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Effects of Alcohol on the Endocrine System

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Abstract

Synopsis—The endocrine system ensures a proper communication between various organs of the body to maintain a constant internal environment. The endocrine system also plays an essential role in enabling the body to respond and appropriately cope with changes in the internal or external environments, such as respond to stress and injury. These functions of the endocrine system to maintain body homeostasis are aided by its communication with the nervous system, immune system and body's circadian mechanism. Chronic consumption of a large amount of alcohol disrupts the communication between nervous, endocrine and immune system and causes hormonal disturbances that lead to profound and serious consequences at physiological and behavioral levels. These alcohol-induced hormonal dysregulations affect the entire body and can result in various disorders such as stress abnormalities, reproductive deficits, body growth defect, thyroid problems, immune dysfunction, cancers, bone disease and psychological and behavioral disorders. This review summarizes the findings from human and animal studies that provide consistent evidence on the various effects of alcohol abuse on the endocrine system.

Keywords

Alcoholism; endocrine disorders; circadian dysfunction and immune abnormalities

Introduction

Alcohol consumption is one of the most serious substance abuse disorders worldwide. Alcohol-related deaths, diseases and disabilities are much higher in men than women and are highest in developed countries, where they range from 8%-18% for males and 2%-4% for females. According to the National Institute of Alcohol Abuse and Alcoholism, each year, approximately 80,000 people die from alcohol-related causes, making it the third leading cause of death in the United States. Approximately 14 million of Americans (7.4%)

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have an alcohol use disorder that is classified as either alcoholism (alcohol dependency) or alcohol abuse ¹. There is considerable evidence, from human genome-wide association studies of individuals with family history of alcoholism and twin studies that several susceptibility genes are linked to the vulnerability and risk of developing alcohol-related disorders. Heritability of alcohol abuse, from twin studies, was estimated to range from 50 to 60% ^{2;3}. However, alcoholism is a multi-factorial and polygenic disorder in which complex gene-to-gene and gene-to-environment interactions occur resulting in a variety of addiction phenotypes. Environmental factors play an equally important role in the development of alcohol-related disorders, i.e. stressful life events have been shown to influence alcohol drinking and relapse behaviors ⁴.

Excessive alcohol drinking has been recognized as having several adverse health consequences. Heavy alcohol drinking increases the risk of cardiovascular and liver disease, metabolic disturbances, nutritional deficiencies, cancers (i.e. mouth, stomach, colon, liver and breast cancer), neurobiological disorders and fetal abnormalities⁵. In contrast to heavy alcohol use, light to moderate drinking, especially of alcoholic beverages rich in polyphenols such as red wine, was reported to lower the risk of coronary heart disease⁶, stroke⁷ and osteoporosis⁸. In this article we will discuss some of the literature surrounding studies done in humans and animal models regarding the effects of both acute and chronic alcohol consumption on one of the body's most important systems, the endocrine system.

The endocrine system

Along with the nervous system, the endocrine system ensures a proper communication between various organs of the body to maintain a constant internal environment, also called homeostasis. The nervous system allows rapid transmission of information between different body regions, whereas, the endocrine system, which is a complex system of glands that produce and secrete hormones directly into the blood circulation, have longer lasting actions. Almost every organ and cell in the body is affected by the endocrine system. The endocrine system controls metabolism and energy levels, electrolyte balance, growth and development and reproduction. The endocrine system also plays an essential role in enabling the body to respond and appropriately cope with changes in the internal or external environments, such as changes in the body's temperature or in the electrolyte composition of the body's fluids as well as respond to stress and injury. Substance abuse, such as chronic alcohol consumption was shown to have serious adverse effects on the different components of the endocrine system⁹. Alcohol's effects induce hormonal disturbances that lead to profound and serious consequences at physiological and behavioral levels. These alcoholinduced hormonal dysregulations affect the entire body and can result in various disorders such as cardiovascular diseases, reproductive deficits, immune dysfunction, certain cancers, bone disease and psychological and behavioral disorders.

The goal in this review is to discuss the effects of both acute and chronic alcohol exposures on the different components of the endocrine system. We will summarize the findings from human and animal studies that provide consistent evidence on the various effects of alcohol abuse on the endocrine system and on how the latter might have a role in the initiation, the development and the maintenance of alcohol drinking disorders and relapse. We will first discuss the impact of alcohol on the primary hormonal center of the endocrine system, the hypothalamic-pituitary axis and review how its different components, the adrenal axis (Hypothalamic Pituitary Adrenal axis; HPA axis), the gonadal axis (Hypothalamic Pituitary Gonadal axis; HPG axis) and the thyroid axis (Hypothalamic Pituitary Thyroid axis; HPT axis) are affected by alcohol consumption. We will also review the role that the HPA axis plays in the alcohol seeking behavior and dependence and discuss the effects of alcohol consumption on the activity of the HPA axis. Second, we will review the recent literature on the effects of alcohol on body growth, circadian mechanism and the pancreas. Lastly, and

since it is currently well documented that there is an overlap between the endocrine and the immune systems, we will discuss how dysregulation in the hypothalamic-pituitary axis can negatively impact the body's immune response.

