



CVID-associated *TACI* mutations affect autoreactive B cell selection and activation

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Common variable immune deficiency (CVID) is an assorted group of primary diseases that clinically manifest with antibody deficiency, infection susceptibility, and autoimmunity. Heterozygous mutations in the gene encoding the tumor necrosis factor receptor superfamily member TACI are associated with CVID and autoimmune manifestations, whereas two mutated alleles prevent autoimmunity. To assess how the number of TACI mutations affects B cell activation and tolerance checkpoints, we analyzed healthy individuals and CVID patients carrying one or two TACI mutations. We found that TACI interacts with the cleaved, mature forms of TLR7 and TLR9 and plays an important role during B cell activation and the central removal of autoreactive B cells in healthy donors and CVID patients. However, only subjects with a single TACI mutation displayed a breached immune tolerance and secreted antinuclear antibodies (ANAs). These antibodies were associated with the presence of circulating B cell lymphoma 6-expressing T follicular helper (Tfh) cells, likely stimulating autoreactive B cells. Thus, TACI mutations may favor CVID by altering B cell activation with coincident impairment of central B cell tolerance, whereas residual B cell responsiveness in patients with one, but not two, TACI mutations enables autoimmune complications.

Introduction

Common variable immune deficiency (CVID) is a heterogeneous group of primary diseases that manifest with antibody deficiency, susceptibility to infections, and autoimmunity (1). Since its identification as a clinical entity, CVID has been principally described as a B cell disorder (2). B cell receptor (BCR) and TLR7 and TLR9 responses are found altered in most CVID patients (3–5). However, CVID patients often have additional immunological abnormalities not apparently attributable to B cells, such as Treg deficiency, a finding associated with clinical autoimmunity (6, 7). Although the majority of CVID cases have no identifiable pathogenic mutation, 7%–10% of patients bear a mutation in the *TNFRSF13B* gene encoding TACI, a tumor necrosis factor receptor superfamily member expressed on B cells (8, 9). TACI can bind two ligands, a proliferation-inducing ligand (APRIL) and B cell activation factor (BAFF), both of which were found elevated in the serum of CVID patients (10–12). Interestingly, elevated serum BAFF concentrations in mice have been reported to interfere with the removal of autoreactive B cells (13, 14). *TACI* mutations in CVID patients are typically found in the heterozygous state, suggesting either that *TACI* mutations exert a dominant-negative effect on the unmutated allele, or that defects induced by *TACI* mutations result from haploinsufficiency (15–17). Yet, the lack of disease in the majority of carriers with *TACI* mutations and their puzzling relative commonness (approximately 1%) in the general population cast doubt

on their role in the pathogenesis of immune deficiency (18). When associated with CVID, a single *TACI* mutation predicts the development of autoantibody-mediated autoimmune disease, whereas patients with two mutated alleles are mostly spared clinical autoimmune conditions, suggesting a complex role for TACI in maintaining B cell tolerance (19, 20).

In healthy controls, most autoreactivity is purged from the repertoire at two distinct B cell tolerance checkpoints (21). The first checkpoint occurs centrally in the bone marrow and is dependent upon B cell intrinsic factors including the BCR and TLR signaling pathways that mediate binding to self-antigens (22–25). In contrast, regulation of the peripheral B cell tolerance checkpoint involves Tregs and potentially plasma BAFF concentrations (26–28). To determine the impact of *TACI* mutations on the establishment of human B cell tolerance, we cloned and expressed in vitro recombinant antibodies from single new emigrant/translational and mature naive B cells from subjects with or without CVID carrying one or two *TACI* mutation(s). We found that *TACI* mutations impaired the removal of autoreactive B cells at the central B cell tolerance checkpoint by imposing BCR and TLR defects in a dose-dependent manner in all subjects, regardless of CVID status. In contrast, only healthy individuals, and not CVID patients, were capable of mitigating central B cell tolerance defects with an effective peripheral B cell tolerance checkpoint, which does not rely on functional TACI. Finally, we report that secreted antinuclear antibodies (ANAs) are common in CVID patients with one *TACI* mutation and correlate with the presence of circulating T follicular helper (Tfh) cells as well as a high incidence of autoim-

Conflict of interest: The authors have declared that no conflict of interest exists.

Citation for this article: *J Clin Invest.* 2013;123(10):4283–4293. doi:10.1172/JCI69854.

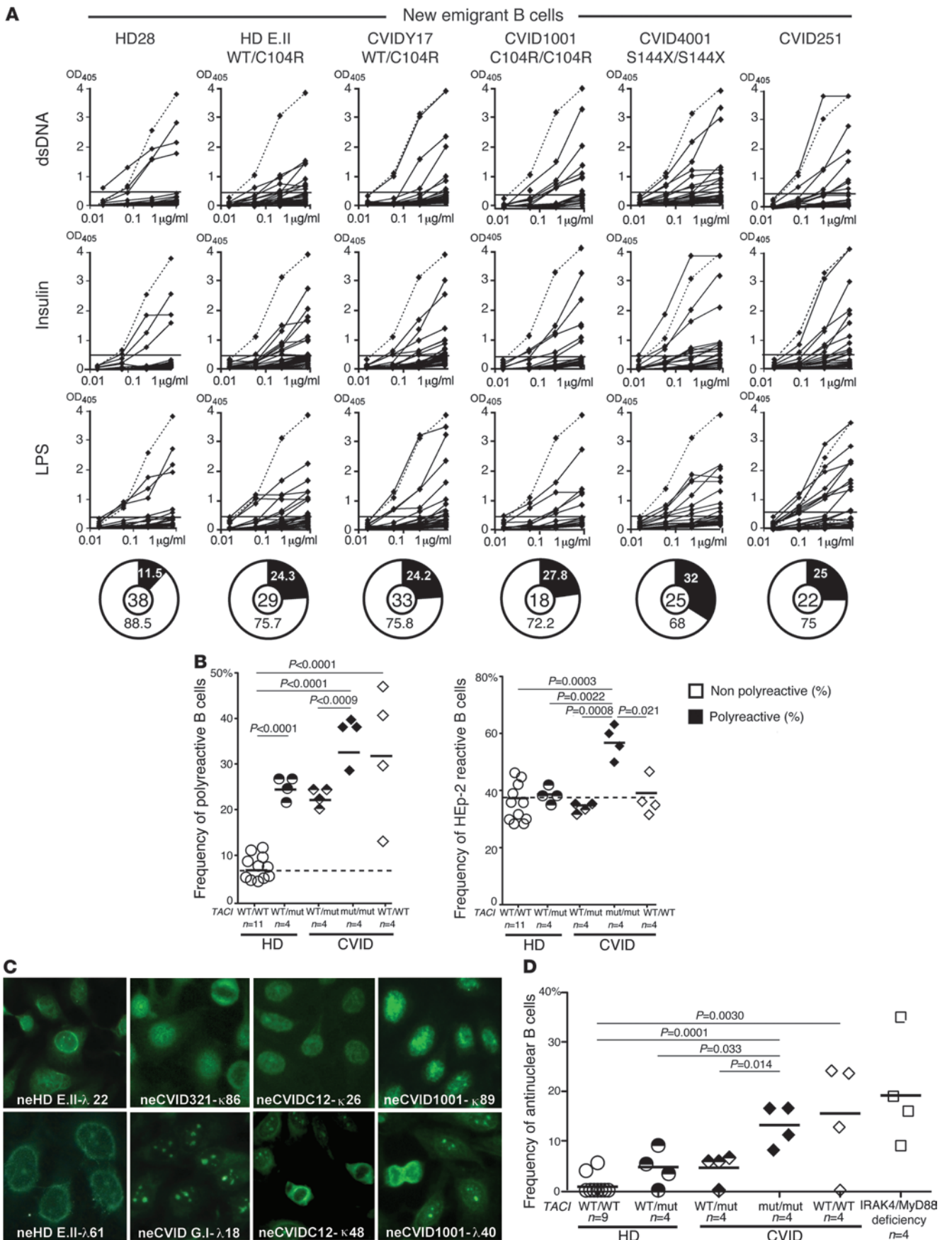




Figure 1

Defective central B cell tolerance checkpoint in individuals carrying *TACI* mutation(s). **(A)** Recombinant antibodies derived from new emigrant/transitional B cells from representative individuals were tested by ELISA for reactivity against dsDNA, insulin, and LPS (21). Antibodies were considered polyreactive when they reacted against all three antigens. Dashed lines show ED38 antibody–positive control, and solid lines show binding for each cloned recombinant antibody. Horizontal lines define the cutoff OD405 for positive reactivity. For each individual, the frequency of polyreactive and nonpolyreactive clones is summarized in pie charts, with the total number of antibodies tested indicated in the center. **(B)** The frequency of polyreactive new emigrant/transitional B cells increased in all individuals carrying *TACI* mutation(s) and most CVID patients without *TACI* mutations, whereas the frequency of HEP-2–reactive new emigrant/transitional B cells only increased in patients with two mutated *TACI* alleles. **(C)** Autoreactive antibodies expressed by new emigrant/transitional B cells from individuals carrying *TACI* mutation(s) included clones with various nuclear staining patterns on HEP-2 cells. Original magnification, $\times 40$. **(D)** The frequency of antinuclear new emigrant/transitional B cells in individuals carrying *TACI* mutation(s) was elevated compared with healthy controls and is reminiscent of patients with MyD88 and IRAK4 deficiency (white squares) (23). Statistical significance by an unpaired Student's *t* test is indicated.

munity, whereas subjects with two *TACI* mutations who are mostly protected from autoimmunity were completely devoid of ANAs and circulating Tfh cells.

