

The Concept of Allosteric Interaction and Its Consequences for the Chemistry of the Brain

Published, JBC Papers in Press, July 22, 2013, DOI 10.1074/jbc.X113.503375

Jean-Pierre Changeux

From the Collège de France, 75005 Paris and the Institut Pasteur, 75724 Paris Cedex 15, France

Throughout this Reflections article, I have tried to follow up on the genesis in the 1960s and subsequent evolution of the concept of allosteric interaction and to examine its consequences within the past decades, essentially in the field of the neuroscience. The main conclusion is that allosteric mechanisms built on similar structural principles operate in bacterial regulatory enzymes, gene repressors (and the related nuclear receptors), rhodopsin, G-protein-coupled receptors, neurotransmitter receptors, ion channels, and so on from prokaryotes up to the human brain yet with important features of their own. Thus, future research on these basic cybernetic sensors is expected to develop in two major directions: at the elementary level, toward the atomic structure and molecular dynamics of the conformational changes involved in signal recognition and transduction, but also at a higher level of organization, the contribution of allosteric mechanisms to the modulation of brain functions.

Theory in Biology

In the physical sciences, there is a long tradition of theory and model building; the advancement of the discipline has relied and still relies on an active interplay between theory and experimental studies and between the development of novel technologies and conceptual invention. In the biological sciences, a similar tradition exists, but it is not as systematic. The frequent enthusiasm for new techniques without explicit theoretical perspectives often hides an impoverished theoretical landscape where original hypotheses or ideas are rare. In my view, the theoretical objectives should precede, or be concomitant with, the technological and experimental means. According to Claude Bernard, “the experimenter who does not know what he is looking for will not understand what he finds” (1).¹ The reasons for the difficulties encountered with the biological sciences possibly lie in the fact that the objects of biology are highly differentiated with multiple, intricate levels of organization. The identification of common conceptual rules further hinges on the considerable structural and functional diversity of the biological objects. Thus, there is the constant dilemma, often split between scientists, about the universality or the singularity of the objects they investigate. In this context, I have selected as a tentative common rule the concept of allosteric interaction and its consequences on the molecular chemistry of proteins, going from bacterial regulatory enzymes up to the nervous system and, tentatively, the higher functions of the brain.

The circumstances of my personal life have been such that as an adolescent, I was already exposed to theory through the teaching of an exceptional professor of natural sciences, Jean Bathellier, who introduced me to the idea of biological evolution. This experience deeply influ-

¹ The full quote (see Ref. 255) is, “Empiricism may serve to accumulate facts, but it will never build science. The experimenter who does not know what he is looking for will not understand what he finds.”

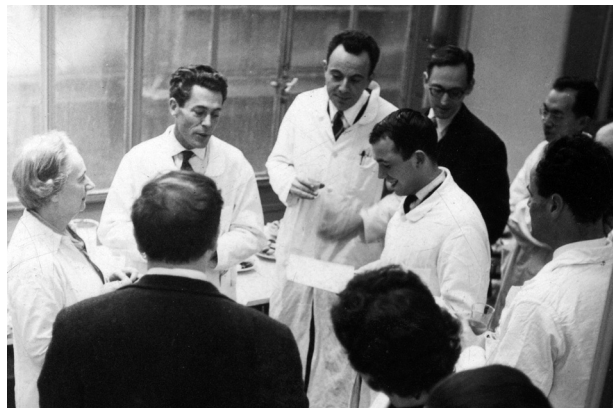


FIGURE 1. The author reading a poem in 1960 at Jacques Monod's 50th birthday. From left to right: Sarah Rapkine, Jacques Monod, François Jacob, the author, and François Gros. The photo was provided by the Fonds d'Archives Madeleine Brunerie at the Institut Pasteur.

enced my scientific career to the extent that until now, as we shall see, my thinking has been framed within the Darwinian evolutionary paradigm. My first laboratory work was in marine biology (2), but I felt rapidly unsatisfied by perspectives essentially limited to descriptive morphology. In the fall of 1958, I visited Jean Brachet's laboratory in Brussels, where I also attended Christian de Duve's lectures and became fascinated by their theoretical perspective that the solution of the great problems of biology had ultimately to be found in elementary biochemical mechanisms. My interest shifted to the more explanatory chemistry of embryonic development. In this context, I imagined a theory in which the enzyme activations that follow the entry of the spermatozoon in the egg were due to a release of enzymes hidden in subcellular particles referred to as lysosomes by Christian de Duve. Back in Paris, I tried to test this hypothesis. Confronted by concrete technical difficulties, I met, by chance, Jacques Monod, who offered to let me enter his laboratory at the Institut Pasteur on the strict condition I abandon my ideas on embryonic development and work with *Escherichia coli*. I drudgingly accepted his demand (3), keeping my theoretical interest in animal biology for the future!

My fascination for theory once again was stimulated in the spring of 1959, when the moment came for the selection of the topic for my Ph.D. thesis. I met Jacques Monod and François Jacob (Fig. 1), who suggested several themes for my research, most of them dealing with their ongoing work on the regulation of gene expression and the operon model. They did not suit me since the theoretical insights and outcomes were already in their hands. An alternative, original topic, mentioned by François Jacob, held my attention. He had heard a talk by Edwin Umberger, who had shown that in bacterial biosynthetic pathways, such as those for the biosynthesis of L-isoleucine or L-valine, the

first enzyme is feedback-inhibited by the end product of the pathway (4). The issue was to understand the molecular mechanism of this "apparently competitive" regulatory interaction, a critical step in the cybernetics of the cell. This topic fitted with my first interest about enzyme activations during fertilization but with a much broader biological perspective. I adopted the theme for my thesis work.

The Concept of Allosteric Interaction (1961–1963)

My first naive idea was that if a special molecular device mediated the feedback inhibition by the enzyme, one should be able to find a way to identify its molecular constituents, for instance, by dissociating the regulatory interaction from the catalytic activity *in vitro*. I first confirmed Umberger's *in vitro* observations of the apparent competitive inhibition of L-threonine deaminase by L-isoleucine and its "bimolecular" cooperative kinetics toward both the substrate and the feedback inhibitor. I noticed that the sensitivity of enzyme preparations to L-isoleucine changed with the time of storage, purification, heating, and exposure to reagents for –SH groups, resulting in a loss of response to L-isoleucine without a significant decline in enzyme activity. Interestingly, the loss of L-isoleucine feedback inhibition was also accompanied by the abolition of the bimolecular kinetics of the enzyme toward its substrate. The paper I presented at the 26th Cold Spring Harbor Symposium on Quantitative Biology entitled "Cellular Regulatory Mechanisms" (5), at the initiative of Jacques Monod, gave me the stimulating opportunity to theorize. I briefly discussed the two plausible models that might account for the apparently competitive antagonism between the feedback inhibitor L-isoleucine and the substrate L-threonine. According to the first model, the binding sites for the substrate and regulatory inhibitor are partially overlapping, so the interaction is a classical competition by steric hindrance. In the second "new" model, referred to as "no-overlapping," the two sites are separated from each other, and the interaction between ligands takes place between topographically distinct sites. I favored the second model, in which the substrate and regulatory effector were to bind topographically distinct sites, specifically on the basis of the argument that loss of feedback inhibition was accompanied by a normalization of the kinetics (5). Following my presentation, the distinguished bacteriologist Bernard Davis mentioned the possible analogy between the properties of hemoglobin and those of threonine deaminase. As we shall see, it was a highly relevant and inspiring comment (6).

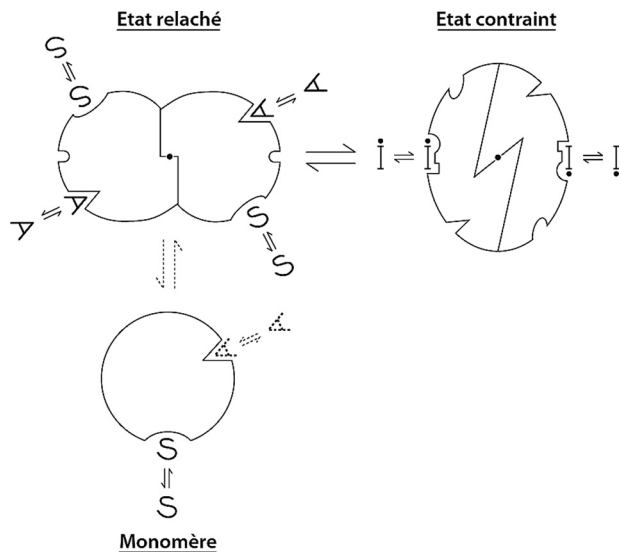


FIGURE 2. The two-state concerted Monod-Wyman-Changeux model (1965) as originally drawn by the author (Ph.D. thesis, 1964) (73). *Etat relâché*, relaxed state; *Etat contraint*, constrained state; *Monomère*, monomer.

In the oral presentation of the “General Conclusions” of the symposium, Jacques Monod reported my results and interpretation in the section dealing with the regulation of enzyme activity. He also wrote, “Closely similar observations have been made independently and simultaneously by Pardee (private communication) on another enzyme sensitive to end-product (aspartate-carbamyl-transferase)” (7). In Monod and Jacob’s subsequently written “General Conclusions,” the word “allosteric” appears for the first time. It is composed of two Greek roots expressing the difference (allo-) in (stereo-) specificity of the two binding sites to qualify and generalize the no-overlapping sites mechanism of indirect interaction between stereospecifically distinct sites mediated by a conformational change of the protein (8).

This was the birth of the word allosteric and of its general definition, which is nowadays widely accepted (8). The introduction of the concept created a major landmark in classical enzymology and the ill-defined notions of “un-competitive” or “non-competitive” inhibition (9) most often (but not always) assumed to take place in the neighborhood (or even at the level) of the active site.

At this stage, the conformational change linking the topographically distinct sites was interpreted by us in terms of the “induced-fit” theory of Daniel Koshland (10, 11) in the sense that the ligand “instructs” rather than “selects” the structural change (12). At the time, Koshland’s concern was not the regulation of enzyme activity by a metabolic signal but the specificity of enzyme action. His theory (11, 13) was that a local steric fit seemed essential for the reaction to occur “only after a change in shape

of the enzyme molecule had been induced by the substrate” (13). We suggested the extension of the idea to a higher level long-range and distant allosteric interaction between active and regulatory sites (12). Without being aware of it, we were following the widely accepted tradition of Karl Landsteiner and Linus Pauling’s empiricist ideology, viewing the local environment as directly instructing structural changes within biological organisms.

The Monod-Wyman-Changeux Model (1965)

A paradigmatic change from instruction to selection occurred with the Monod-Wyman-Changeux (MWC) model (14). Quite surprisingly, in my opinion, it did not emerge from a deliberate shift of theoretical position by any one of us, but from experimental observations. At the end of 1963, I handed Jacques Monod the first typed version of my thesis work (15, 73, 251–254). Of the many observations I had made, he became especially interested by the experiments I did in 1962 on the effects of urea (16). At an adequate concentration, urea reversibly inactivates L-threonine deaminase, an inactivation interpreted as a split of the enzyme into subunits. Interestingly, in this system, allosteric activators, such as L-norleucine, L-valine, and L-*allo*-threonine, facilitated inactivation and thus subunit dissociation; feedback inhibitors, such as isoleucine, protected against inactivation and thus strengthened the assembly of the subunits in the protein. To formally account for the observed effects, I mentioned the possibility of three possible conformational states, one for each experimental condition (presence of activator, presence of inhibitor, and no effector). Jacques Monod pressed me to systematically adopt in my theoretical reasoning the rule to reduce the number of hypotheses to the strict minimum. Two states should suffice! I immediately agreed and further documented the experimental consequences of the two states with a tight (T) or relaxed (R) mode of packing of the subunits, each state pre-existing the binding of ligand and exhibiting different intrinsic affinities for both substrate and allosteric effectors. In my opinion, this was the birth of the *two-state mechanism* of pre-existing conformational states ($R \leftrightarrow T$), a consequence of a selective, rather than an instructive, effect of the ligands. These views were the start of many debates and theoretical developments that, after many successive writings of the paper by Jacques Monod (17, 18), led to the final version of the MWC model (14).

The model was originally planned to conclude my thesis work with the aim to formally establish a causal link between the structural organization of the known regulatory proteins and their signal transduction properties (Fig.

2). The problem was risky because at that time, the only known three-dimensional structures of proteins were those of hemoglobin (19) and myoglobin (20). Emphasis was placed on the cooperative binding of ligands, a property found frequently associated with the ability to transduce regulatory signals. An essential postulate for us was that cooperativity between ligand-binding sites relies upon the structural cooperativity existing between several subunits in a regulatory protein. In other words, the quaternary assembly of the subunits into a cooperative symmetrical oligomers is critical (although exceptions exist where single folded polypeptides that form several binding sites for different ligands may display allosteric interactions) (21). Max Perutz's structural model of hemoglobin (22, 23) revealed the symmetries of the $\alpha_2\beta_2$ -tetramer and the 25–36-Å distance between hemes, together with the evidence that the conformational change that takes place upon O₂ binding primarily concerns the quaternary organization of the molecule. Through several visits, Jeffries Wyman made us aware about his views that the symmetry properties of the O₂ saturation function for hemoglobin reveal the existence of elements of structural symmetry (24). Additional analysis of the assembly of protein oligomers and their symmetry properties was carried out in parallel by Jacques Monod himself and was included in the text.

