Pyridine Extraction of Nocardial Acid Fastness

B. L. BEAMAN AND J. BURNSIDE

Department of Microbiology, Georgetown University Schools of Medicine and Dentistry, Washington, D.C. 20007

Received for publication 19 June 1973

Several strains of *Mycobacterium* and *Nocardia* were stained for acid fastness by using the method of Kinyoun. Duplicate, glutaraldehyde-fixed smears were extracted with fresh pyridine prior to staining. The results clearly demonstrated that both in vivo- and in vitro-grown *Nocardia* differed from *Mycobacterium* by becoming nonacid fast after extraction. This pyridine extraction of nocardial acid fastness should have diagnostic and taxonomic application.

The acid-fast staining property of members of the genus Nocardia is variable. Some strains are reported as being strongly acid fast (5-7), whereas others are readily decolorized with acid-alcohol (5-7). Furthermore, it is known that the acid fastness can be enhanced by growth in certain media such as milk (5). We found that many strains of N. asteroides that were not acid fast when grown in brain-heart infusion (BHI) became acid fast when grown in mice or on Middlebrook 7H10 agar supplemented with 1% glycerol (Beaman, unpublished data). Since nocardial filaments tend to fragment into rods and cocci that may be acid fast, especially in vivo, they may easily be mistaken for mycobacteria (9, 12). In fact, this has resulted in misdiagnoses (9, 12). Therefore, a rapid and simple method for distinguishing between the acid fastness of mycobacterium and that of nocardia would be of clinical and taxonomic value.

It is well established that the acid fastness of the leprosy bacillus, as well as nocardia, is not the same as that observed in most mycobacteria (2-6, 11). Fisher and Barksdale (3) demonstrated that the acid-fast staining of the leprosy bacillus could be removed by extracting the cells with pyridine, although this did not occur with other acid-fast mycobacteria. Later Convit and Pinardi (2) confirmed these findings, and Fisher and Barksdale extensively expanded the number of strains tested (4). Therefore, it appears that the acid fastness of the leprosy bacillus can be distinguished from that of other mycobacteria by using pyridine extraction (2-4).

The present investigation was undertaken to determine whether pyridine removed the acid fastness of nocardia. The bacterial strains used and their source are listed in Table 1. All strains of Nocardia were maintained on BHI agar: the Mycobacterium were grown on Middlebrook 7H10 agar. To obtain in vivo-grown nocardiae, Swiss Webster mice (4 weeks old) were injected intraperitoneally with approximately 10⁸ organisms suspended in saline. The bacteria used for the inoculum were grown in BHI broth for 24 h at 34 C in a Psychotherm environmental incubator (New Brunswick). Acid-fast organisms were obtained in vitro by growing them on Middlebrook 7H10 agar supplemented with 1% glycerol. Smears from either animal lesions or agar slants were air-dried and fixed for 30 min in 3% glutaraldehyde in Kellenberger buffer at pH 6.5 (8). The smears were rinsed in deionized water and dried. Duplicate sets of slides were either extracted in fresh pyridine (Baker analyzed reagent) at room temperature for 4 h or remained unextracted. All slides were stained by the Kinyoun acid-fast method (11) by using 1% concentrated hydrochloric acid in 70% ethanol (vol/vol) as the decolorizing agent. The smears were counterstained for 30 s in aqueous methylene blue. The results are shown in Tables 2 and 3.

Pyridine removed the acid fastness of all members of the genus Nocardia; however, Mycobacterium were not visibly affected by pyridine. Therefore, nocardiae can be distinguished from most mycobacteria by pyridine extraction of the acid fastness of these organisms. We encountered one problem with N. farcinica C. This organism was never acid fast when grown on BHI, but it was strongly acid fast when grown in mice or on Middlebrook 7H10 agar. This acid fastness was not always removed by pyridine extraction. In fact, some of the smears resisted extraction for 18 h, as did all of the strains of Mycobacterium. No other nocardial strain behaved in this manner. Many

Vol. 26, 1973

TABLE 1. Bacterial strains tested for pyridine extraction of acid fastness

Organism	Source
N. rubra 721-A	Isolated from the air (Univ. of Kansas, Lawrence)
N. asteroides-mahvi	Fatal human infection (T. A. Mahvi, South Caro- lina Medical College, Charlston)
N. asteroides-GUH-1	Human infection (V. Garagusi, Georgetown Univ. Hospital, Washing- ton, D.C.)
N. asteroides 10905	ATCC culture obtained from J. Rozanis, Univ. of Western Ontario, Lon- don, Canada.
N. asteroides 14759	ATCC ^a
N. asteroides 19247	ATCC
N. braziliensis 19296	ATCC
N. braziliensis 337	J. Rozanis, University of Western Ontario
N. corallina 4273	ATCC
N. pellegrino 159	J. Rozanis, University of Western Ontario
N. farcinica C	Originally from a bovine in- fection in Africa (From J. Rozanis, University of Western Ontario)
M.rhodochrous 4277	ATCC
M. smegmatis 14468	ATCC
M. smegmatis KU	University of Kansas Stock Culture
M. fortuitum 6841	ATCC
M. intracellulare GU	Georgetown University Stock Culture
M. kansasii GU	Georgetown University Stock Culture
M. tuberculosis H37Rv	ATCC
Tuberculous sputum	Confirmed active tuber- culosis, VA Hospital, Washington, D.C.

