



Published in final edited form as:

*Am J Physiol Regul Integr Comp Physiol*. 2008 April ; 294(4): R1213–R1219. doi:10.1152/ajpregu.00885.2007.

## Effect of Chronic Endothelin Receptor Antagonism on Cerebrovascular Function in Type 2 Diabetes

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### Abstract

Diabetes increases the risk of stroke and contributes to poor clinical outcomes in this patient population. Myogenic tone of the cerebral vasculature, including basilar arteries, plays a key role in controlling cerebral blood flow. Increased myogenic tone is ameliorated with ET receptor antagonism in Type 1 diabetes. However, the role of ET-1 and its receptors in cerebrovascular dysfunction in Type-2 diabetes, a common comorbidity in stroke patients, remains poorly elucidated. Therefore, we hypothesized that 1) cerebrovascular dysfunction occurs in the Goto-Kakizaki (GK) model of Type-2 diabetes, and 2) pharmacological antagonism of ET<sub>A</sub> receptors ameliorates while ET<sub>B</sub> receptor blockade augments vascular dysfunction. GK or control rats were treated with antagonists to either ET<sub>A</sub> (Atrasentan, 5mg/kg/d) or ET<sub>B</sub> (A-192621, 15 or 30 mg/kg/d) receptors for four weeks and vascular function of basilar arteries was assessed using a wire myograph. GK rats exhibited increased sensitivity to ET-1. ET<sub>A</sub> receptor antagonism caused a rightward shift indicating decreased sensitivity in diabetes while it increased sensitivity to ET-1 in control rats. Endothelium-dependent relaxation was impaired in diabetes. ET<sub>A</sub> receptor blockade restored relaxation to control values in the GK animals with no significant effect in Wistars and ET<sub>B</sub> blockade with 30 mg/kg/d A-192621 caused paradoxical constriction in diabetes. These studies demonstrate that cerebrovascular dysfunction occurs and may contribute to altered regulation of myogenic tone and cerebral blood flow in diabetes. While ET<sub>A</sub> receptors mediate vascular dysfunction, ET<sub>B</sub> receptors display differential effects. These results underscore the importance of ET<sub>A</sub>/ET<sub>B</sub> receptor balance and interactions in cerebrovascular dysfunction in diabetes.

### INTRODUCTION

Diabetes exists as an independent risk factor for cardiovascular disease (CVD) (29). Within the diabetic population, CVD is the leading cause of death accounting for 65–75% of all deaths (4). Studies have demonstrated that there is a strong correlation between diabetes and cerebrovascular disorders such as cerebral ischemia and stroke (5, 52). Diabetes not only increases the risk of stroke, but also contributes to increased stroke mortality and impaired recovery after stroke (3, 15, 16).

Regulation of vascular tone is important for maintenance of proper blood flow. In the cerebral circulation, changes in blood flow are buffered by the myogenic response to maintain cerebral blood flow. However, alterations to this system may be detrimental and

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could contribute to cerebrovascular disease. Studies have demonstrated increased myogenic tone in experimental diabetes (8, 9, 54). In addition to increased basal tone, cerebral arteries from diabetic animals exhibit diminished endothelium derived relaxation (2, 9, 34)

It is well established that the potent vasoconstrictor endothelin-1 (ET-1) is upregulated in the circulation in both clinical and experimental diabetes (7, 51). The effects of endothelin are mediated via two G-protein coupled receptors: ET<sub>A</sub> and ET<sub>B</sub> (17). ET<sub>A</sub> receptors reside on the smooth muscle cell (SMC) and produce vasoconstriction and mediate the proliferative effects of ET-1. Functional studies have suggested that different ET<sub>B</sub> receptor subtypes may exist (39, 40, 50). However, Mizuguchi et al demonstrated that the observed heterogeneous responses of the ET<sub>B</sub> receptor are abolished in ET<sub>B</sub> knockout mice suggesting that one gene is responsible for these actions (38). Inasmuch, ET<sub>B</sub> receptors do elicit different responses. ET<sub>B</sub> receptors on endothelial cells promote vasodilation via cGMP while VSMC ET<sub>B</sub> receptors elude ET<sub>A</sub>-like responses. Mounting evidence suggests that ET-1 is involved in the pathology of cerebrovascular disease (26). Studies have demonstrated enhanced contractile responses to ET-1 (1, 33) as well as a reduction of increased myogenic tone after ET receptor antagonism (9) in Type 1 diabetes. However, the relative roles of ET-1 and its receptors in cerebrovascular dysfunction in Type-2 diabetes, which is a common comorbidity in stroke patients, are poorly elucidated. Thus, we hypothesized that 1) cerebrovascular dysfunction occurs in the Goto-Kakizaki (GK) model of Type-2 diabetes, and 2) pharmacological antagonism of ET<sub>A</sub> receptors ameliorates whereas ET<sub>B</sub> receptor blockade augments vascular dysfunction.

## RESEARCH DESIGN AND METHODS

### Animals

All experiments were performed on male Wistar (Harlan, Indianapolis, IN) and GK (in-house bred, derived from the Tampa colony) rats (14, 49). The animals were housed at the Medical College of Georgia animal care facility that is approved by the American Association for Accreditation of Laboratory Animal Care. All protocols were approved by the Institutional Animal Care and Use Committee. Animals were fed standard rat chow and tap water ad-libitum until sacrifice at 18 weeks of age. Blood pressure was monitored either by telemetric method (as previously reported) (21) or via the tail cuff method which we have previously validated on telemetry implanted animals (10).

