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No Evidence for Association of the *MDM2* –309 T/G Promotor Polymorphism with Prostate Cancer Outcomes

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Abstract

Objectives—MDM2 SNP309 polymorphism (T>G) has been correlated with an increased risk of cancer in multiple tumor types. MDM2 overexpression in has shown to be weakly associated with distant tumor metastases, and downregulation of MDM2 via antisense oligonucleotides *in vitro* has resulted in the radiosensitization of prostate cancer cell lines. Based on these results, we decided to evaluate the role of MDM2 SNP309 in the context of histopathologic parameters and clinical outcomes in prostate cancer tumors.

Materials and Methods—The population consisted of 212 consecutive prostate cancer patients who underwent radical prostatectomy between 1997 and 1999 at Vanderbilt University Medical Center. Two hundred eight of the samples were successfully genotyped for the MDM2 SNP309 polymorphism. Correlations between the polymorphism, recurrence, and survival data were analyzed using univariate and multivariate genetic models.

Results—The only prognostic factor predictive of overall survival in our study was Gleason score (P < 0.005). Using chi square analysis, we determined that the MDM2 SNP309 polymorphism had no significant association with race (P = 0.7512), patient's age at diagnosis (P = 0.6820), pre-prostatectomy PSA level (P = 0.8606), Gleason's score (P = 0.4839), surgical margin status (P = 1.0000), extracapsular extension (P = 0.6175) and disease stage (P = 0.4945). In addition, there was no significant difference in 3 year recurrence-free survival (P = 0.218) or 8 year overall survival (P = 0.376).

Conclusions—Our study finds no evidence for association of the MDM2 SNP309 polymorphism with clinicopathologic variables, recurrence risk, and overall survival outcome in prostate cancer.

Keywords

promotor; polymorphism; mdm2; prostate cancer; prognosis

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Introduction

Mouse double-minute 2 (MDM2) is an E3-ubiquitin ligase that binds, inhibits and promotes the degradation of the tumor suppressor protein, p53⁻¹. A strong correlation between overexpression of MDM2, tumor proliferation, and an early onset of tumorigenesis exists ^{2–6}. In fact, the MDM2 gene is amplified in approximately 30% of the osteosarcomas and soft tissue sarcomas ^{7–9}.

*Bond et al*¹⁰ demonstrated that a single nucleotide polymorphism in the promoter region of the MDM2 gene (-309 T/G; SNP309) could result in elevated MDM2 levels with increased p53 degradation. Interestingly, Bond and co-workers found that the effects of this polymorphism on enhanced tumorigenesis occurs in a gender-specific and hormone-dependent manner ¹¹. Based on these findings, we performed genotypic analysis of MDM2 SNP309 in tumor tissues of 212 patients with prostate cancer status post prostatectomy and evaluated the association of this polymorphism with clinicopathologic and prognostic parameters. To our knowledge, the current investigation represents the largest report evaluating the clinical effects of MDM2 SNP 309 in prostate cancer outcome.

Materials and Methods

Study Population

The population consisted of 212 consecutive prostate cancer patients who underwent radical prostatectomy between 1997 and 1999 at Vanderbilt University Medical Center. Ninety-eight percent of the patients were Caucasian with the remainder being 4 African-American patients. The median follow-up for overall survival was 9.1 years (mean 8.3 ± 2.4 years), and for assessment of prostate cancer recurrence was 3.4 years (mean 4.4 ± 3.9 years). Recurrence post-prostatectomy was classified as biochemical, local or distant. All patients had histologically confirmed adenocarcinoma. Two hundred eight of the samples were successfully genotyped for the MDM2 SNP309 polymorphism. This study was approved by the Vanderbilt University Institutional Review Board (IRB No. 030986).

