



A practical synthesis of long-chain iso-fatty acids (iso-C₁₂–C₁₉) and related natural products

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Full Research Paper

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Keywords:
chemoselective reduction; Evans' auxiliary; Grignard addition; homologation; ionic hydrogenation

Beilstein J. Org. Chem. **2013**, *9*, 1807–1812.
doi:10.3762/bjoc.9.210

Received: 11 June 2013
Accepted: 15 August 2013
Published: 04 September 2013

Associate Editor: H. Ritter

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Abstract

A gram-scale synthesis of terminally-branched iso-fatty acids (iso-C₁₂–C₁₉) was developed commencing with methyl undec-10-enoate (methyl undecylenate) (for iso-C₁₂–C₁₄) or the C₁₅ and C₁₆ lactones pentadecanolide (for iso-C₁₅–C₁₇) and hexadecanolide (for iso-C₁₈–C₁₉). Central to the approaches outlined is the two-step construction of the terminal isopropyl group through addition of methylmagnesium bromide to the ester/lactones and selective reduction of the resulting tertiary alcohols. Thus, the C₁₂, C₁₇ and C₁₈ iso-fatty acids were obtained in three steps from commercially-available starting materials, and the remaining C₁₃–C₁₆ and C₁₉ iso-fatty acids were prepared by homologation or recursive dehomologations of these fatty acids or through intercepting appropriate intermediates. Highlighting the synthetic potential of the iso-fatty acids and various intermediates prepared herein, we describe the synthesis of the natural products (*S*)-2,15-dimethylpalmitic acid, (*S*)-2-hydroxy-15-methylpalmitic acid, and 2-oxo-14-methylpentadecane.

Introduction

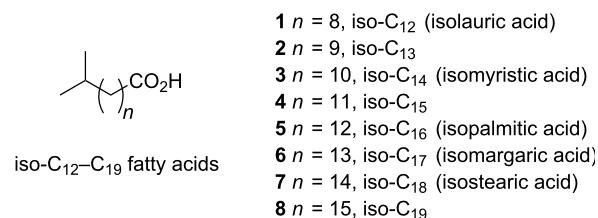
Long-chain iso-fatty acids occur in a broad range of organisms, and are especially abundant in bacteria where, through incorporation into phospholipids, they influence membrane fluidity [1]. Emerging evidence has revealed unexpected roles for certain iso-fatty acids; for example iso-C₁₅ and iso-C₁₇ fatty acids have been shown to be essential in the development of the model eukaryote *Caenorhabditis elegans* [2]. They are present as esters and amides in natural products including

septacidin [3], teicoplanins [4], tunicaminyuracil-based antibiotics [5] (tunicamycins [6], corynetoxins [7], and streptovirudins [8]), the arylomycin glycopeptide antibiotics [9,10], maradolipids [11], plipastatin-type lipopeptides [12], Nod factors [13], glycosylglycerides [14,15], phosphoglycolipids [16], and various sphingolipids [17-19]. The terminal isopropyl group of the iso-fatty acids arises from valine and leucine, which through transamination and decarboxylation reactions

yield isobutyryl-CoA and isovaleryl-CoA [20]. These starter units are elongated by fatty acid synthases to the final iso-fatty acids (even numbered for isobutyryl-CoA; odd-numbered for isovaleryl-CoA) through extension with malonyl-CoA [21,22]. Long-chain iso-fatty acids are important analytical reference compounds owing to the presence of these materials in tobacco [23], wool wax [24], butter fat [25], human sebaceous secretions [26] (adult skin [27], meibum [28], cerumen [29], and newborn vernix caseosa [30,31]), and a wide variety of microbiological samples [1].