### Endothelin receptor blockade

GK rats develop hyperglycemia around 6–8 weeks of age. We have previously shown that by 18 weeks of age, this model displays significant cerebrovascular remodeling and ET<sub>A</sub> receptor antagonism started after 8 weeks of diabetes at week 14 can ameliorate this response (19). Using the same treatment paradigm, at 14 weeks of age, animals were divided into groups and treated for four weeks with highly selective antagonists for each receptor as follows: *ET<sub>A</sub> receptor blockade* – Atrasentan (Abbott Labs) 5 mg/kg/day in drinking water and *ET<sub>B</sub> receptor blockade* – A-192621 (Abbott Labs) 15 or 30 mg/kg/day by oral gavage split into two daily doses or vehicle (10). Vehicle for the A-192621 consisted of 83% deionized water, 10% Polyethylene Glycol-400, 5% ethanol, and 2% Cremaphor EL. Tap water was used as vehicle for the Atrasentan.

### Surgical procedure

Animals were anesthetized with sodium pentobarbital and exsanguinated via cardiac puncture. The brain was quickly excised for isolation of basilar arteries.

## Determination of vascular function

Isometric tension exerted by the vessels was recorded via a force transducer using the wire-myograph technique (Danish Myo Technologies, Denmark). The myograph chambers were filled with Krebs buffer (NaCl 118.3, NaHCO<sub>3</sub> 25, KCl 4.7, MgSO<sub>4</sub> 1.2, KH<sub>2</sub>PO<sub>4</sub> 1.2, CaCl<sub>2</sub> 1.5 and Dextrose 11.1 mM), gassed with 95% O<sub>2</sub> and 5% CO<sub>2</sub> and maintained at 37°C. Vessel segments were mounted in the chamber using 40 μm-thin wires and adjusted to a baseline tension of 0.4g. Cumulative dose response curves to ET-1 (0.1–100 nM) were generated and the force generated was expressed as % change of baseline. In a subset of vehicle-treated animals, basilar arteries were challenged with Sarafotoxin-6-c (S6c, 1–100 nM) with or without preincubation (30 min) with 1 μM BQ-123. In additional vehicle-treated rats, cumulative dose response (1 nM–1 μM) curves to serotonin (5-HT) were generated to determine if hyperreactivity is a general response. Endothelium-dependent relaxation to acetylcholine (ACh, 1 nM–1 μM) was assessed after vessels were constricted to 60% of the baseline tension with 5-HT or directly after ET-1 dose response. Sensitivity (EC<sub>50</sub>) and maximum response (R<sub>max</sub>) values were calculated from the respective dose-response equations.

## Statistical analysis

Results are given as mean ± SEM. For EC<sub>50</sub> and R<sub>max</sub> values, a two-way analysis of variance (ANOVA) was done to analyze disease and treatment effects with a post-hoc Bonferroni test. A repeated measures ANOVA was used to determine group differences (Diabetic vs. Control) across the ET-1 or ACh concentrations. Post-hoc group comparisons at each concentration used a Tukey's adjustment for the multiple comparisons. Graphpad Prism 4.0 was used for all statistical tests performed.

## RESULTS

### Animal data

Metabolic parameters of animals are listed in Table 1. Diabetic animals were significantly smaller than control rats in all treatment groups ( $p < 0.001$  vs control). Plasma ET-1 levels indicated a disease and drug interaction such that ET-1 was higher in diabetic rats for all treatment groups except for the A-192621 30 mg/kg/day group which was similar in control and diabetic rats. In addition, A-192621 (30 mg/kg/day) treatment caused a significant increase in plasma ET-1 levels in both control and diabetic rats when compared to vehicle, Atrasentan or the A-192621 (15 mg/kg/day) group.

### ET-1-mediated contractility

Basilar arteries of GK rats were hypersensitive to ET-1 (EC<sub>50</sub> 2.8 ± 1.6 vs 11.3 ± 1.7 nM in controls,  $p < 0.05$ ). (Fig. 1A, Table 2). Dose response curves in both control and diabetic animals showed significant differences between treatment protocols (Fig. 2A and B, Table 2). In control animals, Atrasentan treatment caused a paradoxical leftward shift that was statistically significant compared to vehicle or ET<sub>B</sub> receptor blockade groups. While A-192621 at 15 mg/kg dose had no effect on responses to ET-1, blockade with 30 mg/kg dose caused a rightward shift of the dose response curve. On the other hand, in the diabetic animals, blockade of the ET<sub>A</sub> receptor with Atrasentan attenuated the contractile response to ET-1 as indicated by the rightward shift. Interestingly, low dose ET<sub>B</sub> receptor blockade with 15 mg/kg A-192621 rendered basilar arteries more sensitive to ET-1. A dose response curve to 5-HT in vehicle treated control and diabetic animals showed no difference between R<sub>max</sub> (102 ± 8 in control vs 103 ± 10 in GK rats) or EC<sub>50</sub> values (15.3 ± 4.2 control vs 13.2 ± 2.5) in GK rats.