Along these lines, the MWC model introduces the important notion that signal transduction requires a particular flexibility of protein structure associated with a quaternary constraint established between the subunits within the oligomer, which affects their tertiary organization (and vice versa) (14, 21, 25). Its principal consequence is the all-or-none character of the molecular switch between discrete conformational states, without intermediate hybrid states, associated with cooperative ligand binding. The statement then enunciated put forward that regulatory proteins spontaneously form closed and symmetrical oligomers (or microcrystals) that exist in a few discrete (all-or-none) and symmetrical (R, T...) conformations in thermal equilibrium in the absence of a regulatory signal. The regulatory ligands do not instruct any new adaptative conformations, but merely shift the spontaneous equilibrium between the conformations, selectively stabilizing the one that displays the biological activity and the highest affinity. The model fundamentally distinguishes a function of state "R," which describes the spontaneous conformational equilibrium (defined by its intrinsic equilibrium constant L_0), and a binding function, "Y," which describes the occupation of the binding sites and distinctly evolves as a function of ligand concentration.

Several of these statements have since then been reformulated and discussed by several groups in terms of "conformational shift" or "shape shifting" (21, 26–29). They created a theoretical landmark in biochemical thinking by opposing a selectionist model to the traditional induced-fit scheme.

A year following the MWC publication, Koshland, Némethy, and Filmer attempted to rehabilitate the Pauling scheme for O₂ cooperativity in hemoglobin (30) by proposing an instructive model (KNF) (31) that implied a gradual induced change of biophysical parameters with abundant intermediate hybrid states accompanied by the superimposition of the state and binding functions. I took the opportunity to test this prediction during a postdoctoral position with John Gerhart and Howard Schachman. First, I extended the formulation of the MWC model to account for nonexclusive binding of regulatory ligands to both R and T states with the help of Merry Rubin, a computational scientist from the laboratory (32). Then making use of the high amounts of purified aspartate transcarbamylase (7, 33) available for binding experiments (34), together with simultaneously collected conformational data (35), we were able to demonstrate unambiguous differences between the state and binding functions. Moreover, the aspartate transcarbamylase data could be fitted with the general equation of the MWC model (Ref. 36; also see Ref. 37 with phosphofruktokinase).

Since the 1960s, abundant structural studies have confirmed the MWC statement that many regulatory devices possess a symmetrical oligomeric structure and undergo the predicted conformational changes (21). The MWC model has also elicited new structural and molecular dynamics investigations in hemoglobin (38–43) and was extended to gene repressors (44, 45), together with the superfamily of nuclear receptors (46–48). We shall see that it was also extended to ion channel-linked (49) and G-protein-linked (50) membrane receptors.

As any scientific theory, the MWC model did not aim to give an exhaustive description of physical reality. It has intrinsic limitations introduced, in particular, by its founding postulates. The formal model was limited to a minimum of two main conformational states, which, as we shall see, are often more numerous (51, 52) and did not include either kinetics and/or molecular dynamics, which was invented after that (29). Also, it did not mention the possible occurrence of regulatory proteins undergoing allosteric transitions as single monomers (21, 29, 52) and the case of some G-protein-coupled receptors (GPCRs) (see "The Nicotinic Receptor: An Authentic Allosteric Membrane Protein"); yet, in my opinion, the major out-

comes of the MWC theory are 2-fold. First, it implements a paradigmatic shift from the cybernetics of biological systems (53, 54) to the molecular mechanism of signal transduction. Second, it has been and still is a strong incentive for empirical research on regulatory proteins using the broad diversity of biophysical methods available.

Cooperativity of Membrane Receptors and the MWC Model

In the conclusion of my Ph.D. thesis in 1964 (15, 73, 251–254), I considered the possibility of extending the MWC model to the “membrane phenomena involved in the recognition of communication signals and their transmission (for example, synaptic transmission)” (73). A few years later, I met Max Delbrück at the California Institute of Technology (Caltech), and he gave me the opportunity to move a step forward in my theoretical reflections and to jump to the level of biological membranes. In September 1966, Max invited me to give a series of three lectures in October at Caltech. Commenting on my last lecture, he was struck by a slide on which I had shown where nearest-neighbor cooperative interactions between allosteric proteins were suggested to take place in a biological membrane. I said that I wanted to develop a mathematical model of such an interaction, and Max suggested that I meet the solid-state physicist Charles Kittel when back in Berkeley. Together with him and a French colleague, Jean Thiéry, the MWC model was re-examined and reformulated. In addition to the classical oligomeric case of membrane receptors, the model was applied to larger, unlimited cooperative assemblies of membrane proteins in a two-dimensional lattice (57). At variance with the MWC model, the thermodynamic formulation was based on the conformational transition of single units (or protomers) between a minimum of two states modulated (or not) by the interaction with other protomers. Depending on the value of the free energy of the interaction between protomers, the model predicts the existence of various classes of responses exhibited by biological systems: from a graded response of a single-receptor protomer or of an oligomeric receptor (MWC model) to an all-or-none phase transition response in large and periodic protein assemblies (56, 57).

The lattice model was rediscovered and successfully developed thirty years later by Dennis Bray and colleagues with the experimental system of bacterial chemoreceptors (58, 59); yet, to date, such cooperative interactions have not been identified in synaptic membranes, only in reconstituted artificial membranes. In contrast, with GPCRs (60–62), the assemblies of homo- and hetero-oligomers

of variable sizes as well as between GPCRs and ion channels or ligand-gated ion channels (also referred to as receptor-receptor interactions) have been abundantly documented (63–66).

Other kinds of supramolecular assemblies might possibly be interpreted in terms of adapted versions of the MWC (14) and Changeux *et al.* (57) models such as the chaperonins (67, 68) and possibly the gene transcription complex (69). These models of allosteric interactions have still largely unexplored consequences of the structural and functional organization of supramolecular structures, where the interactions not only take place between ligand-binding sites but also mobilize multiple protein interfaces. In any case, it was an opportunity to reframe the original model in a new biological context.

The Identification of the Nicotinic Acetylcholine Receptor

The MWC model was a fertile inspiration for deeper research into the structural chemistry of proteins, protein design of new functions, and new modulators (21), but I preferred to extend my reflections about membrane cooperativity to carrying the concept of allostery into neuroscience and, to begin with, test the available models on a neural receptor. The John Newport Langley (70) concept of “receptive substance,” or receptor theory, was a major landmark in the history of biochemical pharmacology in 1905, yet it was soon criticized and even found “unnecessary” (71) by many experimentalists. Also, for decades, it was believed that neuronal communications in the brain were electrical rather than chemical! No neurotransmitter receptor was biochemically identified until nearly 60 years later (72).

Before leaving the Institut Pasteur in mid-1965, I decided to experimentally challenge the plausible extension of the MWC model to a synaptic protein, the enzyme that degrades the neurotransmitter acetylcholine (ACh), acetylcholinesterase (AChE). I discovered that some of the compounds designed by Daniel Bovet when he was working at the Institut Pasteur (in particular, derivatives of Flaxedil) did not behave as steric competitors but as allosteric modulators of AChE activity (55, 74–76), suggesting a possible but superficial analogy with the authentic receptor.

The time had come to test the MWC model with the true physiological receptor for ACh. A prerequisite was to identify it as a protein! It was achieved by starting from the *Electrophorus electricus* electric organ (72, 77) to use the deliberate strategy to follow the *in vivo* physiological and pharmacological response of a single electroplaque to nic-

otinic agents (77) and to keep these features unmodified *in vitro* at all stages of receptor isolation and chemical identification. I learned to dissect and record from the single electroplaque during a visit of a few months to David Nachmansohn's laboratory at Columbia University in 1967. Back at the Institut Pasteur, I developed an important intermediate assay from purified membrane fragments that make closed microsacs from which radioactive Na^+ (or K^+) ion flux responses to nicotine agonists can be measured. The *in vitro* data displayed ligand specificities that closely resembled those recorded with single electroplaques (78, 79). I used the detergent deoxycholate to gently extract, without denaturation, a protein that reversibly bound the radiolabeled nicotinic agonist decamethonium, using the method of equilibrium dialysis formerly used by Walter Gilbert and Benno Müller-Hill (80) to identify the *lac* repressor. Last, the snake venom toxin α -bungarotoxin, discovered by the Taiwanese pharmacologist Chen-Yuan Lee, was discovered to specifically block the *in vivo* neuromuscular transmission in high vertebrates at the postsynaptic level without interacting with AChE (81). α -Bungarotoxin was found to block altogether the electroplaque's *in vivo* electrical response to nicotinic agonists, the microsac's ion flux response *in vitro*, and the binding of radioactive decamethonium to the detergent extract (82). The extract contained a high molecular weight hydrophobic protein that could then be physically separated from AChE (82) and purified to homogeneity (72, 83).

The Nicotinic Receptor: An Authentic Allosteric Membrane Protein

The secure identification of the nicotinic ACh receptor (nAChR) protein opened a new field of empirical biochemical research (72) motivated by the following question: is the newly identified receptor a *bona fide* allosteric protein? Examination by electron microscopy of the nAChR protein purified from *E. electricus* and from purified nAChR-rich membranes from *Torpedo marmorata* revealed ring-like particles (8–9 nm in diameter) with a hydrophilic core made up of several (five to six) subunits associated in a compact bundle (84–94). Cross-linking of *E. electricus* nAChR further suggested a pentameric organization (95) composed of four types of homologous subunits, $2\alpha 1$, $\beta 1$, $\gamma 1$, and $\delta 1$ (96–100), all of which with important sequence identities (99, 101, 102). This finding suggested the possibility, consistent with the MWC model, of a pseudo-symmetrical organization of the nAChR oligomer. The complete sequence of the cDNAs from the several homologous nAChR subunits from the



FIGURE 3. The author working in his laboratory at the Institut Pasteur in 1987. This photo was provided by Martine Franck of Magnum Photos.

electric organs of *Torpedo californica* (103–106) and *T. marmorata* (107, 108), muscle (109), and brain (110–113) revealed a common organization of all subunits (104, 106, 108) that consisted of a large N-terminal hydrophilic domain, four hydrophobic segments (TM1–TM4), and a small hydrophilic domain. Affinity labeling and site-directed mutagenesis demonstrated that the large hydrophilic domain is extracellular and carries the ACh-binding site (114–118) and that the four hydrophobic segments traverse the transmembrane, with the TM2 segment lining the ion channel (Figs. 3 and 4) (119–122).

The nAChR appeared to behave as an authentic allosteric protein but with particularities of its own (49, 123, 124). 1) It possesses a rather odd oligomeric structure of an homo- or heteropentameric organization, with a C5 axis of rotational symmetry (or quasi-symmetry) perpendicular to the plane of the membrane. 2) The topological distinction between the neurotransmitter (ACh) sites (separated by 40 Å between ACh sites) in the extracellular domain and the ion channel (separated by 60 Å from the ACh sites) in the transmembrane domain unambiguously

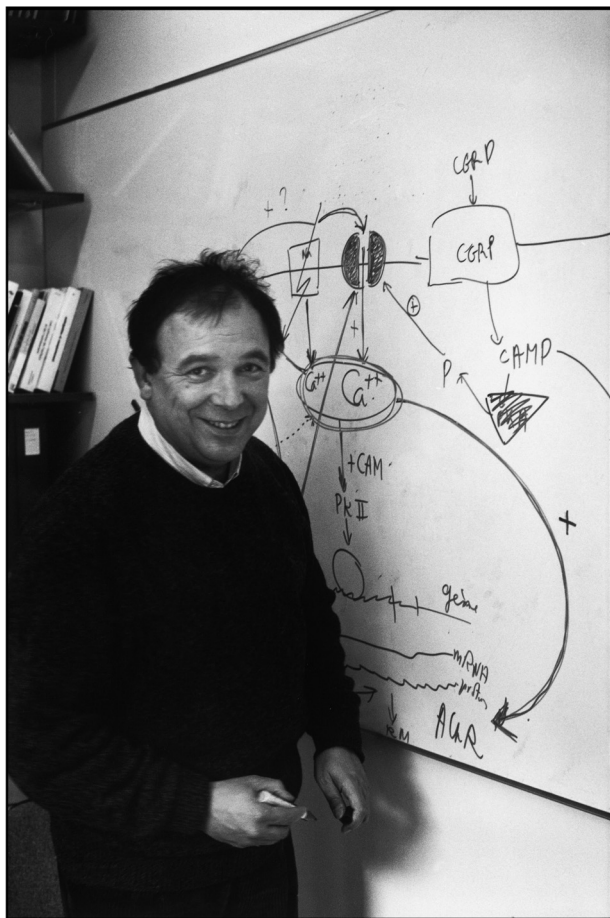


FIGURE 4. The author (1987) discussing models on the themes of his teaching at the Collège de France (Chair of Cellular Communications). This photo was provided by Martine Franck of Magnum Photos.

demonstrates that the interaction between the neurotransmitter (ACh) and the channel is an indirect allosteric interaction. 3) Consistent with the MWC model, several discrete conformational changes spontaneously occur in the absence of ligand, but more than two mediate activation (channel opening). The fast and slow inactivations toward higher affinity closed state(s) are referred to as desensitized (49, 125–131). 4) Multiple allosteric “modulatory sites” are present in the synaptic (e.g. Ca^{2+}) and transmembrane domain. The anthelmintic ivermectin, general anesthetics, and other molecules regulate these transitions (see below). 5) The conformational equilibrium is shifted by orthosteric and allosteric ligands in favor of the state(s) for which they exhibit a preferential affinity (124–127). Last, constitutive mutations stabilize any one of the accessible conformations (active *versus* inactive) of the nAChR (129, 132–136). Such mutations confer predispositions to human diseases such as congenital myasthenia (129, 133) and autosomal dominant frontal lobe epilepsy (134, 137) (Fig. 5).