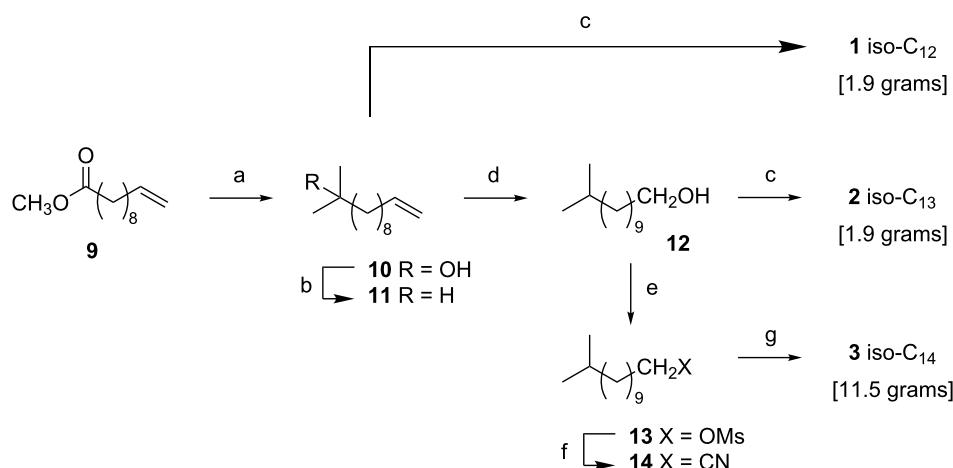
Previous syntheses of iso-fatty acids have typically utilized extended, multi-step sequences. Two main approaches have been used: (1) two-component cross-couplings that include α -ketoester alkylation/decarboxylation [32,33], aldehyde–olefin photoaddition [34], acetylide alkylation (sp^3 – sp) [35,36], Wittig coupling [3,21,37–39], Kolbe electrosynthesis [35,40–42], organocadmium (sp^2 – sp^3) [43–46], organomagnesium (sp^2 – sp^3) [47], or organocupper (sp^3 – sp^3) [48,49] cross-couplings; or (2) bidirectional extension of a central thiophene C₄-fragment [50]. Two fundamentally different approaches worth special mention are the synthesis of the iso-C₁₄ acid **3** by direct hydro-isopropylation of the terminal alkene of methyl undecylenate (available as a pyrolysis product of ricinoleic acid) using isopropyl chloroformate and ethyldichloroaluminium [51], and the synthesis of the iso-C₁₇ acid **6** from methyl ustilate (15,16-dihydroxypalmitate) [52]. Despite the interest in natural products containing iso-fatty acids, these compounds are not readily acquired in multigram quantities due to the complexity of the synthetic routes or limited availability of starting materials. To overcome these problems, we report the scalable, gram-scale syntheses of eight common iso-C₁₂–C₁₉ acids **1**–**8** (Figure 1), from readily available starting materials.

To illustrate the opportunities that our approach provides, we demonstrate the elaboration of the C₁₇-iso-fatty acid **6** and an intermediate, **22**, to several terminal-branched natural products that have not previously been synthesized.