1. Alcohol and the hypothalamic-pituitary-adrenal axis

The HPA axis, also called the stress axis, is a major component of the neuroendocrine system that controls the body's response to environmental stressors (psychological, physical or infectious) and regulates many of the body's physiological processes such as metabolism, reproduction, growth, mood and emotions and the immune function. In response to any type of stress, release of corticotropin-releasing hormone (CRH) and increased peripheral glucocorticoid levels initiate a cascade of biological responses that help counteract the altered homeostatic state. Parvocellular neurons in the paraventricular nucleus (PVN) of the hypothalamus synthesize and secrete CRH into the hypothalamic-pituitary portal network that connects the hypothalamus and the anterior pituitary. At the anterior pituitary, CRH stimulates the synthesis and secretion of a proopiomelanocortin (POMC)-derived peptide called adrenocorticotropic hormone (ACTH) from corticotropic cells. ACTH is then transported through the blood circulation to the adrenal glands where it acts on the zona fasciculata cells in the adrenal cortex to stimulate the production of glucocorticoids, mainly cortisol in humans and corticosterone in rodents. Glucocorticoids then, through a negative feedback loop, act on the hypothalamus, hippocampus and the pituitary to decrease CRH and ACTH production. This regulation is done by a feedback system via glucocorticoid receptors (GR) in the PVN, and mineralocorticoid receptors (MR) and GR in the hippocampus. Glucocorticoid receptors inhibit HPA activity; because at basal levels of cortisol, the MR are occupied while GR are mostly unoccupied. However, when there are elevated levels of plasma cortisol, like during a period of stress, there is an increased occupation of GR. It is the increased occupation of GR that triggers the negative feedback. The glucocorticoids released by the adrenal cortex interact with the GR in the pituitary, hypothalamus and hippocampus; so any over-activity results in feedback causing a reduction in circulating CRH, thereby 'switching off' the stress response. In addition to their main function in restoring homeostasis following exposure to stress, glucocorticoids influence carbohydrate, lipid and nucleic acid metabolism, bone and calcium metabolism, growth and development, it also have immuno-supressive and anti-inflammatory effects ¹⁰. Perturbations of the HPA-axis may, therefore, have serious long-term health consequences.

The other main physiological aspect of the stress response is the autonomic nervous system (ANS). The ANS has the capacity to trigger two opposing responses- the 'flight or flight' of the sympathetic nervous system or the 'tend and mend' of the parasympathic nervous system. The release of ACTH from the pituitary signals not only to induce the synthesis of glucocorticoids, but also the release of catecholamines from the adrenal glands. Catecholamines such as epinephrine and norepinephrine trigger the activation of the sympathetic nervous system (SNS) that results in the familiar feeling of decreased saliva, increased perspiration and heart rate. The SNS is the first line of defense against the stressor, traditionally allowing escape from the immediate threat. However, such a response cannot be physiologically maintained for long periods of time.

As part of the SNS response to stress, the opioid peptide $\,$ -endorphin (BEP) is synthesized by POMC and is implicated in the behavioral as well as the biological response associated with stress stimuli. The most well studied effect of BEP is its ability to modulate pain, but the peptide has also been implicated in the central regulation of ACTH and hypothalamic CRH. Indeed, BEP has been shown to play a critical role in bringing the stress response to a state of homoeostasis. In response to stress, the secretion of CRH and catecholamines stimulate the synthesis of BEP and other POMC-derived peptides from the hypothalamus, which in turn inhibit the activity of the HPA axis. Central BEP binds to $\,$ and $\,$ µ opioid

receptors and modulates the ANS via neurons within the PVN. BEP produced from pituitary POMC that circulates in the periphery is primarily regulated by CRH and AVP and has less impact on ANS function¹¹.

Considerable evidence, from human and animal studies, has shown that alcohol administration affects the HPA-axis activity. An acute exposure to alcohol activates the HPA stress axis leading to a dose-related increase in circulating ACTH and glucocorticoids. In rats, an acute administration of ethanol increases plasma ACTH and corticosterone levels, with females showing a higher response then males¹². These ethanol effects are primarily mediated through an enhanced release of CRH from the hypothalamus. Neutralization of circulating CRH with antibodies interrupts the stimulatory effects of ethanol on ACTH and corticosterone secretion¹³. Lesions of the PVN attenuate, but do not abolish the stimulatory effects of ethanol on ACTH release, suggesting that extra PVN regions, and/or other ACTH secretagogues, such as vasopressin, mediate the ethanol-stimulation of ACTH release. Neutralization of endogenous vasopressin, using antibodies, decreased the ethanol-induced ACTH secretion in both sham-operated and PVN-sectioned rats, indicating that vasopressin from outside the PVN partially mediates the pituitary-adrenocortical response to ethanol¹⁴.

In humans, several studies have also reported the stimulatory effect of alcohol on the HPA-axis. In an early study, Jenkins and Connolly¹⁵ showed that plasma cortisol levels significantly increased in healthy subjects at alcohol doses exceeding 100 mg/100 ml. In a recent study by Thayer and colleagues¹⁶, it was shown that healthy men who were self-reported alcohol consumption, had higher levels of excreted cortisol in urine. These authors also reported that in heavy drinkers, the inhibitory control of the HPA axis was impaired. Richardson and colleagues¹⁷, using an operant self-administration animal model of alcohol dependence, showed that the HPA responses to several weeks of daily 30-min self-administration of alcohol were higher in 'low-responding' non-dependent animals (< 0.2 mg/kg/session), intermediate in non-dependent animals (~0.4 mg/kg/session), and most blunted in dependent animals (~1.0 mg/kg/session). This and a number of other studies have all shown that basal ACTH and corticosterone levels are attenuated in response to chronic exposure to alcohol.

A decrease in CRH mRNA expression in the PVN¹⁸ and a reduced responsiveness of the pituitary to CRH¹⁹ were also observed. Low CRH levels were associated with more intense craving and increased probability of relapse after acute abstinence suggesting that the CRH system plays an important role in the control of long-term alcohol drinking and dependence. In CRH knock-out mice ethanol intake was doubled and the behavioral response to ethanol was diminished²⁰. In rhesus macaques, which had a single nucleotide polymorphism in the promoter region of CRH (-248C--> T) that confers increased stress reactivity, it was shown that release of ACTH, cortisol and suppression of environmental exploration (a behavioral response to social separation stress) were higher in those with the T allele²¹. These animals consumed more alcohol in a limited-access paradigm, suggesting that the CRH promoter variation which conferred increased stress responsivity also increased the risk for alcohol dependence. It has also been demonstrated that in mice lacking a functional CRH1 receptor, repeated stress leads to enhanced and progressively increasing ethanol consumption that seems to persist throughout their life²². This effect was associated with enhanced protein levels of NR2B subunit of the NMDA receptor of the glutamargic system also known to be involved in addiction behavior²³. As reported by Koob and colleagues²⁴, disruptions in the CRH system develop over time and are worsened by prolonged exposure to excessive alcohol drinking. This phenomenon is seen only in rat strains that are prone to selfadminister alcohol; in non-dependent rats, the CRH system does not significantly modulate ethanol consumption^{25, 26}.