Results

Central B cell tolerance is defective in all subjects with TACI mutations. Central B cell tolerance is responsible for the removal of most polyreactive and antinuclear B cells (21). To determine whether this checkpoint is affected by *TACI* mutations, we cloned antibodies expressed by single CD10⁺CD21^{lo}IgM^{hi}CD27⁺CD20⁺ new emigrant/transitional B cells from four representative individuals from the following three subject groups: healthy donors with one *TACI* mutation and CVID patients with one or two *TACI* mutations. We found a significant increase in the frequency of polyreactive clones in new emigrant/transitional B cells from all individuals with *TACI* mutations, which represented 20.0%–26.1% in either healthy individuals or CVID patients with a single mutation, compared with 5.3%–11.5% in healthy controls without mutations (Figure 1, A and B, and Supplemental Tables 1–18; supplemental material available online with this article; doi:10.1172/JCI69854DS1). Polyreactive B cells were even more prevalent in patients with two *TACI* mutations comprising 28.5%–39.9% of their new emigrant/transitional B cells and were also frequent in CVID patients without *TACI* mutations as previously reported (Figure 1, A and B, and ref. 29). This increase in autoreactive clones in patients with two *TACI* mutations compared with subjects with a single *TACI* mutation was further evidenced by the significantly increased frequency of both HEP-2–reactive and nuclear-reactive new emigrant/transitional B cells in these subjects (Figure 1, B–D). Hence, *TACI* mutations interfere in a gene-dosage manner with the establishment of central B cell tolerance in all individuals regardless of their CVID status.

B cell activation after BCR, TLR7, and TLR9 stimulation is TACI gene dosage dependent. The high frequency of ANA-expressing new emigrant B cells in subjects with *TACI* mutations is reminiscent of IRAK4- and MYD88-deficient patients who display similar features, suggesting defective TLR functions in the presence of

mutated *TACI* molecules (23). We therefore assessed the in vitro activation of naive B cells from individuals with and without *TACI* mutations; the cells were stimulated for 2 days through BCR, CD40, TLR7, or TLR9 (Figure 2). We found that independent of disease status, naive B cells carrying a single *TACI* mutation displayed a diminished induction of B cell activation markers CD86 and CD69 when stimulated with F(ab')₂ anti-human IgM, which triggers BCRs, gardiquimod (TLR7 agonist), or CpG (TLR9 agonist). By contrast, B cells with two *TACI* mutations were nearly unresponsive to such stimulation as also evidenced by the defective induction of the coactivation molecule ICOSL (Figure 2A and Supplemental Figure 1A). Similarly to adenosine deaminase (ADA) inhibition (25), *TACI* mutations imposed selective, not global, TLR7- and TLR9-dependent B cell activation defects because the induction of CD25 by TLR agonists was unaffected in B cells carrying *TACI* mutations (Figure 2B and Supplemental Figure 1B). In addition, B cells with *TACI* mutations did not suffer from an intrinsic inability to upregulate CD86 because this molecule was normally induced after CD40 triggering (Figure 2A). Moreover, we observed B cell activation defects in CVID patients without *TACI* mutations that were similar to the defects in B cells carrying *TACI* mutations but with overall greater variability (Figure 2, A and B, Supplemental Figure 1, A and B, and refs. 3, 4). Thus, *TACI* plays an important and selective role in mediating BCR, TLR7, and TLR9 functions during B cell activation.

TACI interacts with TLR7 and TLR9 in B cells. The identification of *TACI*-dependent TLR7 and TLR9 functions in B cells and the rapid induction of *TACI* on the B cell surface after activation by these TLRs suggest an interplay between *TACI*, TLR7, and TLR9, since each uses MyD88 for intracellular signaling (25, 30, 31). To explore the potential interaction between *TACI* and these TLRs, we performed coprecipitation experiments using healthy donor splenic B cells under different stimulating conditions (Figure 3). We found that *TACI* coprecipitated with TLR9 and that the interaction was enhanced upon activation of these receptors by their respective ligands APRIL and CpG, but less so by the TLR7 ligand gardiquimod (Figure 3A). *TACI* did not associate with the 120-kDa unactivated full-length TLR9, but associated only with its cleaved 65-kDa mature form created after the binding of its ligand, DNA (32). Hence, *TACI* only interacts with TLR9 after activation (Figure 3A). Coprecipitation of *TACI* and mature TLR9 was also observed in the human IgD⁺ E2E B cell line under similar activating conditions (Supplemental Figure 2). In contrast to TLR9, the interaction between *TACI* and TLR7 was only detected upon simultaneous activation by APRIL and gardiquimod, respectively (Figure 3B). Once more, *TACI* did not bind to the 120-kDa unactivated full-length TLR7, but was found to bind only to its cleaved 75-kDa mature form, thereby demonstrating that the association of *TACI* with TLR7 follows TLR7 activation by its ligand (Figure 3B and ref. 33).

A role for *TACI* in potentiating TLR7 and TLR9 functions in B cells was further revealed by B cell activation experiments using *TACI* and TLR costimulations. Indeed, *TACI* triggering by its ligand APRIL enhanced TLR7- or TLR9-induced B cell proliferation and IgM secretion in vitro, whereas APRIL alone did not elicit any responses (Supplemental Figure 3). The specificity of the synergy between *TACI* and TLR7 or TLR9 was demonstrated by the addition of *TACI*-Ig, a decoy receptor for soluble APRIL and BAFF that blocked the enhanced B cell responses induced by APRIL and TLR agonists (Supplemental Figure 3 and ref. 33). The in vivo activation of *TACI* likely relies upon the production of its ligands in

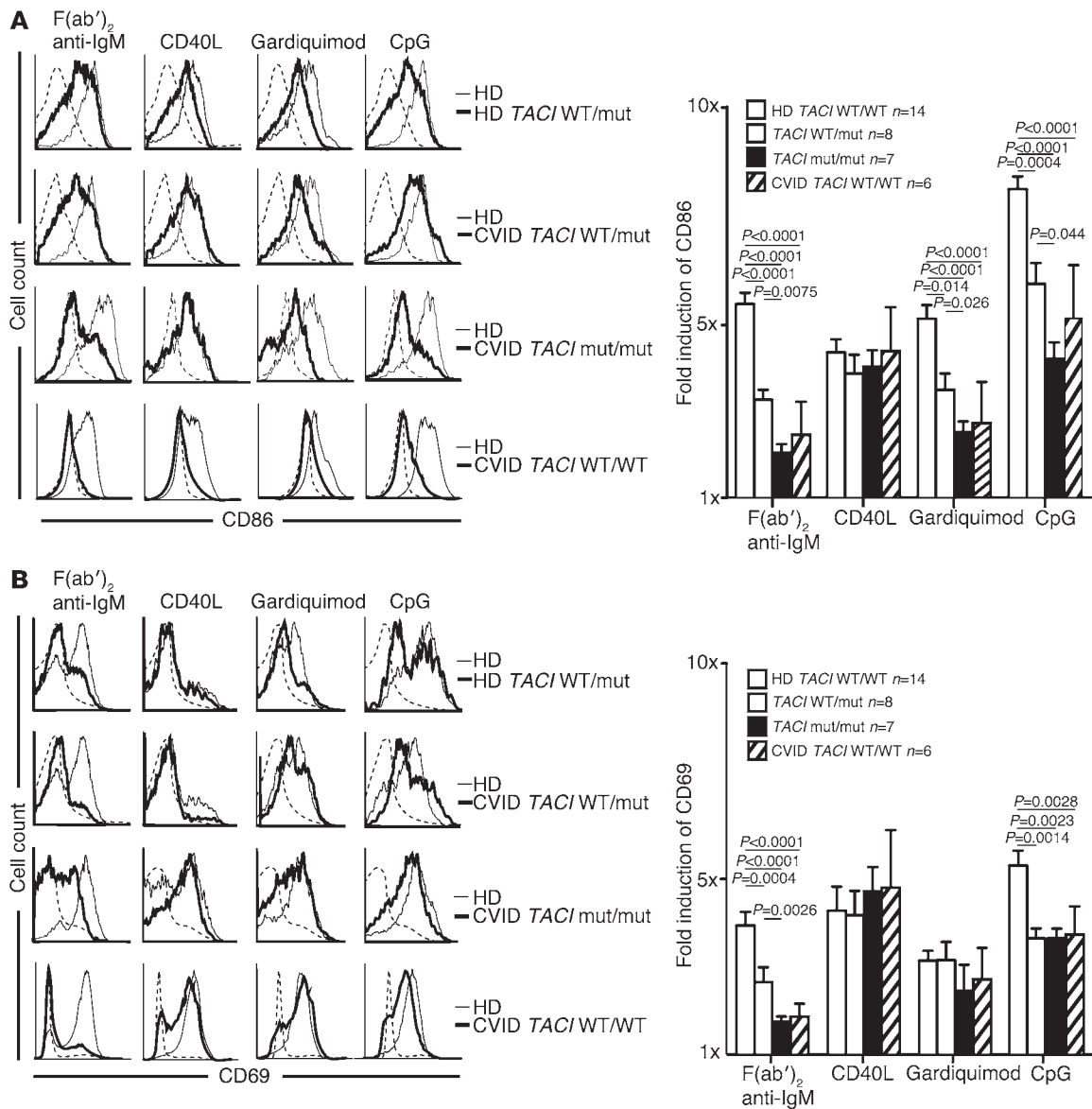


Figure 2

TAC1 mutation(s) result in selective defects for in vitro naive B cell activation. Purified naive B cells from representative individuals with and without *TAC1* mutation(s) (thick lines) displayed decreased CD86 (A) but mostly normal CD69 (B) induction compared with healthy controls (thin lines) after 48 hours of stimulation with F(ab')₂ anti-IgM, CpG, and gardiquimod, but not CD40L. CVID patients without *TAC1* mutation(s) (thick lines) displayed more variable CD86 and CD69 induction under activating conditions. Unstimulated healthy controls (dashed line) are shown for comparison. CVID patients (n = 3) and healthy subjects (n = 5) with a single *TAC1* mutation were pooled because they displayed similar decreased B cell responses. Bar graphs (right) represent the combined data from multiple B cell activation experiments. Error bars represent the mean ± SEM. Statistical significance is indicated by an unpaired Student's *t* test.

