

Computational study of the activated O_H state in the catalytic mechanism of cytochrome *c* oxidase

Vivek Sharma^{a,b,1}, Kenneth D. Karlin^c, and Mårten Wikström^a

^aHelsinki Bioenergetics Group, Institute of Biotechnology, University of Helsinki, FI-00014, Helsinki, Finland; ^bDepartment of Physics, Tampere University of Technology, FI-33101, Tampere, Finland; and ^cDepartment of Chemistry, The Johns Hopkins University, Baltimore, MD 21218

Edited by Harry B. Gray, California Institute of Technology, Pasadena, CA, and approved September 10, 2013 (received for review November 22, 2012)

Complex IV in the respiratory chain of mitochondria and bacteria catalyzes reduction of molecular oxygen to water, and conserves much of the liberated free energy as an electrochemical proton gradient, which is used for the synthesis of ATP. Photochemical electron injection experiments have shown that reduction of the ferric/cupric state of the enzyme's binuclear heme a_3 /Cu_B center is coupled to proton pumping across the membrane, but only if oxidation of the reduced enzyme by O_2 immediately precedes electron injection. In contrast, reduction of the binuclear center in the "as-isolated" ferric/cupric enzyme is sluggish and without linkage to proton translocation. During turnover, the binuclear center apparently shuttles via a metastable but activated ferric/cupric state (O_H), which may decay into a more stable catalytically incompetent form (O) in the absence of electron donors. The structural basis for the difference between these two states has remained elusive, and is addressed here using computational methodology. The results support the notion that Cu_B[III] is either three-coordinated in the O_H state or shares an OH⁻ ligand with heme a_3 in a strained μ -hydroxo structure. Relaxation to state O is initiated by hydration of the binuclear site. The redox potential of Cu_B is expected, and found by density functional theory calculations, to be substantially higher in the O_H state than in state O . Our calculations also suggest that the neutral radical form of the cross-linked tyrosine in the binuclear site may be more significant in the catalytic cycle than suspected so far.

oxygen reduction | electron transfer

The respiratory chain in the mitochondria of all eukaryotes and many prokaryotes terminates in cytochrome *c* oxidase (CcO), which conserves free energy from the reduction of O_2 to water by linking it to proton pumping across the mitochondrial or bacterial membrane (1, 2). Oxygen reduction takes place at a binuclear center (BNC) consisting of a heme iron (heme a_3) and a copper ion (Cu_B) (Fig. 1 *A* and *B*) (1–4). Electrons from cytochrome *c* are supplied to this site in succession via two other metal sites, Cu_A and heme *a* (1, 2). Protons for consumption at the BNC, or to be pumped across the membrane, are taken from the negatively charged *N*-side of the membrane via the proton transfer pathways D and K, named after conserved aspartate and lysine residues, respectively (1–4). The D pathway has been proposed to be responsible for transferring all of the pumped protons, plus 2–3 “chemical” protons for consumption in O_2 reduction, whereas the K pathway is suggested to carry the remaining 1–2 chemical protons to the BNC (5). The D pathway terminates at a highly conserved glutamic acid residue in subunit I, E242 (numbering based on the bovine heart enzyme). From here the protons to be pumped across the membrane are suggested first to be carried to a pump site (also called a proton-loading site, PLS) in a redox-dependent manner (6–10). The proton at the PLS is generally agreed to be ejected to the positively charged P side of the membrane by electrostatic repulsion from uptake of a chemical proton into the BNC, as suggested independently by Morgan et al. (11) and Rich (12).

The main features of the catalytic cycle of CcO are fairly well established (Fig. 1C). Low-temperature work by Chance et al.

(13) identified the O_2 adduct of heme a_3 (compound **A**), and partial reversal of the catalytic reaction in mitochondria (14) helped to establish the subsequent intermediates **P** and **F**, both of which turned out to have heme a_3 in the ferryl state (1, 2, 15, 16). A conserved tyrosine in the BNC is covalently bonded to one of the three histidine ligands of Cu_B, and forms a neutral tyrosine radical in state **P_M** (Fig. 1 *A* and *C*) (2). The **P_R** state differs from **P_M** in having one more electron in the BNC (Fig. 1C and Fig. 2). **P_R** is formed directly after state **A** when the fully reduced (thus the R subscript) enzyme reacts with O_2 , and its structure may formally be written as shown in Fig. 1C (2, 17, 18). The next state of the BNC, called **F** (or **F_H** here, Fig. 1C), differs from **P_R** only by an additional proton, by which the unusual electron paramagnetic resonance (EPR) characteristics of the **P_R** state are lost (2, 19, 20). Recent infrared experiments have suggested that the cross-linked tyrosine in the BNC is anionic in both states **P_R** and **F** (21), on which basis we concluded that the extra proton in the **F** state is used to form water from the OH⁻ ligand of Cu_B (2, 17). Consequently, this water molecule is often formally drawn as a ligand of Cu_B in state **F** (17), but it may well dissociate from the copper (in which case we call it state **F_H** here; Fig. 2), which is one of the issues analyzed in the current work.