A role for ACTH and corticosteroids has also been documented in alcohol consumption behavior. Injections of ACTH analogues to rats exposed to free-choice ethanol consumption (10% ethanol) led to a reduction in ethanol consumption²⁷. In another study by Fahlke and colleagues²⁸, male rats of alcohol-preferring (AA) and alcohol-avoiding (ANA) strains were adrenalectomized then given access to 10% (v/v) ethanol or water for 2 weeks. This study showed that adrenalectomy results in decreased ethanol consumption in AA rats compared to sham-operated AA controls and injection with corticosterone restores the ethanol consumption. In contrast, no alterations in ethanol intake after adrenalectomy and following corticosterone injection were observed in the ANA rats. These data suggest that corticosterone stimulates ethanol consumption in animals with high preference for alcohol²⁸. In a subsequent study the same authors found that intracerebroventricular infusions of corticosterone restore ethanol intake in adrenalectomized animals to levels similar to shamoperated controls ²⁹. These data suggest that differences in the activity of the HPA axis may help determine whether certain individuals are at high risk for developing alcohol-related disorders and alcoholism.

A relationship between the endogenous opioid system, the HPA axis and reward has been proposed³⁰. BEP neurons in the arcuate nucleus of the hypothalamus both inhibit CRH release in the PVN of the hypothalamus³¹ and simultaneously stimulate dopamine release in the nucleus accumbens³². Because the opioids are involved in regulating both of these systems, it is proposed that a deficit in opioid neurotransmission may cause a simultaneous derangement of both physiological processes. In recent years, a body of research has accumulated that provides evidence of deranged HPA axis function in drinking alcoholics³³ in acute withdrawal and more prolonged abstinence¹⁷, as well as in nondependent individuals with a family history of alcoholism³⁴. Also, glucocorticoids, which are released during stress and following ethanol-induced activation of the HPA axis, modulate the activities of the opioidergic, CRH, and mesolimbic dopaminergic systems³⁵ and have been shown to interact with the rewarding properties of alcohol abuse³⁶. It is beleived that diminished opioid activity, which is either the result of alcoholism or genetically linked to the risk of alcoholism, could induce hypercortisolemia, alter mesolimbic DA production, and lead to abnormal ethanol reinforcement^{32, 33}.

2. Alcohol and the hypothalamic-pituitary-gonadal axis

Alcohol abuse and alcoholism are associated with disorders of reproductive function in both men and women. The hypothalamic-pituitary-gonadal axis (HPG) and its hormones are essential for proper functioning of reproductive system. In alcohol abusers, the HPG dysfunction was shown to be associated with a decrease in libido, infertility and gonadal atrophy. Several studies have clearly documented that alcohol has deleterious effects on all three components of the HPG axis, the hypothalamus, pituitary, and gonads. We will review some of these studies on the acute and chronic effects of alcohol on male and female reproductive systems.

2.1. Overview of male and female hypothalamic-pituitary-gonadal axis—The

hypothalamus in a pulsatile way produces and secretes the hormone called luteinizing hormone–releasing hormone (LHRH), also called gonadotropin-releasing hormone (GnRH), into the hypothalamic-pituitary portal network³⁷. At the anterior pituitary, LHRH binds to specific receptors on gonadotroph cells and stimulates a cascade of events that lead to production and secretion of two important gonadotropin hormones, follicle–stimulating hormone (FSH) and luteinizing hormone (LH) into the general circulation. In the ovary, during the follicular phase of each reproductive cycle (28 days in human cycle), FSH stimulates the development of a dominant follicle which, as it matures, produces and secretes increasing amounts of the estrogen called estradiol. Both FSH and LH stimulate

estradiol secretion and this rise in estradiol is responsible for the LH and FSH surge seen in midcycle. LH then stimulates ovulation and the development of the corpus luteum during the luteal phase which then produces and secretes progesterone, an important hormone in the preparation of the uterine wall for the fertilized egg and for the maintenance of the pregnancy. In the testis, LH stimulates testosterone secretion while FSH controls the initiation and maintenance of spermatogenesis. In addition, testosterone, estrogen and progesterone, control their own production through a feedback loop mechanism and can act on the hypothalamus and the pituitary to either inhibit or stimulate the release of LHRH, LH and FSH³⁸.

Although, sex hormones are essential to reproductive processes, they can also act on other organs in the body and trigger numerous physiological processes. For example, estrogen, progesterone and testosterone were shown to play an important role in maintaining normal bone mass^{39, 40}. Estrogen is a vasodilator agent that can induce vascular relaxation⁴¹ and was shown to reduce the risk of atherosclerosis and therefore cardiovascular disease in menopausal women by decreasing circulating low-density lipoproteins (LDL) and inflammatory processes in the vasculature⁴². Testosterone was also shown to impact muscle mass and adiposity in adult men⁴³ and to affect emotional and cognitive behavior⁴⁴. Dysregulation of the hypothalamic-pituitary-gonadal axis, therefore, can lead not only to reproductive dysfunction but also to other serious health problems such as mood and memory disorders, osteoporosis and muscle atrophy.

2.2 Alcohol and hypothalamic-pituitary-gonadal axis in puberty—Normal initiation and progression of puberty is under the control and is mediated by central inputs which stimulate the pulsatile diurnal secretion of LHRH into the hypothalamic-pituitary portal system. LHRH then stimulates the pituitary gonadotropin secretion and subsequent ovarian maturation⁴⁵. This LHRH surge, which is normally inhibited during childhood through hypothalamic inhibitory inputs such as gamma aminobutyric acid and opioid peptides is triggered at puberty by stimulatory agents such as insulin-like growth factor-1 (IGF-1), norepinephrine, leptin, transforming growth factor (TGF)— and the kisspeptins⁴⁶⁻⁴⁸.