trans by non-B cells, including dendritic cells, since B cells did not seem to secrete substantial amounts of BAFF or APRIL (Supplemental Figure 4 and refs. 34, 35). Hence, we identified ligand-dependent interactions between TAC1 and endosomal receptors TLR7 and TLR9, which further reveal that TAC1 plays an important role in mediating human B cell activation by these TLRs.

Peripheral B cell tolerance is defective in CVID patients. A second B cell tolerance checkpoint is responsible for the elimination of additional autoreactive B cells before they enter the CD10⁺C-CD21⁺IgM⁺CD27⁺CD20⁺ mature naive B cell compartment (21). We

assessed the impact of *TAC1* mutations on peripheral B cell tolerance by testing HEP-2 cell lysates for the reactivity of recombinant antibodies cloned from mature naive B cells from healthy subjects and CVID patients with or without *TAC1* mutations (Supplemental Tables 19–35). We observed a significantly elevated frequency of HEP-2 reactive mature naive B cells in all tested CVID patients with one *TAC1* mutation (38.5%–47.6%; *P* < 0.001), those with two *TAC1* mutations (58.5%–65%; *P* < 0.001), and those without *TAC1* mutations (28.6%–56%; *P* < 0.001) compared with healthy controls (16.0%–26.3%), demonstrating an impaired peripheral B cell toler-

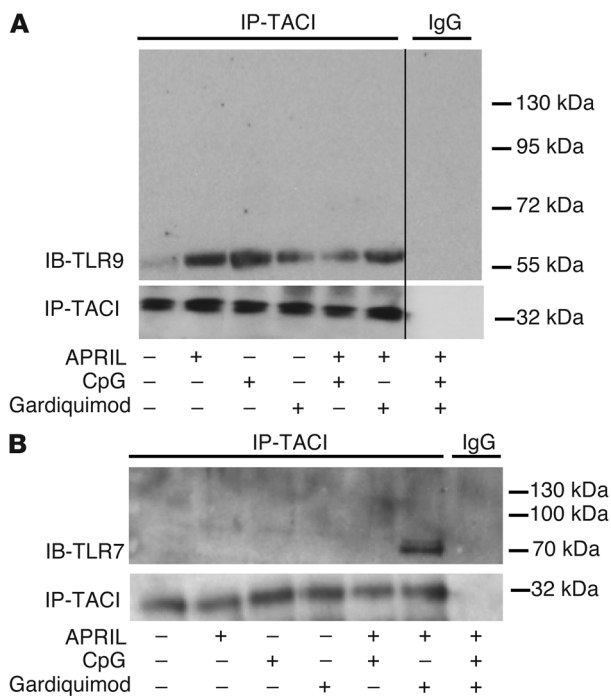


Figure 3

TACI interacts with activated/cleaved TLR7 and TLR9. TACI associates with activated/cleaved 65-kDa TLR9 (A) or 75-kDa TLR7 (B) after the receptors triggered or cotriggered with their respective ligands. Immunoprecipitations were performed using human splenic B cells stimulated or not by the TACI ligand APRIL, TLR9 ligand CpG, and/or TLR7 ligand gardiquimod, as indicated below the blots. The thin black vertical line in A signifies the absence of a single lane from an otherwise contiguous blot that was removed, as it did not further clarify the TACI-TLR9 interaction.

ance checkpoint in these patients (Figure 4, A and B). Peripheral B cell tolerance defects were further evidenced by higher frequencies of polyreactive mature naive clones in CVID patients compared with healthy controls, with the highest frequency being observed in B cells from subjects with two *TACI* mutations that frequently displayed antinuclear reactivity (Supplemental Figure 5 and Supplemental Figure 6, A and B). In contrast, between the new emigrant/transitional and mature naive B cell developmental stages, healthy donors with one *TACI* mutation showed a significant reduction in both HEP-2-reactive (37.7% to 19.3%; $P = 0.005$) and polyreactive (24.9% to 18.2%; $P = 0.046$) clones (Figure 4 and Supplemental Figure 6B). We conclude that a single *TACI* mutation does not affect the peripheral B cell tolerance checkpoint, which is functional in all healthy subjects but defective in CVID patients.

Peripheral B cell tolerance defects in CVID patients correlate with elevated plasma BAFF concentrations and decreased Treg frequencies. Defects in peripheral B cell tolerance have been correlated with elevated plasma BAFF concentrations and lower Treg frequencies, both features that have already been reported in CVID (7, 11, 12, 26–28). We found 4- to 5-fold increased elevations in plasma BAFF concentrations in CVID patients, contrasting with normal BAFF concentrations in healthy donors with a single *TACI* mutation (Figure 5A). Using a quantitative PCR-based assay for enumerating κ -deletion recombination excision circles (KRECs) that estimates the number of divisions undergone by a B cell population (36), we found that mature naive B cells from CVID patients with and without *TACI* mutations displayed significantly increased homeostatic proliferation, with an average of 3.0 divisions *in vivo* compared with 1.9 divisions in B cells from healthy donors both with and without *TACI* mutations (Figure 5B). Both the defective peripheral B cell tolerance checkpoint and the dysregulated homeostatic B cell proliferation observed in CVID patients were characteristic of FOXP3-deficient patients who do not have functional Tregs, suggesting a similarly altered Treg compartment in

CVID (28). Indeed, we found that the frequency of CD4⁺CD25^{hi}CD127^{lo}FOXP3⁺ Tregs was decreased in most CVID patients, independent of the presence and/or number of *TACI* mutations, but not in healthy donors with a single *TACI* mutation (Figure 5C). Impaired Treg function in CVID patients was further suggested by their altered Ki67⁺CD45RO⁺CD25^{lo} phenotype, also reminiscent of nonfunctional Tregs from FOXP3-deficient IPEX patients (Supplemental Figure 7, A and B, and ref. 28). Our direct assessment of Treg suppressive function *in vitro* demonstrated the impaired ability of Tregs from CVID patients with and without *TACI* mutations to inhibit the proliferation of CFSE-labeled CD3⁺CD4⁺CD25⁻ autologous and heterologous T responder (Tresp) cells (Figure 5, D and E). In contrast, Tregs from healthy donors with a single *TACI* mutation were as suppressive as Tregs from controls with no *TACI* mutation, further demonstrating that *TACI* mutations are not inducing Treg defects *per se* (Figure 5, D–F). Thus, *TACI* mutations are not responsible for the elevated plasma BAFF concentrations, the low Treg frequencies, or the poor Treg suppressive function associated with the majority of CVID patients displaying a defective peripheral B cell tolerance checkpoint.

Circulating ANAs and BCL6⁺ Tfh cells are associated with a single TACI mutation. CVID patients with a single *TACI* mutation are prone to developing autoimmune syndromes, whereas carrying two mutations seems to prevent autoimmunity (19, 20). To further investigate this apparent discrepancy, we assessed the plasma of healthy donors and CVID patients with or without *TACI* mutation(s) for IgG ANAs. In agreement with previous reports, 16.2% of healthy donors were found positive for low-titer (1:80 dilution) plasma ANAs, with very few displaying high ANA titers (5.4% at a dilution of 1:320) (37). In contrast, most CVID patients with one *TACI* mutation displayed low-titer plasma ANAs (1:80 dilution), and 30% of them had detectable high-titer ANAs (1:320) (Figure 6, A and B). We found that 40% of healthy donors carrying a single *TACI* mutation also frequently harbored significant high-titer

