (36.1)	1.1 (0.6–2.2)	0.8		
- urine sample	39 (70.9)	58 (59.8)	1.6 (0.8–3.3)	0.2		
- urinary tract infection	42 (76.4)	58 (59.8)	2.2 (1.0–4.6)	0.04		

ICU; intensive care unit, LTCF; long term care facility,

*P value resulting from the analysis of the variable "ward" classified into 3 categories, ie ICU, LTCF and others.

doi:10.1371/journal.pone.0072191.t003

CI = 0.1–0.7) (Table 2), and hospitalisation in ICU (OR = 0.2; 95% CI = 0.05–0.8) (Table 3) were, independently, inversely associated with isolation of *E. coli* ST131 from clinical samples.

Discussion

E. coli ST131 has been shown to be a worldwide predominant clone among extra-intestinal pathogenic isolates but also among the human commensal flora [2,4,10–12]. Interestingly, it was found to be almost the only lineage among clinical isolates of group B2 *E. coli* that produced CTX-M enzymes [1]. It displayed a higher ability to colonize the digestive tract and a lower level of virulence in various animal models in comparison with reference group B2 urinary pathogenic *E. coli* strains (CFT053, J536 and HT7) [13–15]. Therefore, better knowing the epidemiology of clone ST131, which appears to be a very peculiar group B2 lineage, especially among isolates producing CTX-M enzymes, is of interest due to its worldwide success. The present prospective study investigated which factors among 66 studied were associated with those of ST131 *E. coli* clinical isolates that produce CTX-M enzymes. Among the various types of food products analysed, it was found, for the first time, that consumption of poultry meat at

least twice a week is a factor inversely associated with isolation of a CTX-M-producing ST131 *E. coli* clinical isolate among the CTX-M-producing *E. coli* clinical isolates. In other words, it means that consumption of poultry meat was associated with isolation of CTX-M-producing *E. coli* that did not belong to ST131. This finding is of importance with regard to the debate on the potential food-borne source of *E. coli* ST131, notably those producing CTX-M enzymes [8,16]. Poultry meat was suggested as a source of *E. coli* ST131 on the basis of two studies published in 2010 because *E. coli* ST131 has been isolated from poultry meat samples [7,8]. The most recent studies conducted in the Netherlands and in Spain challenged this hypothesis as they failed to isolate CTX-M-producing *E. coli* ST131 from chicken meat samples [3,6]. Overall, the results of our study are in accordance with the fact that *E. coli* ST131 has not been identified among ESBL-producing *E. coli* isolated from retail chicken meat on the contrary to other lineage [3,5,6,17]. Although, there are very few studies on the population structure of ESBL-producing *E. coli* isolates from poultry meat, it is noteworthy that, among the CTX-M-producing non ST131 *E. coli* clinical isolates, some dominant clonal groups (ST10, ST117 and ST354) are commonly identified from chicken meat [3,8,18].

Interestingly, ST167 and ST648, the two highest dominant clonal groups after ST10 among the CTX-M-producing non-ST131 *E. coli* clinical isolates had been identified among ESBL-producing *E. coli* isolates from Spanish poultry farms and from birds of prey from Germany and Mongolia [19,20]. In summary, the dominant non-ST131 clonal groups in our population are clonal groups commonly identified in avian populations.

The only factor positively associated with isolation of CTX-M-producing *E. coli* ST131 from clinical samples was residence in LTCF before inclusion in the study. Rooney *et al.* showed that a high proportion of people living in such settings in England had digestive tract colonization with ESBL-producing *E. coli* ST131 [21]. Of note, the first identification of CTX-M-15-producing *E. coli* ST131 in France was achieved from patients in LTCFs [22,23]. More recently, Banerjee *et al.* conducted a retrospective study in all healthcare settings in Olmsted County (Minnesota) and found that LTCF residence was a factor independently associated with *E. coli* ST131 [24]. Overall, three studies conducted in three different developed countries have found a link between LTCF residence and *E. coli* ST131. This might suggest that human cross-transmission is a key factor in the dissemination of CTX-M-producing *E. coli* ST131.

Although the proportion of hospital-acquired (isolation after 48 h of hospitalisation) CTX-M-producing ST131 and non-ST131 *E. coli* isolates was high and not significantly different (63.6% vs 56.2%; $P=0.2$) and the patients infected by either *E. coli* ST131 or *E. coli* non-ST131 did not differ with regard to Mac Cabe score, we found that presence of invasive devices in the preceding six months and stay in ICU before study inclusion were inversely associated with isolation of CTX-M-producing *E. coli* ST131. It suggests that isolation of CTX-M-producing non-ST131 *E. coli* from clinical samples is more likely to be healthcare-related. Such results seem to be in contradiction with those obtained by Banerjee *et al.* [24]. Indeed, they found that *E. coli* ST131 is linked

to healthcare and hospital acquisition. However, we noted that this link was identified by Banerjee *et al.* in the univariate and not in the multivariate analysis that they carried out.

