

## CARBAPENEM-RESISTANT *ACINETOBACTER BAUMANNII* OUTBREAK AT UNIVERSITY HOSPITAL

E.H. Takagi<sup>1</sup>; N. Lincopan<sup>1</sup>; V.C. Cassettari<sup>2</sup>; L.F. Passadore<sup>3</sup>; E.M. Mamizuka<sup>1,2</sup>; M.B. Martinez<sup>1,2,3</sup>

<sup>1</sup>Laboratório de Microbiologia Clínica, Faculdade de Ciências Farmacêuticas, Universidade de São Paulo, São Paulo, SP, Brasil;

<sup>2</sup>Comitê de Controle de Infecção Nosocomial, Hospital Universitário, Universidade de São Paulo, São Paulo, SP, Brasil;

<sup>3</sup>Serviço de Laboratório Clínico, Hospital Universitário, Universidade de São Paulo, São Paulo, SP, Brasil

Submitted: June 03, 2008; Returned to authors for corrections: July 07, 2008; Approved: February 14, 2009.

---

### SHORT COMMUNICATION

---

#### ABSTRACT

Nineteen clonally related imipenem-resistant *Acinetobacter baumannii* isolates were recovered from eight intensive care unit patients. All isolates harboured *bla*<sub>OXA-51</sub>-like  $\beta$ -lactamase genes and showed the absence of 22 kDa fraction in outer membrane porin profile analysis. It suggests a combination of two mechanisms as responsible for carbapenem-resistant phenotypes.

**Key words:** *Acinetobacter*, *bla*<sub>OXA</sub>-type genes, carbapenemases, nosocomial infection, Brazil

---

#### INTRODUCTION

In Brazilian hospitals, multidrug-resistant (MDR) *Acinetobacter baumannii* constitute a serious cause of nosocomial infection, comprising 8.8% of the total nosocomial bacterial isolates that cause infections in ICU patients, according to the MYSTIC Program Brazil (11). In this respect, carbapenems remain as the widest spectrum therapeutic option for treatment of such infections. However, resistance to these antimicrobial agents has increased, resulting in the use of potentially more toxic agents such as the polymyxins (7). Although high carbapenem resistance rates have been reported among *Acinetobacter* spp. isolated in Brazil, very little is known about their mechanisms of resistance. Recently, it has been reported that IMP-1 metallo-beta lactamase-producing *Acinetobacter* strains emerged in 1998 in some Brazilian hospitals (15). Regarding OXA-type carbapenemases, only the *bla*<sub>OXA-23</sub>-like gene has been associated with imipenem resistance in Brazil (6). We hereby report the combination of the naturally intrinsic harboured *bla*<sub>OXA-51</sub>-like gene and impermeability as mechanism responsible for imipenem-resistant phenotype in clonally related

*A. baumannii* recovered from an outbreak, in a Brazilian teaching hospital.

From September 2005 to February 2006 eleven MDR, *Acinetobacter baumannii* isolates were recovered from six ICU patients hospitalized at the Hospital Universitário da Universidade de São Paulo (HU-USP). Species identification and antimicrobial susceptibility (Table 1) were evaluated using the Vitek system (BioMérieux, Hazelwood, Mo.) and the disk diffusion method, respectively. Molecular typing was performed by Pulsed Field Gel Electrophoresis (PFGE) of *Apa*I-digested genomic DNA of *A. baumannii* isolates (17). PFGE band profiles were identical for all carbapenem-resistant strains. Minimum inhibitory concentrations (MICs) for all isolates were determined by the agar dilution method (4). Additionally, some combinations antibiotic/ $\beta$ -lactamase inhibitors were tested as follows: ceftazidime/clavulanic acid (4.0 mg/L) (4), imipenem/EDTA (320 mg/L) (22), imipenem/NaCl (200 mM) (16). All strains were found to be resistant to more than 3 antimicrobial groups (Table 1), presenting MICs  $\geq 32$  and  $\geq 64$  mg/L for imipenem and ceftazidime, respectively. The inhibitors tested did not affect MIC's values when associated with imipenem or ceftazidime.