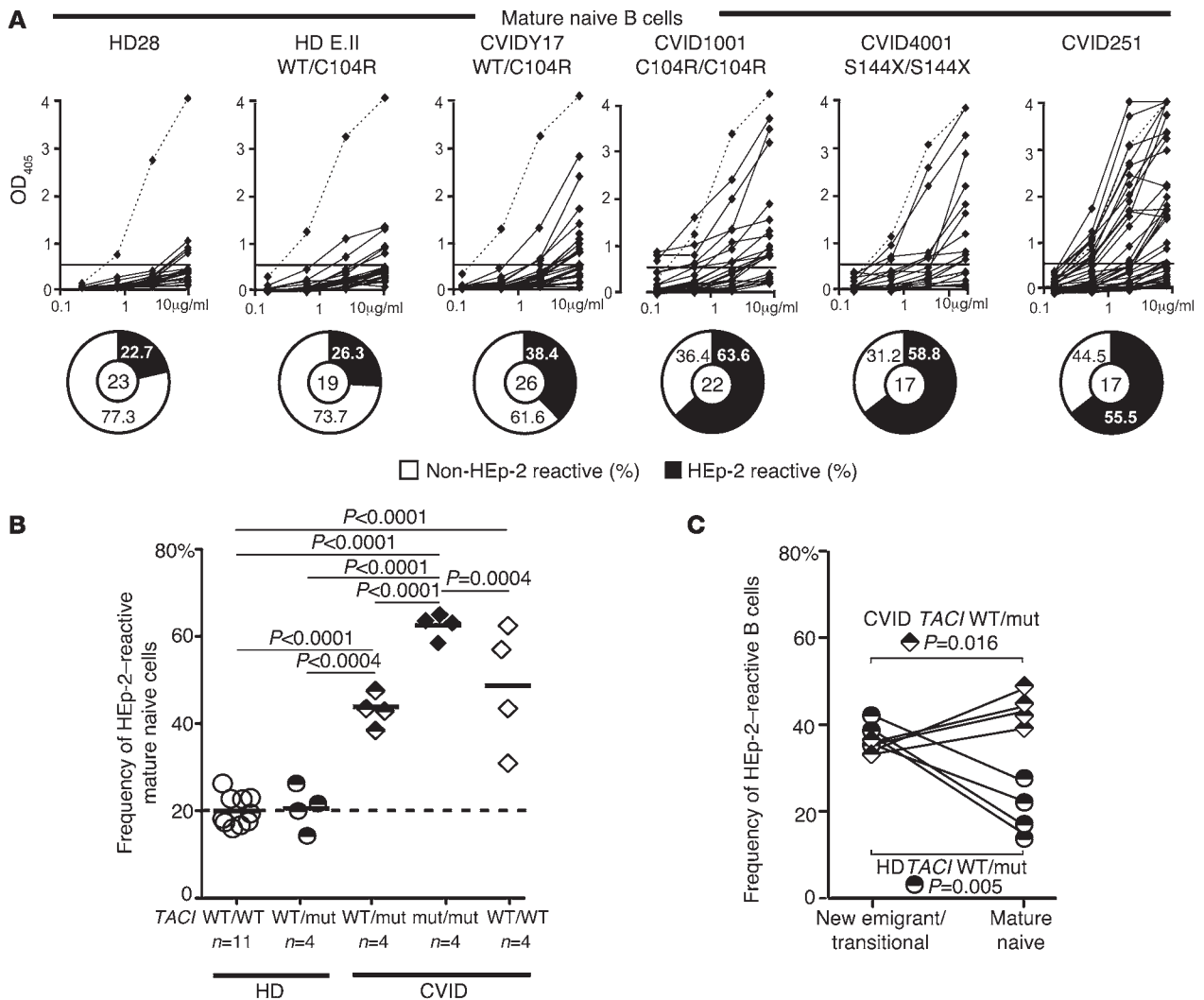


Figure 4

A defective peripheral B cell tolerance checkpoint in CVID patients. (A) Recombinant antibodies derived from mature naive B cells from subjects or CVID patients with or without *TAC1* mutation(s) were tested by ELISA for anti-HEp-2 cell reactivity. Dashed lines show the ED38-positive control (21). Horizontal lines define the cutoff OD405 for positive reactivity. For each individual, the frequencies of HEp-2-reactive and non-HEp-2-reactive clones are summarized in pie charts, with the number of antibodies tested indicated in the center. (B) The frequency of HEp-2-reactive mature naive B cells was higher in all CVID patients than was observed in healthy donors with or without one *TAC1* mutation. (C) The frequency of HEp-2-reactive clones rose between the new emigrant/transitional and mature naive B cell stages in CVID patients with a *TAC1* mutation, whereas it decreased in healthy carriers with one *TAC1* mutation. Statistical significance is indicated by an unpaired or paired (for C) Student's *t* test.

ANAs, thereby suggesting a role for TAC1 in preventing autoantibody secretion (Figure 6, A and B). Remarkably, ANAs were undetectable even at a low 1:80 titer in all seven CVID patients with two *TAC1* mutations, revealing that the absence of functional TAC1 blocks autoreactive B cell activation (Figure 6, A and B).

Since the generation of IgG ANAs is considered T cell dependent, we analyzed effector T cell subsets potentially contributing to autoantibody production, such as CXCR5⁺PD-1^{hi}CD4⁺ circulating Tfh cells (38, 39). These T cells are rare in the peripheral blood of control healthy donors without a *TAC1* mutation and in CVID patients with two *TAC1* mutations, comprising 3.7% and 2.1% of their CD4⁺ T cells, respectively (Figure 6, C and D). In contrast, we found a high average frequency of CXCR5⁺PD-1^{hi}CD4⁺ T cells in healthy donors and CVID patients with a single *TAC1* mutation as well as in some

CVID patients without *TAC1* mutations (healthy donor [HD] TAC1 WT/mut: 10.1%; CVID TAC1 WT/mut: 17.1%; and CVID: 9.9% of CD4⁺ cells) (Figure 6, C and D). These cells resembled tonsillar Tfh cells, which are normally localized in GCs, since they expressed intracellular BCL6, the master transcriptional regulator responsible for Tfh differentiation, as well as the coactivation molecules ICOS and CD40L (Figure 6C and data not shown) (40). Tfh cells were generally most prevalent in the peripheral blood of subjects who had the lowest frequencies of circulating Tregs, whereas in subjects with two *TAC1* mutations, Tfh cells were nearly absent (Figure 6E). In addition, several cytokines secreted by Tfh cells, including IL-4 and IL-10, but not IL-21, were found at elevated concentrations in the plasma of CVID patients but not in that of healthy donors with one *TAC1* mutation (Supplemental Figure 8). Hence, a single