The Metastable O_H State

Further transfer of one electron and one proton to the BNC in the ferryl/cupric state **F** leads, in addition to translocation of one proton across the membrane (2), to formation of the ferric/cupric state. The ferric/cupric form of the BNC of “as-isolated” CcO (often called **O**, for oxidized) cannot be a true catalytic intermediate for several independent reasons (see below in this section), and must therefore be preceded by a metastable activated form of the ferric/cupric site (22, 23), which is denoted **O_H** here. Further reduction of state **O** is sluggish and not coupled to proton translocation (24, 25), whereas the catalytically relevant one-electron reduction of the activated **O_H** state is much faster, and at least 2–3 orders of magnitude faster than the decay of **O_H** into **O** (24, 25). Two more observations show that state **O** cannot be the catalytically active form. First, electron transfer from cytochrome *c* ($E_{m,7} \sim 270$ mV) to the BNC during reduction of

Significance

Density functional theory studies on the O_H state in the catalytic mechanism of cytochrome *c* oxidase are performed. The proposed structure of O_H state and its calculated characteristics are found to comply with the experimental data. The presented hypothesis solves, at least in part, the dilemma of the O_H and O states in the catalytic mechanism of cytochrome oxidase.

Author contributions: V.S. and M.W. designed research; V.S. performed research; V.S., K.D.K., and M.W. analyzed data; and V.S., K.D.K., and M.W. wrote the paper.

The authors declare no conflict of interest.

This article is a PNAS Direct Submission.

¹To whom correspondence should be addressed. E-mail: vivek.sharma@helsinki.fi.

This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10.1073/pnas.1220379110/-DCSupplemental.

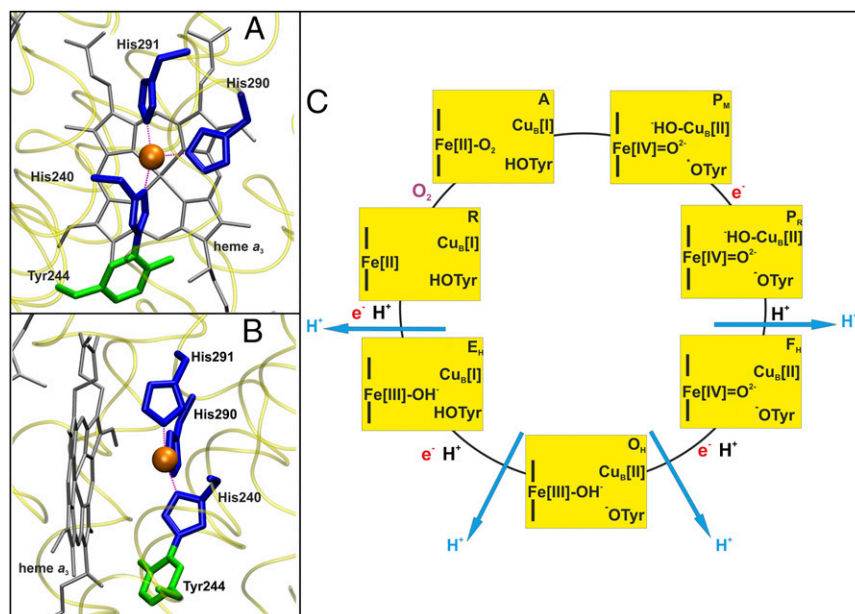


Fig. 1. (A and B) The BNC of CcO from *Bos taurus* (PDB ID code 1V54). The T-shaped three-coordinate structure (A) and near planarity (B) of the Cu_B center are shown. The copper, its histidine ligands, heme, and cross-linked tyrosine are displayed in orange, blue, gray, and green, respectively. (C) The proposed catalytic cycle is drawn clockwise. The oxidation states of Fe, Cu_B , and the cross-linked tyrosine in the BNC are shown, along with protonation states of the cross-linked tyrosine and metal ligands. The name of each state is shown in the upper right corner of each yellow rectangle. Blue arrows show proton pumping, whereas electrons and protons consumed at the BNC are shown in red and black, respectively. O_2 binds to the BNC in the $\text{R} \rightarrow \text{A}$ transition.

the O state could hardly be coupled to effective translocation of 2 charges/ e^- against a proton electrochemical gradient of at least 0.15 V across the membrane due to the low redox potential ($E_{m,7} \leq 0.4$ V) of state O (2, 26). Second, the sum of the individual free energy changes of the partial reactions of the catalytic cycle, determined at equilibrium, falls considerably short of the overall free energy change of the net reaction, viz. that of O_2 reduction to water (2, 26). Again, this shortfall is due to the low $E_{m,7}$ values of the $\text{O} \rightarrow \text{E}$ and $\text{E} \rightarrow \text{R}$ transitions found in equilibrium titrations. All these observations suggest involvement of a metastable activated form of the BNC (O_H) during turnover that has a high $E_{m,7}$, and that decays into a lower energy state O when an electron donor is lacking (e.g., during enzyme isolation).