Little research has been done on the impact of alcohol consumption during puberty in humans. However, numerous studies documented decrease in estrogen levels adolescent girls and that this decrease was sustained for prolonged periods after consumption of a moderate amount of alcohol⁴⁹. Alcohol abuse was also shown to induce alterations in puberty-related hormones in adolescent boys evidenced by significant reductions in testosterone, LH and FSH levels⁵⁰. In the last decade or so, however, the use of animal models such as rodents and monkeys helped in understanding and identifying some of the mechanisms by which alcohol abuse affects puberty-related processes. In early studies on female rats, Bo and colleagues⁵¹ reported that puberty, measured by vaginal opening, was markedly delayed in prepubertal female rats administered alcohol. Naltrexone which is a blocker of the opioid receptors completely prevented the alcohol-induced delay in vaginal opening⁵², which suggests that alcohol-induced pubertal delay might in part be due to an increased opioid restraint on the normal progression of pubertal processes. It was also documented that alcohol decrease hypothalamic secretion of LHRH levels⁵³. Moreover, these same authors reported that alcohol increased hypothalamic growth hormone releasign hormone (GRH) content which was associated with a decrease in circulating growth hormone (GH)⁵⁴, suggesting that alcohol decreased GH secretion as a result of decreased GRH release from the hypothalamus. This alcohol-induced decrease in GH was also associated with a decrease in circulating IGF-I, which could explain the growth impairments observed in animals administered alcohol⁵⁵.

In another study on immature female rhesus macaques, it was shown that alcohol consumption (2 g/kg) for 12 months resulted in the suppression of the night-related increase in circulating growth hormone (GH) that is expected to occur during late juvenile development. This effect of alcohol was associated with a significant decline in circulating IGF-I, LH and estrogen levels which were most pronounced at 32 months of age. FSH and leptin levels however were not altered by alcohol consumption. The authors also found that alcohol affected the monthly pattern of menstruation which was reflected by lengthening of the intervals between menses⁵⁶. In a more recent study, the same group of researchers suggested that the alcohol damaging action on ovaries is partly mediated through an elevated ovarian nitric oxide synthase and suppressed steroidogenic acute regulatory protein, two important intermediates in steroid hormone production⁵⁷. These studies demonstrate that alcohol abuse induces hormonal alterations of HPG axis and GH/IGF-I axis during puberty. HPG activity and GH/IGFI secretion during this stage of development are closely interconnected. In fact, estrogen can stimulate GH secretion⁵⁸ and IGFI can stimulate LHRH secretion⁵⁹ suggesting that HPG axis activation leads to both sexual maturation and a growth spurt mediated through estrogen-induced stimulation of the GH--IGF axis. Therefore, alcohol-induced disturbances in HPG axis activity, during this critical stage of development, could have far reaching consequences on reproductive function as well as growth which might persist through adult life.

2.3 Alcohol and the female hypothalamic-pituitary-gonadal axis—Alcohol use in premenopausal women, even in moderate amounts, was linked to a multitude of reproductive disorders such as irregular menstrual cycles, anovulation, increased risk of spontaneous abortions and early menopause. Jensen et al.⁶⁰ in a study on 430 healthy women age 20 to 35 years who were trying to conceive for the first time, found that alcohol intake, even as few as five or less drinks per week, was associated with decreased fecundability. In another study on 26 healthy nonalcoholic women that were classified as heavy (~7.81 drinks/day), social (~3.84 drinks/day) and occasional (~1.22 drinks/day) alcohol users, Mendelson and Mello found that 50% of social drinkers and 60% of heavy drinkers had significant disturbances of reproductive hormones and of menstrual cycle compared to occasional drinkers and the social drinkers who consumed less than 3 drinks/ day. Social drinkers had anovulatory cycles and heavy drinkers had hyperprolactinemia^{61, 62}. It was reported that alcohol intake increases estradiol levels in humans⁶³ and in rodents⁶⁴. The elevated estradiol level may in part explain the negative effect of alcohol on menstrual cycle and on its irregularity. Long-term moderate alcohol consumption was also shown to decrease ovarian reserve which was associated with increased FSH levels⁶⁵. Faut and colleagues⁶⁶ have suggested that an in-situ metabolism of ethanol to acetaldehyde increases the susceptibility of rat ovarian tissue to oxidative stress and leads to cell damage and ovarian dysfunction.

In postmenopausal women receiving hormone therapy, acute exposure to alcohol induces a temporary increase in estradiol levels which might be due to impaired estradiol metabolism, with decreased conversion of estradiol to estrone. In contrast, alcohol exposure had no effect on estradiol levels in women who were not receiving hormone therapy⁶⁷.

2.4 Alcohol and the male hypothalamic-pituitary-gonadal axis—There are numerous studies in animals and humans that amply document the deleterious effects of alcohol on male reproductive function. Associations between both acute and chronic alcohol consumption and lower testosterone levels have been clearly demonstrated. Muthusami and colleagues, in a study on 66 alcoholic and 30 non-alcoholic men, found that chronic alcohol consumption significantly increased FSH, LH, and estrogen levels, whereas testosterone and progesterone were significantly decreased and prolactin (PRL) unchanged. Semen volume, sperm count, motility, and number of morphologically normal sperm were also significantly

lower in alcoholic group⁶⁸. Alcohol abuse results in hypogonadism even in the absence of liver disease. Alcoholic men with cirrhosis have elevated circulating estradiol and estrone levels⁶⁹. Ethanol increases aromatase activity, an enzyme that converts androgens to estrogens, especially in the liver⁷⁰. A decrease in IGF-I bioavailability as a result of liver disease, contributes at least in part to the development of hypogonadism associated with cirrhosis⁷¹ since IGF-I is known to stimulate testosterone synthesis and spermatogenesis⁷².

Both acute and chronic exposure of pubertal young male rats to alcohol induced a profound decrease in testosterone concentration that was associated with lower or normal LH and FSH levels, which is abnormal given that low testosterone in normal conditions signals to the hypothalamus to produce LHRH which in turn stimulates LH and FSH secretion and therefore stimulation of testis to produce testosterone⁷³. However, in an another study these authors found that chronic ethanol feeding in pubertal male rats significantly decreased testosterone but did not produce any changes in circulating FSH and LH levels⁷⁴. These authors also found that chronic ethanol exposure induced an increase in free radical damage at the pituitary level resulting in inappropriate serum levels of FSH and LH in response to lower testosterone⁷⁴.