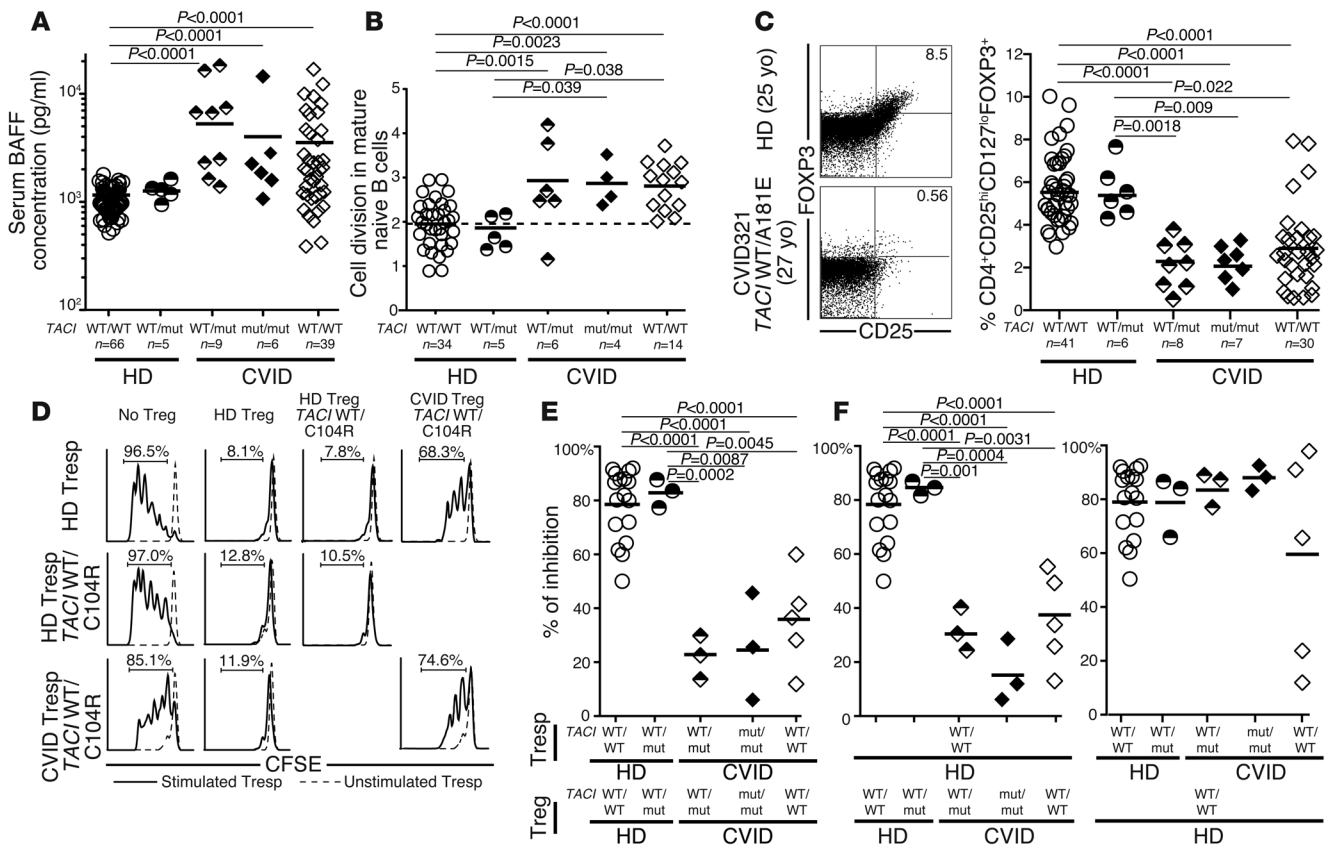


Figure 5

Increased plasma BAFF concentrations, homeostatic naive B cell proliferation, and an impaired Treg compartment are features common to CVID patients with and without *TAC1* mutations. *TAC1* mutations did not affect plasma BAFF concentrations (pg/ml) measured by ELISA (A) or mature naive B cell expansion detected by KREC analysis (B). Both were greater in CVID patients compared with healthy controls. (C–F) CVID status, but not *TAC1* mutations, affected Treg frequency and function. (C) Dot plots represent CD4⁺ gated CD25^{hi}FOXP3⁺ T cells of a healthy donor control and an age-matched CVID patient with one *TAC1* mutation. Scatter plots reveal that decreased CD4⁺CD25^{hi}CD127^{lo}FOXP3⁺ Treg frequency only correlated with CVID. yo, year-old. (D) Representative histograms of Treg-mediated suppression of autologous and heterologous CFSE-labeled Tresp cells on day 4.5 from a CVID patient and a healthy relative both carrying a *TAC1* mutation were compared with a healthy control. Dashed lines display nonstimulated Tresp cells. The autologous and heterologous suppressive activities of Tregs from healthy donor controls, healthy carriers with a single *TAC1* mutation, CVID patients with one or two mutated *TAC1* alleles, as well as CVID patients without *TAC1* mutations are summarized in E and F, respectively. Statistical significance is indicated by an unpaired Student's *t* test.

TAC1 mutation correlates with an increased frequency of circulating CD4⁺CXCR5⁺PD-1^{hi}ICOS^{hi}BCL6⁺ Tfh cells and the production of high-titer IgG ANAs, thereby revealing an additional, more distal B cell tolerance breach often associated with autoimmune syndromes. In contrast, a second *TAC1* mutation resulting in a lack of functional TAC1 counteracts the effects of a single *TAC1* mutation, creating a scenario of scant circulating Tfh cells, absent ANAs, and a reduced susceptibility to autoantibody-mediated disease.

Discussion

We report herein several roles played by *TAC1* mutations in the development of CVID. Previously, the association between *TAC1* mutation(s) and CVID was unclear because many people carrying *TAC1* mutations are healthy and without infectious susceptibility or decreased serum immunoglobulins (18, 41). However, we found that all individuals with *TAC1* mutations failed to remove developing autoreactive B cells in the marrow. This defective central B cell tolerance checkpoint is the first evidence of the full pene-

trance of *TAC1* mutations in B cell development. This effect is not influenced by the nature of the mutation itself or the CVID status of the carrier. Indeed, healthy carriers with a single C104R *TAC1* mutation abrogating ligand binding displayed abnormalities similar to those in CVID patients carrying the A181E mutation affecting the TAC1 transmembrane domain. Likewise, patients homozygous for the allele encoding the C104R mutation showed profound B cell defects and elevated autoreactive B cell frequencies similar to those in the S144X/S144X individual in whom no TAC1 protein is expressed (8). Hence, various mutated *TAC1* alleles encode similarly nonfunctional molecules. It remains to be determined whether these mutations result in the expression of interfering dominant-negative products or whether their impact on B cell activation and central selection is solely due to haploinsufficiency.

How do *TAC1* mutations affect the removal of developing autoreactive B cells? Previous studies have suggested that the regulation of central B cell tolerance is intrinsic to B cells and involves the sensing of self-antigens through binding with BCRs

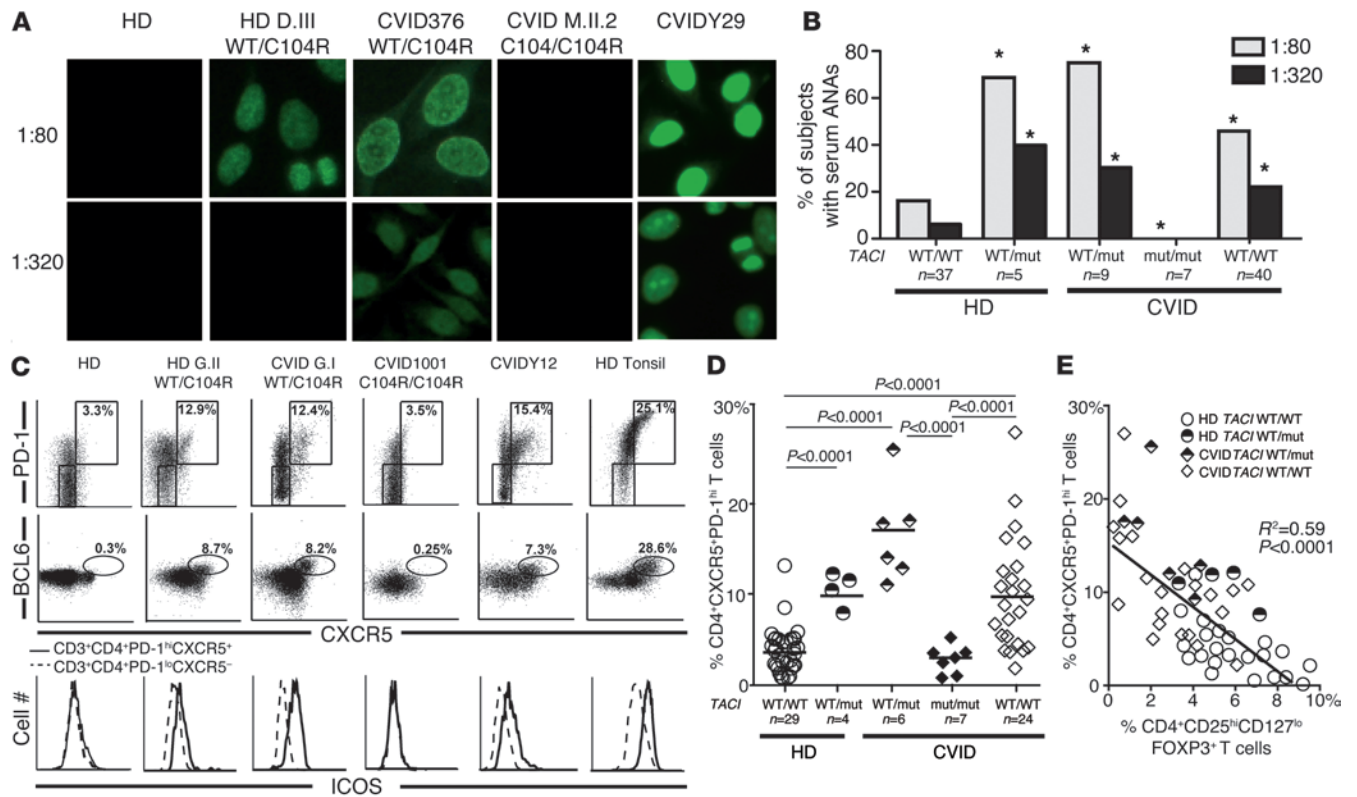


Figure 6 Heterozygous *TAC1* mutations correlate with secreted ANAs and circulating Tfh cells. (A and B) ANAs with diverse nuclear staining patterns at low (1:80) and high (1:320) dilutions were common in individuals with one *TAC1* mutation, but were absent in patients with two mutated alleles. **P* < 0.05 (statistical differences compared with healthy controls by χ^2 testing). (C and D) Expanded CD4+PD-1^{hi}CXCR5⁺ Tfh cells in the peripheral blood of individuals with one, but not two, *TAC1* mutations and some CVID patients without *TAC1* mutations. In addition to PD-1 and CXCR5 expression, Tfh cells in carriers of a single *TAC1* mutation were further evidenced by increased intracellular BCL6 expression in and increased cell surface ICOS expression on CD4+PD-1^{hi}CXCR5⁺ T cells (solid line in lower panel of C) compared with CD4+PD-1^{lo}CXCR5⁻ T cells (dashed line in lower panel of C). (E) The frequency of circulating Tfh cells was inversely correlated with the frequency of circulating Tregs in enrolled subjects with or without single *TAC1* mutations. Statistical significance is indicated using χ^2 testing (B), an unpaired Student's *t* test (D), or linear regression analysis (E).

and potentially TLRs (23). Our data from subjects with *TAC1* mutations further support such a model, as *TAC1* is expressed primarily by B cells and regulates BCR, TLR7, and TLR9 functions. Indeed, B cells carrying a single *TAC1* mutated allele display decreased CD86 upregulation in vitro when BCR, TLR7, or TLR9 are triggered, whereas B cells with two mutated *TAC1* alleles fail to upregulate this costimulatory molecule. Interestingly, defective CD86 induction after BCR and TLR9 stimulation as well as impaired TLR responses are features already reported for B cells from CVID patients and may represent one mechanism by which *TAC1* mutations favor the development of CVID (5, 42). *TAC1*-deficient patients as well as subjects with a single *TAC1* mutation share a similar defective central B cell tolerance checkpoint that is also characterized by a failure to remove antinuclear clones. The presence of antinuclear clones in the new emigrant/transitional B cell compartment is reminiscent of IRAK4-deficient, MYD88-deficient, and ADA-deficient severe combined immune deficiency patients who have revealed the IRAK4/MYD88 and ADA pathways to be essential for the counterselection of such reactivity (23, 25). Since TLR7 and TLR9 bind nucleic acids and require IRAK4, MYD88, ADA, and *TAC1* to function in B cells, these receptors may therefore play an important role in establish-

ing central tolerance against RNA- and DNA-containing antigens. In addition, *TAC1*'s intracellular domain binds MYD88 (30), and our data show that *TAC1* can interact with both TLR7 and TLR9 upon coengagement of these receptors. Hence, polyreactive and antinuclear BCRs that bind self-antigens in the bone marrow may internalize and deliver them to endosomal compartments. TLR7 and TLR9 may then recognize RNA- and DNA-containing complexes, leading to TLR proteolytic activation and interaction with *TAC1*, which in turn may allow the amplification of IRAK4/MYD88-dependent signals to mediate central tolerance mechanisms. We conclude that *TAC1* plays an essential role in the establishment of central B cell tolerance in humans.