The recent discovery that orthologs of nAChRs are present in prokaryotes (138) and behave as ligand-gated ion channels (139) has led to high resolution crystal structures of the receptors from two bacterial species, *Gloeobacter violaceus* (140) and *Erwinia chrysanthemi* (141, 142). They confirm a pentameric symmetrical structure that is strikingly conserved from prokaryotes to eukaryotes (49, 143), thereby opening the exploration of the allosteric transition at the atomic level. Comparison of the x-ray core structures of *Erwinia* and *Gloeobacter* receptors, which are present in closed *versus* open conformations (140–142), revealed a conformational change associated with channel gating manifested by a global quaternary twist around the 5-fold axis of rotational symmetry yet with additional tertiary deformations. An *in silico* normal mode analysis, initially applied to a model of $\alpha 7$ -nAChR (144–146), accounts for at least 29% of the allosteric transition deduced from *Erwinia* *versus* *Gloeobacter* receptor structures (140–142). The latest ongoing structural developments disclose a novel locally closed structure of the *Gloeobacter* receptor (51), pointing to additional intermediate conformations possibly relevant to the multistep process of desensitization (147).

The isolation of the nicotinic receptor and the demonstration of its allosteric properties were followed by the identification of many members of the pentameric receptor family in the brain (including nAChRs; 5-hydroxytryptamine type 3, GABA_A , GABA_C , and glycine receptors; and some glutamate, histamine, and 5-hydroxytryptamine-activated anionic receptors) (49, 148–150), as well as other classes of receptors, including the GPCRs (151–153) and channelrhodopsin light-gated cation channels (154).

These studies also pioneered the elucidation at the atomic level of the signal transduction mechanism these receptors mediate and their molecular dynamics (155). In a general manner, the available experimental data favor, but with possible important exceptions, the MWC scheme of pre-existing conformational equilibria (21, 156, 157). However, in several instances such as bias agonism in GPCRs (158), equilibria between multiple conformations (rather than two) have to be taken into consideration (52).

Allosteric Modulation: A New Receptor Pharmacology

Another important consequence of the idea of allostery was the recent re-evaluation of the ancient concept of allosteric modulation (21, 123). It opened new avenues in the pharmacology of the brain and resulted in important applications to drug discovery (158, 159). Early studies



FIGURE 5. The author in his laboratory at the Institut Pasteur in 1995 with (from left to right) Alain Bessis, Anne Devillers-Thiéry, Clément Léna, Jean-Luc Eiselé, and Jean-Luc Galzi. This photo was provided by the Institut Pasteur.

carried out in the 1970s in my laboratory with the aim to identify the binding specificity of various ligands for the nAChR already revealed that channel blockers, like local anesthetics, do not directly displace nicotinic ligands from the ACh-binding site but reversibly bind to a different allosteric site (160–162). The interaction between ACh and local anesthetic sites examined by fluorescence recording was shown to be positive and reciprocal, thus introducing the concept of an allosteric modulatory ligand. Since then, in addition to the main allosteric interaction between neurotransmitter (orthosteric) sites and the ion channel (which binds local anesthetics), multiple allosteric sites to which ligands bind and thus regulate signal transduction have been identified (Fig. 6). One of them was for external Ca^{2+} ions (163, 164), which bind to the extracellular domain of the $\alpha 7$ -receptor (165).

Another important type of site for allosteric modulation was discovered in a collaborative work carried out with Daniel Bertrand from the University Medical Center in Geneva (166) using ivermectin. In the micromolar range, this compound strongly enhances the ACh-evoked response of $\alpha 7$ -receptors with increased apparent affinity, cooperativity, and maximal response. Based on the altera-

tion of the ivermectin effect caused by mutations in the transmembrane domain (166), it was suggested that ivermectin, like PNU-120596, LY-2087101, and others, acts on $\alpha 7$ -nAChRs at the level of a new type of allosteric modulatory site (167, 168), tentatively identified by mutagenesis within the transmembrane domain.

Amazingly, general anesthetics such as propofol and desflurane, which behave as negative modulators of the prokaryotic *Gloeobacter* receptor (169), possess a common binding site identified by x-ray analysis of the *Gloeobacter* receptor within the upper part of the transmembrane domain of each protomer inside a cavity delimited by TM1, TM3, and TM2 within each subunit (169). The cavity of the general anesthetics is accessible from the lipid bilayer, and its entrance is obstructed by a lipid alkyl chain that clashes with propofol binding. Thus, lipids might be the endogenous ligands of this membrane allosteric site (169). These sites appear to be homologous to important modulatory sites of pentameric receptors, such as for ethanol (170, 171), anticonvulsants, anesthetics, and diuretics acting on the glycine or GABA_A receptor or on nAChR, as well present in the transmembrane domain both within and between subunits (172–174).

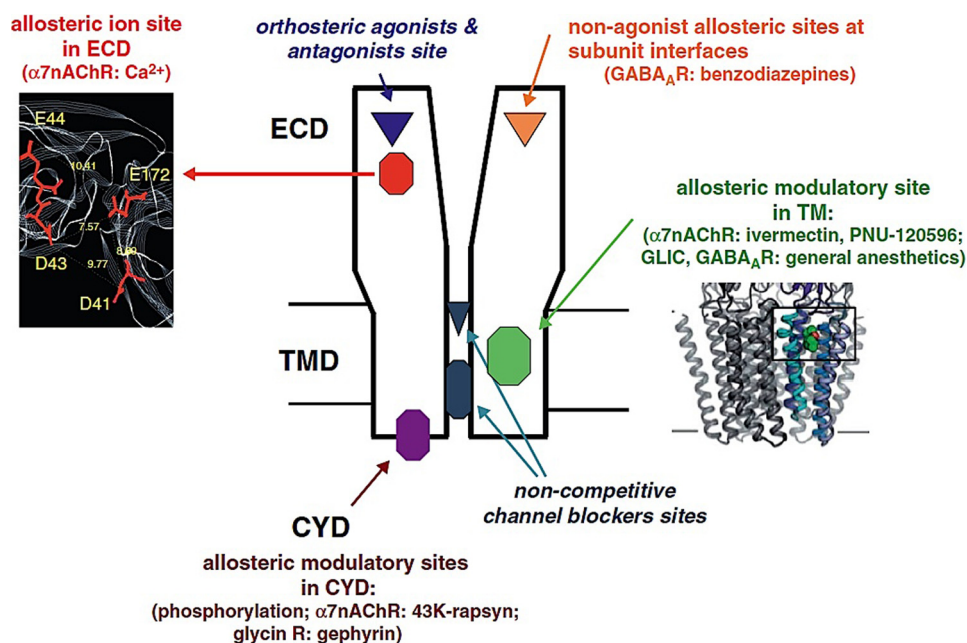


FIGURE 6. **Schematic representation of the various classes of orthosteric and allosteric sites in pentameric ligand-gated ion channels.** Orthosteric sites (shown in *italic blue type*) include the neurotransmitter-binding site and the ion channel. Allosteric sites (shown in various colors) are present in all three domains. *Inset, upper left*, model of the allosteric Ca^{2+} site in α -nAChR (from Ref. 146); *lower right*, x-ray structure of the *Gloeobacter* receptor intrasubunit site for general anesthetics (green) in the transmembrane domain (Ref. 169; from Ref. 187). ECD, extracellular domain; TMD, transmembrane domain; CYD, cytoplasmic domain; GABA_AR, GABA_A receptor; GLIC, *Gloeobacter* ligand-gated ion channel.

In homopentameric pentamers such as $\alpha 7$ -nAChR, there are five active sites per pentamer located at subunit interfaces in the extracellular domain. In heteropentameric neuronal nAChRs such as $\alpha 4\beta 2$ -nAChR, not all five homologous sites bind ACh. The non-agonist-binding interface may accommodate, using ACh-binding protein as a model, galantamine, strychnine, cocaine, and morphine at micromolar concentrations (175–177). The possible binding of allosteric modulators at interfaces in nAChRs that do not normally bind nicotinic ligands might be homologous (178) to the important binding site through which benzodiazepines allosterically potentiate GABA_A receptors. Considerable biochemical, pharmacological, and modeling evidence has since then demonstrated that benzodiazepine ligands bind to intersubunit sites in the extracellular domain of the GABA_A receptor that are homologous to the GABA site but do not bind GABA (172–174). Benzodiazepines, after their discovery by Leo Sternbach in 1955, have become the most commonly prescribed psychopharmaceutical drug and the eleventh-most prescribed drug overall, with 46 million prescriptions written for them in 2010 in the United States! Other allosteric modulatory sites are present on the cytoplasmic domain and may play important roles in the clustering, stabilization, and modulation of receptor functions (21, 179, 180, 187).

Without a doubt, the concept of allosteric modulation has created a major landmark in the strategies of drug design for ligand-gated ion channels but also GPCRs, resulting in the successful development of new classes of drugs used in the clinic, such as Gleevec (allosteric inhibitor of Abl tyrosine kinase), cinacalcet (allosteric activator of the calcium-sensing receptor), and maraviroc (allosteric inhibitor of chemokine receptor) (158, 159).

Allosteric Models of Short- and Long-term Synaptic Plasticity

Since my early days in Jacques Monod's laboratory, where I was assigned to work on bacterial proteins, I faithfully remained attached to my adolescent wishes to explore the chemistry of higher organisms. I sought to exploit the knowledge acquired with the chemistry and allosteric properties of the nAChR to progress in the exploration of the well recognized plasticity of brain functions. I realized that to bridge the gap from molecules to cognition, multiple nested levels of organization had to be taken into consideration. Additional theoretical reflections on allosteric receptors were needed, which integrate the molecular, synaptic, neuronal assemblies and cognitive levels.

The first attempt dealt with the elementary process of synapse formation during the postnatal development period, which is especially long in humans (up to 15 years).

In 1970, I had read Jacques Monod's *Chance and Necessity* with great interest but found Monod's position on brain development much too biased by innate influences. As a well informed admirer of Torsten Wiesel and David Hubel's work on the effects of experience on the postnatal development of the visual cortex, I did not share their functional validation hypothesis of preformed innate patterns of nerve connections. The proposal I suggested was that, as an alternative to the classical nativist *versus* empiricist instructive views, an epigenetic "Darwinian" mechanism of synapse selection took place (181–183). It was suggested that overproduced and variable distributions of connections would become transiently established in the course of synapse formation. Then, at a critical or sensitive period, a synapse selection would take place through some kind of trial-and-error process under the control of the electrical and chemical activity of the network (182, 183). As a consequence, some synapses would be stabilized, whereas others, in agreement with earlier observations (184), would regress. The model that was mathematically formalized together with Philippe Courrège and Antoine Danchin (182) had several major consequences. First, it predicted a phenotypic variability of the brain that is found in monozygotic twins. Also, it was an incentive for the search for synapse elimination (frequently called synaptic pruning) and its control by activity. It has since then been recognized in many systems (185, 186)² and becomes of critical importance with human babies because of the extended interactions taking place with the social and cultural environment.

The model aroused many and constructive debates (188, 189) but also concrete biochemical investigations, for example, on the differentiation and plasticity of the postsynaptic domain (190, 191), and the discovery that distinct DNA elements (190) and transcription factors (192) control the targeting of receptor transcription under the neuromuscular synapse and its repression by electrical activity outside the end plate, followed by the identification of these various components (124). Finally, post-transcriptional mechanisms, including the assembly of the nAChR by the cytoplasmic 43K-rapsyn protein (87, 193) into supramolecular aggregates, revealed a progressive compartmentalization of the dispersed nAChR at the sub-junctional level (194, 195). These studies unveiled the contribution of endogenous trophic factors and electrical activity in the assembly of the subsynaptic membrane (124) and offered a plausible mechanism of long-term synaptic plasticity at the gene expression level.

² J.-P. Changeux, manuscript in preparation.

In the chapter "Mental Objects" of *Neuronal Man: The Biology of Mind* (196), inspired by the investigations of the allosteric modulation of the nAChR, I attempted to extend the theory of synapse selection to short-term learning in the brain. In 1982, Thierry Heidmann and I proposed a model for coincident reading of synaptic signals by an allosteric receptor and thus of short-term synaptic efficacy to be able to eventually link individual neurons into cooperative Hebbian assemblies (197). It is based upon the ability of post-synaptic receptors to integrate extra- and intracellular signals through their transmembrane organization via the allosteric modulatory sites and to shift the equilibrium between desensitized and activatable states. This would then determine the amplitude of the change of synaptic efficacy. This mechanism differs from that suggested for the glutamate NMDA receptor, which is based on the voltage-sensitive blocking of a "rigid" ion channel by Mg^{2+} ions; yet it strikingly resembles the proposal by Eric Kandel to account for data from *Aplysia* learning, although the allosteric mechanisms involved were not examined (198, 199).

This allosteric learning mechanism served as an elementary building block (200–202) in the subsequent modeling, realized together with Stanislas Dehaene, of cognitive functions such as the well known delayed-response tasks, which, in mammals, mobilize the prefrontal cortex (203). We proposed that the prefrontal cortex produces variable "anticipations," or hypotheses, as transient patterns of spontaneous neuronal firings (200–202, 204–206). These firings could be selected (*versus* destabilized) at the level of allosteric receptors through the release of a positive (or negative) reward signal evoked by a successful (or unsuccessful) interaction with the outside world (generator of diversity). The neuronal systems specializing in reward and punishment are known to engage defined neurotransmitters and co-existing peptides, including dopamine and serotonin, or ACh/nicotine, together with their receptors, including the nAChR. More elaborate reward-learning algorithms based on predictive Hebbian learning were subsequently reported by other groups (207) and implemented both *in silico* and *in vivo* with monkeys (208, 209). An important consequence of these models, which rely on allosteric receptors, was to establish plausible biochemical links with concrete receptor pharmacology.