### Endothelium-dependent relaxation

Basilar arteries of GK rats exhibited impaired endothelium relaxation following pre-contraction with 5-HT. Basilar arteries of GKs relaxed only  $26 \pm 4\%$  in response to acetylcholine whereas the control rat arteries relaxed  $56 \pm 5\%$  (Fig. 1B, Table 2). In the control group, dose- response curves indicated significant differences between treatment arms (Fig. 3A). While Atrasentan had no effect on relaxation response, ET<sub>B</sub> receptor antagonism with A-192621 30 mg/kg significantly improved relaxation and A-192621 15 mg/kg group displayed about 40% relaxation across the Ach concentration range. In diabetic rats, however, Atrasentan, significantly improved relaxation. Interestingly, treatment with A-192621 30 mg/kg caused paradoxical constriction while treatment with A-192621 15 mg/kg had no effect (Fig. 3B).

### S6c-mediated reactivity

In order to determine whether there is an up-regulation of VSMC ET<sub>B</sub> receptors, contractile responses to S6c, an ET<sub>B</sub> receptor selective ligand, were determined (Fig. 4). Since previous studies suggested that a possible ET<sub>A</sub>-ET<sub>B</sub> receptor interaction prevents ET<sub>B</sub>-receptor responses to S6c, these studies were repeated in basilar artery segments preincubated with 1  $\mu$ M BQ-123. There was a small but significant increase in S6c-mediated constriction in the presence of BQ-123 in diabetic animals suggesting a possible involvement of VSMC ET<sub>B</sub> receptors in the contractile response.

## DISCUSSION

There are three major findings in the current study: first, basilar arteries were hypersensitive to ET-1 in a Type-2 diabetic rat model; second, acetylcholine induced relaxation of basilar arteries was impaired in diabetes; lastly, these effects are primarily mediated by the ET<sub>A</sub> receptor. These results build upon the past reports that showed cerebrovascular dysfunction in different models of diabetes and provide evidence for the involvement of ET-1 in cerebrovascular dysfunction in a non-obese, spontaneous model of Type 2 diabetes.

While there are many models of diabetes, most are either chemically induced Type-1 models or have co-morbid conditions such as hypertension, obesity and hyperlipidemia. The spontaneously diabetic GK rat is a non-obese, normotensive rat model originally developed from selective inbreeding of glucose intolerant Wistar rats (18). Previous studies have demonstrated that GK rats retain greater than 40% of their beta cell mass and have a reduction of post-prandial glucose when given the insulin secretagogue nateglinide (28, 30, 47, 48). In addition, our laboratory has previously shown that these rats have impaired glucose tolerance as compared to control Wistars (21). Therefore, the GK rat serves as an excellent model for studying the effects of hyperglycemia alone on cerebrovascular function in Type-2 diabetes.

We and others have observed elevated plasma ET-1 levels in both clinical and experimental diabetes (13, 21, 32, 45). Several studies have demonstrated enhanced contractile responses to ET-1 in aorta and peripheral arteries in diabetes (22, 27, 31). In addition, McIntyre et al reported increased sensitivity to ET-1 in subcutaneous resistance arteries from patients with Type-1 diabetes (37). In the present study, we have demonstrated an exacerbated contractile response of the rat basilar artery to ET-1 in diabetes. Matsumoto et al as well as Alabadi et al have both demonstrated similar results in the rat and rabbit basilar arteries respectively (1, 33). In contrast, Mayhan reported no differences in contractile responses to ET-1 in isolated rat basilar arteries (35). However, these discrepancies may be attributable to the differences in methodologies (e.g. in-vivo vs. in-vitro). Collectively, these previous studies were all performed in chemically induced Type-1 models of diabetes. To our knowledge, the present

study is the first demonstration of enhanced cerebrovascular contractile response to ET-1 in a model of Type-2 diabetes.

Previous studies using models of Type-2 diabetes have demonstrated that endothelium derived relaxation is impaired in aortas and the peripheral vasculature (6, 33, 42, 53). However, Type-1 models, such as the STZ rat and alloxan rabbit, have predominated in studies of the cerebral vessels (24, 34, 36). Only in recent years have investigators begun to study the effects of Type-2 diabetes on the cerebral circulation and thus far, there are but a handful of these studies (8, 11, 12, 25, 46). Schwaninger and Karagiannis both reported diminished vasodilation in response to acetylcholine in obese Zucker rats (OZR) (25, 46). However, the OZR is a model of Type-2 diabetes known to have co-morbid conditions such as hypertension and hyperlipidemia which may contribute to altered vascular states. More recently, diabetic *db/db* mice, characterized by hyperinsulinemia, severe hyperglycemia and obesity, were shown to exhibit similar reductions in endothelium dependent relaxation (8). In the present study, we found that Type-2 diabetes significantly impairs acetylcholine induced relaxation in serotonin precontracted basilar arteries. ET<sub>A</sub> receptor antagonism completely restored ACh induced relaxation in diabetic basilar arteries indicating that an activated endothelin system contributes to this impairment. This data, in accordance with previous studies done in varying models of diabetes, suggests that diabetes impairs vascular relaxation without regard to the etiology of the disease or other co-morbid conditions.