Alcohol metabolism, by generating highly toxic reactive oxygen species, such as anion superoxide, hydrogen peroxide, hydroxyl radicals, can induce cell damage in the testes. It was shown that acetaldehyde, a metabolite of alcohol, was more toxic than alcohol alter testosterone production by inhibiting protein kinase C, a key enzyme in testosterone synthesis^{75, 76}. Nitric oxide (NO), also synthesized in the testes, has been suggested as another player in the alcohol's reduced production of testosterone. Inhibition of the NO synthase (NOS), enzyme responsible for NO synthesis, prevents the decrease in testosterone associated with alcohol drinking⁷⁷.

2.5. Alcohol and Pituitary Prolactin—Prolactin's action on the mammary gland and in the maintenance of lactation has long been known and, indeed, is the source of the name for this hormone. It is one of the most abundant hormones in the pituitary gland and is produced and secreted by lactrope cells. Common manifestations of elevation of plasma PRL (hyperprolactinemia) in women include amenorrhea (lack of menstrural cycles) and galactorrhea (excessive secretion of milk). Men with hyperprolactinemia typically show hypogonadism, with decreased sex drive, low sperm production and impotence. Such men often show breast enlargement (gynecomastia), but very rarely produce milk. A pituitary microadenoma or hyperplasia is the cause of hyperprolactinemia in most patients⁷⁸.

There are several reports showing evidence for the existence of hyperprolactinemia in chronic alcoholic men and women. In a study conducted by European scientists, persistent hyperprolactinemia was observed in 16 alcoholic women during a 6-week treatment trial⁷⁹. These patients reported daily alcohol intake of 170 g for a 2- to 16-year period but had no clinical evidence of alcoholic liver cirrhosis. In a study reported by Japanese scientists, 22 of 23 women admitted for alcoholism treatment had PRL levels above normal, ranging between 27 and 184 ng/ml. These women reported drinking an average of 84 g of alcohol each day for at least 7 years. None of these patients showed liver cirrhosis, but 10 had hepatitis and the rest had fatty liver⁸⁰. Studies conducted in a Massachusetts hospital reported hyperprolactinemia (22-87 ng/ml) in 6 of 12 alcohol-dependent women who had a history of drinking 75-247 g of alcohol per day for a minimum period of 7 years⁸¹. Alcoholinduced hyperprolactinemia was also reported in healthy, well-nourished women during residence on a clinical research ward for 35 days⁶². Sixty percent of women in the heavy drinker category (blood alcohol level, BAC: 109-199 mg/dl) and 50 percent of moderate drinkers (BAC: 48-87 mg/dl) showed elevated PRL levels, and many of these drinkers had elevated PRL several days after cessation of drinking. Alcohol-induced hyperprolactinemia

was also evident in 66 postmenopausal women⁸². Alcoholic men also showed elevated plasma levels of PRL^{83, 84}. Thus, it appears that chronic alcohol intake in humans promotes hyperprolactinemia.

Alcohol-induced hyperprolactinemia has also been demonstrated in nonhuman primates and laboratory animals. Studies conducted in macaque female monkeys showed that the PRL levels were elevated after chronic self-administration of alcohol (3.4 g/kg/day)^{85, 86}. Interestingly, histological examination of the pituitary gland of one monkey showed apparent pituitary hyperplasia ⁸⁶. Using the laboratory rats, we have shown that ethanol increases plasma PRL levels and pituitary weight in cyclic female rats and ovariectomized rats⁸⁷, and promotes estradiol-induced development of prolactinomas⁸⁸. Therefore, the clinical data as well as animal data suggest that ethanol consumption is a positive risk factor for prolactinomas and hyperprolactinemia.

3. Alcohol and the hypothalamic-pituitary-thyroid axis

The hypothalamic-pituitary-thyroid axis is responsible for the regulation of metabolism in every cell in the body. When circulating levels of thyroid hormones, thyroxin (T4) and triiodothyronine (T3) are low, the hypothalamus responds by releasing thyrotropin-releasing hormone (TRH), which then stimulates thyrotrope cells in the anterior pituitary to produce and secrete thyroid-stimulating hormone (TSH). TSH then stimulates the synthesis and secretion of T4 and T3 from the thyroid gland. T3 is the active form of thyroid hormones and although T4 and T3 are both secreted following TSH stimulation, 80% of circulating T3 comes from liver conversion of T4 by enzymes called deiodinases. T4, but primarily T3 by negative feedback at the hypothalamus and the pituitary, can control their own release by inhibiting TRH and TSH release⁸⁹.

Alcoholic individuals often show dysregulations of the hypothalamic-pituitary-thyroid axis. A significant reduction in T4 and T3 concentrations was observed in the alcoholic groups during withdrawal and early abstinence, compared to non-alcoholic healthy groups ⁹⁰. In addition a blunted response of TSH to TRH has been consistently reported in alcoholics and during early withdrawal and was positively correlated with severity of withdrawal symptoms ⁹¹. It was also documented that, after longer periods of abstinence, thyroid dysfunction recovers and thyroid hormones and TSH response to TRH return to normal levels ⁹¹. However, in individuals who relapsed and returned to their alcohol drinking behavior, lower T4 and T3 and a blunted TSH response to TRH were again observed ⁹².

Similar results were found in studies done on animal models. For example, chronic alcohol administration to adult male rats in a liquid diet containing 10% (w/v) ethanol for 40 days induced a significant decrease in total T4 and T3, free T4 and T3 as well as basal TSH when compared to control animals fed an isocaloric diet⁹³. In addition, chronic ethanol treatment was shown to induce an increase in TRH mRNA content in neurons of the PVN, however the peripheral stimulation of thyroid hormones by cold exposure was absent in rats⁹⁴, suggesting that chronic exposure to ethanol induces thyroid gland dysfunction, which no longer is able to properly respond to TRH stimulation. Furthermore, down-regulation of TRH receptors in the pituitary has been proposed as a mechanism by which chronic alcohol induces the blunted TSH response to TRH.

Other mechanisms for ethanol action on HPT axis have been proposed, which focused on thyroid hormone metabolism and on the activity of enzymes that catalyze the conversion of T4 to T3 (5 II deiodinase) and inactivate T3 to 3,3 -T2 (5-II deiodinase). Baumgartner et al. 95 in a study in "behaviorally dependent" and ethanol-exposed but "not dependent" rats, found that the activity of the 5 II deiodinase isoenzyme was elevated in the frontal cortex in both groups of rats. The activity of the 5-II deiodinase isoenzyme, however, was selectively

inhibited in the amygdala of the rats who were "behaviorally dependent" on ethanol, but was normal in the "non-dependent" rats. These authors suggested that increases in intracellular concentrations of T3 in the amygdala may be involved in the development of dependence behaviors to alcohol.