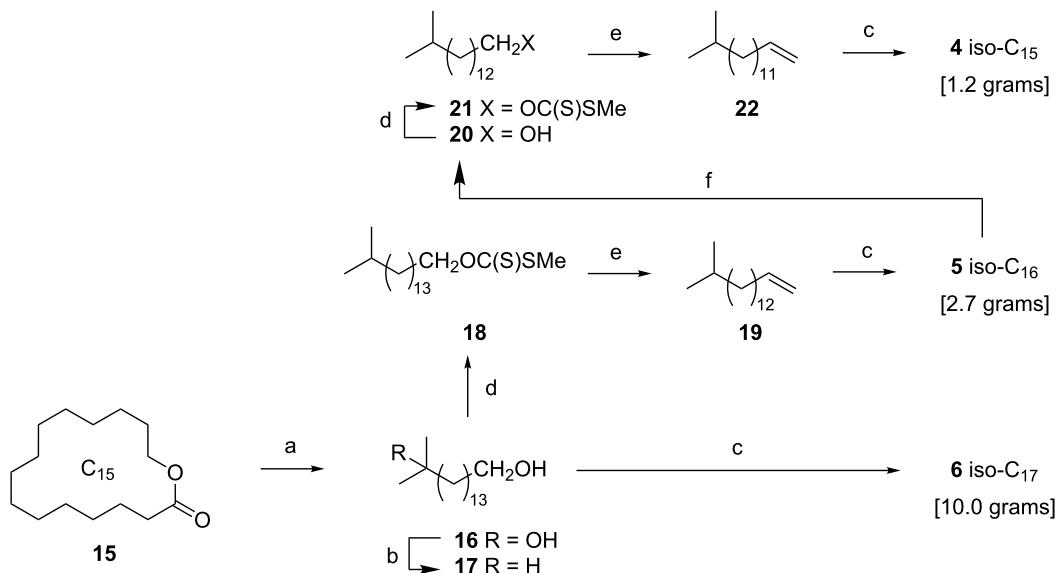


Results and Discussion

Our approach to the iso-C₁₂–C₁₄ fatty acids **1**–**3** commenced from methyl undec-10-enoate (methyl undecylenate) **9**. Reaction of **9** with methylmagnesium bromide afforded the tertiary alcohol **10** in 98% yield (Scheme 1). Selective reduction of the tertiary alcohol of **10** was achieved by ‘ionic hydrogenation’ with triethylsilane and $BF_3\cdot Et_2O$ [53], affording **11**. Oxidative cleavage of **11** with $KMnO_4/Bu_4NBr$ [54] afforded iso-C₁₂ acid **1**. Alternatively, anti-Markovnikov hydration of **11**, using $I_2/NaBH_4$ then hydrogen peroxide [55], afforded the alcohol **12**, and oxidation of **12** with $KMnO_4/Bu_4NBr$ afforded iso-C₁₃ acid **2**. Alternatively, alcohol **12** could be intercepted and converted to the mesylate **13** using $MsCl/Et_3N$ [56] and thence the nitrile **14** (KCN in DMSO/THF). Finally, hydrolysis of the nitrile **14** with $NaOH$ in $H_2O/EtOH$ afforded iso-C₁₄ acid **3**.



Scheme 1: Synthesis of iso-C₁₂ **1**, iso-C₁₃ **2**, and iso-C₁₄ **3** fatty acids from methyl undecylenate (**9**). Reagents and conditions: (a) $MeMgBr$, THF, 98%; (b) $BF_3\cdot Et_2O$, Et_3SiH , CH_2Cl_2 , 99%; (c) $KMnO_4$, Bu_4NBr , $AcOH$, H_2O , 88% for **1**, 96% for **2**; (d) i) I_2 , $NaBH_4$, THF, ii) H_2O_2 , 95%; (e) $MsCl$, Et_3N , CH_2Cl_2 , 98%; (f) KCN , DMSO, THF, 72%; (g) aq $NaOH$, $EtOH$, 96%.

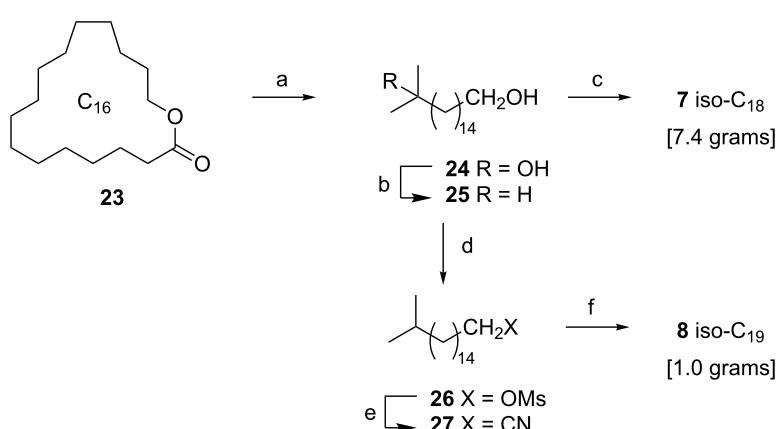


Scheme 2: Synthesis of iso-C₁₅ **4**, iso-C₁₆ **5**, and iso-C₁₇ **6** fatty acids from pentadecanolide (**15**). Reagents and conditions: (a) MeMgBr, THF, 98%; (b) BF₃·Et₂O, Et₃SiH, CH₂Cl₂, 96%; (c) KMnO₄, Bu₄NBr, AcOH, H₂O, 95% for **6**, 93% for **5**; 86% for **4**; (d) NaH, CS₂ then MeI; (e) reflux, 89% for **19** over 2 steps from **17**, 98% for **22** over 2 steps from **20**; (f) BH₃-SMe₂, THF, 78%.