---

\*Corresponding Author. Mailing address: Laboratory of Clinical Microbiology, Faculdade de Ciências Farmacêuticas, Universidade de São Paulo, Av. Professor Lineu Prestes 580, Cidade Universitária. CEP 05508-000, São Paulo, SP, Brasil. E-mail: mbmartin@usp.br

**Table 1.** Antibiotic susceptibilities of the *Acinetobacter baumannii* strains in this study.

Case	Origin (Specimen)	Isolation date	Hospital unit	Antimicrobial susceptibility
Case 1	Urine	05/09/2005	Adult ICU	none
Case 2	Tracheal secretion	11/01/2006	Adult ICU	ART
Case 3	Blood	12/01/2006	Adult ICU	SAM, ART
	Blood	19/01/2006	Adult ICU	SAM
	Catheter tip	19/01/2006	Adult ICU	SAM
Case 4	Catheter tip	19/01/2006	Adult ICU	SAM
	Abdominal secretion	20/01/2006	Adult ICU	SAM
	Tracheal secretion	24/01/2006	Adult ICU	SAM
Case 5	Tracheal secretion	31/01/2006	Adult ICU	SAM
	Blood	02/02/2006	Adult ICU	SAM, FEP
Case 6	Vaginal secretion	10/02/2006	Adult ICU	SAM, FEP

Table captions: All strains were tested by Kirby Bauer method for: PIP, piperacillin; TZP, piperacillin/tazobactam; CAZ, ceftazidime; CTX, cefotaxime; FEP, cefepime; ATM, aztreonam; IMP, imipenem; MEM, meropenem; CIP, ciprofloxacin; AMK, Amikacin; GEN, Gentamicin; SXT, Trimethoprim/Sulfamethoxazole; ART, aztreonam; SAM, ampicillin/sulbactam; ICU, intensive care unit.

Research on carbapenemase production and outer membrane porin profile was performed using imipenem-susceptible and non-susceptible isolates recovered from HU-USP.

Carbapenemase activity was evaluated using a bioassay (9). This test involved satellite growth of *Staphylococcus aureus* ATCC 25923 around the putative carbapenemase-producing *A. baumannii* strains growing on Muller-Hinton agar plates containing  $10^8$  CFU of ATCC strain/mL and imipenem at a concentration of 0.06 or 0.12 mg/L. Imipenemase activity was confirmed in all imipenem resistant isolates.

Metallo- $\beta$ -lactamase (MBL) production was then screened by a double disk synergy test using ceftazidime and imipenem as substrates and EDTA and thiol compounds as  $\beta$ -lactamase inhibitors (1,12). Imipenemase activity was not inhibited by EDTA or thiol compounds, suggesting that a serine-type  $\beta$ -lactamase was responsible for the hydrolysis of imipenem.

Imipenem-susceptible and resistant *A. baumannii* were screened by PCR for the *bla*<sub>IMP</sub>, *bla*<sub>VIM</sub>, *bla*<sub>SPM-1</sub>, *bla*<sub>OXA-23</sub>-like, *bla*<sub>OXA-24</sub>-like, *bla*<sub>OXA-51</sub>-like and *bla*<sub>OXA-58</sub>-like genes, as previously described (21,23). The *bla*<sub>OXA-51</sub>-like gene was the only one detected, even in imipenem-susceptible strain, confirming global reports of the intrinsic presence of class D carbapenemase in *A. baumannii* (10,20). ISAbal was also found by PCR, but the insertion sequence was not upstream of the *bla*<sub>OXA-51</sub>-like gene in any isolate (21).

Alterations in permeability were evaluated by outer membrane porin (OMP) analysis in imipenem susceptible (MIC 0.5 mg/L) and resistant *A. baumannii* isolates. The OMP fractions were prepared by the *N*-Lauryl-sarcosinate method (5). Total OMP concentration was measured according to Bradford (3). OMP profiles were analyzed by SDS-PAGE and

showed absence of an expected 22 kDa fraction in the imipenem-resistant isolates.

*A. baumannii* is recognized as playing a significant role in the colonization and infection of hospitalized patients, especially those in critical care environments. The carbapenems, such as imipenem, have been widely used to treat infections caused by MDR *A. baumannii* clinical isolates, nevertheless, regrettably, carbapenem-resistant *A. baumannii* clinical isolates have become more prevalent. In this respect, impermeability or drug inactivation by carbapenemases belonging to metallo-beta-lactamase class B or some class D OXA-type enzyme subgroups have been described as major causes of resistance.

Additionally, a decrease in outer membrane permeability has been associated with resistance to carbapenems in *A. baumannii* clinical strains (15). It was associated to with the loss of 29 kDa OMP (13), 31-36kDa (5), 25/29 kDa corresponding to the so-called CarO (2,5,18,14). Thus, isolates with weak OXA carbapenemases could be required to bear additional co-determinants of resistance, in particular, the absence of outer-membrane proteins as demonstrated by Costa *et al.* (5), whose resistant isolates had acquired two  $\beta$ -lactamases and had also lost a protein of 31-36 kDa. Bou *et al.* (2) report a multiresistant isolate that produce OXA-24 with reduced expression of two proteins 22kDa, the same lacked protein in our isolate, and 33kDa.