TAC1 mutations potentially favor CVID development by impairing B cell activation. However, many individuals carrying a *TAC1* mutation do not develop CVID (18, 41). By comparing such subjects with CVID patients with *TAC1* mutation(s), we found that disease correlated with an abnormal peripheral B cell tolerance checkpoint independently of *TAC1* mutations. Indeed, all asymptomatic individuals with a single *TAC1* mutation managed to prevent the accumulation of autoreactive clones in their mature naive B cell compartment. This demonstrates that peripheral B cell tolerance can be established despite a defective central B cell tolerance checkpoint. The increased



frequency of autoreactive mature naive B cells in patients with autoimmune diseases such as rheumatoid arthritis, lupus erythematosus, and type 1 diabetes — all of whom display defective central B cell tolerance — is therefore likely due to a defective peripheral B cell tolerance checkpoint and not just an accumulation in the periphery of autoreactive clones produced by the marrow (24, 43, 44). The defect in peripheral tolerance of CVID patients is associated with an impaired Treg compartment, which plays an important role in regulating this checkpoint (28). In addition, decreased Treg functions may also lead to increased proliferative events in the mature naive B cell compartment of CVID patients compared with healthy donors with or without *TACI* mutations (28). The elevated BAFF concentration in the plasma of CVID patients may also favor B cell homeostatic expansion and provide survival signals to autoreactive clones, thereby preventing their elimination in the periphery (14, 26–28, 45). Hence, one *TACI* mutation is not sufficient to induce the development of CVID and is not responsible for the defective peripheral B cell tolerance checkpoint observed in CVID patients. It is unclear at this point whether another susceptibility gene is responsible for CVID development, potentially affecting Treg function, or whether nongenetic environmental factors are more important in the development of this disease.

It was previously reported that CVID patients with a single *TACI* mutation are prone to autoimmune cytopenias, whereas patients devoid of functional TACI are protected from autoimmunity (20). This apparent discrepancy was initially puzzling, as we found that subjects with two mutated *TACI* alleles had increased frequencies of autoreactive clones in all naive B cell compartments compared with CVID patients with a single *TACI* mutation. However, in the absence of functional TACI, this increase in autoreactive B cells is accompanied by profound impairments of B cell activation after BCR, TLR7, and TLR9 triggering that are significantly worse than in subjects with a single *TACI* mutation. Moreover, TLRs play an essential role in the development of autoimmunity (46). Hence, severe TLR function impairment in TACI-deficient individuals is likely to be protective against autoimmunity despite profoundly defective autoreactive B cell counterselection, leaving these individuals with an immunodeficient phenotype. In agreement with this hypothesis, we could detect ANAs in the plasma of CVID patients and asymptomatic individuals with a single *TACI* mutation, but not in subjects with two mutated *TACI* alleles.

High-titer plasma ANAs in individuals with a single *TACI* mutation correlated with the presence of circulating CD4⁺CXCR5⁺P-D-1^{hi}ICOS^{hi}BCL6⁺ Tfh cells, which normally reside in GCs and provide B cell help. B cells in GCs also produce reciprocal signals for Tfh differentiation. The absence of functional TACI resulting from two mutated *TACI* alleles impairs costimulatory ICOSL and CD86 expression and thereby precludes the development of Tfh cells and limits ANA secretion. Indeed, another group of CVID patients who are spared autoimmune disease and devoid of circulating Tfh cells are those with ICOS deficiency, whose lymph node GCs display a disturbed architecture (47–49). Thus, the absence of circulating Tfh cells in patients with two *TACI* mutations may illustrate dysfunctional GC reactions and decreased isotype-switched antibody secretion. In agreement with this hypothesis, murine models have demonstrated that either CD86 or ICOSL deficiency abolish GC formation and impair the generation of Tfh cells (50–52). However, expanded Tfh cells driven by overexpression of ICOSL on B cells have been reported in *Taci* knockout mice (53). Such differences between mouse and man may reconcile

the development of an SLE-like autoimmune condition in TACI-deficient mice with the absence of autoimmune manifestations in CVID patients with two *TACI* mutations (54). On the other hand, as in *Taci*^{-/-} mice, Tfh cells are enriched in CVID patients with a single *TACI* mutation and likely contribute to the activation of autoreactive B cells through cell-cell interactions and the production of cytokines such as IL-4 and IL-10, leading to ANA secretion. It is unknown at this point why Tfh cell frequency is increased in CVID patients with a single *TACI* mutation or without *TACI* mutations, but their low Treg numbers and poor Treg suppressive function may fail to downregulate Tfh production in GCs (55, 56). Nevertheless, increased Tfh cells may favor autoreactive B cell activation in CVID patients and contribute to the development of autoimmune conditions characteristic of these patients.

In summary, *TACI* mutations favor CVID development by altering B cell activation after BCR, TLR7, and TLR9 stimulation. However, additional genetic or environmental factors are required for the development of CVID. Since TACI regulates the function of BCR, TLR7, and TLR9, which are likely involved in sensing self-antigens in the marrow, subjects with *TACI* mutation(s) not only suffer from B cell activation defects but also from an impaired removal of developing autoreactive B cells. B cells from CVID patients with a single *TACI* mutation may respond to self-antigen stimulation and receive additional T cell help due to the presence of Tfh cells, leading to autoantibody secretion and an increased risk for autoimmune syndromes. In contrast, autoreactive B cells from subjects with two mutated *TACI* alleles lack the functional TACI necessary for B cell activation, resulting in diminished immune and autoimmune humoral responses.

Methods

Patients. We obtained peripheral blood from 21 individuals with *TACI* mutations including 9 CVID patients with one *TACI* mutation and 7 immune-deficient patients with two *TACI* mutations, 6 with CVID, and 1 with hypogammaglobulinemia (Supplemental Table 1). Patient A.II.1, a *TACI* S144X homozygote who has a complete absence of measurable TACI-specific mRNA and protein, has no increased susceptibility to infections and does not require Ig replacement therapy, yet is nonetheless hypogammaglobulinemic (serum IgG 410 mg/dl) (8). Although not a classical CVID phenotype, this patient was grouped here with other CVID patients carrying two *TACI* mutations to simplify subject categorization. Healthy relatives of these patients made up the remaining 5 subjects; each carried one *TACI* mutation. *TACI* alleles present in our study cohort included common missense variants associated with CVID (C104R and A181E) as well as rare mutations such as insertions (204insA, 571insG) and truncations (S144X and S194X). For purposes of comparison, we obtained peripheral blood samples from 50 healthy controls and 50 CVID patients without *TACI* mutations. The study groups were controlled for age and sex. In addition, healthy controls analyzed for their autoreactive B cell frequencies were previously reported and included a 50-year-old male of mixed European descent previously splenectomized after accidental blunt-force trauma (HD28) (25). We also included HD29, the 29-year-old healthy brother of CVID patient CVID332. All subjects were genotyped at the *TACI* gene loci by Sanger sequencing using published primer sets (exons 1–4) (8). For exon 5, the following primer pair was used: 5'-CCTAGTGCAGGGC-CAGGCCTG-3' and 5'-CCGACCTCTGCTCTATCT-3'. All samples were collected in accordance with Yale University's IRB-approved protocol HIC0906005336.



Cell staining and sorting, cDNA, RT-PCR, antibody production, ELISAs, and indirect fluorescence assays. Peripheral B cells were purified from the peripheral blood of research subjects by positive selection using CD20 magnetic beads (Miltenyi Biotec). Single CD19⁺CD21^{lo}CD10⁺IgM^{hi}CD27⁻ new emigrant/transitional and CD19⁺CD21⁺CD10⁺IgM⁺CD27⁻ peripheral mature naive B cells from patients and control donors were sorted on a FACSVantage cell sorter (BD) into 96-well PCR plates. RT-PCR reactions, primer sequences, cloning strategy, expression vectors, recombinant antibody expression, recombinant antibody purification, and recombinant antibody reactivity determination were as previously described (21). For indirect immunofluorescence assays, HEp-2 cell-coated slides (Bion Enterprises) were incubated in a moist chamber at room temperature with patient plasma samples at 1:80 and 1:320 dilutions in PBS according to the manufacturer's instructions. FITC-conjugated goat anti-human IgG was used as a detection reagent. The following antibodies were used for flow cytometric stainings: CD19 APC-Cy7, CD27 PerCP-Cy5.5, CD10 PE-Cy7, IgM FITC, CD21 APC, CD25 FITC, CD69 PE, CD80 FITC, CD86 PE, CD4 APC-Cy7, CD25 PE-Cy7, CD127 PerCP-Cy5.5, CD45RO Pacific Blue, TACI PE, CD303 FITC, CD14 Pacific Blue, CD4 APC-Cy7, CXCR5 PerCP-Cy5.5, PD-1 PE-Cy7, ICOSL PE, CD25 PE, and CD40 APC (all from Biolegend), CD3 eFluor 605NC, BCL6 PE (both from eBioscience), and CD21 BD Horizon V450 (BD). Intracellular staining for FOXP3 Alexa Fluor 488 (clone PCH101; eBioscience) was performed using the FOXP3/Transcription Factor Staining Buffer Set (eBioscience) in accordance with the manufacturer's instructions.