The iso-C₁₅–C₁₇ fatty acids **4**–**6** were prepared from the readily available C₁₅ lactone pentadecanolide (exaltolide, **15**) [57], a natural product that is produced industrially for use as a musk-odored perfumery fixative. Reaction of **15** with methylmagnesium bromide afforded the tertiary alcohol **16** in 98% yield (Scheme 2). Selective reduction of the tertiary alcohol of **16** was achieved using triethylsilane/BF₃·Et₂O [53], yielding **17**. Finally, oxidation of **17** with KMnO₄/Bu₄NBr [54] afforded the iso-C₁₇ acid **6**. The iso-C₁₅ acid **4** and iso-C₁₆ acid **5** were prepared by recursive dehomologation through intercepting the alcohol **17**. Preparation of the xanthate ester **18** (NaH, CS₂, then

MeI) [58] followed by Chugaev elimination afforded the terminal alkene **19**. Oxidative cleavage of **19** using KMnO₄/Bu₄NBr [54] afforded iso-C₁₆ acid **5**. Reduction of **5** (BH₃·Me₂S) [59] afforded the alcohol **20** that when subjected to the same transformations as before, via the xanthate ester **21**, delivered the terminal alkene **22**. Finally, oxidative cleavage (KMnO₄/Bu₄NBr) [54] of **22** afforded iso-C₁₅ acid **4**.

The iso-C₁₈ **7** and iso-C₁₉ **8** fatty acids were synthesized through similar approaches from the related C₁₆ lactone hexadecanolide **23** [60] (Scheme 3). Reaction of **23** with methylmag-



Scheme 3: Synthesis of iso-C₁₈ **7** and iso-C₁₉ **8** fatty acids from hexadecanolide **23**. Reagents and conditions: (a) MeMgBr, THF, 97%; (b) BF₃·Et₂O, Et₃SiH, CH₂Cl₂, 95%; (c) KMnO₄, Bu₄NBr, AcOH, H₂O, 82%; (d) MsCl, Et₃N, 96%; (e) KCN, DMSO, THF, 75%; (f) aq NaOH, EtOH, 86%.

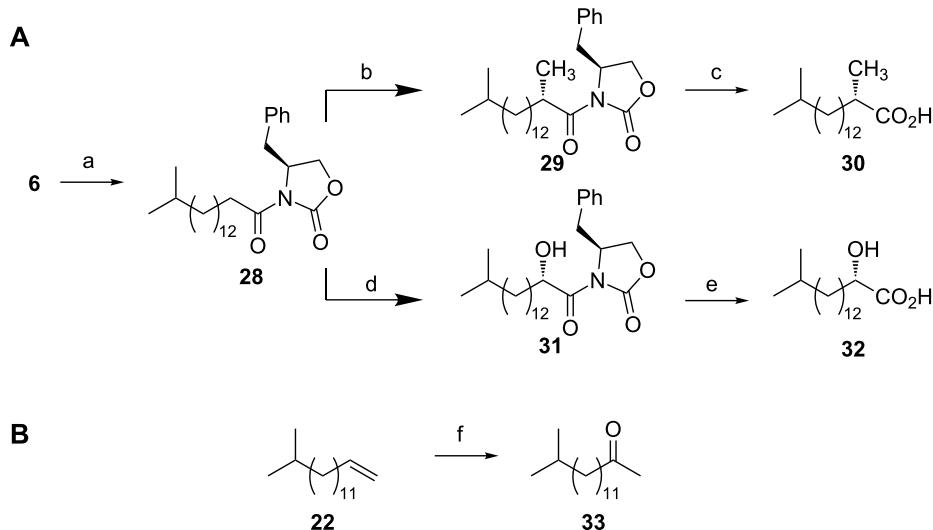
nesium bromide afforded **24**; triethylsilane/BF₃·Et₂O [53] reduction gave **25**; and KMnO₄/Bu₄NBr oxidation afforded iso-C₁₈ acid **7** (Scheme 3). The iso-C₁₉ acid **8** was readily prepared by a three-step homologation through intercepting the alcohol **25**. Thus, mesylation of **25** (MsCl/Et₃N [56]) afforded **26**; substitution (KCN/DMSO) afforded the nitrile **27**; and hydrolysis (NaOH in H₂O/EtOH) of **27** afforded **8**.