It has turned out to be difficult to trap the metastable O_H state to study its structure, lifetime, and how it differs from state O (27, 28). However, electron photo-injection experiments indeed showed that Cu_B has a raised redox potential in the O_H state: the $E_{m,7}$ is at least 100 mV higher than the corresponding potentials of hemes a and a_3 (25, 29). This observation is interesting, because according to anaerobic redox titrations the $E_{m,7}$ values of heme a , Cu_B , and ferric/ferrous heme a_3 for adding the first electron to enzyme in state O are all similar, and do not exceed 0.4 V (30) (see also ref. 2). It matches the expectation that the O_H state has a raised redox potential and suggests that this is due to Cu_B . The lifetime of the O_H state (in the absence of an electron donor) can be estimated from such experiments, but not very accurately due to technical difficulties, giving a range from 0.2 s to 5 min.

Resonance Raman spectroscopy studies by Rousseau and co-workers (31) have established an intermediate beyond the F state, but before state O . The observed stretching frequency at 450 cm^{-1} is indicative of a distal heme iron-bound hydroxide that is strongly hydrogen-bonded. The data also showed that the ferric heme iron is high spin in this state, which is consistent with the strongly hydrogen-bonded distal hydroxide. A state with hydroxide-liganded high-spin ferric heme a_3 is therefore a possible candidate for state O_H (17), as concluded originally (31), and

this is the rationale for the subscript H (but see footnote in Table 1). The half-life of the 450 cm^{-1} species was ~ 1 ms (31), which is too short for the O_H state. However, this short lifetime may only be apparent, because of exchange of the isotopically labeled hydroxide ligand with water (31). Preliminary data by Han et al. (31) indicated that the ferric hydroxide species was no longer present on a seconds time scale, which would considerably reduce the upper lifetime limit for O_H if identical to the ferric hydroxide species, and also tends to exclude a hydroxide ligand of heme iron in the O state. Early spectroscopic data indicated that heme a_3 iron is six-coordinated high-spin ferric in state O (32), which suggests that the distal ligand is water.

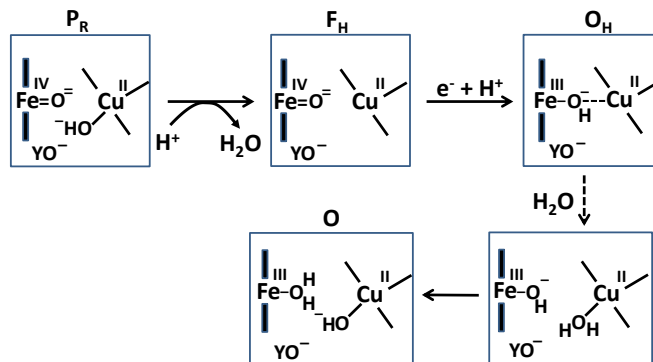


Fig. 2. Sequence of events leading to the formation of states O_H and O . The protonation of the $-\text{OH}$ ligand of Cu_B in P_R leads to the formation of an activated state F_H , in which Cu is three-coordinated. Subsequent reduction and protonation of the BNC forms state O_H , which has a strained μ -hydroxo-bridged structure (dashed line indicates a weak bonding between Cu_B and the $-\text{OH}$ ligand of Fe; Table S2). Diffusion of a water molecule toward Cu_B , structural relaxation (expansion) of the active site, and hydrogen bonding rearrangements (all depicted as a dashed vertical arrow) would lead to the formation of O from O_H .

Table 1. Energies (E) of various model systems in different spin states (in kcal/mol)

Models	E	Models	E
O_H^1 *	16.1	O^1	0.2
O_H^2	28.2	O^2	0
O_H^3	19.4	O^3	3.8
$F_H^{1/2}$	5.0	$F^{1/2}$	0
$F_H^{3/2}$	5.0	$F^{3/2}$	0
$F_H^{5/2}$	17.8	$F^{5/2}$	14.71
$O_{H,R}^{1/2}$	6.9	$O_R^{1/2}$	0
$O_{H,R}^{3/2}$	29.2	$O_R^{3/2}$	25.1
$O_{H,R}^{5/2}$	13.8	$O_R^{5/2}$	9.2
$F_{H,R}^1$	2.5	F_R^1	1.7
$F_{H,R}^2$	14.6	F_R^2	24.3
$F_{H,R}^3$	47.8	F_R^3	0

The energies (E) reported in the table are obtained at the B3LYP (Becke, three-parameter, Lee-Yang-Parr)/def2-TZVP (triple zeta valence plus polarization)/dielectric constant $\epsilon = 4$ level of theory. The energies of all O_H and O , F_H , and F , $O_{H,R}$ and O_R , and $F_{H,R}$ and F_R spin states are relative to the corresponding lowest energy O , F , O_R , and F_R states (i.e., O^2 , $F^{1/2}$, $O_R^{1/2}$, and F_R^3). *The H subscript in O_H state denotes the hydroxyl ligand of heme a_3 Fe (see *The Metastable O_H State*). For all other states, subscript H is retained for consistency and to distinguish them from “inactivated” four-coordinated states (O , F , O_R , and F_R).