Nicotine Addiction, General Anesthesia, and Cognitive Functions

Smoking is the most important preventable cause of mortality and morbidity worldwide. Nicotine, the principal if not sole addictive component of tobacco smoke, exerts reinforcing effects through its action on brain nAChRs

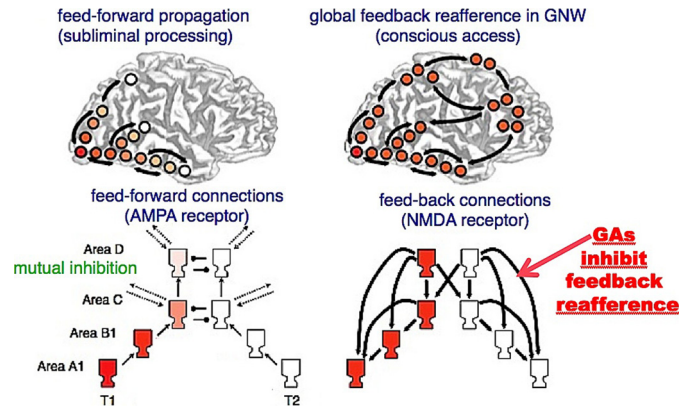


FIGURE 7. Schematic representation of the hypothesized events leading to conscious access according to the global neuronal workspace model and the action of general anesthetics on cortical refference (232, 237). *GNW*, global neuronal workspace; *GAs*, general anesthetics.

together with the laying down of long-term traces in the brain. Their identification and the eventual contribution of allosteric mechanisms to these alterations hinge upon the difficulty that nAChR oligomers are most often composed of several different subunits (sampled among the seventeen ones coded in the human genome) with distinct allosteric properties and are distributed over several brain areas. To identify the patterns of nAChR subunits that contribute to nicotine addiction, new strategies were developed in mice, including nAChR subunit gene deletions, targeted knock-in gene mutations, and re-expression of a deleted gene using stereotaxic injection of a lentiviral vector carrying the missing gene or the relevant siRNA (210–213). The data obtained *in vivo* with these genetically modified mice (214–217) provided evidence that within the dopaminergic neurons from the ventral tegmental area, nAChRs containing combinations of $\alpha 4$ -, $\alpha 3$ -, $\alpha 5$ -, $\alpha 6$ -, and $\beta 2$ -subunits mediate the rewarding effects of nicotine administration (215, 218–220). In all of these instances, the different kinds of oligomers involved display characteristic allosteric properties for channel activation, fast and slow desensitization, allosteric modulation, or up-regulation (221–224). These properties thus offer a novel panel of conformational targets for long-term smoking cessation therapies and possibly for alcohol consumption (225).

Addiction is viewed as the end point of a series of transitions from initial voluntary drug use to the “loss of control” over this behavior. Theoretical models (226, 227) and experimental approaches have been suggested for the neuronal bases of drug-elicited loss of control. Smokers with brain damage involving the insula are more likely to quit smoking than smokers with brain damage not involving the insula (228). Also, diffusion tensor imaging studies have revealed that prenatal and adolescent exposure to tobacco smoke alters the development of the microstruc-

ture of the white matter, with increased fractional anisotropy in the right and left frontal regions and in the genu of the corpus callosum (229). There is also electrophysiological evidence supporting a direct action of nicotine on axon conduction at the level of nAChRs present at the nodes of Ranvier (230) and at the white matter level (231). These observations support the proposal that nAChRs control a “gating circuit,” which itself modulates, in a top-down manner and through the long-range cortical connections of the “global neuronal workspace” (206, 232, 233), nicotine uptake and addiction (227, 234) (Fig. 7). In a general manner, they underline the contribution of ACh and more specifically nAChRs to global “conscious” control (232–235).

In this framework, general anesthesia offers the most vivid evidence for an allosteric modulation of higher brain functions (236, 237). Administration of general anesthetics to patients causes a reversible loss of consciousness manifested, in particular, by a loss of response to oral commands. At appropriate levels of general anesthetics, a deeply unconscious state follows in which the electroencephalogram pattern resembles slow-wave sleep. Parallel biophysical studies on the detailed neural architecture accounting for conscious access called masking experiments (232, 238) suggested the contribution of two distinct sets of connections (239, 240). Bottom-up feedforward short connections link primary sensory areas up to the prefrontal cortex in a cascade. Complementary long-distance top-down connections project from the prefrontal cortex to all preceding areas, creating a top-down global “cortical refference” (Fig. 7). Quantitative electroencephalogram analysis in humans under propofol anesthesia showed that upon induction, the dominant feedback connectivity of the base-line conscious state was differentially and reversibly inhibited in the frontoparietal network (241). Such selective inhibition would plausibly



FIGURE 8. The author at the 2009 meeting “Darwin: 200 Years” at Collège de France. This photo was provided by the Collège de France.

result from a positive allosteric modulation of the cortical GABA_A receptor (among other molecular targets) (242). As mentioned above, these allosteric sites for general anesthetics have been identified in their transmembrane domain (Fig. 6) (169, 172–174). In other words, the loss of consciousness caused by general anesthetic infusion would ultimately result from a modulation of brain allosteric receptors and/or ion channels. The mode of action of general anesthetics on the brain may serve as a concrete illustration of how drugs may allosterically modulate higher brain functions.

Conclusion: The Brain as a Chemical Machine?

In *Neuronal Man: The Biology of Mind* (p. 114 of Ref. 196), I wrote “We must not underestimate the importance of the diversity of the neurons...the multiplicity of the chemical mediators... (to the extent that) the chemical makeup of the cells that participate in a sensation...is closer to a canvas by Georges Seurat than a composition by Piet Mondrian.” This is still very true. The many neurotransmitters and coexisting neuropeptides (243, 244),

their multiple receptors (49, 72, 196, 245), which may reach hundreds per cell (246), and their possible combinations are immensely rich and diverse. Also, any individual neuron differs from any other neuron within a given category by its connectivity, its biochemistry, and its physiology due to the developmental variability consecutive to the synapse selection mechanism (182, 196). Confronted with such a complexity, new ideas and modeling enterprises are needed (Fig. 8).

Such a multidisciplinary program is expected to contribute to our understanding of the brain but also to be an incentive for the design and development of technical applications. One of them is optogenetics, which allows the control of the activity of neuronal networks underlying behavior through the millisecond allosteric transition of channelrhodopsin or related photoactivatable molecules (247). Another one of biomedical importance is the design of drugs directed against brain diseases. For instance, it is now recognized that receptors may be present under diverse conformations that spontaneously exist in the absence of ligand. From a drug design perspective, the model suggests that the design of drugs should be targeted to site(s) present on these defined conformation(s) rather than to a single category of rigid binding sites, resulting in the production of agonistic *versus* antagonistic ligands. These studies have also led to the successful discovery of novel allosteric modulators that regulate the activity of ligand-gated ion channels, as well as of other receptors such as GPCRs and tyrosine kinase receptors. Among them are the benzodiazepines, which are the most commonly prescribed psychopharmaceutical drug in the world and which behave as allosteric modulators of GABA_A receptors. The model also predicts that by altering the unliganded equilibrium between discrete conformational states, gene mutations may cause constitutive receptor activation (or inhibition) with important pathological consequences (124, 248). Also, these modeling approaches have enlarged the field of short- and long-term learning to still largely unexplored conformational mechanisms. The “molecular selectionist” schemes of short- and long-term learning, which take place within the genetic envelope of the adult and developing brain, are expected to have important consequences on higher brain functions and their pathologies. Last but not least, the analysis of the patterns of genes expressed in the course of brain development led to the suggestion of a new strategy for designing drugs as orthosteric and allosteric ligands, targeting the transcription factors regulating the “predisposition” genes rather than the proteins they encode (249).

One should not forget in this respect that transcription factors are *bona fide* allosteric proteins.

These theoretical reasonings and data further document and enrich what may be referred to as a “chemical theory of higher brain functions” (196, 250). This creates a striking landmark in the thinking of brain sciences by causally and reciprocally linking the molecular to the cognitive levels both within the individual brain and between brains in the social and cultural environment, thus suggesting new bridges between brain sciences and humanities. From a philosophical point of view, it appears legitimate to say that the future understanding of the mind-brain relationships and the relevant mental processes is likely to rest upon the biochemical world of the allosteric transitions that mediate interneuronal communications through the multiple levels of organization spanning the human brain.

Acknowledgments—I gratefully thank Drs. Tamas Bartfai, Henri Korn, Uwe Maskos, and Ákos Némethy for comments and suggestions.

Address correspondence to: changeux@noos.fr.

REFERENCES

- Robinson, E. D. (1979) Claude Bernard. Pioneer of regulatory biology. *JAMA* **242**, 1283–1284
- Changeux, J.-P. (1958) Some biological characters of a parasitic copepod from holothurians: *Allantogynus delamarei* n.g.n.sp. *C. R. Hebd. Seances Acad. Sci.* **247**, 961–964
- Changeux, J.-P. (1960) On the biochemical expression of genetic determinants of *Escherichia coli* introduced to *Salmonella typhimurium*. *C. R. Hebd. Seances Acad. Sci.* **250**, 1575–1577
- Umbarger, H. E. (1956) Evidence for a negative-feedback mechanism in the biosynthesis of isoleucine. *Science* **123**, 848
- Changeux, J.-P. (1961) The feedback control mechanisms of biosynthetic L-threonine deaminase by L-isoleucine. *Cold Spring Harb. Symp. Quant. Biol.* **26**, 313–318
- Davis, B. (1961) Commentary to Changeux's presentation. *Cold Spring Harbor Symp. Quant. Biol.* **26**, 318
- Gerhart, J. C., and Pardee, A. B. (1962) The enzymology of control by feedback inhibition. *J. Biol. Chem.* **237**, 891–896
- Changeux, J.-P. (2011) 50th anniversary of the word “allosteric.” *Protein Sci.* **20**, 1119–1124
- Haldane, J. B. S. (1930) *Enzymes*, Longmans, Green and Co., London
- Koshland, D. E., Jr. (1956) Molecular geometry in enzyme action. *J. Cell. Physiol.* **47**, 217–234
- Koshland, D. E., Jr. (1959) Enzyme flexibility and enzyme action. *J. Cell. Comp. Physiol.* **54**, 245–258
- Monod, J., Changeux, J.-P., and Jacob, F. (1963) Allosteric proteins and cellular control systems. *J. Mol. Biol.* **6**, 306–329
- Koshland, D. E., Jr. (1963) The role of flexibility in enzyme action. *Cold Spring Harbor Symp. Quant. Biol.* **28**, 473–482
- Monod, J., Wyman, J., and Changeux, J.-P. (1965) On the nature of allosteric transitions: a plausible model. *J. Mol. Biol.* **12**, 88–118
- Changeux, J.-P. (1964) On the allosteric properties of L-threonine deaminase. I. Methods of studying biosynthetic L-threonine deaminase. *Bull. Soc. Chim. Biol.* **46**, 927–946
- Changeux, J.-P. (1963) Allosteric interactions on biosynthetic L-threonine deaminase from *E. coli* K12. *Cold Spring Harb. Symp. Quant. Biol.* **28**, 497–504
- Buc, H. (2006) Interactions between Jacques Monod and Jeffries Wyman (or the burdens of co-authorship). *Rendiconti Lincei* **17**, 31–49
- Buc, H. (2013) The design of an enzyme: a chronology on the controversy. *J. Mol. Biol.* **425**, 1407–1409
- Perutz, M. F., Rossmann, M. G., Cullis, A. F., Muirhead, H., Will, G., and North, A. C. T. (1960) Structure of haemoglobin: a three-dimensional Fourier synthesis at 5.5-Å resolution, obtained by x-ray analysis. *Nature* **185**, 416–422
- Kendrew, J. C. (1959) Structure and function in myoglobin and other proteins. *Fed. Proc.* **18**, 740–751
- Changeux, J.-P. (2012) Allosteric and the Monod-Wyman-Changeux model after 50 years. *Annu. Rev. Biophys.* **41**, 103–133
- Perutz, M. F. (1963) X-ray analysis of hemoglobin. *Science* **140**, 863–869
- Muirhead, H., and Perutz, M. F. (1963) Structure of haemoglobin. A three-dimensional Fourier synthesis of reduced human haemoglobin at 5–5 Å resolution. *Nature* **199**, 633–638
- Wyman, J., Jr. (1948) Heme Proteins. *Adv. Protein Chem.* **4**, 407–531
- Changeux, J.-P. (2013) The origins of allostery: from personal memories to material for the future. *J. Mol. Biol.* **425**, 1396–1406
- Boehr, D. D., Nussinov, R., and Wright, P. E. (2009) The role of dynamic conformational ensembles in biomolecular recognition. *Nat. Chem. Biol.* **5**, 789–796
- Deupi, X., and Kobilka, B. K. (2010) Energy landscapes as a tool to integrate GPCR structure, dynamics, and function. *Physiology* **25**, 293–303
- Hammes, G. G., Chang, Y. C., and Oas, T. G. (2009) Conformational selection or induced fit: a flux description of reaction mechanism. *Proc. Natl. Acad. Sci. U.S.A.* **106**, 13737–13741
- Cui, Q., and Karplus, M. (2008) Allostery and cooperativity revisited. *Protein Sci.* **17**, 1295–1307
- Pauling, L. (1935) The oxygen equilibrium of hemoglobin and its structural interpretation. *Proc. Natl. Acad. Sci. U.S.A.* **21**, 186–191
- Koshland, D. E., Jr., Némethy, G., and Filmer, D. (1966) Comparison of experimental binding data and theoretical models in proteins containing subunits. *Biochemistry* **5**, 365–385
- Rubin, M. M., and Changeux, J.-P. (1966) On the nature of allosteric transitions: implications of non-exclusive ligand binding. *J. Mol. Biol.* **21**, 265–274
- Gerhart, J. C., and Pardee, A. B. (1964) Aspartate transcarbamylase, an enzyme designed for feedback inhibition. *Fed. Proc.* **23**, 727–735
- Changeux, J.-P., Gerhart, J. C., and Schachman, H. K. (1968) Allosteric interactions in aspartate transcarbamylase. I. Binding of specific ligands to the native enzyme and its isolated subunits. *Biochemistry* **7**, 531–538
- Gerhart, J. C., and Schachman, H. K. (1968) Allosteric interactions in aspartate transcarbamylase. II. Evidence for different conformational states of the protein in the presence and absence of specific ligands. *Biochemistry* **7**, 538–552
- Changeux, J.-P., and Rubin, M. M. (1968) Allosteric interactions in aspartate transcarbamylase. III. Interpretations of experimental data in terms of the model of Monod, Wyman, and Changeux. *Biochemistry* **7**, 553–561
- Blangy, D., Buc, H., and Monod, J. (1968) Kinetics of the allosteric interactions of phosphofructokinase from *Escherichia coli*. *J. Mol. Biol.* **31**, 13–35
- Perutz, M. F., Wilkinson, A. J., Paoli, M., and Dodson, G. G. (1998) The stereochemical mechanism of the cooperative effects in hemoglobin revisited. *Annu. Rev. Biophys. Biomol. Struct.* **27**, 1–34
- Edelstein, S. J. (1971) Extensions of the allosteric model for haemoglobin. *Nature* **230**, 224–227
- Cammarata, M., Levantino, M., Wulff, M., and Cupane, A. (2010) Unveiling the timescale of the R-T transition in human hemoglobin. *J. Mol. Biol.* **400**, 951–962
- Fischer, S., Olsen, K. W., Nam, K., and Karplus, M. (2011) Unsuspected pathway of the allosteric transition in hemoglobin. *Proc. Natl. Acad. Sci. U.S.A.* **108**, 5608–5613
- Levantino, M., Spilotros, A., Cammarata, M., Schirò, G., Ardiccioni, C., Vallone, B., Brunori, M., and Cupane, A. (2012) The Monod-Wyman-Changeux allosteric model accounts for the quaternary transition dynamics in wild type and a recombinant mutant human hemoglobin. *Proc. Natl. Acad. Sci. U.S.A.* **109**, 14894–14899
- Tekpinar, M., and Zheng, W. (2013) Coarse-grained and all-atom modeling of structural states and transitions in hemoglobin. *Proteins* **81**, 240–252
- Lewis, M. (2005) The *lac* repressor. *C. R. Biol.* **328**, 521–548
- Lewis, M. (2011) A tale of two repressors. *J. Mol. Biol.* **409**, 14–27
- Gronemeyer, H., Gustafsson, J. A., and Laudet, V. (2004) Principles for modulation of the nuclear receptor superfamily. *Nat. Rev. Drug Discov.* **3**, 950–964
- Chandra, V., Huang, P., Hamuro, Y., Raghuram, S., Wang, Y., Burris, T. P., and Rastinejad, F. (2008) Structure of the intact PPAR-γ-RXR-α nuclear receptor complex on DNA. *Nature* **456**, 350–356
- Huang, P., Chandra, V., and Rastinejad, F. (2010) Structural overview of the