The vascular effects of ET-1 are mediated by two distinct receptor subtypes: ET<sub>A</sub> and ET<sub>B</sub>. In the present study, we have demonstrated hypersensitivity of the rat basilar artery to ET-1 in diabetes. ET<sub>A</sub> receptor blockade induced a significant rightward shift of the dose response curve indicating a decrease in sensitivity. Interestingly, in control animals, ET<sub>A</sub> receptor antagonism increased sensitivity to ET-1. Schilling et al. previously reported decreased sensitivity to ET-1 upon acute blockade of this receptor subtype suggesting that acute vs chronic blockade of the receptors yield different responses (44). Unexpectedly, higher dose blockade of the ET<sub>B</sub> receptor also produced a rightward shift of the dose response curve in a manner similar to ET<sub>A</sub> blockade. Previous studies in diabetic rabbit and rat basilar arteries have produced similar results using in-vitro inhibition of ET<sub>B</sub> receptors (1, 31, 33). Interestingly, Matsumoto et al showed that endothelial denudation of the diabetic basilar artery produced a significant leftward shift of the ET-1 dose response curve (33). In the current study, lower dose ET<sub>B</sub> blockade also caused a leftward shift possibly suggesting that removal of endothelial ET<sub>B</sub> receptors alone affects the dilatory actions of ET in diabetes, while complete blockade (both endothelial and VSMC ET<sub>B</sub>) alters constriction. It has been suggested that ET-1 mediated contraction in the cerebrovasculature may be, in part, mediated through crosstalk of the ET receptor subtypes. Zuccarello et al previously demonstrated that ET<sub>B</sub> mediated vasoconstriction relies on activation of the smooth muscle as well as the endothelial ET<sub>B</sub> receptors (55).

Vascular reactivity comprises both constriction and relaxation of blood vessels. In this study, ET<sub>A</sub> receptor blockade with Atrasentan caused a significant improvement of endothelium-dependent dilatation in diabetic rats and no significant effect in control animals suggesting a greater involvement of ET-1 in the regulation of vascular relaxation in diabetes. More interestingly, ET<sub>B</sub> receptor antagonism at 30 mg/kg dose resulted in completely opposite results in control vs diabetic rats, i.e, improving relaxation in control animals and causing paradoxical constriction in diabetic animals. Complete reversal of the relaxation response in A-192621(30 mg/kg)-treated GK rats suggests that endothelial ET<sub>B</sub> receptors are upregulated as a compensatory mechanism to offset impaired relaxation in diabetes and that blockade of these receptors ultimately results in decreased relaxation. Alternatively, these receptors are involved to a greater extent in the regulation of Ach-mediated dilatation

in diabetes and again antagonism of this receptor subtype causes paradoxical constriction to Ach.

Collectively, these studies strongly suggest involvement of both endothelial and VSMC ET<sub>B</sub> receptors in the regulation of vascular function in diabetes. Indeed, it has been demonstrated that diabetes does upregulate vascular ET<sub>B</sub> receptor expression. Mumtaz et al found increased ET<sub>B</sub> receptor density in the diabetic rabbit urinary bladder while Ikeda showed a significantly increased ET<sub>B</sub> gene expression in STZ diabetic rat adrenal glands (23). In 10 week old NOD mice, a Type-1 diabetes model, aortic ET<sub>B</sub> gene expression was significantly increased while ET<sub>A</sub> expression was unchanged (41). Conceivably, blockade of ET<sub>B</sub> receptors in a system which may be shifted more toward contractile responses, would produce effects similar to ET<sub>A</sub> blockade. It is also possible that ET<sub>A</sub>-ET<sub>B</sub> heterodimerization can affect the reactivity studies. Both homo and heterodimerization of the two ET receptors have been reported by functional as well as fluorescence resonance studies (19, 20, 43). It is suggested that when heterodimerized, the ET<sub>A</sub> receptor overrides ET<sub>B</sub> receptor activation and thus ET<sub>B</sub> receptor antagonism provides an ET<sub>A</sub> blockade-like effect. Harada et al. reported that due to ET<sub>A</sub> and ET<sub>B</sub> receptor heterodimerization, ET<sub>B</sub> receptors do not independently recognize ligands such as ET-1 and S6c unless ET<sub>A</sub> receptors are blocked with BQ-123. To test this hypothesis, we examined vascular responses to the ET<sub>B</sub>-selective agonist S6c in the presence and absence of ET<sub>A</sub> blockade with BQ-123. These studies demonstrated that S6c can induce vasoconstriction in the diabetes group when ET<sub>A</sub> receptors are antagonized thus indicating increased presence of VSMC ET<sub>B</sub> receptors.

Though our studies correlate with previously published data, certain limitations must be addressed. First, we examined reactivity of the basilar artery to ET-1 in all groups and to 5-HT only vehicle treated control and GK rats but not in ET receptor antagonist treatment groups. Others have found similar responses to ET-1 in diabetic rat and rabbit basilar arteries (1, 33). Second, due to the limited availability of ET<sub>B</sub> receptor antagonist as well as the basilar artery segments that can be obtained from one animal, especially in the low dose A-192621 group we had limited number of animals. We treated 5 animals in this group but in 2 animals vessels were completely unresponsive to any stimulus. For the same reasons, reactivity experiments could not be performed with endothelium denuded vessels.