The above routes enable the acquisition of (multi)gram quantities of the iso-C_{12–19} acids **1–8**, and provide opportunities for their use as starting materials for the preparation of more complex fatty acids. To illustrate their potential we undertook the synthesis of several representative natural products (Scheme 4). 2,15-Dimethylpalmitic acid has been isolated from a microaerophilic subsurface microbial community [61], and is a component of human newborn vernix caseosa [31], although the absolute configuration of natural samples has not been determined. Conversion of iso-C₁₇ acid **6** to the *N*-acyloxazolidinone **28** was achieved using pivalyl chloride/LiCl [62] and (*S*)-4-benzyloxazolidinone. Diastereoselective methylation [63] of the chelated Z-enolate, derived from deprotonation of **28**, using NaHMDS, followed by addition of iodomethane, yielded **29** as a single diastereoisomer (as determined by ¹H NMR) in 80% yield. Cleavage of the chiral auxiliary using LiOH/H₂O₂ (which occurs without racemization at the α -position) [64] afforded (*S*)-2,15-dimethylpalmitic acid (**30**) in 98% yield. 2-Hydroxy-15-methylpalmitic acid has been identified from a range of sources [1] including the myxobacterium *Stigmatella aurantiaca* [21,65], and the oral bacterium *Veillonella parvula* [66],

although the absolute configuration has not been reported. Diastereoselective hydroxylation [67] of the chelated Z-enolate derived from **28** using the Davis oxaziridine [68] afforded the 2-hydroxy compound **31** as a single diastereoisomer (as determined by ¹H NMR) in 71% yield. Esterification with MeOMgCl [69] (which has been shown not to cause epimerization at the α -position [67]) and saponification [70] afforded (*S*)-2-hydroxy-15-methylpalmitic acid (**32**). The ketone **33** was isolated from *Xanthomonas campestris* pv. *vesicatoria* 85-10 [71]. A direct one step synthesis of **33** was achieved in 51% yield by Wacker oxidation using Pd/O₂ [72] of the alkene **22**, intercepted from the synthesis of the iso-C₁₅ acid **4**.

Conclusion

We have accomplished a highly practical synthesis of the homologous iso-fatty acids **1–8**. The iso-C₁₂ **1**, iso-C₁₇ **6** and iso-C₁₈ **7** acids were prepared from commercially-available starting materials through three-step sequences and produced more than 1 g of each of the three iso-fatty acids in just 2 days each. The remaining five fatty acids were each prepared on >1 g scale by homologation or dehomologation reactions, or through the elaboration of intermediates in the synthesis of **1**. Under-scoring the practicability of this approach, the iso-fatty acids or appropriate intermediates were used for the preparation of three natural products, enantiopure acids **30** and **32**, and the ketone **33**. The simplicity of our approach suggests that it will be of great utility in the preparation of iso-fatty acids for incorporation into more complex molecules.



Scheme 4: Synthesis of (A) 2-methyl- and 2-hydroxy-iso-fatty acids **30** and **32**, and (B) the ketone **33**. Reagents and conditions: (a) Et₃N, PivCl, LiCl, DMAP, (*S*)-4-benzyloxazolidinone, 71%; (b) NaHMDS, Mel, THF, 80%; (c) LiOH, H₂O₂, THF, H₂O, 98%; (d) NaHMDS, Davis oxaziridine, THF, 71%; (e) i) iPrMgCl, MeOH, 76%, ii) NaOH, MeOH, 83%; (f) O₂, PdCl₂, DMA, H₂O, 51%.

Supporting Information

Supporting Information File 1

Experimental part.

[<http://www.beilstein-journals.org/bjoc/content/supplementary/1860-5397-9-210-S1.pdf>]

Supporting Information File 2

NMR spectra.

[<http://www.beilstein-journals.org/bjoc/content/supplementary/1860-5397-9-210-S2.pdf>]

Acknowledgements

The authors thank the Australian Research Council for financial support.