B cell activation and proliferation. Naive B cells were enriched from the blood of healthy donors by negative selection using the Naive B Cell Isolation Kit II (Miltenyi Biotec) and plated for 48 hours at 150,000–200,000 cells per well in a 96-well plate in RPMI 10% FBS and either 2.5 µg/ml polyclonal F(ab')₂ rabbit anti-human IgM (Jackson ImmunoResearch), multimeric soluble recombinant human CD40L 1.0 µg/ml (Alexis Biochemicals), 0.5 µg/ml CpG (TLR9 agonist; Invitrogen), or 1.0 µg/ml gardiquimod (TLR7 agonist; InvivoGen). In proliferation and antibody secretion experiments, total human splenic B cells isolated by negative selection using EasySep were stimulated for 5 days with CpG and gardiquimod in the presence or absence of 500 ng/ml APRIL (R&D Systems). APRIL was blocked with 30 µg/ml of TACI-Ig (Anell). Mouse IgG1 with irrelevant binding activity was used as a control for TACI-Ig. Cell proliferation was assessed by CFSE staining using the CellTrace CFSE Cell Proliferation Kit (Invitrogen) according to the manufacturer's instructions. Supernatants were collected and analyzed to test the presence of BAFF and APRIL using a soluble human BAFF ELISA Kit (Adipogen) and a LEGEND MAX Human APRIL/TNFSF13 ELISA Kit (BioLegend). IgM secretion was detected by ELISA. In brief, M96-Nunc ELISA plates were coated with anti-human IgM (SouthernBiotech). After blocking with 1% BSA in PBS, samples were added to the plate overnight. Purified anti IgM-HRP antibody (Cappel) and TMB substrate solution (BD) were used according to the manufacturer's instructions.

Coinmunoprecipitation experiments. FACS-sorted human CD19⁺ splenic B cells were treated with 0.2 µg/ml recombinant human APRIL (R&D Systems), 0.5 µg/ml CpG (Invitrogen), and/or 1.0 µg/ml gardiquimod (InvivoGen) for 2 hours. For each condition, 10⁷ cells were used. Cells were homogenized and solubilized in lysis buffer (50 mM Tris-HCl, pH 7.5, 150 mM NaCl, 1 mM EDTA, 1 mM EGTA, 0.5% Triton, 0.2 mg/ml BSA, and protease inhibitors) using a Dounce homogenizer. An equal amount of each protein lysate was incubated with anti-TACI biotinylated antibody (Abcam) overnight at 4°C, followed by incubation with 50 µl of streptavidin-sepharose beads (GE Healthcare) for 3 hours. Different buffers were used to wash streptavidin-antigen-antibody complexes (wash buffer 1: 50 mM Tris-HCl, pH 7.5, 500 mM NaCl, 0.1% Triton; wash buffer 2: 10 mM Tris-HCl, pH 7.5, 0.1% Triton). Denatured proteins were analyzed by SDS-polyacrylamide gel electrophoresis. The membranes were immunoblotted with anti-TLR9 (Pierce), anti-TLR7 (Abcam), and anti-TACI (Abcam) antibodies.

KREC assay. The ratio of KREC joints (signal joint) to the Jκ-Cκ recombination genomic joints (coding joint) was determined as previously described (36). In brief, genomic DNA was isolated from sorted B cell fractions by lysing cell pellets in 10 mM Tris-HCl, pH 8.0, containing 100 µg/ml proteinase K (Roche), incubating for 1 hour at 56°C, and heat inactivating the enzyme at 95°C for 10 minutes. Two separate RT-PCR reactions were performed, one reaction to amplify the signal joint and another to amplify the coding joint, as previously detailed. The number of cell divisions was calculated by subtracting the cycle threshold of the PCR detecting the coding joint from that detecting the signal joint.

In vitro Treg suppression assay. CD4⁺ T cells were enriched using the EasySep Human CD4⁺T cell enrichment kit (STEMCELL Technologies). CD4⁺CD25^{hi}CD127^{lo/-} Tregs were sorted by flow cytometry, whereas CD3⁺CD4⁺CD25⁻ Tresp cells were obtained after the depletion of CD25⁺ cells with anti-human CD25 microbeads (Miltenyi Biotec) and then labeled with CellTrace CFSE (InvivoGen) at 5 µM. Treg and Tresp cells were cocultured at a 1:1 ratio in the presence of beads loaded with anti-CD2, anti-CD3, and anti-CD28 (Treg suppression inspector human; Miltenyi Biotec). On days 3.5 to 4.5, cocultures were stained for viability with the LIVE/DEAD kit (Invitrogen), and proliferation of the viable Tresp was analyzed by CFSE dilution.

Cytokine detection. Plasma BAFF concentrations were determined by ELISA according to the manufacturer's instructions (R&D Systems). Cytokines (IL-4, IL-10) in plasma were measured with the High Sensitivity Human Cytokine Magnetic Bead Kit (Millipore) using a Luminex 200. IL-21 was measured by ELISA with the Human IL-21 ELISA Ready-Set-Go Kit (eBioscience).

Statistics. Statistical analysis was performed using GraphPad Prism software, version 5.0 (GraphPad Software). Differences between groups of research subjects were analyzed for statistical significance with unpaired or paired (when within groups), two-tailed Student's *t* tests except for plasma indirect immunofluorescence assay comparisons that used χ^2 testing. For these statistical tests, α was set to $\alpha = 0.05$; accordingly, a *P* value less than or equal to 0.05 was considered significant. The relationship between Tfh cells and Tregs was statistically analyzed using linear regression analysis.

Acknowledgments

We thank J. Bussel, F. Kantor, P. Askenase, C. Randolph, J. Sproviero, F. Hsu, C. Price, and O. Pagovich for providing blood samples from CVID patients, and S. Rudchenko, L. Devine, and C. Wang for cell sorting. This work was supported by grants from the NIH-NIAID (AI071087, AI082713, and AI095848, to E. Meffre, and AI061093, to E. Meffre, A. Cerutti, and C. Cunningham-Rundles); and the NIH-NICHHD (K12HD0141401-10, to N. Romberg); and by a CIS/Talecris Fellowship Award (to N. Romberg). T. Kinnunen was supported by the Sigrid Juselius Foundation, the Finnish Medical Foundation, and the Saastamoinen Foundation. L. Menard was supported by NIH-NIAID grant T32 AI089704. T. Cantaert received support from the Rubicon program, Netherlands Organization for Scientific Research, and H. Morbach received support from the German Research Foundation (DFG, MO 2160/2-1).

Received for publication March 14, 2013, and accepted in revised form July 25, 2013.