## PERSPECTIVES AND SIGNIFICANCE

Diabetes induces vascular dysfunction in isolated rat basilar arteries in the form of increased sensitivity to ET-1 and diminished relaxation capacity to acetylcholine. Inasmuch, these effects appear to be, at least in part, due to an increased contribution of the ET<sub>A</sub> and smooth muscle ET<sub>B</sub> receptor activation. These alterations of cerebrovascular function may potentially underlie the increased propensity for cerebrovascular disease in diabetes.

## Acknowledgments

This work was supported by grants from NIH (HL076236, DK074385), American Heart Association Established Investigator Award and Philip Morris Research Award to Adviye Ergul and American Heart Association SouthEast Affiliate Pre-Doctoral Fellowship to Alex Harris.

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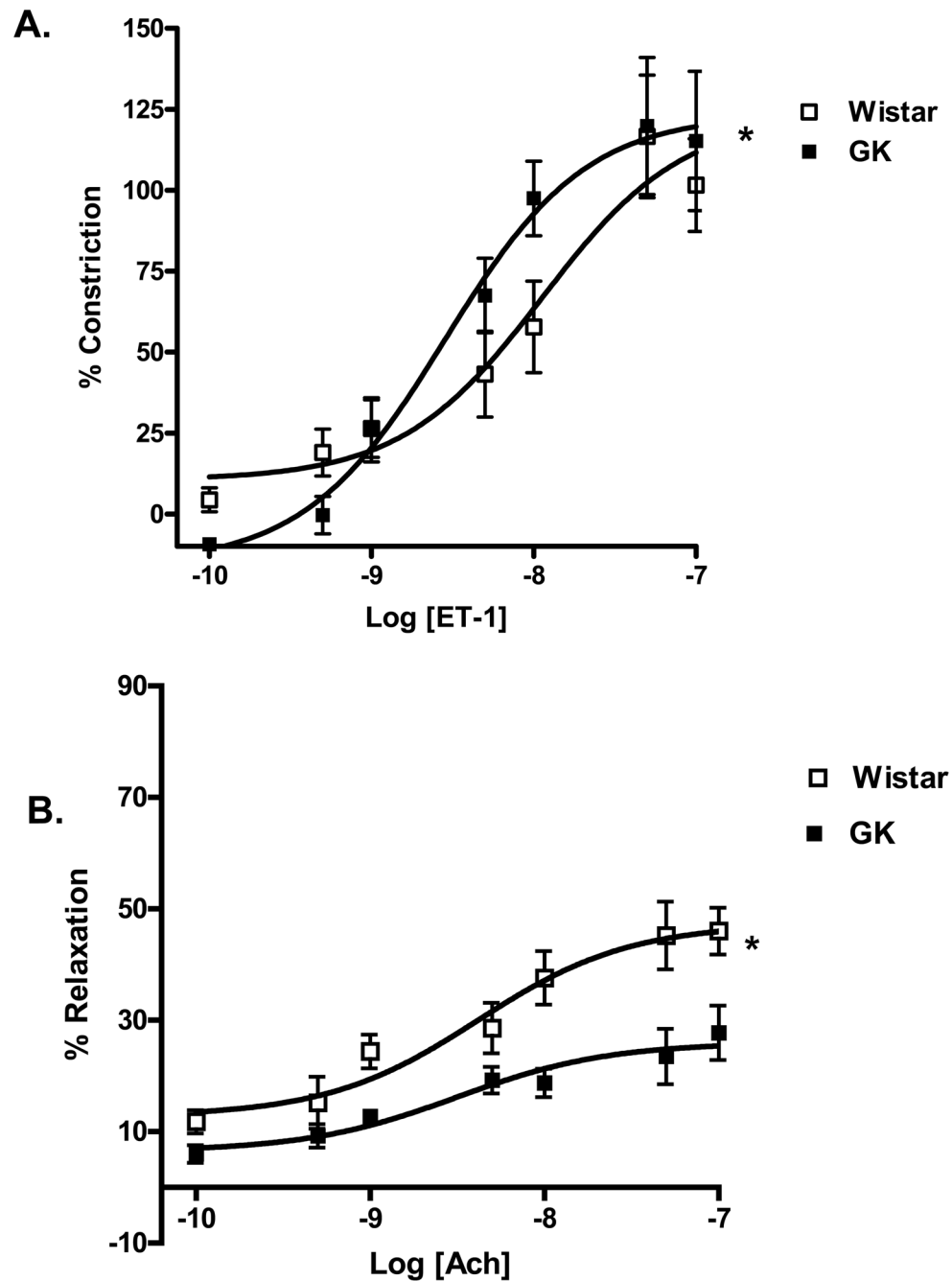
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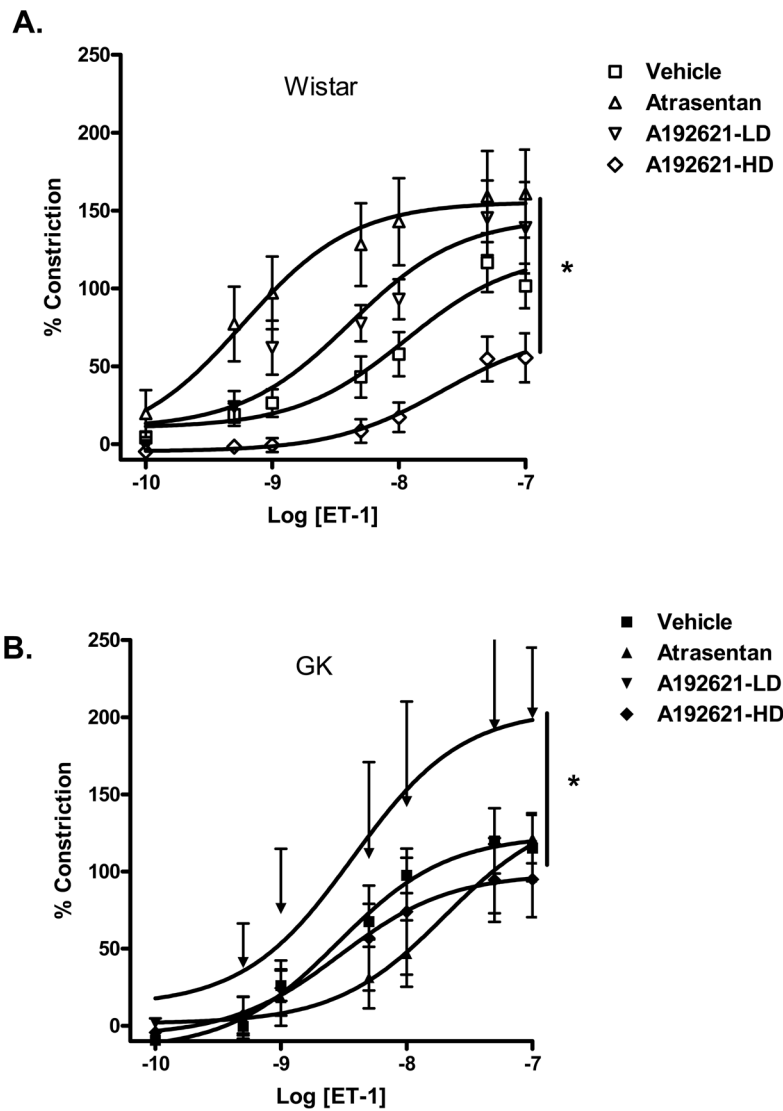
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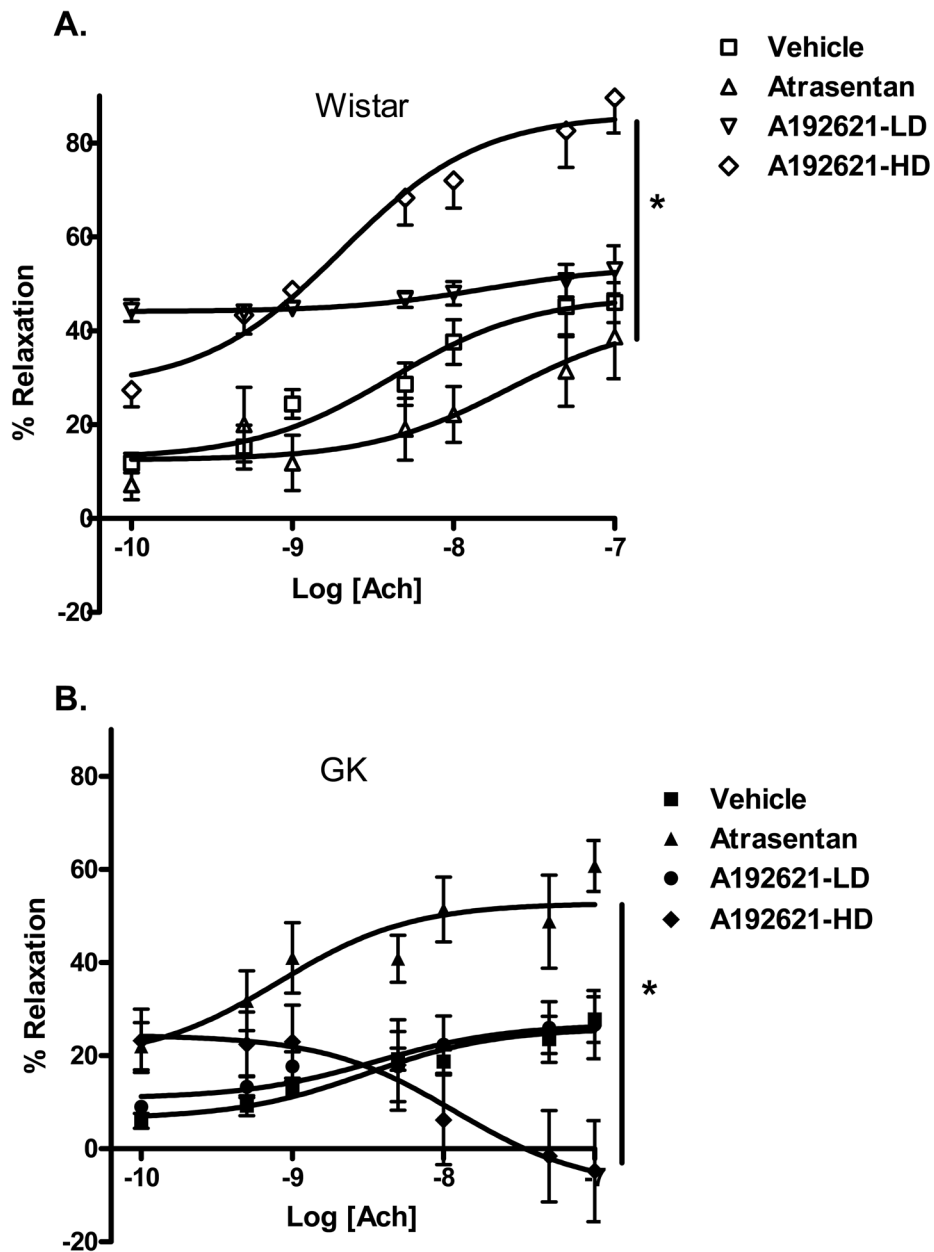
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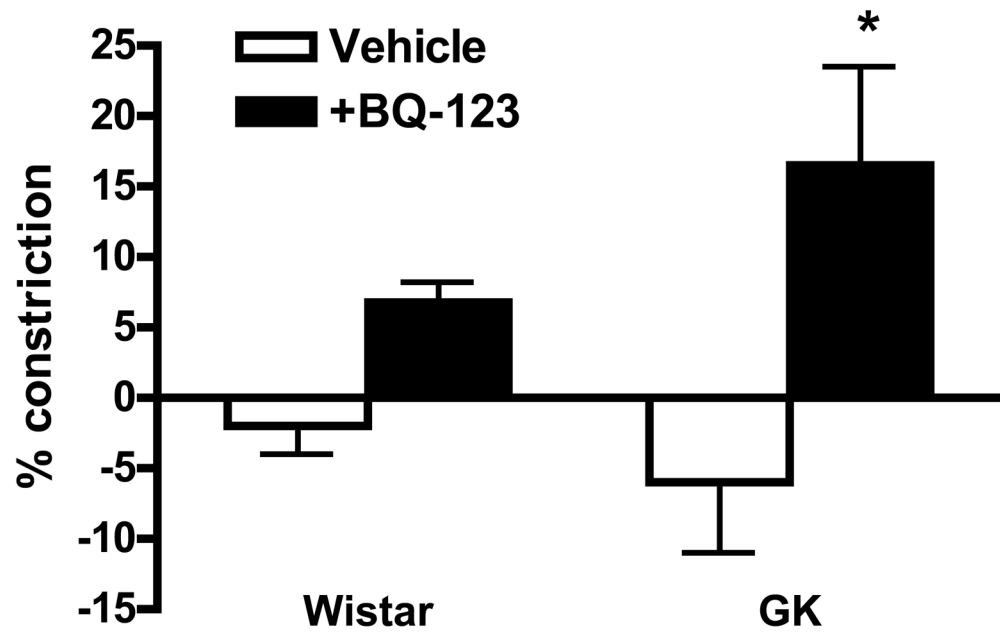
**FIG. 1.** Diabetes mediates vascular dysfunction of basilar arteries. **(A)** GK rats display hypersensitivity (EC<sub>50</sub>) to ET-1, although the magnitude of constriction did not differ. **(B)** Dose-response curves to acetylcholine demonstrated impaired endothelium dependent relaxation as compared to controls. Results are shown as mean  $\pm$  SEM, control = 8 and GK=6. \* $p$ <0.01