4. Alcohol and growth hormone insulin-like growth factor-1 axis

In the hypothalamic-pituitary-growth hormone-IGF-1 axis, growth hormone releasing hormone (GHRH) is secreted from cells in the arcuate and ventromedial nuclei of the hypothalamus into the hypophyseal portal system. GHRH then acts on somatotropic cells in the anterior pituitary where it stimulates the production and release of GH, which is then released in the general circulation and stimulates IGF-1 mainly in the liver. IGF-I through negative feedback at the hypothalamus and pituitary reduces GH synthesis and release. Another hormone called somatostatin, which is secreted in the PVN of the hypothalamus also acts on the pituitary and inhibits GH secretion. Between GHRH, somatostatin and IGF-I, the amount of GH secreted by the anterior pituitary is tightly regulated. Together GH and IGF-I regulate important physiological processes in the body such as, postnatal growth and development, carbohydrate and lipid metabolism⁹⁶. Numerous studies in both humans and experimental animals have shown that acute and chronic alcohol exposure reduces circulating GH and IGF-I levels. Acute exposure of healthy men to ethanol (1.5g/kg) was shown to reduce the nightly peak of GH secretion⁹⁷. This effect did not seem to be mediated through a direct action of ethanol on the pituitary rendering it less sensitive to GHRH since intravenous injection of exogenous GHRH induced an increase in GH secretion in both groups ethanol-treated (1g/kg) and control men⁹⁸. However, an acute exposure of healthy women to ethanol had no significant effects on GH secretion⁹⁹. Sonntag and Boyd¹⁰⁰ found that, in rats exposed to 5% ethanol in a liquid diet for 4.5 months ethanol induced a significant decrease in circulating IGF-1 levels. Soszynski and colleagues reported, that a 6 day administration of 5% ethanol to chronically cannulated unanesthetized rats resulted in 75 to 90 % decrease in spontaneous GH secretion in rats¹⁰¹. In addition, IGF-I serum levels and GHRH mRNA levels were significantly decreased with no change in somatostatin or GH mRNA levels following ethanol treatments ¹⁰². These results suggest that chronic ethanol affects GH secretion primarily at the hypothalamic level where it induces impairments in GHRH gene expression.

Chronic alcohol consumption was shown to induce muscle wasting ¹⁰³ and since IGF-I is a potent anabolic agent that regulates muscle protein balance, this next study showed that rats fed ethanol for 8 weeks had significantly lower IGF-I in plasma, liver and skeletal muscle compared to pair-fed control ¹⁰⁴. The IGF-I mRNA level was also lower in liver and skeletal muscle. However, the concentration of IGF binding protein (IGFBP)-1 was increased in plasma, liver and muscle; and no changes in glucocorticoids or insulin or GH were reported in these ethanol-fed rats compared with controls ¹⁰⁴. These results suggest that the ethanol mediated-changes in IGF-I and IGFBP-I concentrations are mediated through mechanisms that are independent of classical hormonal regulators of the IGF system (i.e. GH, high glucocorticoids and low insulin-induced hypoglycemia).

5. Alcohol and circadian rhythm

In all organisms, especially in mammals, a time-keeping system called the master circadian pacemaker located in the suprachiasmatic nucleus (SCN) of the anterior hypothalamus synchronizes biological rhythms in response to external cues. The circadian rhythm generated in the SCN, in response to external cues such as day light, is converted into neuronal or hormonal signals that affect the entire body's physiological and metabolic processes, thereby optimizing the organism's interaction with changing environmental conditions. The circadian clockwork involves the interaction of specific clock genes,

including Period (*Per*1, *Per*2, *Per*3), Clock, Bmal1, and Cryptochrome (*Cry*1, *Cry*2) genes within two tightly interlocked transcriptional and translational feedback loops that sustain a near 24 h period of cellular activity and regulates the expression of downstream clock-controlled genes. These processes occur not only in the master pacemaker, the suprachiasmatic nucleus, but also in other regions of the brain as well as in the peripheral cells in the entire body^{105, 106}.

Numerous studies in both, humans and animal models, showed that chronic exposure to alcohol induces alterations in the activity of clock genes resulting in severe desynchronizations of physiological clock systems such as sleep, body temperature, blood pressure and hormonal secretions; and vice versa clock genes influence alcohol use and abuse behavior¹⁰⁷. Our own work have shown that chronic exposure of prenatal or postnatal rats to ethanol induces alterations in the circadian expression of POMC and the clock genes r*Per*1, r*Per*2 and r*Per*3 in the arcuate nucleus and r*Per*1 and r*Per*2 levels in SCN during the adult period¹⁰⁸.