References

1. Kaneda, T. *Microbiol. Rev.* **1991**, *55*, 288–302.
2. Kniazeva, M.; Crawford, Q. T.; Seiber, M.; Wang, C. Y.; Han, M. *PLoS Biol.* **2004**, *2*, E257. doi:10.1371/journal.pbio.0020257
3. Acton, E. M.; Ryan, K. J.; Luetzow, A. E. *J. Med. Chem.* **1977**, *20*, 1362–1371. doi:10.1021/jm00221a002
4. Borghi, A.; Antonini, P.; Zanol, M.; Ferrari, P.; Zerilli, L. F.; Lancini, G. C. *J. Antibiot.* **1989**, *42*, 361–366. doi:10.7164/antibiotics.42.361
5. Cockrum, P. A.; Edgar, J. A. *J. Chromatogr.* **1983**, *268*, 245–254. doi:10.1016/S0021-9673(01)95411-1
6. Ito, T.; Takatsuki, A.; Kawamura, K.; Sato, K.; Tamura, G. *Agric. Biol. Chem.* **1980**, *44*, 695–698. doi:10.1271/bbb1961.44.695
7. Edgar, J. A.; Frahn, J. L.; Cockrum, P. A.; Anderton, N.; Jago, M. V.; Culvenor, C. C. J.; Jones, A. J.; Murray, K.; Shaw, K. J. *J. Chem. Soc., Chem. Commun.* **1982**, 222–224. doi:10.1039/C39820000222
8. Eckardt, K.; Wetzstein, H.; Thrum, H.; Ihn, W. *J. Antibiot.* **1980**, *33*, 908–910. doi:10.7164/antibiotics.33.908
9. Kulanthaivel, P.; Kreuzman, A. J.; Stroge, M. A.; Belvo, M. D.; Smitka, T. A.; Clemens, M.; Swartling, J. R.; Minton, K. L.; Zheng, F.; Angleton, E. L.; Mullen, D.; Jungheim, L. N.; Klimkowski, V. J.; Nicas, T. I.; Thompson, R. C.; Peng, S.-B. *J. Biol. Chem.* **2004**, *279*, 36250–36258. doi:10.1074/jbc.M405884200
10. Liu, J.; Luo, C.; Smith, P. A.; Chin, J. K.; Page, M. G. P.; Paetzel, M.; Romesberg, F. E. *J. Am. Chem. Soc.* **2011**, *133*, 17869–17877. doi:10.1021/ja207318n
11. Penkov, S.; Mende, F.; Zagoriy, V.; Erkut, C.; Martin, R.; Pässler, U.; Schuhmann, K.; Schwudke, D.; Gruner, M.; Mäntler, J.; Reichert-Müller, T.; Shevchenko, A.; Knöller, H.-J.; Kurzchalia, T. V. *Angew. Chem., Int. Ed.* **2010**, *49*, 9430–9435. doi:10.1002/anie.201004466
12. Esumi, Y.; Suzuki, Y.; Itoh, Y.; Chijimatsu, M.; Uramoto, M.; Kimura, K.-i.; Nakayama, S.; Yoshihama, M.; Ichikawa, T.; Haramo, T.; Fujishige, J. *J. Antibiot.* **2003**, *56*, 716–720. doi:10.7164/antibiotics.56.716
13. Poinsot, V.; Bélanger, E.; Laberge, S.; Yang, G.-P.; Antoun, H.; Cloutier, J.; Treilhou, M.; Dénaire, J.; Promé, J.-C.; Debelle, F. *J. Bacteriol.* **2001**, *183*, 3721–3728. doi:10.1128/JB.183.12.3721-3728.2001
14. Hunter, S. W.; McNeil, M. R.; Brennan, P. J. *J. Bacteriol.* **1986**, *168*, 917–922.
15. Orgambide, G. G.; Hollingsworth, R. I.; Dazzo, F. B. *Carbohydr. Res.* **1992**, *233*, 151–159. doi:10.1016/S0008-6215(00)90927-3
16. Fujimoto, Y.; Mitsunobe, K.; Fujiwara, S.; Mori, M.; Hashimoto, M.; Suda, Y.; Kusumoto, S.; Fukase, K. *Org. Biomol. Chem.* **2013**, *11*, 5034–5041. doi:10.1039/c3ob40899j