**FIG. 2.**

Effects of chronic ET receptor antagonism on ET-1-mediated contractility in control Wistar (A) and diabetic GK (B) rats. While  $ET_A$  receptor antagonism improved sensitivity to ET-1 in control animals ( $n=10$ ), it decreased ET-1 sensitivity in GKs ( $n=6$ ) as demonstrated by a rightward shift of the dose response curve.  $ET_B$  blockade with 30 mg/kg A-192621 caused a rightward shift of the dose response almost identical to that caused by Atrasentan. A-192621 15 and 30 mg/kg/day groups are shown as LD ( $n=3$  for both control and GK) and HD ( $n=5-6$  in control and GK), respectively. Results are shown as mean  $\pm$  SEM,  $*p<0.001$  repeated measures of ANOVA. Significant posthoc comparisons ( $P<0.05$ ) for control Wistar groups are vehicle vs atrasentan, vehicle vs HD, atrasentan vs LD, atrasentan vs HD, and LD vs HD. For GK groups, vehicle vs atrasentan, vehicle vs LD, atrasentan vs LD, LD vs HD were significantly different.



**FIG. 3.** Effects of chronic ET receptor antagonism on endothelium-dependent relaxation in control Wistar (A) and diabetic GK (B) rats.  $ET_A$  receptor blockade restored maximum ACh induced relaxation in GK basilar arteries to greater than control values. While high dose blockade caused paradoxical vasoconstriction in diabetic animals, it improved the dilatory response in control rats. Results are shown as mean  $\pm$  SEM, \* $p < 0.001$  repeated measures of ANOVA. Significant posthoc comparisons ( $P < 0.05$ ) for control Wistar groups are vehicle vs LD, vehicle vs HD, atrasentan vs LD, atrasentan vs HD and LD vs HD. For GK groups, vehicle vs atrasentan, atrasentan vs LD, and atrasentan vs HD were significantly different.