Specimen preparation and deoxyribonucleic acid extraction

Utilizing a standard microtome with disposable blades, we cut 5-µm thick sections of representative areas of normal prostate glands from paraffin-embedded blocks and stained these tissues with hematoxylin and eosin. We subsequently examined the slides under a microscope to verify the absence of prostate cancer. A 5-µm thick section (approximately 1 µg) from each patient was used for DNA extraction. The sections were deparaffinized twice with xylene at room temperature for 30 minutes, washed twice with 100% ethanol, and after complete ethanol evaporation, the tissue was completely lysed with proteinase K. The QIAamp DNA Mini Kit (QIAGEN Inc., Valencia, CA) was used to extract and purify DNA from the tissues according to the manufacturer's protocol.

Genotyping

Genomic DNA samples were obtained from patient tumor specimens and processed as described previously ^{12,13}. MDM2 SNP309 (rs2279744) genotyping was performed using PyrosequencingTM technology (Biotage AB, Uppsala, Sweden).¹⁴ The primers were: 5 - GGGGTGGTTCGGAG GTCT-3 (sense); and 5 -Biotin -GTGACCCGACAG GCACCT-3 (antisense). A 113 bp fragment was amplified from 10 ng genomic DNA. Single stranded DNA was isolated from the PCR reactions using the Pyrosequencing Vacuum Prep Workstation and transferred into a 96-well plate. A sequencing primer, 5 GGGCTGCGGGGCCGCT, was annealed to the single stranded DNA. The plate was then transferred to the Pyrosequencing PSQ96MA by dispensing the nucleotides in the following

order [G/T]CGGCGCGGGAGGTCCGGATGATCGC. The genotype was determined using SNP Software (Biotage AB).

Statistical Analysis

Testing of adherence to Hardy-Weinberg equilibrium was performed using a Chi-square goodness of fit test. Overall survival was determined as the time between the date of surgery and the date of death or last follow-up. Recurrence-free survival was calculated from the date of surgery to the date of recurrence (biochemical, local or distant) or last follow-up, as described previously ^{12,13}. The data were censored for live (or recurrence-free) patients as of their last follow-up visits. Biochemical recurrence was defined as a prostate-specific antigen (PSA) detection of > 0.1 ng/ml in at least two consecutive lab draws. Kaplan-Meier survival curves were calculated for each genotype and were compared using the log rank test. Association of the genotypes to various clinicopathologic characteristics was assessed using the Fisher's exact test. Multivariate analysis was performed using the Cox proportional hazards model. MDM2SNP 309 had no evidence of association with our endpoints, but we used the model to validate known variables predictive of our studied outcomes.

Results

The MDM2 SNP309 genotype frequencies among the prostate cancer cohort were in Hardy-Weinberg equilibrium (P= 0.705). Two hundred and six of the 212 samples gave analyzable results. The frequencies of the SNP309 genotypes were: 16.5% (35 of 212) for G/G; 48.6% (103 of 212) for G/T; and 32.5% (69/212) for T/T. Log-rank univariate analyses of known prognostic factors, including pre-prostatectomy PSA levels (P< 0.045), Gleason score (P< 0.001), surgical margin status (P< 0.001), extracapsular extension status (P< 0.001) and T stage (P< 0.001) were predictive of relapse-free survival.

The only one of these prognostic factors predictive of overall survival in our study was Gleason score (P < 0.005). Using chi square analysis, we determined that the MDM2 SNP309 polymorphism had no significant association with race (P = 0.7512), patient's age at diagnosis (P = 0.6820), pre-prostatectomy PSA level (P = 0.8606), Gleason's score (P = 0.4839), surgical margin status (P = 1.0000), extracapsular extension (P = 0.6175) and disease stage (P = 0.4945) (Results in Table 1). We utilized the Kaplan-Meier method to determine the recurrence and survival rates, and the log-rank test was used to test the difference in these rates between the different genotypes. There was no significant difference in 3 year recurrence-free survival (Fig. 1A, P = 0.218) or 8 year overall survival (Fig. 1B, P = 0.376).