Rationale

Taken together, the experimental evidence suggests that the BNC in the catalytically relevant O_H state may have high-spin ferric heme a_3 with a strongly hydrogen-bonded distal hydroxide ligand, and cupric Cu_B with a raised redox potential. The cross-linked tyrosine is unprotonated (21) so that the cupric Cu_B will have either a water molecule as the fourth ligand or no fourth ligand at all. The reason for this is that the oxidation of the fully reduced enzyme is coupled to net uptake of no more than two protons, which occurs in the steps $P \rightarrow F$ and $F \rightarrow O$ (Fig. 1C). The current work was spurred by the observation of a near-planar T-shaped geometry for Cu_B (Fig. 1A and B) with its three nitrogenous histidine ligands (3, 4), by the known rather T-shaped tris-histidyl copper (Cu_H) coordination in the enzyme peptidyl-glycine α -hydroxylating monooxygenase (33), and by the recognition of the fact that inorganic cuprous complexes prefer a three-coordinated planar geometry, whereas cupric complexes prefer a higher coordination number (34–36). Due to this preference, a near-planar three-coordinated form of $Cu_B[II]$ would be expected to have a much higher $E_{m,7}$ than the four-coordinated variant, a prediction that is reminiscent of an earlier “rack” hypothesis (36). If so, the requirements of a raised $E_{m,7}$ of the O_H state and the experimental observation of an elevated $E_{m,7}$ of Cu_B in that state would all be satisfied. We therefore decided to test this hypothesis using computational methodologies.

Results

Based on the above rationale, the O_H/O difference might simply relate to the absence/presence of a fourth oxygenous ligand of Cu_B . To test this possibility we constructed model systems corresponding to the two states (O_H and O), and their precursors F_H and F , as described in *Methods* (SI Methods). We then systematically compared the properties of the binuclear site structures in these states. In addition, we calculated redox potentials—that is, we gathered information on the relative electron affinities of these states by analyzing the corresponding structures with an additional electron (i.e., F_R , O_R vs. $F_{H,R}$, $O_{H,R}$), but before

subsequent protonation. These latter states may be considered putative intermediates between states $F(F_H)$ and $O(O_H)$, and between $O(O_H)$ and $E(E_H)$, in the catalytic cycle (Fig. 1C). The total charge and spin of the model systems tested are listed in Table S1, and Table 1 lists the calculated energies.

The constructed O_H states are consistently found to be 12–19 kcal/mol higher in energy than the corresponding O states (Table 1). Neglecting zero-point vibrational contributions, and assuming an entropy loss of 3 kcal/mol upon binding of a water molecule to $Cu_B[II]$ (37) in state O (Fig. 2 and Fig. S1), the O_H state is still 9–17 kcal/mol higher in free energy than the O state.

Geometric analysis of the optimized structures of the different redox states (Table S2) shows a root mean square deviation of ≤ 0.7 Å after superposition, which suggests high overall structural similarity. In most of the states where Cu_B lacks a fourth oxygenous ligand (O_H , F_H , $O_{H,R}$, $F_{H,R}$), the copper attains near-planar geometry with the dihedral angle $N_{\text{His}244}-N_{\text{His}291}-N_{\text{His}290}-Cu_B$ approaching 10° , which is about 10° smaller than what is observed in the four-coordinate O , F , O_R , and F_R states (Table S2). The Fe–O bond lengths in the O and O_H states, and in the F and F_H states (Table S2), are also in agreement with previously reported bond lengths in similar systems (38–40).

Redox potentials calculated for the lowest energy states are listed in Table 2. Even though such density functional theory (DFT) calculations are challenging (41, 42), our data systematically suggests a higher redox potential of the BNC in the activated O_H and F_H states relative to the corresponding O and F states. It is noteworthy that this effect is much larger for the $O/O_H \rightarrow O_R/O_{H,R}$ transitions (~ 0.43 V) than for the corresponding transitions of states F/F_H into $F_R/F_{H,R}$ (~ 0.18 V). Analysis of the optimized geometries of the O states provides additional reasons for their lower redox potential. In the low energy O^2 and O^3 states, the hydrogen on the water molecule ligating the Cu_B ion (H2) shifts toward the distal hydroxyl ligand of Fe_{a3} (Fig. S1, see also Table S2) in such a way that the formal structure should

Table 2. Redox potentials (E°) of different redox couples (in volts)

Redox couple*	E°
$O_H^1/O_{H,R}^{5/2}$	0.037
$O_H^1/O_{H,R}^{1/2}$	0.338
$O_H^3/O_{H,R}^{5/2}$	0.180
$O_H^3/O_{H,R}^{1/2}$	0.481
$O^1/O_R^{5/2}$	−0.452
$O^1/O_R^{1/2}$	−0.053
$O^2/O_R^{5/2}$	−0.460
$O^2/O_R^{1/2}$	−0.060
$O^3/O_R^{5/2}$	−0.293
$O^3/O_R^{1/2}$	0.105
$F_H^{1/2}/F_{H,R}^1$	0.144
$F_H^{3/2}/F_{H,R}^1$	0.146
$F^{1/2}/F_R^1$	−0.037
$F^{3/2}/F_R^1$	−0.036
$F^{1/2}/F_R^3$	0.034
$F^{3/2}/F_R^3$	0.036

*Redox potentials (E°) are evaluated for the low energy spin states of reduced/oxidized forms in Table 1.