- nuclear receptor superfamily: insights into physiology and therapeutics. *Annu. Rev. Physiol.* **72**, 247–272
49. Corringer, P. J., Poitevin, F., Prevost, M. S., Sauguet, L., Delarue, M., and Changeux, J.-P. (2012) Structure and pharmacology of pentameric receptor channels: from bacteria to brain. *Structure* **20**, 941–956
 50. Canals, M., Lane JR, Wen, A., Scammells, P. J., Sexton, P. M., and Christopoulos, A. (2012) A Monod-Wyman-Changeux mechanism can explain G protein-coupled receptor (GPCR) allosteric modulation. *J. Biol. Chem.* **287**, 650–659
 51. Prevost, M. S., Sauguet, L., Nury, H., Van Renterghem, C., Huon, C., Poitevin, F., Baaden, M., Delarue, M., and Corringer, P. J. (2012) A locally closed conformation of a bacterial pentameric proton-gated ion channel. *Nat. Struct. Mol. Biol.* **19**, 642–649
 52. Nygaard, R., Zou, Y., Dror, R. O., Mildorf, T. J., Arlow, D. H., Manglik, A., Pan, A. C., Liu, C. W., Fung, J. J., Bokoch, M. P., Thian, F. S., Kobilka, T. S., Shaw, D. E., Mueller, L., Prosser, R. S., and Kobilka, B. K. (2013) The dynamic process of β_2 -adrenergic receptor activation. *Cell* **152**, 532–542
 53. Novick, A., and Szilard, L. (1954) *Dynamics of Growth Processes*, pp. 21–32, Princeton University Press, Princeton, NJ
 54. Liu, Y.-Y., Slotine, J.-J., and Barabási, A.-L. (2013) Observability of complex systems. *Proc. Natl. Acad. Sci. U.S.A.* **110**, 2460–2465
 55. Changeux, J.-P. (1966) Responses of acetylcholinesterase from *Torpedo marmorata* to salts and curarizing drugs. *Mol. Pharmacol.* **2**, 369–392
 56. Changeux, J.-P. (1969) Symmetry and cooperative properties of biological membranes. in *Nobel Symposium 11: Symmetry and Function of Biological Systems at the Macromolecular Level* (Engström, A., and Strandberg, B., eds) pp. 235–256, Almquist & Wiksell, Stockholm
 57. Changeux, J.-P., Thiéry, J., Tung, Y., and Kittel, C. (1967) On the cooperativity of biological membranes. *Proc. Natl. Acad. Sci. U.S.A.* **57**, 335–341
 58. Duke, T. A., and Bray, D. (1999) Heightened sensitivity of a lattice of membrane receptors. *Proc. Natl. Acad. Sci. U.S.A.* **96**, 10104–10108
 59. Shimizu, T. S., Le Novère, N., Levin, M. D., Bevil, A. J., Sutton, B. J., and Bray, D. (2000) Molecular model of a lattice of signalling proteins involved in bacterial chemotaxis. *Nat. Cell Biol.* **2**, 792–796
 60. Gurevich, V. V., and Gurevich, E. V. (2008) How and why do GPCRs dimerize? *Trends Pharmacol. Sci.* **29**, 234–240
 61. Breitwieser, G. E. (2004) G protein-coupled receptor oligomerization: implications for G protein activation and cell signaling. *Circ. Res.* **94**, 17–27
 62. Pin, J.-P., Comps-Agrar, L., Maurel, D., Monnier, C., Rives, M. L., Trinquet, E., Kniazeff, J., Rondard, P., and Prézeau, L. (2009) G-protein-coupled receptor oligomers: two or more for what? Lessons from mGlu and GABAB receptors. *J. Physiol.* **587**, 5337–5344
 63. Ferré, S., Fredholm, B. B., Morelli, M., Popoli, P., and Fuxe, K. (1997) Adenosine dopamine receptor-receptor interactions as an integrative mechanism in the basal ganglia. *Trends Neurosci.* **20**, 482–487
 64. Agnati, L. F., Fuxe, K., Zini, I., Lenzi, P., and Hökfelt, T. (1980) Aspects on receptor regulation and isoreceptor identification. *Med. Biol.* **58**, 182–187
 65. Ferré, S., Baler, R., Bouvier, M., Caron, M. G., Devi, L. A., Durroux, T., Fuxe, K., George, S. R., Javitch, J. A., Lohse, M. J., Mackie, K., Milligan, G., Pfleger, K. D. G., Pin, J.-P., Volkow, N. D., Waldhoer, M., Woods, A. S., and Franco, R. (2009) Building a new conceptual framework for receptor heteromers. *Nat. Chem. Biol.* **5**, 131–134
 66. Fuxe, K., Canals, M., Torvinen, M., Marcellino, D., Terasmaa, A., Genedani, S., Leo, G., Guidolin, D., Diaz-Cabiale, Z., Rivera, A., Lundstrom, L., Langel, U., Narvaez, J., Tanganelli, S., Lluis, C., Ferré, S., Woods, A., Franco, R., and Agnati, L. F. (2007) Intramembrane receptor-receptor interactions: a novel principle in molecular medicine. *J. Neural Transm.* **114**, 49–75
 67. Horwich, A. L., Fenton, W. A., Chapman, E., and Farr, G. W. (2007) Two families of chaperonin: physiology and mechanism. *Annu. Rev. Cell Dev. Biol.* **23**, 115–145
 68. Bai, F., Branch, R. W., Nicolau, D. V., Jr., Pilizota, T., Steel, B. C., Maini, P. K., and Bery, R. M. (2010) Conformational spread as a mechanism for cooperativity in the bacterial flagellar switch. *Science* **327**, 685–689
 69. Garcia, H. G., Kondev, J., Orme, N., Theriot, J. A., and Phillips, R. (2011) Thermodynamics of biological processes. *Methods Enzymol.* **492**, 27–59
 70. Langley, J. N. (1905) On the reaction of cells and of nerve-endings to certain poisons, chiefly as regards the reaction of striated muscle to nicotine and to curari. *J. Physiol.* **33**, 374–413
 71. Dale, H. H. (1942) Wartime arrangements for international biological standards. *Br. Med. J.* **2**, 385–387
 72. Changeux, J.-P. (2012) The nicotinic acetylcholine receptor: the founding father of the pentameric ligand-gated ion channel superfamily. *J. Biol. Chem.* **287**, 40207–40215
 73. Changeux, J.-P. (1965) On the allosteric properties of biosynthetic L-threonine deaminase. VI. General discussion. *Bull. Soc. Chim. Biol.* **47**, 281–300
 74. Morel, N., Bon, S., Greenblatt, H. M., Van Belle, D., Wodak, S. J., Sussman, J. L., Massoulié, J., and Silman, I. (1999) Effect of mutations within the peripheral anionic site on the stability of acetylcholinesterase. *Mol. Pharmacol.* **55**, 982–992
 75. Rosenberry, T. L., Johnson, J. L., Cusack, B., Thomas, J. L., Emani, S., and Venkatasubban, K. S. (2005) Interactions between the peripheral site and the acylation site in acetylcholinesterase. *Chem. Biol. Interact.* **157–158**, 181–189
 76. Silman, I., and Sussman, J. L. (2008) Acetylcholinesterase: how is structure related to function? *Chem. Biol. Interact.* **175**, 3–10
 77. Schoffeniels, E., and Nachmansohn, D. (1957) An isolated single electroplax preparation. I. New data on the effect of acetylcholine and related compounds. *Biochim. Biophys. Acta* **26**, 1–15
 78. Kasai, M., and Changeux, J.-P. (1970) Demonstration of the excitation by cholinergic agonists from fractions of purified membranes, *in vitro*. *C. R. Acad. Sci. Hebd. Seances Acad. Sci. D* **270**, 1400–1403
 79. Kasai, M., and Changeux, J.-P. (1971) *In vitro* excitation of purified membrane fragments by cholinergic agonists. *J. Memb. Biol.* **6**, 1–23
 80. Gilbert, W., and Müller-Hill, B. (1966) Isolation of the *lac* repressor. *Proc. Natl. Acad. Sci. U.S.A.* **56**, 1891–1898
 81. Lee, C.-Y., and Chang, C. (1966) Modes of actions of purified toxins from elapid venoms on neuromuscular transmission. *Mem. Inst. Butantan* **33**, 555–572
 82. Changeux, J.-P., Kasai, M., and Lee, C.-Y. (1970) Use of a snake venom toxin to characterize the cholinergic receptor protein. *Proc. Natl. Acad. Sci. U.S.A.* **67**, 1241–1247
 83. Olsen, R. W., Meunier, J.-C., and Changeux, J.-P. (1972) Progress in purification of the cholinergic receptor protein from *Electrophorus electricus* by affinity chromatography. *FEBS Lett.* **28**, 96–100
 84. Cartaud, J., Benedetti, E. L., Cohen, J. B., Meunier, J.-C., and Changeux, J.-P. (1973) Presence of a lattice structure in membrane fragments rich in nicotinic receptor protein from the electric organ of *Torpedo marmorata*. *FEBS Lett.* **33**, 109–113
 85. Nickel, E., and Potter, L. T. (1973) Ultrastructure of isolated membranes of *Torpedo electric* tissue. *Brain Res.* **57**, 508–517
 86. Cartaud, J., Bon, S., and Massoulié, J. (1978) *Electrophorus* acetylcholinesterase. Biochemical and electron microscope characterization of low ionic strength aggregates. *J. Cell Biol.* **7**, 315–322
 87. Cartaud, J., Sobel, A., Rousselet, A., Devaux, P. F., and Changeux, J.-P. (1981) Consequences of alkaline treatment for the ultrastructure of the acetylcholine-receptor-rich membranes from *Torpedo marmorata* electric organ. *J. Cell Biol.* **90**, 418–426
 88. Ross, M. J., Klymkowsky, M. W., Agard, D. A., and Stroud, R. M. (1977) Structural studies of a membrane-bound acetylcholine receptor from *Torpedo californica*. *J. Mol. Biol.* **116**, 635–659
 89. Kistler, J., Stroud, R. M., Klymkowsky, M. W., Lalancette, R. A., and Fairclough, R. H. (1982) Structure and function of an acetylcholine receptor. *Biophys. J.* **37**, 371–383
 90. Kistler, J., and Stroud, R. M. (1981) Crystalline arrays of membrane-bound acetylcholine receptor. *Proc. Natl. Acad. Sci. U.S.A.* **78**, 3678–3682
 91. Bon, F., Lebrun, E., Gommel, J., van Rapenbusch, R., Cartaud, J., Popot, J. L., and Changeux, J.-P. (1982) Relative orientation of the two oligomers composing the heavy form of the acetylcholine receptor from *Torpedo marmorata*. *C. R. Seances Acad. Sci. III* **295**, 199–204
 92. Bon, F., Lebrun, E., Gommel, J., Van Rapenbusch, R., Cartaud, J., Popot, J. L., and Changeux, J.-P. (1984) Image analysis of the heavy form of the acetylcholine receptor from *Torpedo marmorata*. *J. Mol. Biol.* **176**, 205–237
 93. Brisson, A., and Unwin, P. N. (1985) Quaternary structure of the acetylcholine receptor. *Nature* **315**, 474–477
 94. Unwin, N. (2005) Refined structure of the nicotinic acetylcholine receptor at 4 Å resolution. *J. Mol. Biol.* **346**, 967–989
 95. Hucho, F., and Changeux, J.-P. (1973) Molecular weight and quaternary structure of the cholinergic receptor protein extracted by detergents from *Electrophorus electricus* electric tissue. *FEBS Lett.* **38**, 11–15
 96. Weill, C. L., McNamee, M. G., and Karlin, A. (1974) Affinity labeling of purified acetylcholine receptor from *Torpedo californica*. *Biochem. Biophys. Res. Commun.* **61**, 997–1003
 97. Karlin, A. (1980) Molecular properties of nicotinic acetylcholine receptors. in *Cell Surface Reviews* (Poste, G., Nicolson, G. L., and Cotman, C. W., eds) pp. 191–260, Elsevier Science Ltd., Amsterdam
 98. Raftery, M. A., Vandlen, R., Michaelson, D., Bode, J., Moody, T., Chao, Y.,