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- Al-Herz W, et al. Primary immunodeficiency diseases: an update on the classification from the international union of immunological societies expert committee for primary immunodeficiency. *Front Immunol*. 2011;2:54.
- Sanford JP, Favour CB, Tribeman MS. Absence of serum gamma globulins in an adult. *N Engl J Med*. 1954; 250(24):1027–1029.
- Bryant A, Calver NC, Toubi E, Webster AD, Farrant J. Classification of patients with common variable immunodeficiency by B cell secretion of IgM and IgG in response to anti-IgM and interleukin-2. *Clin Immunol Immunopathol*. 1990;56(2):239–248.
- Yu JE, et al. Toll-like receptor 7 and 9 defects in common variable immunodeficiency. *J Allergy Clin Immunol*. 2009;124(2):349–356.
- Denz A, Eibel H, Illges H, KiENZle G, Schlesier M, Peter HH. Impaired up-regulation of CD86 in B cells of “type A” common variable immunodeficiency patients. *Eur J Immunol*. 2000; 30(4):1069–1077.
- Horn J, et al. Decrease in phenotypic regulatory T cells in subsets of patients with common variable immunodeficiency. *Clin Exp Immunol*. 2009; 156(3):446–454.
- Arumugakani G, Wood PM, Carter CR. Frequency of Treg cells is reduced in CVID patients with autoimmunity and splenomegaly and is associated with expanded CD21lo B lymphocytes. *J Clin Immunol*. 2010; 30(2):292–300.
- Salzer U, et al. Mutations in TNFRSF13B encoding TACI are associated with common variable immunodeficiency in humans. *Nat Genet*. 2005; 37(8):820–828.
- Castigli E, et al. TACI is mutant in common variable immunodeficiency and IgA deficiency. *Nat Genet*. 2005;37(8):829–834.
- Wu Y, et al. Tumor necrosis factor (TNF) receptor superfamily member TACI is a high affinity receptor for TNF family members APRIL and BlyS. *J Biol Chem*. 2000;275(45):35478–35485.
- Knight AK, Radigan L, Marron T, Langs A, Zhang L, Cunningham-Rundles C. High serum levels of BAFF, APRIL, and TACI in common variable immunodeficiency. *Clin Immunol*. 2007;124(2):182–189.
- Kreuzaler M, et al. Soluble BAFF levels inversely correlate with peripheral B cell numbers and the expression of BAFF receptors. *J Immunol*. 2012; 188(1):497–503.
- Lesley R, et al. Reduced competitiveness of autoantigen-engaged B cells due to increased dependence on BAFF. *Immunity*. 2004;20(4):441–453.
- Thien M, et al. Excess BAFF rescues self-reactive B cells from peripheral deletion and allows them to enter forbidden follicular and marginal zone niches. *Immunity*. 2004;20(6):785–798.
- Gariyban L, Lobito AA, Siegel RM, Call ME, Wucherpfennig KW, Geha RS. Dominant-negative effect of the heterozygous C104R TACI mutation in common variable immunodeficiency (CVID). *J Clin Invest*. 2007;117(6):1550–1557.
- Chinen J, et al. Transmembrane activator and CAML interactor (TACI) haploinsufficiency results in B-cell dysfunction in patients with Smith-Magenis syndrome. *J Allergy Clin Immunol*. 2011; 127(6):1579–1586.
- Lee JJ, et al. The C104R mutant impairs the function of transmembrane activator and calcium modulator and cyclophilin ligand interactor (TACI) through haploinsufficiency. *J Allergy Clin Immunol*. 2010; 126(6):e1234–e1232.
- Pan-Hammarstrom Q, et al. Reexamining the role of TACI coding variants in common variable immunodeficiency and selective IgA deficiency. *Nat Genet*. 2007;39(4):429–430.
- Zhang L, et al. Transmembrane activator and calcium-modulating cyclophilin ligand interactor mutations in common variable immunodeficiency: clinical and immunologic outcomes in heterozygotes. *J Allergy Clin Immunol*. 2007; 120(5):1178–1185.
- Salzer U, et al. Relevance of biallelic versus monoallelic TNFRSF13B mutations in distinguishing disease-causing from risk-increasing TNFRSF13B variants in antibody deficiency syndromes. *Blood*. 2009; 113(9):1967–1976.
- Wardemann H, Yurasov S, Schaefer A, Young JW, Meffre E, Nussenzweig MC. Predominant autoantibody production by early human B cell precursors. *Science*. 2003;301(5638):1374–1377.
- Ng Y-S, Wardemann H, Chelnis J, Cunningham-Rundles C, Meffre E. Bruton’s tyrosine kinase (Btk) is essential for human B cell tolerance. *J Exp Med*. 2004; 200(7):927–934.
- Isnardi I, et al. IRAK-4- and MyD88-dependent pathways are essential for the removal of developing autoreactive B cells in humans. *Immunity*. 2008; 29(5):746–757.
- Menard L, et al. The PTPN22 allele encoding an R620W variant interferes with the removal of developing autoreactive B cells in humans. *J Clin Invest*. 2011;121(9):3635–3644.
- Sauer AV, Morbach H, Brigidia I, Ng YS, Aiuti A, Meffre E. Defective B cell tolerance in adenosine deaminase deficiency is corrected by gene therapy. *J Clin Invest*. 2012;122(6):2141–2152.
- Herve M, et al. CD40 ligand and MHC class II expression are essential for human peripheral B cell tolerance. *J Exp Med*. 2007;204(7):1583–1593.
- Meyers G, et al. Activation-induced cytidine deaminase (AID) is required for B-cell tolerance in humans. *Proc Natl Acad Sci U S A*. 2011; 108(28):11554–11559.
- Kinnunen T, et al. Accumulation of peripheral autoreactive B cells in the absence of functional human regulatory T cells. *Blood*. 2013;121(9):1595–1603.
- Romberg N, Ng YS, Cunningham-Rundles C, Meffre E. Common variable immunodeficiency patients with increased CD21-*lo* B cells suffer from altered receptor editing and defective central B cell tolerance. *Blood*. 2011;118(22):5977–5978.
- He B, et al. The transmembrane activator TACI triggers immunoglobulin class switching by activating B cells through the adaptor MyD88. *Nat Immunol*. 2010; 11(9):836–845.
- Kawai T, Adachi O, Ogawa T, Takeda K, Akira S. Unresponsiveness of MyD88-deficient mice to endotoxin. *Immunity*. 1999;11(1):115–122.
- Ewald SE, et al. The ectodomain of Toll-like receptor 9 is cleaved to generate a functional receptor. *Nature*. 2008;456(7222):658–662.
- Ewald SE, Engel A, Lee J, Wang M, Bogoy M, Barton GM. Nucleic acid recognition by Toll-like receptors is coupled to stepwise processing by cathepsins and asparagine endopeptidase. *J Exp Med*. 2011; 208(4):643–651.
- Moore PA, et al. BlyS: member of the tumor necrosis factor family and B lymphocyte stimulator. *Science*. 1999;285(5425):260–263.
- Dillon SR, Gross JA, Ansell SM, Novak AJ. An APRIL to remember: novel TNF ligands as therapeutic targets. *Nat Rev Drug Discov*. 2006;5(3):235–246.
- van Zelm MC, Szczepanski T, van der Burg M, van Dongen JJ. Replication history of B lymphocytes reveals homeostatic proliferation and extensive antigen-induced B cell expansion. *J Exp Med*. 2007; 204(3):645–655.
- Tan EM, et al. Range of antinuclear antibodies in “healthy” individuals. *Arthritis Rheum*. 1997; 40(9):1601–1611.
- Mietzner B, et al. Autoreactive IgG memory antibodies in patients with systemic lupus erythematosus arise from nonreactive and polyreactive precursors. *Proc Natl Acad Sci U S A*. 2008; 105(28):9727–9732.
- Morita R, et al. Human blood CXCR5(+)CD4(+) T cells are counterparts of T follicular cells and contain specific subsets that differentially support antibody secretion. *Immunity*. 2011;34(1):108–121.
- Nurieva RI, et al. Bcl6 mediates the development of T follicular helper cells. *Science*. 2009; 325(5943):1001–1005.
- Martinez-Gallo M, Radigan L, Almejún MB, Martínez-Pomar N, Matamoros N, Cunningham-Rundles C. TACI mutations and impaired B-cell function in subjects with CVID and healthy heterozygotes. *J Allergy Clin Immunol*. 2013;131(2):468–476.
- Cunningham-Rundles C, Radigan L, Knight AK, Zhang L, Bauer L, Nakazawa A. TLR9 activation is defective in common variable immunodeficiency. *J Immunol*. 2006;176(3):1978–1987.
- Samuels J, Ng YS, Coupillaud C, Paget D, Meffre E. Human B cell tolerance and its failure in rheumatoid arthritis. *Ann NY Acad Sci*. 2005;1062:116–126.
- Yurasov S, et al. Defective B cell tolerance checkpoints in systemic lupus erythematosus. *J Exp Med*. 2005; 201(5):703–711.
- Mackay F, et al. Mice transgenic for BAFF develop lymphocytic disorders along with autoimmune manifestations. *J Exp Med*. 1999;190(11):1697–1710.
- Sadanaga A, et al. Protection against autoimmune nephritis in MyD88-deficient MRL/lpr mice. *Arthritis Rheum*. 2007;56(5):1618–1628.
- Grimbacher B, et al. Homozygous loss of ICOS is associated with adult-onset common variable immunodeficiency. *Nat Immunol*. 2003;4(3):261–268.
- Bossaller L, et al. ICOS deficiency is associated with a severe reduction of CXCR5+CD4 germinal center Th cells. *J Immunol*. 2006;177(7):4927–4932.
- Warnatz K, et al. Human ICOS deficiency abrogates the germinal center reaction and provides a monogenic model for common variable immunodeficiency. *Blood*. 2006;107(8):3045–3052.
- Borriello F, et al. B7-1 B7-2 have overlapping, critical roles in immunoglobulin class switching germinal center formation. *Immunity*. 1997;6(3):303–313.
- Mak TW, et al. Costimulation through the inducible costimulator ligand is essential for both T helper and B cell functions in T cell-dependent B cell responses. *Nat Immunol*. 2003;4(8):765–772.
- Salek-Ardakani S, et al. B cell-specific expression of B7-2 is required for follicular Th cell function in response to vaccinia virus. *J Immunol*. 2011; 186(9):5294–5303.
- Ou X, Xu S, Lam KP. Deficiency in TNFRSF13B (TACI) expands T-follicular helper germinal center B cells via increased ICOS-ligand expression but impairs plasma cell survival. *Proc Natl Acad Sci U S A*. 2012; 109(38):15401–15406.
- Seshasayee D, Valdez P, Yan M, Dixit VM, Tumas D, Grewal IS. Loss of TACI causes fatal lymphoproliferation and autoimmunity, establishing TACI as an inhibitory BlyS receptor. *Immunity*. 2003; 18(2):279–288.
- Chung Y, et al. Follicular regulatory T cells expressing Foxp3 and Bcl-6 suppress germinal center reactions. *Nat Med*. 2011;17(8):983–988.
- Linterman MA, et al. Foxp3+ follicular regulatory T cells control the germinal center response. *Nat Med*. 2011;17(8):975–982.