†Energies used in evaluating the redox potentials are calculated at the B3LYP/def2-TZVP/ $\epsilon = 4$ level of theory. It is especially noteworthy that these are the midpoint redox potentials without charge-compensating protonation.

be described as $\text{Fe}[\text{III}]\text{-OH}_2$, $\text{Cu}_B[\text{II}]\text{-OH}^-$. The OH^- ligand of Cu_B is the likely cause for the lowered redox potentials (Table 2). Such a scenario is not possible in the O_H states, where no water molecule ligates the Cu_B ion, which thereby maintains planarity and a high redox potential (but see below in this section). In support of our computational results, elevated $\text{Cu}(\text{II})/\text{Cu}(\text{I})$ redox potentials, ≥ 0.7 V vs. normal hydrogen electrode (NHE), are in fact known for rare examples where synthetic coordination complexes with copper ion are found to be ligated by three neutral nitrogenous ligand donors in a three-coordinate near-planar coordination geometry (35, 43, 44).

Analysis of the electronic structures of the O_H and O states also reveals critical differences. The extensive delocalized spin density on the cross-linked tyrosine in the O_H states is markedly different from the near absence of such spin density on the same moiety in the O states (Fig. S2 and Table S3). Consequently (although not shown in Fig. 1C), the Cu_B system in the O_H state may have significant characteristics of a cuprous/neutral tyrosyl radical state, which may contribute to its higher free energy and higher redox potential. Overall, analyses of energies (Table 1), redox potentials (Table 2), and geometrical data (Table S2) together suggest that the likely reasons for the higher free energy and higher redox potential of the O_H state relative to state O are geometrical as well as electronic in nature.

To understand why the metastable O_H state would preferentially form in the catalytic cycle, instead of the lower energy O state, we also analyzed the preceding F state of the catalytic cycle, as well as its one-electron reduced derivative (F_R). In contrast to the large energy differences obtained between the O_H and O states, the difference between the F and F_R states is much smaller, ca. 5 kcal/mol (Table 1), and only around 2–3 kcal/mol when entropic effects are included (37). Due to near degeneracy of the F_H and F states, they would be equally favored from a thermodynamic viewpoint. However, the higher redox potential of the F_H state (Table 2), and hence the higher driving force, may render it kinetically more competent than F in accepting the electron. Geometric analysis (Table S2) suggests that the water molecule is indeed weakly bonded to the Cu_B ion in different spin states of F , with $d(\text{Cu-O}) \sim 2.2$ Å (45, 46). In contrast, in forming the O state, the copper-bound water is effectively stabilized as a strongly bonded hydroxide ligand due to proton transfer to the distal OH^- ligand of iron, as depicted in Fig. 2, Fig. S1, and Table S2.

The spin density analysis of the O/F and O_H/F_H states and their one-electron-reduced counterparts, O_R/F_R and $\text{O}_{H,R}/\text{F}_{H,R}$, revealed some further interesting details. Upon one-electron reduction of O_H to $\text{O}_{H,R}$, the electron transfer primarily occurs to the neutral tyrosine radical, whereas it takes place to $\text{Cu}_B[\text{II}]$ in the case of reduction of state O (Table S3). This is likely one of the reasons for the large redox potential difference observed between the redox couples $\text{O}_H/\text{O}_{H,R}$ and O/O_R (Table 2). Similarly, there is more spin density on the active site tyrosine in the two low-energy F_H states, relative to the corresponding spin states of F (Table S3). Hence, F_H may at least in part be characterized as $\text{Fe}[\text{IV}] = \text{O Cu}[\text{I}]$, Tyr-O^* , where the copper is reduced and the tyrosine is in the form of the neutral radical. When this state receives an electron to yield $\text{F}_{H,R}$, the lowest energy states all have contributions from $\text{Cu}[\text{I}]$ and Tyr-O^- , indicating that a significant part of this process has been the transfer of the electron to the tyrosine (Table S3). In contrast, the lowest energy F_R state is one where part of the reduction has occurred on the heme iron ($\text{Fe}[\text{III}]\text{-OH}^- \text{Cu}[\text{II}]\text{-OH}^-$, TyrO^-), allowing local protonation of the oxo ligand by the water ligand of the copper.

The above results are in good agreement with the expected structure of state O_H , but a detailed structural analysis yields an unexpected result. In the O_H^3 state the heme iron is slightly pulled out of the heme plane in the distal direction, and the

copper is also pulled toward the iron, so that a weak bond may be formed between the ferric OH ligand and the copper (Fig. 2 and Table S2). Such a μ -hydroxo bridge between iron and copper has been anticipated in the past for the BNC, and model compounds of this kind have been produced (47, 48). Note that this state no longer conforms to the idea of three-coordinate cupric copper, nor to a planar copper geometry, and that the distance between the proximal histidine nitrogen and heme iron has lengthened considerably (Table S2 and Discussion).