To determine independent risk factors for recurrence, multivariate analyses using the Cox proportional hazards regression model were performed. In this model the recessive genotypes of SNP309 were not found to be independent predictors of recurrence (G/T: hazard ratio of 3.480 (1.031 – 11.746, 95% CI), P= 0.045; T/T: hazard ratio of 2.159 (0.581 – 8.030, 95% CI), P=0.251) or overall survival (G/T: hazard ratio of 2.478 (0.736 – 8.347, 95% CI), P= 0.143; T/T: hazard ratio of 1.564 (0.400 – 6.116, 95% CI), P= 0.521).

Discussion

MDM2 SNP309 polymorphism (T>G) is associated with an increased risk of cancer in multiple tumor types including renal cancer ¹⁵, non-small cell lung cancer ¹⁶, oral squamous cancer¹⁷, and gastric cancer ¹⁸. In B cell lymphoma a significant association between MDM2 SNP309 polymorphism and relapse-free and overall survival rates was observed ¹⁹. MDM2 overexpression in prostate cancers has also been shown to be weakly associated with distant metastases in a small trial ³. Furthermore downregulation of MDM2 via antisense

oligonucleotides *in vitro* has resulted in the radiosensitization of prostate cancer cell lines ²⁰. Based on these results we evaluated the role of MDM2 SNP309 in the context of histopathologic parameters and clinical outcomes in prostate cancer tumors and found no statistically significant association.

Recently there has been a report evaluating the risk of MDM2 SNP309 in prostate cancer in a case control design ²¹. The study included 145 affected men who underwent radical prostatectomy, as well as 124 patient controls. They concluded that there was no correlation between the MDM2 SNP 309 polymorphism and an increased risk of cancer. The authors also analyzed histological, pathological and recurrence parameters from which no significant association was found. The major weakness of their study was that only 65 of the 145 affected prostate patients were evaluable for prognostic parameters, and that over 10% of the pathologic data was missing. In contrast, the present study consists of a larger sample size, a greater median follow-up, and a more complete database for all patient endpoints than the only other published study of its kind.

Our study similarly revealed no association between the MDM2 SNP309 polymorphism and clinicopathologic outcomes in prostate cancer. Of note several limitations from this present study may obscure the understanding of MDM2 polymorphism in association with tumorigenesis. First the population studied was primarily non-Jewish Caucasian (98%), thus limiting the extrapolation to other races. However, the frequency of G allele of MDM2 SNP309 is much lower in African-Americans and intermediate in Caucasians²². Notably our study patients had clinical low risk features, but more intermediate pathologic features. Although 53% of these patients had PSA values in the normal range (<4 ng/ml), ~27% of our patients had pathologic T3 disease. This degree of clinical understaging was in line with historical rates at VUMC (30% for open retropubic radical prostactectomies)²³.

Unlike this study, previous studies have examined MDM2 SNP309 polymorphism in the context of p53 status. MDM2 SNP309 polymorphism has been shown to enhance tumorigenesis when co-existent with mutated p53, as in patients with Li-Fraumeni Syndrome ¹⁰. Not only does the T > G allelic change in MDM2 SNP309 lead to increased occurrence of tumors in a lifetime for these patients, but also leads to an earlier onset of cancer ²⁴. As such several studies have pooled genotypes according to p53 mutation status, and have either shown an increased risk of tumorigenesis or poor prognosis with MDM2 SNP309 polymorphism ^{13,18}. Although the present data did not take into consideration p53 mutation status, it is worth noting that p53 alterations represent a rare event in prostate cancer tumorigenesis in which p53 mutation occurs with only the most aggressive, metastatic prostate cancer transitioning from androgen-dependent to androgen-independent growth ²⁵. Likewise, p53 alterations may be of less relevance in prostate carcinogenesis than in other cancers.

It has been recently suggested that MDM2 SNP309 polymorphism and subsequent changes in the affinity of the promoter to hormones such as estrogen could affect breast tumorigenesis ^{11,26,27}, although other data have noted no such association ^{28,29}. One study showed that the G-allele of MDM2 SNP309 accelerates colorectal formation in women and not in men and hypothesized the effects of female-specific hormones such as estrogen on the observed finding ¹¹. In the same way, prostate tumors with MDM2 overexpression may be of value in tumorigenesis only when the tumor retains androgen-sensitivity. We have not attempted to evaluate this association in our study, as we have not seen a significant correlation of MDM2 SNP309 with any of our endpoints.