17. Minamino, M.; Sakaguchi, I.; Naka, T.; Ikeda, N.; Kato, Y.; Tomiyasu, I.; Yano, I.; Kobayashi, K. *Microbiology* **2003**, *149*, 2071–2081. doi:10.1099/mic.0.25922-0
18. Nakayama, M. *Seikatsu Eisei* **1998**, *42*, 135–148.
19. Yano, I.; Tomiyasu, I.; Yabuuchi, E. *FEMS Microbiol. Lett.* **1982**, *15*, 303–307. doi:10.1111/j.1574-6968.1982.tb00239.x
20. Schulz, S.; Dickschat, J. S. *Nat. Prod. Rep.* **2007**, *24*, 814–842. doi:10.1039/b507392h
21. Dickschat, J. S.; Bode, H. B.; Kroppenstedt, R. M.; Müller, R.; Schulz, S. *Org. Biomol. Chem.* **2005**, *3*, 2824–2831. doi:10.1039/b504889c
22. Dickschat, J. S.; Bruns, H.; Riclea, R. *Beilstein J. Org. Chem.* **2011**, *7*, 1697–1712. doi:10.3762/bjoc.7.200
23. Kolattukudy, P. E. *Plant Physiol.* **1968**, *43*, 375–383. doi:10.1104/pp.43.3.375
24. Weitkamp, A. W. *J. Am. Chem. Soc.* **1945**, *67*, 447–454. doi:10.1021/ja01219a027
25. Hansen, R. P.; Shorland, F. B. *Biochem. J.* **1951**, *50*, 207–210.
26. James, A. T.; Wheatley, V. R. *Biochem. J.* **1956**, *63*, 269–273.
27. Nicolaides, N.; Apon, J. M. B. *Biomed. Mass Spectrom.* **1977**, *4*, 337–347. doi:10.1002/bms.1200040604
28. Harvey, D. J.; Tiffany, J. M.; Duerden, J. M.; Pandher, K. S.; Mengher, L. S. J. *Chromatogr.* **1987**, *414*, 253–263. doi:10.1016/0378-4347(87)80051-8
29. Harvey, D. J. *Biomed. Environ. Mass Spectrom.* **1989**, *18*, 719–723. doi:10.1002/bms.1200180912
30. Nicolaides, N. *Lipids* **1971**, *6*, 901–905. doi:10.1007/BF02531172
31. Nicolaides, N.; Apon, J. M. *Lipids* **1976**, *11*, 781–790. doi:10.1007/BF02533404
32. Arosenius, K. E.; Stallberg, G.; Stenhammar, E.; Tagtstrom-Eketorp, B. *Ark. Kemi, Mineral. Geol.* **1948**, *26A*, 20.
33. Weitzel, G.; Wojahn, J. *Hoppe-Seyler's Z. Physiol. Chem.* **1951**, *287*, 65–89. doi:10.1515/bchm2.1951.287.1-6.65
34. Buu-Hoi, N. G. P. *Recl. Trav. Chim. Pays-Bas* **1953**, *72*, 84–87. doi:10.1002/recl.19530720110
35. Hougen, F. W.; Ilse, D.; Sutton, D. A.; de Villiers, J. P. *J. Chem. Soc.* **1953**, 98–102. doi:10.1039/JR9530000098
36. Silvius, J. R.; McElhaney, R. N. *Chem. Phys. Lipids* **1979**, *24*, 287–296. doi:10.1016/0009-3084(79)90034-3
37. Bergelson, L. D.; Vaver, V. A.; Bezzubov, A. A.; Shemyakin, M. M. *Zh. Obshch. Khim.* **1962**, *32*, 1807–1811.
38. Carballera, N.; Thompson, J. E.; Ayanoglu, E.; Djerassi, C. *J. Org. Chem.* **1986**, *51*, 2751–2756. doi:10.1021/jo00364a024
39. Shioiri, T.; Terao, Y.; Irako, N.; Aoyama, T. *Tetrahedron* **1998**, *54*, 15701–15710. doi:10.1016/S0040-4020(98)00984-3
40. Milburn, A. H.; Truter, E. V. *J. Chem. Soc.* **1954**, 3344–3351. doi:10.1039/jr9540003344
41. Norén, B.; Odham, G. *Lipids* **1973**, *8*, 573–583. doi:10.1007/BF02532714