The above analysis does not address the issue pertaining to the lifetime of the O_H state. To be coupled to proton translocation, the O_H state must receive an electron to form E_H before it relaxes to the O state. The $\text{O}_H \rightarrow \text{O}$ relaxation would hence have to have an activation energy barrier of at least ~ 16 kcal/mol at 310 K. According to our model of O_H , the relaxation barrier would comprise at the least hydrogen-bonding rearrangements as well as diffusion of a water molecule (49) toward Cu_B . Structural rearrangements (50) may further be required, and the calculated optimum structure of the O_H^3 state indeed exhibits a much tighter BNC than those found in the crystal structures (3, 4, 51), the Fe–Cu distance being only 4.01 Å (Table S2). The thermodynamic costs associated with the H-bonding and structural rearrangements are likely to contribute to the lifetime of O_H (SI Methods), which would be significantly longer if a water molecule is recruited from outside the catalytic cavity (49). Finally, when considering the lifetime of the O_H state and its relaxation to state O , it should be noted that the latter could be a catalytically relevant state in which the proton-translocating efficiency is reduced, for example under conditions of a high protonmotive force (8).

Discussion

The structural basis of the heterogeneity of the oxidized C_cO , especially the difference between the form encountered immediately after oxidation of the reduced enzyme by O_2 (O_H) and the as-isolated form (O), has so far escaped recognition. A previous spectroscopic study on the bovine heart enzyme revealed no difference between the two states (27). However, in that study the observed rate of reduction of the BNC in the presumed O_H state was much slower than reported in ref. 52 for the bovine and in ref. 29 for the *Paracoccus* enzyme.

Based on the known chemistry of inorganic copper complexes and the available experimental observations, we have here considered the simple hypothesis that state O_H may be a dehydrated form of state O such that $\text{Cu}_B[\text{III}]$ is three-coordinated planar in the former case, but has a fourth oxygenous ligand in the latter. This hypothesis was tested by DFT calculations.

With the exception of unlikely $S = 0$ states, and after correction for an entropy change due to water dissociation, the calculated free energy of the O_H states lies ca. 9–17 kcal/mol higher than for states O (Table 1), in good agreement with O_H being an “activated” metastable form of the ferric/cupric center. Moreover, due to trigonal near-planar geometry of Cu_B , the cuprous form is favored over the cupric, and the redox potential of the $\text{O}_H/\text{O}_{H,R}$ couple is much higher than for the four-coordinated O/O_R couple (Table 2). It is important to note that these calculated redox potentials differ substantially from the experimentally determined values (26) due to the fact that the latter are equilibrium potentials where charge compensation by protonation has occurred. The calculated $\text{O}_H/\text{O}_{H,R}$ redox potentials are consistently ~ 0.4 V higher than for the O/O_R redox couples, which is in accord with the experimental observations of an elevated redox potential of Cu_B in the O_H state (29, 52). The 0.4 V higher redox potential of the activated form solves the dilemma, at least in part, of an apparently far too low redox potential of the $\text{O} \rightarrow \text{E}$ transition to be consistent with proton pumping, and with the overall energetics of the catalytic cycle (2) (see *The Metastable O_H State*).

The notion of a trigonal near-planar Cu_B in the F_H and O_H states has additional interesting corollaries. Electron transfer to the BNC in the P_M state is primarily expected to yield P_R (Fig. 1C), even though this has not been explicitly shown in experiments (2, 17). Instead, P_R is known to form directly from intermediate **A** in experiments where the fully reduced enzyme reacts with O_2 (opening paragraphs); at the same time, the PLS is loaded by an internal proton transfer from E242 (2) (not shown in Fig. 1C). The notion of a neutral radical form of the cross-linked tyrosine is strong for state P_M (17), and the electron transfer to this state reduces the radical to the tyrosinate anion in P_R : Resonance Raman, EPR, and infrared spectroscopic data prove that P_R has ferryl heme iron, cupric copper, and tyrosinate, respectively (see references in refs. 2, 17). P_R is well known to be followed by proton uptake from the solution to form state **F**. Here, our DFT calculations suggest that if the water molecule formed in this reaction dissociates from Cu_B (forming F_H ; Figs. 1C and 2), the tyrosine tends once again to have some characteristics of a neutral radical (Table S3). The next electron input would in part occur to this radical (to yield $\text{F}_{H,R}$), followed by net proton uptake to yield the O_H state. Even in state O_H at least part of the population seems to have reduced Cu_B and a neutral tyrosine radical (Table S3). Hence, the O_H state may differ from **O**, not only with regard to the coordination of Cu_B but also with respect to the redox nature of the tyrosine and copper. Altogether, our results suggest a more central role in catalysis of the neutral radical state of the cross-linked tyrosine than previously expected.

FTIR data (21) suggested that the cross-linked tyrosine is a deprotonated tyrosinate in both the **F** and **O** states. The proton taken up into the BNC upon reduction of state **F** is thus likely to form a ferric heme a_3 hydroxide (Fig. 2), which agrees very well with the resonance Raman data of Han et al. (31), according to which the hydroxide ligand is strongly hydrogen-bonded to yield the Raman resonance at 450 cm^{-1} and a high-spin ferric heme ($S = 5/2$). We assume that the hydrogen bonding is to one or two water molecules between the hydroxide and the tyrosinate (51) (Fig. S1). Interestingly, the most likely states corresponding to our view of O_H have high-spin ferric heme iron ($S = 5/2$) and $\text{Cu}_B[\text{II}]$ ($S = 1/2$), for which we have calculated the ferromagnetically coupled form ($S = 3$; Table S3), but the antiferromagnetically coupled form ($S = 2$) is likely to have very similar energetics and structural properties (Table S4). Unexpectedly, this form of O_H appears to be a μ -hydroxo-bridged compound with a very compact BNC. Thus, Cu_B actually has a weak fourth ligand that it shares with the high-spin heme iron, the copper geometry is no longer planar, the iron is pulled out of the heme plane on the distal side, and the distance between Fe and the proximal histidine nitrogen is very long (Table S2). In this case our original hypothesis of a three-coordinated Cu_B in the O_H state fails, and this state is instead “strained” with a very short distance between Fe and Cu. The heme iron appears to be five-coordinated as