^a American Type Culture Collection, Rockville, Md.

investigators believe that N. farcinica is really a mycobacterium because of the kinds of mycolic acids it contains (10), and most researchers agree that it is at best a nomen dubium (1, 7, 10, 13). Even though the strain we used exhibited typical nocardial morphology, from the data presented in Tables 2 and 3 we take it to be more closely related to mycobacterium. Mycobacterium rhodochrous 4277 behaved more like a typical nocardia, and indeed, most studies have shown that many strains of M. rhodochrous were Nocardia (1, 10, 13). We found that many of the rapidly growing, soft colony nocardiae represented by N. corallina and N. pellegrino were not acid fast by any of

TABLE 2. Effect of pyridine on the acid fastness of Nocardia and Mycobacterium grown on Middlebrook

NOTES

7H10 agar supplemented with 1% glycerol

Organism	Unex- tracted (acid fastness)	4-h Pyridine extraction (acid fastness)
N. rubra 721-A(R)	+	-
N. asteroides-mahvi	+	-
N. asteroides GUH-1	+	-
N. asteroides 10905	+	-
N. asteroides 14759	+	-
N. asteroides 19247	+	-
N. braziliensis 19296	+	-
N. braziliensis 337	+	-
N. corallina 4273	(±) ^a	-
N. pellegrino 159	-	-
<i>N. farcinica</i> C	+	(±) ^a
M. rhodochrous 4277	+	-
M. smegmatis 14468	+	+
M. smegmatis KU	+	+
M. fortuitum 6841	+	+
M. intracellulare GU	+	+
M. kansasii GU	+	+
M. tuberculosis H37Rv	+	+

^a Sometimes acid fast but frequently not.

 TABLE 3. Effect of pyridine on the acid fastness of Nocardia and Mycobacterium grown in vivo

Organism	Unex- tracted (acid fastness)	4-h Pyridine extraction (acid fastness)
N. asteroides GUH-1 (mouse)	+	_
N. asteroides-mahvi (mouse)	+	-
N. asteroides 10905 (mouse)	+	_
N. braziliensis 337 (mouse)	+	-
N. rubra 721-A (S) (mouse)	+	-
N. farcinica C (mouse)	+	(±) ^a
Nocardia sp. DCV (human		
sputum) ^b	+	-
M. tuberculosis (human		
sputum)	+	+

^a The removal of acid fastness by pyridine was variable in both smears and paraffin sections.

[•] The patient had undefined pulmonary problems with the production of sputum containing acid-fast organisms. An organism having most of the physiological properties of *N. asteroides* was repeatedly isolated from this sputum; however, we were never able to demonstrate acid fastness in vitro. No *Mycobacterium* were isolated (supplied by D.C. Village Hospital, Washington, D.C.).

the methods employed. They served, therefore, as negative controls.

The data presented here seem to indicate that

pyridine extraction offers a simple method by which acid-fast nocardiae may be distinguished from most cultivatable mycobacteria. This should have both clinical and taxonomic application. We are presently investigating more nocardial strains, and we are attempting to determine the nature of the substance(s) removed by pyridine.

The expert assistance of B. O'Donnell during this study is greatly appreciated.

This investigation was supported primarily by a grant from the National Tuberculosis and Respiratory Disease Association. Portions also received support from Public Health Service grant AI-10542-02 from the National Institute of Allergy and Infectious Diseases.

LITERATURE CITED

- Bradley, S. G. 1973. Relationships among mycobacteria and nocardiae based upon deoxyribonucleic acid reassociation. J. Bacteriol. 113:645-651.
- Convit, J., and M. E. Pinardi. 1972. A simple method for the differentiation of *Mycobacterium leprae* from other mycobaeteria through routine staining techniques. Int. J. Lepr. 40:130-132.
- Fisher, C. A., and L. Barksdale. 1971. Elimination of the acid fastness but not the gram positivity of leprosy bacilli after extraction with pyridine. J. Bacteriol. 106:707-708.

- Fisher, C. A., and L. Barksdale. 1973. Cytochemical reactions of human leprosy bacilli and mycobacteria: ultrastructural implications. J. Bacteriol. 113:1389-1399.
- Georg, L. K., L. Ajello, C. McDurmont, and T. S. Hosty. 1961. The identification of Nocardia asteroides and Nocardia braziliensis. Amer. Rev. Resp. Dis. 84:337-347.
- Gordon, R. E., and J. M. Mihm. 1959. A comparison of Nocardia asteroides and Nocardia braziliensis. J. Gen. Microbiol. 20:129-135.
- Gordon, R. E., and J. M. Mihm. 1962. The type species of the genus Nocardia. J. Gen. Microbiol. 27:1-10.
- Kay, D., ed. 1967. Techniques for electron microscopy, 2nd ed. Blackwell Scientific Publications, Oxford, Great Britain.
- 9. Langevin, R. W., and S. Katz. 1964. Fulminating pulmonary nocardiosis. Dis. Chest 46:310-316.
- Lechevalier, M. P., A. C. Horan, and H. Lechevalier. 1971. Lipid composition in the classification of nocardiae and mycobacteria. J. Bacteriol. 105:313-318.
- Luna, L. G. 1968. Manual of histologic staining method of the Armed Forces Institute of Pathology. 3rd ed. McGraw Hill Book Co., New York.
- Neu, H. C., M. Silva, E. Hazen, and S. H. Rosenheim. 1967. Necrotizing nocardial pneumonitis. Ann. Intern. Med. 66:274-284.
- Ridell, M., and M. Norlin. 1973. Serological study of nocardia by using mycobacterial precipitation reference systems. J. Bacteriol. 113:1-7.