In conclusion there is no evidence in our study that MDM2 SNP309 polymorphism is significantly associated with clinicopathologic variables, recurrence risk, and overall

survival endpoints in prostate cancer. It is possible that the study is underpowered, and additional patient numbers could detect a difference in our evaluated endpoints. Additionally it is possible that a statistical difference in local recurrence and/or overall survival rates may have been detectable with longer follow-up. However consistent with other recently reported studies MDM2 SNP309, we have found no association of this biomarker with prostate cancer risk or prognosis.

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Figure 1A



MDM2_309

years after Surgery

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Figure 1B



Fig. 1.

Kaplan-Meier estimates of recurrence-free survival (RFS) and overall survival (OS) following radical prostatectomy as treatment for localized prostate cancer. (A) RFS curves were plotted for the MDM2 SNP309 genotypes. (B) OS curves were plotted for the MDM2 SNP309 genotypes.

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FACTORS	T/T (N=68)	T/G (N=103)	G/G (N=35)	Missing (N=6)	TOTAL (N=212)	P Value
Age at diagnosis (yrs), n(%)						
65	47 (69.12)	73 (70.87)	22 (62.86)	6 (100.00)	148 (69.81)	0.6820
> 65	21 (30.88)	30 (29.13)	13 (37.14)	0 (00.00)	64 (30.19)	
Race, n(%)						
Black	1 (1.47)	1 (0.97)	1 (2.86)	1 (16.67)	4 (1.89)	0.7512
White	67 (98.53)	102 (99.03)	34 (97.14)	5 (83.33)	208 (98.11)	
PSA at diagnosis (ng/ml), n(%)						
4	8 (11.76)	10 (9.71)	4 (11.43)	0 (00.00)	22 (10.38)	
4-10	30 (44.12)	60 (58.25)	21 (60.00)	2 (33.33)	113 (53.30)	0.8606
> 10	14 (20.59)	20 (19.42)	7 (20.00)	2 (33.33)	43 (20.28)	
Missing	16 (23.53)	13 (12.62)	3 (8.57)	2 (33.33)	34 (16.04)	
Gleason score, n(%)						
6	44 (64.71)	60 (58.25)	21 (60.00)	5 (83.33)	130 (61.32)	0.4020
7	22 (32.35)	37 (35.92)	10 (28.57)	1 (16.67)	70 (33.02)	0.400
>7	2 (2.94)	6 (5.83)	4 (11.43)	0 (00.00)	12 (5.66)	
Pathologic Stage [*] , n(%)						0.4945
Not palpable (T1)	5 (7.35)	3 (2.91)	0 (0.00)	0 (0.00)	8 (3.77)	
Confined to prostate (T2)	42 (61.76)	64 (62.14)	23 (65.71)	5 (83.33)	134 (63.21)	
Locally advanced (T3)	17 (25.00)	28 (27.18)	11 (31.43)	1 (16.67)	57 (26.89)	
Missing Data	4 (5.88)	8 (7.77)	1 (2.86)	0 (0.00)	13 (6.13)	
Extracapsular extension, n(%)						
Negative	38 (55.88)	64 (62.14)	19 (54.29)	4 (66.67)	125 (58.96)	
Positive	20 (29.41)	32 (31.07)	14 (40.00)	1 (16.67)	67 (31.60)	
Missing	10 (14.71)	7 (6.80)	2 (5.71)	1 (16.67)	20 (9.43)	0.6175
Surgical Margin, n(%)						
Negative	44 (64.71)	65 (63.11)	44 (64.71)	2 (33.33)	133 (62.74)	1.0000
Positive	24 (35.29)	37 (35.92)	24 (35.29)	4 (66.67)	78 (36.79)	

 $\overset{*}{}_{\rm Based}$ on the 2006 American Joint Committee on Cancer TNM Staging

Abbreviations: PSA = prostate-specific antigen.