- Reed, K., Deutsch, J., and Duguid, J. (1974) The biochemistry of an acetylcholine receptor. *J. Supramol. Struct.* **2**, 582–592
99. Raftery, M. A., Hunkapiller, M. W., Strader, C. D., and Hood, L. E. (1980) Acetylcholine receptor: complex of homologous subunits. *Science* **208**, 1454–1456
100. Lindstrom, J., Merlie, J., and Yogeewaran, G. (1979) Biochemical properties of acetylcholine receptor subunits from *Torpedo californica*. *Biochemistry* **18**, 4465–4470
101. Devillers-Thiery, A., Changeux, J.-P., Paroutaud, P., and Strosberg, A. D. (1979) The amino-terminal sequence of the 40,000 molecular weight subunit of the acetylcholine receptor protein from *Torpedo marmorata*. *FEBS Lett.* **104**, 99–105
102. Hunkapiller, M. W., Strader, C. D., Hood, L., and Raftery, M. A. (1979) Amino terminal amino acid sequence of the major polypeptide subunit of *Torpedo californica* acetylcholine receptor. *Biochem. Biophys. Res. Commun.* **91**, 164–169
103. Noda, M., Takahashi, H., Tanabe T., Toyosato, M., Furutani, Y., Hirose, T., Asai, M., Inayama, S., Miyata, T., and Numa, S. (1982) Primary structure of α -subunit precursor of *Torpedo californica* acetylcholine receptor deduced from cDNA sequence. *Nature* **299**, 793–797
104. Noda, M., Takahashi, H., Tanabe, T., Toyosato, M., Kikyotani, S., Furutani, Y., Hirose, T., Takashima, H., Inayama, S., Miyata, T., and Numa, S. (1983) Structural homology of *Torpedo californica* acetylcholine receptor subunits. *Nature* **302**, 528–532
105. Noda, M., Takahashi, H., Tanabe, T., Toyosato, M., Kikyotani, S., Hirose, T., Asai, M., Takashima, H., Inayama, S., Miyata, T., and Numa, S. (1983) Primary structures of β - and γ -subunit precursors of *Torpedo californica* acetylcholine receptor deduced from cDNA sequences. *Nature* **301**, 251–255
106. Claudio, T., Ballivet, M., Patrick, J., and Heinemann, S. (1983) Nucleotide and deduced amino acid sequences of *Torpedo californica* acetylcholine receptor γ -subunit. *Proc. Natl. Acad. Sci. U.S.A.* **80**, 1111–1115
107. Sumikawa, K., Houghton, M., Smith, J. C., Bell, L., Richards, B. M., and Barnard, E. A. (1982) The molecular cloning and characterisation of cDNA coding for the α -subunit of the acetylcholine receptor. *Nucleic Acids Res.* **10**, 5809–5822
108. Devillers-Thiery, A., Giraudat, J., Bentaboulet, M., and Changeux, J.-P. (1983) Complete mRNA coding sequence of the acetylcholine binding α -subunit of *Torpedo marmorata* acetylcholine receptor: a model for the transmembrane organization of the polypeptide chain. *Proc. Natl. Acad. Sci. U.S.A.* **80**, 2067–2071
109. Boulter, J., Evans, K., Goldman, D., Martin, G., Treco, D., Heinemann, S., and Patrick, J. (1986) Isolation of a cDNA clone coding for a possible neural nicotinic acetylcholine receptor α -subunit. *Nature* **319**, 368–374
110. Boulter, J., Evans, K., Martin, G., Mason, P., Stengelin, S., Goldman, D., Heinemann, S., and Patrick, J. (1986) Isolation and sequence of cDNA clones coding for the precursor to the γ subunit of mouse muscle nicotinic acetylcholine receptor. *J. Neurosci. Res.* **16**, 37–49
111. Boulter, J., O'Shea-Greenfield, A., Duvoisin, R. M., Connolly, J. G., Wada, E., Jensen, A., Gardner, P. D., Ballivet, M., Deneris, E. S., and McKinnon, D. (1990) $\alpha 3$, $\alpha 5$, and $\beta 4$: three members of the rat neuronal nicotinic acetylcholine receptor-related gene family form a gene cluster. *J. Biol. Chem.* **265**, 4472–4482
112. Couturier, S., Erkman, L., Valera, S., Runger, D., Bertrand, S., Boulter, J., Ballivet, M., and Bertrand, D. (1990) $\alpha 5$, $\alpha 3$, and non- $\alpha 3$: three clustered avian genes encoding neuronal nicotinic acetylcholine receptor-related subunits. *J. Biol. Chem.* **265**, 17560–17567
113. Couturier, S., Bertrand, D., Matter, J.-M., Hernandez, M.-C., Bertrand, S., Millar, N., Valera, S., Barkas T., and Ballivet, M. (1990) A neuronal nicotinic acetylcholine receptor subunit ($\alpha 7$) is developmentally regulated and forms a homo-oligomeric channel blocked by α -Btx. *Neuron* **5**, 847–856
114. Kao, P. N., Dwork, A. J., Kaldany, R. R., Silver, M. L., Wideman, J., Stein, S., and Karlin, A. (1984) Identification of the α -subunit half-cystine specifically labeled by an affinity reagent for the acetylcholine receptor binding site. *J. Biol. Chem.* **259**, 11662–11665
115. Langenbuch-Cachat, J., Bon, C., Mulle, C., Goeldner, M., Hirth, C., and Changeux, J.-P. (1988) Photoaffinity labeling of the acetylcholine binding sites on the nicotinic receptor by an aryl diazonium derivative. *Biochemistry* **27**, 2337–2345
116. Dennis, M., Giraudat, J., Kotzyba-Hibert, F., Goeldner, M., Hirth, C., Chang, J.-Y., and Changeux, J.-P. (1986) A photoaffinity ligand of the acetylcholine-binding site predominantly labels the region 179–207 of the α -subunit on native acetylcholine receptor from *Torpedo marmorata*. *FEBS Lett.* **207**, 243–249
117. Dennis, M., Giraudat, J., Kotzyba-Hibert, F., Goeldner, M., Hirth, C., Chang, J.-Y., Lazure, C., Chrétien, M., and Changeux, J.-P. (1988) Amino acids of the *Torpedo marmorata* acetylcholine receptor subunit labeled by a photoaffinity ligand for the acetylcholine binding site. *Biochemistry* **27**, 2346–2357
118. Galzi, J.-L., Revah, F., Black, D., Goeldner, M., Hirth, C., and Changeux, J.-P. (1990) Identification of a novel amino acid α -tyrosine 93 within the cholinergic ligands-binding sites of the acetylcholine receptor by photoaffinity labeling. Additional evidence for a three-loop model of the cholinergic ligands-binding sites. *J. Biol. Chem.* **265**, 10430–10437
119. Giraudat, J., Dennis, M., Heidmann, T., Chang, J.-Y., and Changeux, J.-P. (1986) Structure of the high affinity site for noncompetitive blockers of the acetylcholine receptor: serine 262 of the δ -subunit is labeled by [3 H]chlorpromazine. *Proc. Natl. Acad. Sci. U.S.A.* **83**, 2719–2723
120. Giraudat, J., Dennis, M., Heidmann, T., Haumont, P. Y., Lederer, F., and Changeux, J.-P. (1987) Structure of the high affinity binding site for noncompetitive blockers of the acetylcholine receptor: [3 H]chlorpromazine labels homologous residues in the β and δ chains. *Biochemistry* **26**, 2410–2418
121. Hucho, F., Oberthür, W., and Lottspeich, F. (1986) The ion channel of the nicotinic acetylcholine receptor is formed by the homologous helices M II of the receptor subunits. *FEBS Lett.* **205**, 137–142
122. Imoto, K., Busch, C., Sakmann, B., Mishina, M., Konno, T., Nakai, J., Bujo H., Mori Y., Fukuda, K., and Numa, S. (1988) Rings of negatively charged amino acids determine the acetylcholine receptor channel conductance. *Nature* **335**, 645–648
123. Changeux, J.-P. (1990) The TiPS lecture. The nicotinic acetylcholine receptor: an allosteric protein prototype of ligand-gated ion channels. *Trends Pharmacol. Sci.* **11**, 485–492
124. Changeux, J.-P., and Edelman, S. (2005) *Nicotinic Acetylcholine Receptors: From Molecular Biology to Cognition*, Odile Jacob, New York
125. Heidmann, T., and Changeux, J.-P. (1979) Fast kinetic studies on the interaction of a fluorescent agonist with the membrane-bound acetylcholine receptor from *Torpedo marmorata*. *Eur. J. Biochem.* **94**, 255–279
126. Heidmann, T., and Changeux, J.-P. (1980) Interaction of a fluorescent agonist with the membrane-bound acetylcholine receptor from *Torpedo marmorata* in the millisecond time range: resolution of an "intermediate" conformational transition and evidence for positive cooperative effects. *Biochem. Biophys. Res. Commun.* **97**, 889–896
127. Neubig, R. R., and Cohen, J. B. (1980) Permeability control by cholinergic receptors in *Torpedo* postsynaptic membranes: agonist dose-response relations measured at second and millisecond times. *Biochemistry* **19**, 2770–2779
128. Edelman, S. J., Schaad, O., Henry, E., Bertrand, D., and Changeux, J.-P. (1996) A kinetic mechanism for nicotinic acetylcholine receptors based on multiple allosteric transitions. *Biol. Cybern.* **75**, 361–379
129. Edelman, S. J., Schaad, O., and Changeux, J.-P. (1997) Myasthenic nicotinic receptor mutant interpreted in terms of the allosteric model. *C. R. Acad. Sci. III* **320**, 953–961
130. Edelman, S. J., Schaad, O., and Changeux, J.-P. (1997) Single binding versus single channel recordings: a new approach to study ionotropic receptors. *Biochemistry* **36**, 13755–13760
131. Purohit, P., and Auerbach, A. (2009) Unliganded gating of acetylcholine receptor channels. *Proc. Natl. Acad. Sci. U.S.A.* **106**, 115–120
132. Revah, F., Bertrand, D., Galzi, J.-L., Devillers-Thiery, A., Mulle, C., Hussy, N., Bertrand, S., Ballivet, M., and Changeux, J.-P. (1991) Mutations in the channel domain alter desensitization of a neuronal nicotinic receptor. *Nature* **353**, 846–849
133. Ohno, K., and Engel, A. G. (1998) Congenital myasthenic syndromes: gene mutation. *Neuromuscul. Disord.* **8**, XII–XIII
134. Steinlein, O. K., Mulley, J. C., Propping, P., Wallace, R. H., Phillips, H. A., Sutherland, G. R., Scheffer, I. E., and Berkovic, S. F. (1995) A missense mutation in the neuronal nicotinic acetylcholine receptor $\alpha 4$ subunit is associated with autosomal dominant nocturnal frontal lobe epilepsy. *Nat. Genet.* **11**, 201–203
135. Jadey, S. V., Purohit, P., Bruhova, I., Gregg, T. M., and Auerbach A. (2011) Design and control of acetylcholine receptor conformational change. *Proc. Natl. Acad. Sci. U.S.A.* **108**, 4328–4333
136. Auerbach, A. (2012) Thinking in cycles: MWC is a good model for acetylcholine receptor-channels. *J. Physiol.* **590**, 93–98
137. Steinlein, O. K. (2010) Animal models for autosomal dominant frontal lobe epilepsy: on the origin of seizures. *Expert Rev. Neurother.* **10**, 1859–1867
138. Tasneem, A., Iyer, L. M., Jakobsson, E., and Aravind, L. (2005) Identification of the prokaryotic ligand-gated ion channels and their implications for the mechanisms and origins of animal Cys-loop ion channels. *Genome Biol.* **6**, R4
139. Bocquet, N., Prado de Carvalho, L., Cartaud, J., Neyton, J., Le Poupon, C., Taly,