The circadian clock and clock gene activity not only controls the sleep/wake and feeding/ fasting behavior, but it also influences hormone release. The different endocrine axes function is under tight circadian control and dysregulations in the activity of clock genes might therefore affect the oscillatory characteristic of hormone secretion. Jimenez and colleagues ¹⁰⁹, in a study on peripubertal male rats, showed that chronic alcohol exposure for 4 weeks not only induced significant changes in hypothalamic-pituitary hormone levels but also affected their 24 hour secretory pattern. Two peaks of FSH during the inactive phase of the daily cycle appeared instead of one, no LH plasma surge during the first part of the inactive phase, a second peak of testosterone and prolactin during the second part of the inactive phase and a second peak of plasma TSH during the first part of the active phase were observed. In addition, an interaction between circadian clock and HPA axis responsiveness to stress has been consistently linked to alcohol abuse and dependence. Stress is regarded as a major environmental risk factor for both, heavy drinking 110 and disturbed circadian rhythmicity^{110, 111}. In a study on 268 young adults (126 males, 142 females) 19 years of age, a positive association between the major A allele of *PER2* rs56013859 and alcohol consumption was found in homozygote individuals¹¹². Moreover, under conditions of stress carriers of the G allele drunk less than those homozygous for the A allele. These findings suggest a role of a PER2 gene variant in both the drinking patterns as well as the stress responsiveness. In another study, *Per*3 gene was also linked to stress/ anxiety traits and its basal expression correlated with addiction-related phenotypes. Exposure to alcohol increased *Per3* expression in the hippocampus, and this was shown to affect stress response¹¹³. In animal studies, it was shown that *Per*2 mutant mice had impairments in the glutamate transporter Eaat1 resulting in decreased glutamate uptake by astrocytes and its accumulation in the extracellular milieu; this was associated with increased alcohol intake in these Per2 mutant mice¹¹⁴. These researchers found that treating these Per2 mutant mice with acamprosate, a drug used in relapse prevention of alcoholism which is thought to attenuate the hyper-glutamatergic state, reduced the increased glutamate levels and normalized alcohol intake in these mice. A significant association between a single nucleotide polymorphism variation in the human Per2 gene, PER2 SNP 10870, (A/G substitution) and regulation of alcohol consumption was shown in a clinical sample of severe alcoholics¹¹⁴. In addition to this interaction between the stress and circadian rhythm, a large number of data suggest that the disruption in circadian clocks mechanism negatively impact immune system functions and promotes cancer susceptibility 115, 116. Taken together, these studies suggest the important role that clock genes play in the alcohol-mediated dysregulation of the circadian nature of endocrine function as well as in the alcohol preference and abuse behaviors.

6. Alcohol and pancreatic function

The pancreas is one of the most important organs of the endocrine system that is involved in the tight control of blood glucose concentration through synthesis and secretion of a peptide hormone called insulin from β-cells. Diabetes mellitus (DM) is a syndrome of dysregulated metabolism with high blood glucose levels (hyperglycemia) due either to an abnormal insulin secretion and/or signaling in peripheral tissues. DM is characterized by either a β-cell deficit such as in insulin-dependent type 1 diabetes or reduced peripheral insulin sensitivity as in type 2 diabetes. Type 2 diabetes is recognized clinically as a complication which often occurred in alcoholics^{117, 118}. However, it was shown that the relationship between alcohol consumption and the risk of type 2 diabetes is a "U" shaped.

Low or moderate alcohol consumption shows protective effects against type 2 diabetes in some patients through enhanced peripheral insulin sensitivity¹¹⁹. A 30% reduced risk of type 2 diabetes was observed in patients with moderate alcohol consumption, whereas no risk reduction was observed in consumers of amounts of alcohol equal to or over 48 g/day. Whether moderate alcohol consumption affects insulin secretion is still very controversial. Some studies show that moderate alcohol consumption improves insulin action without affecting its secretion¹²⁰, whereas others show a reduced basal insulin secretion rate associated with a lower fasting plasma glucagon concentration¹²¹. Avogaro et al.¹²⁰ have also shown, in the same study, that the enhanced insulin sensitivity observed may be in part due to the inhibitory effect of alcohol on lipolysis. The beneficial metabolic effects of moderate alcohol consumption on insulin sensitivity and glucose tolerance may explain the significant reduction in the development of type 2 diabetes and the risk of cardiovascular disorders reported in several epidemiological studies^{120, 122}.

Heavy alcohol consumption, on the other hand, is an independent risk factor for the development of Type 2 diabetes mellitus¹²³. In addition to its effects on peripheral tissues, such as adipose tissue and liver, where it induces insulin resistance, heavy alcohol consumption was also proposed to negatively impact pancreatic \(\beta \)-cell function. Patto and colleagues ¹²⁴, in a study on 16 healthy volunteer non-alcohol consumers and 10 chronic alcohol consumers, found that there was a decrease in insulin response in the chronic alcohol consumers compared to the control group. They measured total integrated response (TIR) values for Insulin and c-peptide following oral or intravenous glucose loads in these two groups. They found that the insulin and c-peptide TIR values were both significantly lower in the alcoholic group compared to the control group. In addition, in both groups the insulin TIR values following the oral glucose load were significantly higher than those following the intravenous glucose load, suggesting an enhancing incretin effect on insulin secretion. These authors concluded that the decreased insulin response observed in alcoholics was due to a ß-cell dysfunction, not to an enteroinsular axis dysfunction because the decrease was also observed when a glucose load was given intravenously. In another study by Kim and colleagues¹²⁵ it was shown that in mice exposed for 8 to 10 weeks to chronic alcohol, there was a significant increase in the impairment of fasting glucose and an increase in -cell apoptosis, which were associated with a reduction in insulin secretion. These effects of chronic ethanol seemed to be mediated through a down-regulation and inactivation by tyrosine nitration of the enzyme glucokinase, a critical player in glucose metabolism that leads to an increased ATP production and therefore to insulin secretion by beta-cells. These effects were also associated with a decrease in Glut2 and insulin expression, which exacerbates alcohol action. In an in-vitro study on RINm5F -cell line, it was suggested that ethanol generates reactive oxygen species (ROS) and induces -cell apoptosis 126. All of these studies suggest that heavy alcohol consumption has deleterious effects on pancreatic cell function and on glucose homeostasis.

7. Alcohol and immune system

The neuroendocrine and immune systems are highly interrelated. A bidirectional interaction between both systems has long been recognized; especially the sensitivity of the immune system to stress and its interaction with the HPA axis which was primarily evidenced by the immunosuppressive actions of glucocorticoids especially cortisol which upon binding to its specific receptor (GR) can interfere with signaling pathways of other transcription factors such as NFkB and AP-1 to repress transcription of many inflammatory molecules. In addition, CRH and ACTH were also shown to have immuno-potentiating and proinflammatory properties whereas for example interleukins and cytokines produced by activated macrophages, in an adaptive feedback mechanism, can act on the HPA axis and induce CRH and ACTH secretion. For example, IL-2 was shown to stimulate cholinergic neurons and activate neural nitric oxide synthase (nNOS) which releases nitric oxide (NO). After release, NO diffuses into CRH producing neurons and induces CRH secretion which ultimately leads to ACTH release from anterior pituitary and cortisol from adrenal glands. This bidirectional interaction between HPA axis and immune system is essential for survival and for maintenance of the body's homeostasis. However, in alcohol abuse conditions HPA axis and immune system function is compromised and contributes to a worsened state. Glucocorticoids, at excessive amounts have serious negative effects; first through their immunosuppressive action and second through induced acute and chronic metabolic abnormalities 127,128.