Finally, we were not able to link travel abroad, notably in Africa and India, to isolation of CTX-M-producing *E. coli* ST131 from clinical samples probably because of the lack of power regarding this association in our study [25].

In conclusion, this study provides more insight into the epidemiological features of ST131 and non-ST131 *E. coli* producing CTX-M enzymes. It shows, for the first time, that isolation of CTX-M-producing *E. coli* ST131 from clinical samples was not linked to consumption of specific foods and confirms that residence in long term care facilities is linked to these isolates. Further studies are required to know whether our results are also relevant for *E. coli* ST131 not producing CTX-M enzymes.

Acknowledgments

Coli β study group: Anani Akpabie (Hôpital Emile Roux, AP-HP, Limeil Brévannes, France), Catherine Doit (Hôpital Robert Debré, AP-HP, Paris, France), Salah Gallah (Hôpital Charles Foix, AP-HP, Ivry, France), Najiby Kassis-Chikhani (Hôpital Paul Brousse, AP-HP, Villejuif, France), Béatrice Larroque (Hôpital Beaujon, AP-HP, Clichy, France), Véronique Leflon-Guibout (Hôpital Beaujon, AP-HP, Clichy, France), Estelle Marcon (Hôpital Beaujon, AP-HP, Clichy, France), Didier Moissenet (Hôpital Trousseau, AP-HP, Paris, France), Isabelle Podglajen (Hôpital Georges Pompidou, AP-HP, Paris, France), Charlotte Verdet (Hôpital Tenon, AP-HP, Paris, France), Corine Vincent (Université D Diderot, Paris, France), and Jean-Ralph Zahar (Hôpital Necker, AP-HP, Paris, France).

Author Contributions

Conceived and designed the experiments: MHNC VJ FM. Performed the experiments: MHNC SB. Analyzed the data: MV CL FM MHNC JR. Contributed reagents/materials/analysis tools: MHNC SB. Wrote the paper: MHNC JR VJ SB CL.