- A., Grutter, T., Changeux, J.-P., and Corringer, P. J. (2007) A prokaryotic proton-gated ion channel from the nicotinic acetylcholine receptor family. *Nature* **445**, 116–119
140. Bocquet, N., Nury, H., Baaden, M., Le Poupon, C., Changeux, J.-P., Delarue, M., and Corringer, P. J. (2009) X-ray structure of a pentameric ligand-gated ion channel in an apparently open conformation. *Nature* **457**, 111–114
 141. Hilf, R. J., and Dutzler, R. (2008) X-ray structure of a prokaryotic pentameric ligand-gated ion channel. *Nature* **452**, 375–379
 142. Hilf, R. J., and Dutzler, R. (2009) A prokaryotic perspective on pentameric ligand-gated ion channel structure. *Curr. Opin. Struct. Biol.* **19**, 418–424
 143. Hibbs, R. E., and Gouaux, E. (2011) Principles of activation and permeation in an anion-selective Cys-loop receptor. *Nature* **474**, 54–60
 144. Taly, A., Delarue, M., Grutter, T., Nilges, M., Le Novère, N., Corringer, P. J., and Changeux, J.-P. (2005) Normal mode analysis suggests a quaternary twist model for the nicotinic receptor gating mechanism. *Biophys. J.* **88**, 3954–3965
 145. Taly, A., Corringer, P. J., Grutter, T., Prado de Carvalho, L., Karplus, M., and Changeux, J.-P. (2006) Implications of the quaternary twist allosteric model for the physiology and pathology of nicotinic acetylcholine receptors. *Proc. Natl. Acad. Sci. U.S.A.* **103**, 16965–16970
 146. Le Novère, N., Grutter, T., and Changeux, J.-P. (2002) Models of the extracellular domain of the nicotinic receptors and of agonist- and Ca^{2+} -binding sites. *Proc. Natl. Acad. Sci. U.S.A.* **99**, 3210–3215
 147. Heidmann, T., Bernhardt, J., Neumann, E., and Changeux, J.-P. (1983) Rapid kinetics of agonist binding and permeability response analyzed in parallel on acetylcholine receptor-rich membranes from *Torpedo marmorata*. *Biochemistry* **22**, 5452–5459
 148. Pfeiffer, F., Graham, D., and Betz, H. (1982) Purification by affinity chromatography of the glycine receptor of rat spinal cord. *J. Biol. Chem.* **257**, 9389–9393
 149. Sigel, E., Stephenson, F. A., Mamalaki, C., and Barnard, E. A. (1983) γ -Aminobutyric acid/benzodiazepine receptor complex of bovine cerebral cortex. *J. Biol. Chem.* **258**, 6965–6971
 150. Stephenson, F. A., and Mukhopadhyay, R. (2012) How the glycine and GABA receptors were purified. *J. Biol. Chem.* **287**, 40835–40837
 151. Levitzki, A., Atlas, D., and Steer, M. L. (1974) The binding characteristics and number of β -adrenergic receptors on the turkey erythrocyte. *Proc. Natl. Acad. Sci. U.S.A.* **71**, 2773–2776
 152. Bartfai, T., Anner, J., Schultzberg, M., and Montelius, J. (1974) Partial purification and characterization of a muscarinic acetylcholine receptor from rat cerebral cortex. *Biochem. Biophys. Res. Commun.* **59**, 725–733
 153. Cherezov, V., Rosenbaum, D. M., Hanson, M. A., Rasmussen, S. G., Thian, F. S., Kobilka, T. S., Choi, H. J., Kuhn, P., Weis, W. I., Kobilka, B. K., and Stevens, R. C. (2007) High-resolution crystal structure of an engineered human β_2 -adrenergic G protein-coupled receptor. *Science* **318**, 1258–1265
 154. Kato, H. E., Zhang, F., Yizhar, O., Ramakrishnan, C., Nishizawa, T., Hirata, K., Ito, J., Aita, Y., Tsukazaki, T., Hayashi, S., Hegemann, P., Maturana, A. D., Ishitani, R., Deisseroth, K., and Nureki, O. (2012) Crystal structure of the channelrhodopsin light-gated cation channel. *Nature* **482**, 369–374
 155. Nury, H., Poitevin, F., Van Renterghem, C., Changeux, J.-P., Corringer, P. J., Delarue, M., and Baaden, M. (2010) One-microsecond molecular dynamics simulation of channel gating in a nicotinic receptor homologue. *Proc. Natl. Acad. Sci. U.S.A.* **107**, 6275–6280
 156. Xu, Y., Colletier, J. P., Weik, M., Jiang, H., Moulton, J., Silman, I., and Sussman, J. L. (2008) Flexibility of aromatic residues in the active-site gorge of acetylcholinesterase: X-ray versus molecular dynamics. *Biophys. J.* **95**, 2500–2511
 157. Changeux, J.-P., and Edelman, S. (2011) Conformational selection or induced fit? Fifty years of debate resolved. *Fl1000 Biol. Rep.* **3**, 19
 158. Kenakin, T., and Miller, L. J. (2010) Seven transmembrane receptors as shape-shifting proteins: the impact of allosteric modulation and functional selectivity on new drug discovery. *Pharmacol. Rev.* **62**, 265–304
 159. Melancon, B. J., Hopkins, C. R., Wood, M. R., Emmitte, K. A., Niswender, C. M., Christopoulos, A., Conn, P. J., and Lindsley, C. W. (2012) Allosteric modulation of seven transmembrane spanning receptors: theory, practice, and opportunities for central nervous system drug discovery. *J. Med. Chem.* **55**, 1445–1464
 160. Weber, M., and Changeux, J.-P. (1974) Binding of *Naja nigricollis* [^3H] α -toxin to membrane fragments from *Electrophorus* and *Torpedo* electric organs. III. Effects of local anaesthetics on the binding of the tritiated α -neurotoxin. *Mol. Pharmacol.* **10**, 35–40
 161. Cohen, J. B., Weber, M., and Changeux, J.-P. (1974) Effects of local anesthetics and calcium on the interaction of cholinergic ligands with the nicotinic receptor protein from *Torpedo marmorata*. *Mol. Pharmacol.* **10**, 904–932
 162. Grünhagen, H. H., and Changeux, J.-P. (1976) Studies on the electrogenic action of acetylcholine with *Torpedo marmorata* electric organ. Quinacrine: a fluorescent probe for the conformational transitions of the cholinergic receptor protein in its membrane bound state. *J. Mol. Biol.* **106**, 497–516
 163. Mulle, C., Léna, C., and Changeux, J.-P. (1992) Potentiation of nicotinic receptor response by external calcium in rat central neurons. *Neuron* **8**, 937–945
 164. Vernino, S., Amador, M., Luetje, C. W., Patrick, J., and Dani, J. A. (1992) Calcium modulation and high calcium permeability of neuronal nicotinic acetylcholine receptors. *Neuron* **8**, 127–134
 165. Galzi, J.-L., Bertrand, S., Corringer, P. J., Changeux, J.-P., and Bertrand, D. (1996) Identification of calcium binding sites that regulate potentiation of a neuronal nicotinic acetylcholine receptor. *EMBO J.* **15**, 5824–5832
 166. Krause, R. M., Buisson, B., Bertrand, S., Corringer, P. J., Galzi, J.-L., Changeux, J.-P., and Bertrand, D. (1998) Ivermectin: a positive allosteric effector of the $\alpha 7$ neuronal nicotinic acetylcholine receptor. *Mol. Pharmacol.* **53**, 283–294
 167. Bertrand, D., and Gopalakrishnan, M. (2007) Allosteric modulation of nicotinic acetylcholine receptors. *Biochem. Pharmacol.* **74**, 1155–1163
 168. Gill, J. K., Chatzidaki, A., Ursu, D., Sher, E., and Millar, N. S. (2013) Contrasting properties of $\alpha 7$ -selective orthosteric and allosteric agonists examined on native nicotinic acetylcholine receptors. *PLoS ONE* **8**, e55047
 169. Nury, H., Van Renterghem, C., Weng, Y., Tran, A., Baaden, M., Dufresne, V., Changeux, J.-P., Sonner, J. M., Delarue, M., and Corringer, P. J. (2011) X-ray structures of general anaesthetics bound to a pentameric ligand-gated ion channel. *Nature* **469**, 428–431
 170. Murail, S., Wallner, B., Trudell, J. R., Bertaccini, E., and Lindahl, E. (2011) Microsecond simulations indicate that ethanol binds between subunits and could stabilize an open-state model of a glycine receptor. *Biophys. J.* **100**, 1642–1650
 171. Sauguet, L., Howard, R. J., Malherbe, L., Lee, U. S., Corringer, P. J., Harris, R. A., and Delarue, M. (2013) Structural basis for potentiation by alcohols and anaesthetics in a ligand-gated ion channel. *Nat. Commun.* **4**, 1697
 172. Li, G.-D., Chiara, D. C., Cohen, J. B., and Olsen, R. W. (2010) Numerous classes of general anesthetics inhibit etomidate binding to γ -aminobutyric acid type A (GABA_A) receptors. *J. Biol. Chem.* **285**, 8615–8620
 173. Forman, S. A. (2012) Monod-Wyman-Changeux allosteric mechanisms of action and the pharmacology of etomidate. *Curr. Opin. Anaesthesiol.* **25**, 411–418
 174. Forman, S. A. (2013) A paradigm shift from biophysical to neurobiological: the fading influence of Claude Bernard's ideas about general anesthesia. *Anesthesiology* **118**, 984–985
 175. Hansen, S. B., and Taylor, P. (2007) Galanthamine and non-competitive inhibitor binding to ACh-binding protein: evidence for a binding site on non- α -subunit interfaces of heteromeric neuronal nicotinic receptors. *J. Mol. Biol.* **369**, 895–901
 176. Nemečz, Á., and Taylor, P. (2011) Creating an $\alpha 7$ nicotinic acetylcholine recognition domain from the acetylcholine-binding protein. Crystallographic and ligand selectivity analyses. *J. Biol. Chem.* **286**, 42555–42565
 177. Hamouda, A. K., Kimm, T., and Cohen, J. B. (2013) Physostigmine and galanthamine bind in the presence of agonist at the canonical and noncanonical subunit interfaces of a nicotinic acetylcholine receptor. *J. Neurosci.* **33**, 485–494
 178. Galzi, J.-L., and Changeux, J.-P. (1994) Neurotransmitter-gated ion channels as unconventional allosteric proteins. *Curr. Opin. Cell Biol.* **4**, 554–565
 179. Triller, A., and Choquet, D. (2008) New concepts in synaptic biology derived from single-molecule imaging. *Neuron* **59**, 359–374
 180. Zhang, H., Etherington, L. A., Hafner, A. S., Bellelli, D., Coussen, F., Delagrèze, P., Chaouloff, F., Spedding, M., Lambert, J. J., Choquet, D., and Groc, L. (2013) Regulation of AMPA receptor surface trafficking and synaptic plasticity by a cognitive enhancer and antidepressant molecule. *Mol. Psychiatry* **18**, 471–484
 181. Changeux, J.-P. (1972) The brain and the event. in *Communications*, pp. 18–37, Ecole des Hautes Etudes en Sciences Sociales, Paris
 182. Changeux, J.-P., Courrège, P., and Danchin, A. (1973) A theory of the epigenesis of neuronal networks by selective stabilization of synapses. *Proc. Natl. Acad. Sci. U.S.A.* **70**, 2974–2978
 183. Changeux, J.-P., and Danchin, A. (1976) Selective stabilisation of developing synapses as a mechanism for the specification of neuronal networks. *Nature* **264**, 705–712
 184. Redfern, P. A. (1970) Neuromuscular transmission in new-born rats. *J. Physiol.* **209**, 701–709
 185. Luo, L., and O'Leary, D. D. (2005) Axon retraction and degeneration in development and disease. *Annu. Rev. Neurosci.* **28**, 127–156