42. Streibl, M.; Jarolimek, P.; Wollrab, V. *Collect. Czech. Chem. Commun.* **1964**, *29*, 2522–2527. doi:10.1135/cccc19642522
43. Akiya, S.; Nakazawa, Y. *Yakugaku Zasshi* **1956**, *76*, 1403–1405.
44. Balzer, T.; Budzikiewicz, H. *Z. Naturforsch., B: Chem. Sci.* **1987**, *42*, 1367–1368.
45. Cason, J. *J. Am. Chem. Soc.* **1942**, *64*, 1106–1110. doi:10.1021/ja01257a029
46. Stein, J.; Budzikiewicz, H. *Z. Naturforsch., B: Chem. Sci.* **1987**, *42*, 1017–1020.
47. Fordyce, C. R.; Johnson, J. R. *J. Am. Chem. Soc.* **1933**, *55*, 3368–3372. doi:10.1021/ja01335a054
48. Edmunds, A. J. F.; Aluja, M.; Diaz-Fleischer, F.; Patrian, B.; Hagmann, L. *Chimia* **2010**, *64*, 37–42. doi:10.2533/chimia.2010.37
49. Takikawa, H.; Nozawa, D.; Kayo, A.; Muto, S.-e.; Mori, K. *J. Chem. Soc., Perkin Trans. 1* **1999**, 2467–2477. doi:10.1039/A904258J
50. McGhie, J. F.; Ross, W. A.; Evans, D.; Tomlin, J. E. *J. Chem. Soc.* **1962**, 350–355. doi:10.1039/JR9620000350
51. Biermann, U.; Metzger, J. O. *J. Am. Chem. Soc.* **2004**, *126*, 10319–10330. doi:10.1021/ja048904y
52. Crossley, A. T.; Craig, B. M. *Can. J. Chem.* **1955**, *33*, 1426–1432. doi:10.1139/v55-171
53. Orfanopoulos, M.; Smonou, I. *Synth. Commun.* **1988**, *18*, 833–839. doi:10.1080/00397919808057852
54. Herriott, A. W.; Picker, D. *Tetrahedron Lett.* **1974**, *15*, 1511–1514. doi:10.1016/S0040-4039(01)93123-5
55. Prasad, A. S. B.; Kanth, J. V. B.; Periasamy, M. *Tetrahedron* **1992**, *48*, 4623–4628. doi:10.1016/S0040-4020(01)81236-9
56. Crossland, R. K.; Servis, K. L. *J. Org. Chem.* **1970**, *35*, 3195–3196. doi:10.1021/jo00834a087
57. Kerschbaum, M. *Ber. Dtsch. Chem. Ges.* **1927**, *60*, 902–909. doi:10.1002/cber.19270600411
58. Barton, D. H. R.; McCombie, S. W. *J. Chem. Soc., Perkin Trans. 1* **1975**, 1574–1585. doi:10.1039/P19750001574
59. Adams, R. M.; Braun, L. M.; Braun, R. A.; Crissman, H. R.; Opperman, M. *J. Org. Chem.* **1971**, *36*, 2388–2389. doi:10.1021/jo00815a047
60. Bougault, J. C. R. *Acad. Sci.* **1910**, *150*, 874–876.
61. Hedrick, D.; Peacock, A.; Long, P.; White, D. *Lipids* **2008**, *43*, 843–851. doi:10.1007/s11745-008-3206-1
62. Ho, G.-J.; Mathre, D. J. *J. Org. Chem.* **1995**, *60*, 2271–2273. doi:10.1021/jo00112a060
63. Evans, D. A.; Ennis, M. D.; Mathre, D. J. *J. Am. Chem. Soc.* **1982**, *104*, 1737–1739. doi:10.1021/ja00370a050
64. Evans, D. A.; Britton, T. C.; Ellman, J. A. *Tetrahedron Lett.* **1987**, *28*, 6141–6144. doi:10.1016/S0040-4039(00)61830-0
65. Fautz, E.; Rosenfelder, G.; Grotjahn, L. *J. Bacteriol.* **1979**, *140*, 852–858.
66. Leuckfeld, I.; Paster, B. J.; Kristoffersen, A. K.; Olsen, I. *APMIS* **2010**, *118*, 230–242. doi:10.1111/j.1600-0463.2009.02584.x
67. Evans, D. A.; Morrissey, M. M.; Dorow, R. L. *J. Am. Chem. Soc.* **1985**, *107*, 4346–4348. doi:10.1021/ja00300a054
68. Davis, F. A.; Chattopadhyay, S.; Towson, J. C.; Lal, S.; Reddy, T. *J. Org. Chem.* **1988**, *53*, 2087–2089. doi:10.1021/jo00244a043
69. Verma, R.; Ghosh, S. *J. Chem. Soc., Perkin Trans. 1* **1999**, 265–270. doi:10.1039/A808840C
70. Theodorou, V.; Skobridis, K.; Tzakos, A. G.; Ragoussis, V. *Tetrahedron Lett.* **2007**, *48*, 8230–8233. doi:10.1016/j.tetlet.2007.09.074

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doi:10.3762/bjoc.9.210