anticipated from μ -hydroxo-bridged models of the BNC (47, 48). However, the Raman bands typical of five-coordinated high-spin hemes (53) are not observed in the spectra of the ferric heme a_3 hydroxide intermediate described by Han et al. (31), which suggest six-coordination in that state. The true structure of state O_H may thus be a compromise where the bond still persists between the iron and the proximal histidine nitrogen.

The calculations presented here do not address the existence of a similar activated **E** state. The experimental evidence for E_H is not as strong as for O_H (24, 25), and part of the driving force for the proton pumping $\text{E}/\text{E}_H \rightarrow \text{R}$ reaction is likely to emerge from the exergonic $\text{R} \rightarrow \text{P}_M$ transition (~ 10 kcal/mol based on ref. 54).

In summary, the primary finding here is that a subtle change in the hydration of the BNC could account for the difference between the metastable catalytically active O_H state and its low-energy counterpart, state **O**. The high energy of O_H and high redox potential of Cu_B in that state may stem from a three-coordinated near-planar geometry of $\text{Cu}_B[\text{II}]$. Alternatively, the BNC may be compact in state O_H with a very short Fe–Cu distance, a μ -hydroxo ligand bridging the two metals, and Cu pulled out from the plane of its nitrogen ligands. Finally, our DFT data suggest that a contributing reason for the high redox potential of the F_H and O_H states may be that the tyrosine has some characteristics of the neutral free radical, and that this state of the cross-linked tyrosine is more important than considered previously. Validation or falsification of our hypothesis requires further spectroscopic studies of the O_H state. It may be of particular interest to search for Raman evidence for a lengthened Fe–N bond to the proximal histidine, to test whether the lifetime of the ferric heme a_3 -hydroxide species (31) extends beyond the 100 ms domain, and whether a partial tyrosine radical/cuprous copper feature can be distinguished by EPR. An extended X-ray absorption fine structure (EXAFS) study of the ligands of Cu_B would also be of interest, but would require a heme-copper oxidase variant in which the Cu_A center is lacking.

Methods

The details of DFT calculations are presented in *SI Methods*. Figures in *Supporting Information* show DFT-optimized structures and spin-density distribution in relevant redox states. Tables in *Supporting Information* comprise description of model systems, their energies, geometric data and spin-density analysis.

ACKNOWLEDGMENTS. M.W. acknowledges helpful discussions with Drs. Gerhard Hummer [National Institute of Diabetes and Digestive and Kidney Diseases, National Institutes of Health (NIH)] and Denis Rousseau (Albert Einstein College of Medicine). Center for Scientific Computing, the Finnish IT Center for Science, is acknowledged for computational resources. This work was supported by grants from the Academy of Finland, Biocentrum Helsinki, and the Sigrid Jusélius Foundation. V.S. acknowledges funding from the European Research Council project CROWDED-PRO-LIPIDS. K.D.K. acknowledges financial support from NIH (R01GM060353).