186. Wu X., Fu, Y., Knott, G., Lu, J., Di Cristo, G., and Huang, Z. J. (2012) GABA signaling promotes synapse elimination and axon pruning in developing cortical inhibitory interneurons. *J. Neurosci.* **32**, 331–343
187. Changeux, J.-P. (2013) The concept of allosteric modulation: an overview. *Drug Discov. Today Technol.* **10**, e223–e228
188. Edelman, G. M. (1987) *Neural Darwinism: The Theory of Neural Group Selection*, Basic Books, New York
189. Quartz, S. R., and Sejnowski, T. J. (1997) The neural basis of cognitive development: a constructivist manifesto. *Behav. Brain Sci.* **20**, 537–556
190. Duclert, A., and Changeux, J.-P. (1995) Acetylcholine receptor gene expression at the developing neuromuscular junction. *Physiol. Rev.* **75**, 339–368
191. Sanes, J. R., and Lichtman, J. W. (2001) Induction, assembly, maturation and maintenance of a postsynaptic apparatus. *Nat. Rev. Neurosci.* **2**, 791–805
192. Schaeffer, L., Duclert, N., Huchet-Dymanus, M., and Changeux, J.-P. (1998) Implication of a multisubunit Ets-related transcription factor in synaptic expression of the nicotinic acetylcholine receptor. *EMBO J.* **17**, 3078–3090
193. Rousset, A., Cartaud, J., Devaux, P. F., and Changeux, J.-P. (1982) The rotational diffusion of the acetylcholine receptor in *Torpedo marmorata* membrane fragments studied with a spin-labelled α -toxin: importance of the 43,000 protein(s). *EMBO J.* **1**, 439–445
194. Cartaud, J., and Changeux, J.-P. (1993) Post-transcriptional compartmentalization of acetylcholine receptor biosynthesis in the subneural domain of muscle and electrocyte junctions. *Eur. J. Neurosci.* **5**, 191–202
195. Marchand, S., and Cartaud, J. (2002) Targeted trafficking of neurotransmitter receptors to synaptic sites. *Mol. Neurobiol.* **26**, 117–135
196. Changeux, J.-P. (1983) *Neuronal Man: The Biology of Mind*, Fayard, Paris
197. Heidmann, T., and Changeux, J.-P. (1982) Molecular model of the regulation of chemical synapse efficiency at the postsynaptic level. *C. R. Seances Acad. Sci. III* **295**, 665–670
198. Hawkins, R. D., Abrams, T. W., Carew, T. J., and Kandel, E. R. (1983) A cellular mechanism of classical conditioning in *Aplysia*: activity-dependent amplification of presynaptic facilitation. *Science* **219**, 400–405
199. Goelet, P., Castellucci, V. F., Schacher, S., and Kandel, E. R. (1986) The long and the short of long-term memory: a molecular framework. *Nature* **322**, 419–422
200. Dehaene, S., Changeux, J.-P., and Nadal, J. P. (1987) Neural networks that learn temporal sequences by selection. *Proc. Natl. Acad. Sci. U.S.A.* **84**, 2727–2731
201. Dehaene, S., and Changeux, J.-P. (1989) A simple model of prefrontal cortex. Function in delayed-response tasks. *J. Cogn. Neurosci.* **1**, 244–261
202. Dehaene, S., and Changeux, J.-P. (1991) The Wisconsin Card Sorting Test: theoretical analysis and modelling in a neuronal network. *Cereb. Cortex* **1**, 62–79
203. Fuster, J. M. (2008) *The Prefrontal Cortex*, 4th Ed, Academic Press, London
204. Dehaene, S., and Changeux, J.-P. (1997) A hierarchical neuronal network for planning behaviour. *Proc. Natl. Acad. Sci. U.S.A.* **94**, 13293–13298
205. Dehaene, S., and Changeux, J.-P. (2000) Reward-dependent learning in neuronal networks for planning and decision making. *Prog. Brain Res.* **126**, 217–229
206. Dehaene, S., Kerszberg, M., and Changeux, J.-P. (1998) A neuronal model of a global workspace in effortful cognitive tasks. *Proc. Natl. Acad. Sci. U.S.A.* **95**, 14529–14534
207. Montague, P. R., Dayan, P., and Sejnowski, T. J. (1996) A framework for mesencephalic dopamine systems based on predictive Hebbian learning. *J. Neurosci.* **16**, 1936–1947
208. Schultz, W., Dayan, P., and Montague, P. R. (1997) A neural substrate of prediction and reward. *Science* **275**, 1593–1599
209. Schultz, W. (2012) Risky dopamine. *Biol. Psychiatry* **71**, 180–181
210. Picciotto, M. R., Zoli, M., Léna, C., Bessis, A., Lallemand, Y., Le Novère, N., Vincent, P., Pich, E. M., Brûlet, P., and Changeux, J.-P. (1995) Abnormal avoidance learning in mice lacking functional high-affinity nicotine receptor in the brain. *Nature* **374**, 65–67
211. Picciotto, M. R., Zoli, M., Rimondini, R., Léna, C., Marubio, L. M., Pich, E. M., Fuxe, K., and Changeux, J.-P. (1998) Acetylcholine receptors containing the $\beta 2$ subunit are involved in the reinforcing properties of nicotine. *Nature* **391**, 173–177
212. Maskos, U., Molles, B. E., Pons, S., Besson, M., Guiard, B. P., Guilloux, J. P., Evrard, A., Cazala, P., Cormier, A., Mameli-Engvall, M., Dufour, N., Clözé-Tayarani, I., Bemelmans, A. P., Mallet, J., Gardier, A. M., David, V., Faure, P., Granon, S., and Changeux, J.-P. (2005) Nicotine reinforcement and cognition restored by targeted expression of nicotinic receptors. *Nature* **436**, 103–107
213. Avale, M. E., Faure, P., Pons, S., Robledo, P., Deltheil, T., David, D. J., Gardier, A. M., Maldonado, R., Granon, S., Changeux, J.-P., and Maskos, U. (2008) Interplay of $\beta 2^*$ nicotinic receptors and dopamine pathways in the control of spontaneous locomotion. *Proc. Natl. Acad. Sci. U.S.A.* **105**, 15991–15996
214. Mameli-Engvall, M., Evrard, A., Pons, S., Maskos, U., Svensson, T. H., Changeux, J.-P., and Faure P. (2006) Hierarchical control of dopamine neuron-firing patterns by nicotinic receptors. *Neuron* **50**, 911–921
215. Tolu, S., Eddine, R., Marti, F., David, V., Graupner, M., Pons, S., Baudonnat, M., Husson, M., Besson, M., Reperant, C., Zemdeg, J., Pagès, C., Hay, Y. A., Lambolze, B., Caboche, J., Gutkin, B., Gardier, A. M., Changeux, J.-P., Faure, P., and Maskos, U. (2013) Co-activation of VTA DA and GABA neurons mediates nicotine reinforcement. *Mol. Psychiatry* **18**, 382–393
216. Granon, S., Faure, P., and Changeux, J.-P. (2003) Executive and social behaviors under nicotinic receptor regulation. *Proc. Natl. Acad. Sci. U.S.A.* **100**, 9596–9601
217. Maubourguet, N., Lesne, A., Changeux, J.-P., Maskos, U., and Faure P. (2008) Behavioral sequence analysis reveals a novel role for $\beta 2^*$ nicotinic receptors in exploration. *PLoS Comput. Biol.* **4**, e1000229
218. Changeux, J.-P. (2010) Nicotine addiction and nicotinic receptors: lessons from genetically modified mice. *Nat. Rev. Neurosci.* **11**, 389–401
219. Jackson, K. J., and Imad Damaj, M. (2013) $\beta 2$ -containing nicotinic acetylcholine receptors mediate calcium/calmodulin-dependent protein kinase-II and synapsin I protein levels in the nucleus accumbens after nicotine withdrawal in mice. *Eur. J. Pharmacol.* **701**, 1–6
220. Picciotto, M. R., and Kenny, P. J. (2013) Molecular mechanisms underlying behaviors related to nicotine addiction. *Cold Spring Harb. Perspect. Med.* **3**, a012112
221. Sallette, J., Bohler, S., Benoit, P., Soudant, M., Pons, S., Le Novère, N., Changeux, J.-P., and Corringer, P. J. (2004) An extracellular microdomain controls up-regulation of neuronal nicotinic acetylcholine receptors by nicotine. *J. Biol. Chem.* **279**, 18767–18775
222. Sallette, J., Pons, S., Devillers-Thiery, A., Soudant, M., Prado de Carvalho, L., Changeux, J.-P., and Corringer, P. J. (2005) Nicotine upregulates its own receptors through enhanced intracellular maturation. *Neuron* **46**, 595–607
223. Govind, A. P., Walsh, H., and Green, W. N. (2012) Nicotine-induced upregulation of native neuronal nicotinic receptors is caused by multiple mechanisms. *J. Neurosci.* **32**, 2227–2238
224. Baker, L. K., Mao, D., Chi, H., Govind, A. P., Vallejo, Y. F., Iacoviello, M., Herrera, S., Cortright, J. J., Green, W. N., McGehee, D. S., and Vezina, P. (2013) Intermittent nicotine exposure upregulates nAChRs in VTA dopamine neurons and sensitizes locomotor responding to the drug. *Eur. J. Neurosci.* **37**, 1004–1011
225. Liu, L., Hendrickson, L. M., Guildford, M. J., Zhao-Shea, R., Gardner, P. D., and Tapper, A. R. (2013) Nicotinic acetylcholine receptors containing the $\alpha 4$ subunit modulate alcohol reward. *Biol. Psychiatry* **73**, 738–746
226. Bechara, A., Tranel, D., and Damasio, H. (2000) Characterization of the decision-making deficit of patients with ventromedial prefrontal cortex lesions. *Brain* **123**, 2189–2202
227. Changeux, J.-P., and Lou, H. C. (2011) Emergent pharmacology of conscious experience: new perspectives in substance addiction. *FASEB J.* **25**, 2098–2108
228. Naqvi, N. H., Rudrauf, D., Damasio, H., and Bechara, A. (2007) Damage to the insula disrupts addiction to cigarette smoking. *Science* **315**, 531–534
229. Jacobsen, L. K., Picciotto, M. R., Heath, C. J., Frost, S. J., Tsou, K. A., Dwan, R. A., Jackowski, M. P., Constable, R. T., and Mendel, W. E. (2007) Prenatal and adolescent exposure to tobacco smoke modulates the development of white matter microstructure. *J. Neurosci.* **27**, 13491–13498
230. Léna, C., Changeux, J.-P., and Mulle, C. (1993) Evidence for “preterminal” nicotinic receptors on GABAergic axons in the rat interpeduncular nucleus. *J. Neurosci.* **13**, 2680–2688
231. Kawai, H., Lazar, R., and Metherate, R. (2007) Nicotinic control of axon excitability regulates thalamocortical transmission. *Nat. Neurosci.* **10**, 1168–1175
232. Dehaene, S., and Changeux, J.-P. (2011) Experimental and theoretical approaches to conscious processing. *Neuron* **70**, 200–227
233. Avale, M. E., Chabout, J., Pons, S., Serreau, P., De Chaumont, F., Olivo-Marin, J. C., Bourgeois, J. P., Maskos, U., Changeux, J.-P., and Granon, S. (2011) Prefrontal nicotinic receptors control novel social interaction between mice. *FASEB J.* **25**, 2145–2155
234. Rømer Thomsen, K., Joensson, M., Lou, H. C., Møller, A., Gross, J., Kringelbach, M. L., and Changeux, J.-P. (2013) Altered paralimbic interaction in behavioral addiction. *Proc. Natl. Acad. Sci. U.S.A.* **110**, 4744–4749
235. Koch, C. (2004) Thinking about the conscious mind. *Science* **306**, 979–980
236. Franks, N. P. (2008) General anaesthesia: from molecular targets to neuronal pathways of sleep and arousal. *Nat. Rev. Neurosci.* **9**, 370–386
237. Changeux, J.-P. (2012) Conscious processing: implications for general anes-

- thesia. *Curr. Opin. Anaesthesiol.* **25**, 397–404
238. Breitmeyer, B. (2006) *Visual Masking: Time Slices through Conscious and Unconscious Vision*, Oxford University Press, New York
239. Dehaene, S., Sergent, C., and Changeux, J.-P. (2003) A neuronal network model linking subjective reports and objective physiological data during conscious perception. *Proc. Natl. Acad. Sci. U.S.A.* **100**, 8520–8525
240. Lamme, V. A., and Roelfsema, P. R. (2000) The distinct modes of vision offered by feedforward and recurrent processing. *Trends Neurosci.* **23**, 571–579
241. Ku, S. W., Lee, U., Noh, G. J., Jun, I. G., and Mashour, G. A. (2011) Preferential inhibition of frontal-to-parietal feedback connectivity is a neurophysiologic correlate of general anesthesia in surgical patients. *PLoS ONE* **6**, e25155
242. van Loon, A. M., Scholte, H. S., van Gaal, S., van der Hoort, B. J., and Lamme, V. A. (2012) GABA_A agonist reduces visual awareness: a masking-EEG experiment. *J. Cogn. Neurosci.* **24**, 965–974
243. Hökfelt, T., Rehfeld, J. F., Skirboll, L., Ivemark, B., Goldstein, M., and Markey, K. (1980) Evidence for coexistence of dopamine and CCK in meso-limbic neurones. *Nature* **285**, 476–478
244. Agnati, L. F., Benfenati, F., Solfrini, V., Biagini, G., Fuxe, K., Guidolin, D., Carani, C., and Zini, I. (1992) Brain aging and neuronal plasticity. *Ann. N.Y. Acad. Sci.* **673**, 180–186
245. Amunts, K., Lenzen, M., Friederici, A. D., Schleicher, A., Morosan, P., Palomero-Gallagher, N., and Zilles, K. (2010) Broca's region: novel organizational principles and multiple receptor mapping. *PLoS Biol.* **8**, e1000489
246. Bartfai, T., Buckley, P. T., and Eberwine, J. (2012) Drug targets: single-cell transcriptomics hastens unbiased discovery. *Trends Pharmacol. Sci.* **33**, 9–16
247. Pastrana, E. (2010) Optogenetics: controlling cell function with light. *Nat. Methods* **8**, 24–25
248. Changeux, J.-P., and Edelman, S. J. (1998) Allosteric receptors after 30 years. *Neuron* **21**, 959–980
249. Tsigelny, I. F., Kouznetsova, V. L., Baitaluk, M., and Changeux, J.-P. (2013) A hierarchical coherent-gene-group model for brain development. *Genes Brain Behav.* **12**, 147–165
250. Changeux, J.-P. (2012) *The Good, the True and the Beautiful: A Neuronal Approach*, Odile Jacob/Yale University Press, New Haven, CT
251. Changeux, J.-P. (1964) On the allosteric properties of biosynthetic L-threonine deaminase. II. Kinetics of action of biosynthetic L-threonine deaminase with respect to the natural substrate and inhibitor. *Bull. Soc. Chim. Biol.* **46**, 947–961
252. Changeux, J.-P. (1964) On the allosteric properties of biosynthetic L-threonine deaminase. III. Interpretation of the inhibitory effect of L-isoleucine: steric hindrance or allosteric effect. *Bull. Soc. Chim. Biol.* **46**, 1151–1173
253. Changeux, J.-P. (1965) On the allosteric properties of biosynthetic L-threonine deaminase. IV. The desensitization phenomenon. *Bull. Soc. Chim. Biol.* **47**, 113–139
254. Changeux, J.-P. (1965) On the allosteric properties of biosynthetic L-threonine deaminase. V. The allosteric transition. *Bull. Soc. Chim. Biol.* **47**, 267–280
255. Grande, F., and Visscher, M. B. (eds) (1967) Introduction. in *Claude Bernard and Experimental Medicine*, Schenkman Publishing Co., Cambridge, MA