References

1. Brisse S, Diancourt L, Laouenan C, Vigan M, Caro V, et al. (2012) Phylogenetic distribution of CTX-M- and non-extended-spectrum- β -lactamase-producing *Escherichia coli* isolates: group B2 isolates, except clone ST131, rarely produce CTX-M enzymes. *J Clin Microbiol* 50: 2974–2981.
2. Gibreel TM, Dodgson AR, Cheesbrough J, Fox AJ, Bolton FJ, et al. (2012) Population structure, virulence potential and antibiotic susceptibility of uropathogenic *Escherichia coli* from Northwest England. *J Antimicrob Chemother* 67: 346–356.
3. Kluytmans JA, Overdeest ITM, Willemsen I, Kluytmans-van den Bergh MF, van der Zwaluw K, et al. (2012) Extended-spectrum β -lactamase-producing *Escherichia coli* from retail chicken meat and humans: comparison of strains, plasmids, resistance genes, and virulence factors. *Clin Infect Dis* 56: 478–487.
4. Nicolas-Chanoine M-H, Gruson C, Bialek-Davenet S, Bertrand X, Thomas-Jean F, et al. (2013) 10-Fold increase (2006–11) in the rate of healthy subjects with extended-spectrum β -lactamase-producing *Escherichia coli* faecal carriage in a Parisian check-up centre. *J Antimicrob Chemother* 68: 562–568.
5. Overdeest I, Willemsen I, Rijnsburger M, Eustace A, Xu L, et al. (2011) Extended-spectrum β -lactamase genes of *Escherichia coli* in chicken meat and humans, the Netherlands. *Emerg Infect Dis* 17: 1216–1222.
6. Egea P, Lopez-Cerero L, Torres E, Gomez-Sanchez Mdel C, Serrano L, et al. (2012) Increased raw poultry meat colonization by extended-spectrum β -lactamase-producing *Escherichia coli* in the south of Spain. *Int J Food Microbiol* 159: 69–73.
7. Mora A, Herrera A, Mamani R, Lopez C, Alonso MP, et al. (2010) Recent emergence of clonal group O25b:K1:H4-B2-ST131 *ibcA* strains among *Escherichia coli* poultry isolates, including CTX-M-9-producing strains, and comparison with clinical human isolates. *Appl Environ Microbiol* 76: 6991–6997.
8. Vincent C, Boerlin P, Daignault D, Dozois CM, Dutil L, et al. (2010) Food reservoir for *Escherichia coli* causing urinary tract infections. *Emerg Infect Dis* 16: 88–95.
9. Mora A, Lopez C, Herrera A, Viso S, Mamani R, et al. (2012) Emerging avian pathogenic *Escherichia coli* strains belonging to clonal groups O111:H4-D-ST2085 and O111:H4-D-ST117 with high virulence-gene content and zoonotic potential. *Vet Microbiol* 156: 347–352.
10. Nicolas-Chanoine MH, Jarlier V, Robert J, Arlet G, Drieux L, et al. (2012) Patient's origin and lifestyle associated with CTX-M-producing *Escherichia coli*: a case-control study. *PLoS ONE* 7: e30498.
11. Johnson J, R, Menard E, Johnston B, Kuskowski MA, Nichol KA, et al. (2009) Epidemic clonal groups of *Escherichia coli* as a cause of antimicrobial-resistant urinary tract infections in Canada, 2002 to 2004. *Antimicrob Agents Chemother* 53: 2733–2739.
12. Rogers BA, Sidjabat HE, Paterson DL (2011) *Escherichia coli* O25b-ST131: a pandemic multiresistant, community-associated strain. *J Antimicrob Chemother* 66: 1–14.
13. Johnson JR, Porter SB, Zhanel G, Kuskowski MA, Denamur E (2012) Virulence of *Escherichia coli* clinical isolates in a murine sepsis model in relation to sequence type ST131 status, fluoroquinolone resistance, and virulence genotype. *Infect Immun* 80: 1554–1562.
14. Lavigne JP, Vergunst AC, Goret L, Sotto A, Combesure C, et al. (2012) Virulence potential and genomic mapping of the worldwide clone *Escherichia coli* ST131. *PLoS ONE* 7: e34294.
15. Vimont S, Boyd A, Bleibtreu A, Bens M, Goujon JM, et al. (2012) The CTX-M-15-producing *Escherichia coli* clone O25b: H4-ST131 has high intestine colonization and urinary tract infection abilities. *PLoS ONE* 7: e46547.
16. Manges AR, Johnson JR (2012) Food-borne origins of *Escherichia coli* causing extra-intestinal infections. *Clin Infect Dis* 55: 712–719.
17. Dhanji H, Murphy NM, Doumith M, Durmus S, Lee SS, et al. (2010) Cephalosporin resistance mechanisms in *Escherichia coli* isolated from raw chicken imported into the UK. *J Antimicrob Chemother* 65: 2534–2537.
18. Cohen Stuart J, van den Munckhof T, Voets G, Scharringa J, Fluit A, et al. (2012) Comparison of ESBL contamination in organic and conventional retail chicken meat. *Int J Food Microbiol* 154: 212–214.
19. Cortes P, Blanc V, Mora A, Dahbi G, Blanco JE, et al. (2010) Isolation and characterization of potentially pathogenic antimicrobial-resistant *Escherichia coli* strains from chicken and pig farms in Spain. *Appl Environ Microbiol* 76: 2799–2805.
20. Guenther S, Aschenbrenner K, Stamm I, Bethé A, Semmler T, et al. (2012) Comparable high rates of extended-spectrum- β -lactamase-producing *Escherichia coli* in birds of prey from Germany and Mongolia. *PLoS ONE* 7: e33039.

21. Rooney PJ, O'Leary MC, Loughrey AC, McCalmont M, Smyth B, et al. (2009) Nursing homes as a reservoir of extended-spectrum β -lactamase (ESBL)-producing ciprofloxacin-resistant *Escherichia coli*. *J Antimicrob Chemother* 64: 635–641.
22. Kassis-Chikhani N, Vimont S, Asselat K, Trivalle C, Minassian B, et al. (2004) CTX-M β -lactamase-producing *Escherichia coli* in long-term care facilities, France. *Emerg Infect Dis* 10: 1697–1698.
23. Leflon-Guibout V, Jurand C, Bonacorsi S, Espinasse F, Guelfi MC, et al. (2004) Emergence and spread of three clonally related virulent isolates of CTX-M-15-producing *Escherichia coli* with variable resistance to aminoglycosides and tetracycline in a French geriatric hospital. *Antimicrob Agents Chemother* 48: 3736–3742.
24. Banerjee R, Johnston B, Lohse C, Porter SB, Clabots C, et al. (2013) *Escherichia coli* sequence type 131 is a dominant, antimicrobial-resistant clonal group associated with healthcare and elderly hosts. *Infect Control Hosp Epidemiol* 34: 361–369.
25. Peirano G, Laupland KB, Gregson DB, Pitout JD (2011) Colonization of returning travellers with CTX-M-producing *Escherichia coli*. *J Travel Med* 18: 299–303.