In recent years, a body of evidence from human as well as animal studies established a link between long-term alcohol use and alterations of both HPA axis and immune system functions. Alcohol abuse disorders are often associated with chronic systemic inflammation and high circulating proinflammatory cytokine levels as well as high circulating cortisol levels. Two mechanisms through which alcohol induces inflammation have been proposed; first, the gut microflora-derived lipopolisacharides (LPS) were suggested as key players in alcohol-mediated inflammation 129 and second, alcohol metabolism through production of reactive-oxygen species (ROS) and cell damage triggers the production of pro-inflammatory cytokines such as TNF- and IL-6¹³⁰. An alcohol-induced systemic inflammation that persists, in the case of alcohol abuse, has far reaching damaging actions on every organ of the body. In the brain, alcohol has neurotoxic effects that result in neuronal death and neurodegeneration ^{131, 132}. Alcoholics have been shown to have reduced brain mass, cortical neuronal loss and impaired cognitive functions ¹³³, ¹³⁴. Studies from our laboratory as well as from others have clearly shown that ethanol exposure during the developmental period induces neurotoxicity and permanent impairments in the stress axis as well as the immune function ^{131, 135, 136}. Microglial cells, the macrophage cells in the brain, were shown to play an important role in these neurotoxic effects of alcohol on neuronal cells ¹³⁷⁻¹³⁹.

Acute or chronic alcohol exposures have both been shown to induce immunosuppression through dysregulation in all branches of the immune system. Alcohol exposure reduces neutrophil (macrophage) infiltration and migration to sites of infection as well as production of new neutrophils in response to infection and their phagocitic activity ¹⁴⁰, ¹⁴¹. Chronic alcohol exposure also decreases monocyte phagocytic activity even though the number of these cells is increased ¹⁴². Furthermore, chronic alcohol exposure decreases the number and activity of dendritic and natural-killer cell (NK-cells) ¹⁴³⁻¹⁴⁵. Alcohol's effect on cytokine production depends on the length of exposure. For example, acute exposure to ethanol is associated with suppression of cytokine production (i.e. TNF and IL1ß) ¹⁴⁶. However, chronic exposure to alcohol induces an increase in pro-inflammatory cytokine production such as that of TNF ¹⁴⁷, ¹⁴⁸. In addition, alcohol exposure suppresses chemokine production, such as that of MIP2 (macrophage inflammatory protein 2) and IFN (interferon gamma) which were shown to be suppressed following acute exposure to ethanol ¹⁴⁹, ¹⁵⁰. All these studies clearly suggest that ethanol disruption of cytokines and inflammation

contributes in a multitude of ways to a diversity of alcoholic disorders. These alcoholinduced suppressive effects on the immune function were shown to be primarily due to a blunted response of the HPA axis to external stressors such as infections. In this context it is notable that stress axis dysfunction is positively connected with not only immune incompetence but also promotion of various cancers \$^{11,151}\$. More studies are needed to further understand and identify the mechanisms that underlie the bidirectional interactions between immune and endocrine system in the case of alcohol-related disorders which will help in the management and treatment of these disorders.

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Abbreviation used

AA alcohol preferring

ACTH adrenocorticotropic hormone

ANA alcohol non-preferring

ANS autonomic nervous system

AP-1 activator protein 1

BAC blood alcohol concentration

BEP β-endorphin

CRH corticotropin-releasing hormone

DA dopamine

DM diabetes mellitus

FSH follicle-stimulating hormone

GH growth hormone

GnRH gonadotropin-releasing hormone

GR glucocorticoid receptor

GRH growth hormone-releasing hormone

HPA hypothalamic pituitary adrenalHPG hypothalamic pituitary gonadalHPT hypothalamic pituitary thyroid

IFN interferon gamma

IGF-1 Insulin-like growth factor-1

IGFBP-1 IGF binding protein 1

IL-1ß interleukin-1ß
IL-6 interleukin-6

LDL low density lipoproteins
LH luteinizing hormone

LHRL Luteinizing hormone-releasing hormone

LPS lipopolisaccharides

MIP2 macrophage inflammatory protein 2

MR mineralocorticoid receptors

NFkB nuclear factor kB
NK cells natural killer cells

NMDA N-methyl-D-aspartate

NO nitric oxide

NOS nitric oxide synthase
NOS nitric oxide synthase
NR2B NMDA receptor 2B

Per period

POMC proopiomelanocortin

PRL prolactin

PVN praventricular nucleus
ROS reactive oxygen species
SCN suprachiasmatic nucleus

SNS sympathetic nervous system

T3 triiodothyronine

T4 thyroxin

TGF- transforming growth factor alpha

TIR total integrated response

TNF- tumor necrosis factor-alpha

TSH thyroid-stimulating hormone

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KEY POINTS

 Chronic consumption of a large amount of alcohol disrupts the communication between nervous, endocrine and immune system and causes hormonal disturbances that lead to profound and serious consequences at physiological and behavioral levels.

These alcohol-induced hormonal dysregulations affect the entire body and can
result in various disorders such as stress abnormalities, reproductive deficits,
body growth defect, thyroid problems, immune dysfunction, cancers, bone
disease and psychological and behavioral disorders.

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Table 1

Summary of hormonal changes induced by acute and chronic alcohol exposure.

Endocrine Gland	Hormone	Acute	Acute alcohol	Chroni	Chronic alcohol
		Male	Female	Male	Female
	CRH				
	LHRH				
Hypothalamus	TRH				
	GHRH				
	Somatostatin				
	ACTH				
	ГН				
7	FSH				
Antenor Fituntary Gland	HSH				
	НЭ				
	Prolactin				
Adrenal cortex	Cortisol				
Testes	Testosterone				
	Estrogen				
Ovanes	Progesterone				
T TO THE	T4				
Thyrona Orana	Т3				
Pancreas	Insulin				

Table legends: Uncreased hormone release; Decreased hormone release; Unchanged hormone release

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