**FIG. 4.** S6c-mediated vascular reactivity. Basilar arteries were stimulated with S6c (1 nM- 1 $\mu$ M) with or without preincubation with 1 $\mu$ M BQ-123 for 30 min. Selective blockade of ET<sub>A</sub> receptors induced a significant increase in ET-1-mediated constriction in the GK group suggesting presence of ET<sub>B</sub> receptors on VSMC. P<0.01

TABLE 1

Metabolic parameters in treatment groups

	Wistar + Vehicle (n=8)	Wistar + Atrasentan (n=10)	Wistar + A-192621 15 mg/kg (n=5)	Wistar + A-192621 30 mg/kg (n=5)
Weight (g)	506 ± 13	474 ± 29	500 ± 13	406 ± 2 <sup>b</sup>
Glucose (mg/dl)	114 ± 5	89 ± 3	109 ± 4	104 ± 5
Insulin (ng/ml)	2.7 ± 0.6	2.1 ± 0.3	2.2 ± 0.5	2.2 ± .72
ET-1 (fmol/ml)	0.3 ± 0.1	0.4 ± 0.2	0.3 ± 0.2	1.6 ± 0.5 <sup>b</sup>
MAP (mmHg)	103 ± 5	105 ± 5	107 ± 2	121 ± 3 <sup>c</sup>
	GK+ Vehicle (n=8)	GK + Atrasentan (n=8)	GK + A-192621 15 mg/kg (n=5)	GK + A-192621 30 mg/kg (n=5)
Weight (g)	362 ± 8 <sup>a</sup>	357 ± 10 <sup>a</sup>	354 ± 6 <sup>a</sup>	322 ± 10 <sup>a</sup>
Glucose (mg/dl)	207 ± 33 <sup>a</sup>	172 ± 17 <sup>a</sup>	304 ± 17 <sup>a</sup>	173 ± 31 <sup>a</sup>
Insulin (ng/ml)	1.0 ± 0.3 <sup>a</sup>	0.7 ± 0.1 <sup>a</sup>	0.9 ± 0.2 <sup>a</sup>	0.6 ± 0.1 <sup>a</sup>
ET-1 (fmol/ml)	0.9 ± 0.2 <sup>a</sup>	1.2 ± 0.1 <sup>a</sup>	1.4 ± 0.2 <sup>a</sup>	2.3 ± 0.3 <sup>a,b</sup>
MAP (mmHg)	103 ± 4	106 ± 3	108 ± 3	116 ± 2 <sup>c</sup>

<sup>a</sup> p<0.001 vs Wistar,<sup>b</sup> p<0.001 vs vehicle, Atrasentan or A-192621 15 mg/kg,<sup>c</sup> p<0.01 vs vehicle, Atrasentan or A-192621 15 mg/kg

TABLE 2

Sensitivity ( $EC_{50}$ ) and magnitude ( $R_{max}$ ) of vascular responses to ET-1 and ACh in the absence or presence of ET receptor antagonists in basilar arteries of control Wistar and diabetic GK rats.

	Wistar + Vehicle (n=8)	Wistar + Atrasentan (n=10)	Wistar + A-192621 15 mg/kg (n=3)	Wistar + A-192621 30 mg/kg (n=5)
<b>ET-1 constriction</b>				
$EC_{50}$ (nM)	11.3 ± 1.7	0.6 ± 0.2 <sup>a,*</sup>	4.1 ± 1.8	32 ± 2.1 <sup>a,b</sup>
$R_{max}$ (%baseline)	99 ± 15	159 ± 28	139 ± 29 <sup>a</sup>	56 ± 16
<b>Ach relaxation</b>				
$EC_{50}$ (nM)	4.7 ± 1.5	8.1 ± 1.8	15.9 ± 4.0	2.0 ± 1.6
$R_{max}$ (% 5HT)	56 ± 5	46 ± 8	53 ± 5	90 ± 8 <sup>a,*</sup>
	GK+ Vehicle (n=6)	GK + Atrasentan(n=6)	GK + A-192621 15 mg/kg (n=3)	GK + A-192621 30 mg/kg (n=6)
<b>ET-1 constriction</b>				
$EC_{50}$ (nM)	2.8 ± 1.6 <sup>c</sup>	20 ± 2.2 <sup>c,d*</sup>	2.2 ± 3.6	2.6 ± 3.1
$R_{max}$ (%baseline)	115 ± 22	128 ± 14	203 ± 43 <sup>e</sup>	95 ± 25 <sup>f</sup>
<b>Ach relaxation</b>				
$EC_{50}$ (nM)	2.4 ± 2.0	3.8 ± 1.4	3.6 ± 4.3	4.3 ± 4.6
$R_{max}$ (% 5HT)	26 ± 4 <sup>c</sup>	64 ± 3 <sup>c</sup>	27 ± 7 <sup>c</sup>	-2 ± 14 <sup>d,g,*</sup>

<sup>a</sup> p<0.01 vs vehicle Wistar,

<sup>b</sup> p<0.001 vs Atrasentan or A-192621 15 mg/kg Wistar,

<sup>c</sup> p<0.01 vs vehicle Wistar,

<sup>d</sup> p<0.001 vs GK treatment groups,

<sup>e</sup> p<0.05 vs vehicle GK,

<sup>f</sup> p<0.05 vs A-192621 15 mg/kg GK,

<sup>h</sup> p<0.001 vs Wistar,

\* p<0.0001-disease-treatment interaction in Wistar vs GK rats.