At the ICU from HU-USP, the outbreak involved eight cases of infection by a single RAPD-PCR clone. Carbapenem resistant phenotype was related to the lack of a 22 kDa OMP and the presence of *bla*<sub>OXA-51</sub>-like  $\beta$ -lactamase genes. Although *bla*<sub>OXA-51</sub>-like  $\beta$ -lactamase genes were the only ones identified, further studies are necessary to understand the role of these genes like resistance mechanism.

## ACKNOWLEDGEMENTS

FAPESP and CNPq research grants are gratefully acknowledged. E. H. Takagi thanks CNPq for an undergraduate fellowship.

## RESUMO

### Caracterização de cepas de *Acinetobacter baumannii* durante um surto de infecção hospitalar

Foram isoladas 19 cepas monoclonais de 8 pacientes da unidade de terapia intensiva, resistentes aos carbapenêmicos. Todas as cepas apresentaram o gene *bla*<sub>OXA-51</sub>-like e por análise do perfil de proteínas de membrana notou-se ausência da fração de 22 kDa, sugerindo a combinação de dois mecanismos de resistência aos carbapenêmicos.

**Palavras-chaves:** *Acinetobacter*, *bla*<sub>OXA</sub>-type genes, carbapenemases, infecção hospitalar

## REFERENCES

1. Arakawa, Y.; Shibata, N.; Shibayama, K.; Kurokawa, H.; Yagi, T.; Fujiwara, H.; Goto, M. (2000). Convenient test for screening metallo- $\beta$ -lactamase-producing gram-negative bacteria by using thio compounds. *J. Clin. Microbiol.*, 38, 40-43.
2. Bou, G.; Cerveró, G.; Domínguez, M.A.; Quereda, C.; Martínez-Beltrán, J. (2005). Characterization of nosocomial outbreak caused by a multiresistant *Acinetobacter baumannii* strain with a carbapenem-hydrolyzing enzyme: high-level carbapenem resistance in *A. baumannii* is not due solely to the presence of  $\beta$ -lactamases. *J. Clin. Microbiol.*, 38, 3299-3305.
3. Bradford, M.M. (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.*, 7, 248-254.
4. Clinical and Laboratory Standard Institute (2006). *Performance standards for antimicrobial disk susceptibility tests*. Approved standard M2-A9. Wayne, PA.
5. Costa, S.F.; Woodcock, M.G.; Wise, R.; Barone, A.A.; Caiaffa, H.; Levin, A.S.S. (2000) Outer-membrane proteins pattern and detection of  $\beta$ -lactamases in clinical isolates of imipenem resistant *Acinetobacter baumannii* from Brazil. *Int. J. Antimicrob. Agents.*, 13, 175-182.
6. Dalla-Costa, L.M.; Coelho, J.M.; Souza, H.A.P.H.M.; Castro, M.E.S.; Stier, C.J.N.; Bragagnolo, K.L.; Rea-Neto, A.; Penteadó-Filho, S.R.; Livermore, D.M. (2003) Outbreak of Carbapenem-Resistant *Acinetobacter baumannii* producing the OXA-23 enzyme in Curitiba, Brazil. *J. Clin. Microbiol.*, 41, 3403-3406.
7. Falagas, M.E.; Kasiakou, S.K.; Kofteridis, R.; Samonis, G. (2006). Effectiveness and nephrotoxicity of intravenous colistin for treatment of patients with infections due to polymyxin-only-susceptible (POS) gram-negative bacteria. *Eur. J. Clin. Microbiol. Infect. Dis.*, 25, 596-599.
8. Fernández-Cuenca, F.; Martínez-Martínez, L.; Conejo, M.C.; Ayala, J.A.; Perea, E.J.; Pascual, A. (2003) Relationship between  $\beta$ -lactamase production, outer membrane protein and penicillin-binding protein profiles on the activity of carbapenems against clinical isolates of *Acinetobacter baumannii*. *J. Antimicrob. Chemother.*, 53, 565-574.
9. Gots, J.S. (1945) The detection of penicillinase-producing properties of microorganisms. *Science.*, 102, 309.
10. Heritier, C.; Poirel, L.; Fournier, P.E.; Claverie, J.M.; Raoult, D.; Nordmann, P. (2005). Characterization of the naturally occurring oxacillinase of *Acinetobacter baumannii*. *Antimicrob. Agents Chemother.*, 49, 4174-4179.
11. Kiffer, C.; Hsiung, A.; Oplustiu, C.; Sampaio, J.; Sakayami, E.; Turner, P.; Mendes, C. (2005). Mystic Brazil Group. Antimicrobial susceptibility of gram-negative bacteria in Brazilian hospitals: the MYSTIC program Brazil 2003. *Braz. J. Infect. Dis.*, 9, 216-224.
12. Lee, K.; Lim, Y.S.; Yong, D.; Yum, J.H.; Chong, Y. (2003). Evaluation of the Hodge Test and the imipenem-EDTA double-disk synergy test for differentiating metallo- $\beta$ -lactamase-producing isolates of *Pseudomonas* spp. and *Acinetobacter* spp. *J. Clin. Microbiol.*, 41, 4623-4629.
13. Limansky, A.S.; Mussi, M.A.; Viale, A.M.; (2002). Loss of a 29-kilodalton outer membrane protein in *Acinetobacter baumannii* is associated with imipenem resistance. *J. Clin. Microbiol.*, 40, 4776-4778.
14. Martí, S.; Sánchez-Céspedes, J.; Oliveira, E.; Bellido, D.; Giralt, E.; Vila, J. (2006) Proteomic analysis of a fraction enriched in cell envelope proteins of *Acinetobacter baumannii*. *Proteomics.*, 6, 82-87.
15. Peleg, A.Y.; Seifert, H.; Paterson, D. (2008). *Acinetobacter baumannii*: emergence of a successful pathogen. *Clin. Microbiol. Rev.*, 21, 538-582.
16. Pournaras, S.; Markogiannakis A.; Ilkonomidis, A.; Kondyli, L.; Benthimounti, K.; Maniatis, A.N.; Legakis, N.J.; Tsakris, A. (2006). Outbreak of multiple clone of imipenem-resistant *Acinetobacter baumannii* isolates expressing OXA-58 carbapenemase in an intensive care unit. *J. Antimicrob. Chemother.*, 57, 555-561.
17. Seifert, H.; Dolzani, L.; Bressan, R.; Reijden, T.; Strijen, B.; Stefanik, D.; Heersma, H.; Dijkshoorn, L. (2005). Standardization and interlaboratory reproducibility assessment of Pulsed-Field gel electrophoresis-generated fingerprints of *Acinetobacter baumannii*. *J. Clin. Microbiol.*, 43, 4328-4335.
18. Siroy, A.; Molle, V.; Lemaître-Guillier, C.; Vallenet, D.; Pestel-Caron, M.; Cozzone, A.J.; Jouenne, T.; Dé, E. (2005) Channel formation by CarO, the carbapenem resistance-associated outer membrane protein of *Acinetobacter baumannii*. *Antimicrob. Agents Chemother.*, 49, 4876-4883.
19. Tognim, M.C.; Gales, A.C.; Penteadó, A.P.; Silbert, S.; Sader, H.S. (2006) Dissemination of IMP-1 metallo-beta-lactamase-producing *Acinetobacter* species in a Brazilian teaching hospital. *Infect Control Hosp. Epidemiol.*, 27, 742-747.
20. Turton, J.F.; Glover, J.; Yarde, S.; Kaufmann, M.E.; Pitt, T.L. (2006). Identification of *Acinetobacter baumannii* by detection of the *bla*<sub>OXA-51</sub>-like carbapenemase gene intrinsic to this species. *J. Clin. Microbiol.*, 44, 2974-2976.
21. Turton, J.F.; Ward, M.G.; Woodford, N.; Kaufmann, M.G.; Pike, R.; Livermore, D.M.; Pitt, T.L. (2006). The role of IsAba1 in expression of OXA carbapenemase genes in *Acinetobacter baumannii*. *FEMS Microbiol. Lett.*, 258, 72-77.
22. Walsh, R.T.; Bolmström, A.; Qwärnström, A.; Gales, A. Evaluation of a new Etest for detecting metallo- $\beta$ -lactamases in routine clinical testing. *J. Clin. Microbiol.*, 40, 2755-2759.
23. Woodford, N.; Ellington, M.J.; Coelho, J.M.; Turton, J.F.; Ward, M.E.; Brown, S.; Amyes, S.G.B.; Livermore, D.M. (2006). Multiplex PCR for genes encoding prevalent OXA carbapenemases in *Acinetobacter* spp. *Int. J. Antimicrob. Agents.*, 27, 351-353.