- Ferguson-Miller S, Babcock GT (1996) Heme-copper terminal oxidases. *Chem Rev* 96(7):2889–2908.
- Kaila VRI, Verkhovskiy MI, Wikström M (2010) Proton-coupled electron transfer in cytochrome oxidase. *Chem Rev* 110(12):7062–7081.
- Yoshikawa S, et al. (1998) Redox-coupled crystal structural changes in bovine heart cytochrome c oxidase. *Science* 280(5370):1723–1729.
- Iwata S, Ostermeier C, Ludwig B, Michel H (1995) Structure at 2.8 Å resolution of cytochrome c oxidase from *Paracoccus denitrificans*. *Nature* 376(6542):660–669.
- Konstantinov AA, Siletsky S, Mitchell D, Kaulen A, Gennis RB (1997) The roles of the two proton input channels in cytochrome c oxidase from *Rhodobacter sphaeroides* probed by the effects of site-directed mutations on time-resolved electrogenic intraprotein proton transfer. *Proc Natl Acad Sci USA* 94(17):9085–9090.
- Wikström M, Krab K (1979) Proton-pumping cytochrome c oxidase. *Biochim Biophys Acta* 549(2):177–22.
- Wikström M, Verkhovskiy MI, Hummer G (2003) Water-gated mechanism of proton translocation by cytochrome c oxidase. *Biochim Biophys Acta* 1604(2):61–65.
- Wikström M (2004) Cytochrome c oxidase: 25 years of the elusive proton pump. *Biochim Biophys Acta* 1655(1–3):241–247.
- Popović DM, Stuchebrukhov AA (2004) Electrostatic study of the proton pumping mechanism in bovine heart cytochrome c oxidase. *J Am Chem Soc* 126(6):1858–1871.
- Popović DM, Stuchebrukhov AA (2004) Proton pumping mechanism and catalytic cycle of cytochrome c oxidase: Coulomb pump model with kinetic gating. *FEBS Lett* 566(1–3):126–130.
- Morgan JE, Verkhovskiy MI, Wikström M (1994) The histidine cycle: A new model for proton translocation in the respiratory heme-copper oxidases. *J Bioenerg Biomembr* 26(6):599–608.
- Rich PR (1995) Towards an understanding of the chemistry of oxygen reduction. *Aust J Plant Physiol* 22(3):479–486.
- Chance B, Saronio C, Leigh JS, Jr. (1975) Functional intermediates in reaction of cytochrome oxidase with oxygen. *Proc Natl Acad Sci USA* 72(4):1635–1640.
- Wikström M (1981) Energy-dependent reversal of the cytochrome oxidase reaction. *Proc Natl Acad Sci USA* 78(7):4051–4054.
- Weng LC, Baker GM (1991) Reaction of hydrogen peroxide with the rapid form of resting cytochrome oxidase. *Biochemistry* 30(23):5727–5733.
- Proshlyakov DA, et al. (1994) Selective resonance Raman observation of the “607 nm” form generated in the reaction of oxidized cytochrome c oxidase with hydrogen peroxide. *J Biol Chem* 269(47):29385–29388.

17. Wikström M (2012) Active site intermediates in the reduction of O₂ by cytochrome oxidase, and their derivatives. *Biochim Biophys Acta* 1817(4):468–475.
18. Brzezinski P, Gennis RB (2008) Cytochrome c oxidase: Exciting progress and remaining mysteries. *J Bioenerg Biomembr* 40(5):521–531.
19. Morgan JE, Verkhovskiy MI, Palmer G, Wikström M (2001) Role of the P_R intermediate in the reaction of cytochrome c oxidase with O₂. *Biochemistry* 40(23):6882–6892.
20. Blair DF, Witt SN, Chan SI (1985) Mechanism of cytochrome c oxidase-catalyzed dioxygen reduction at low temperatures. Evidence for two intermediates at the three-electron level and entropic promotion of the bond-breaking step. *J Am Chem Soc* 107(25):7389–7399.
21. Gorbikova EA, Wikström M, Verkhovskiy MI (2008) The protonation state of the cross-linked tyrosine during the catalytic cycle of cytochrome c oxidase. *J Biol Chem* 283(50):34907–34912.
22. Antonini E, Brunori M, Colosimo A, Greenwood C, Wilson MT (1977) Oxygen “pulsed” cytochrome c oxidase: Functional properties and catalytic relevance. *Proc Natl Acad Sci USA* 74(8):3128–3132.
23. Belevich I, Verkhovskiy MI (2008) Molecular mechanism of proton translocation by cytochrome c oxidase. *Antioxid Redox Signal* 10(1):1–29.
24. Verkhovskiy MI, Jasaitis A, Verkhovskaya ML, Morgan JE, Wikström M (1999) Proton translocation by cytochrome c oxidase. *Nature* 400(6743):480–483.
25. Bloch D, et al. (2004) The catalytic cycle of cytochrome c oxidase is not the sum of its two halves. *Proc Natl Acad Sci USA* 101(2):529–533.
26. Wikström M, Verkhovskiy MI (2006) Towards the mechanism of proton pumping by the haem-copper oxidases. *Biochim Biophys Acta* 1757(8):1047–1051.
27. Jancura D, et al. (2006) Spectral and kinetic equivalence of oxidized cytochrome C oxidase as isolated and “activated” by reoxidation. *J Biol Chem* 281(41):30319–30325.
28. Brudvig GW, Stevens TH, Morse RH, Chan SI (1981) Conformations of oxidized cytochrome c oxidase. *Biochemistry* 20(13):3912–3921.
29. Belevich I, Bloch DA, Belevich N, Wikström M, Verkhovskiy MI (2007) Exploring the proton pump mechanism of cytochrome c oxidase in real time. *Proc Natl Acad Sci USA* 104(8):2685–2690.
30. Gorbikova EA, Vuorilehto K, Wikström M, Verkhovskiy MI (2006) Redox titration of all electron carriers of cytochrome c oxidase by Fourier transform infrared spectroscopy. *Biochemistry* 45(17):5641–5649.
31. Han S, Takahashi S, Rousseau DL (2000) Time dependence of the catalytic intermediates in cytochrome c oxidase. *J Biol Chem* 275(3):1910–1919.
32. Wikström M, Krab K, Saraste M (1981) *Cytochrome Oxidase: A Synthesis* (Academic, London).
33. Chufán EE, et al. (2010) Differential reactivity between two copper sites in peptidylglycine α -hydroxylating monooxygenase. *J Am Chem Soc* 132(44):15565–15572.
34. Himes RA, Park GY, Siluvai GS, Blackburn NJ, Karlin KD (2008) Structural studies of copper(I) complexes of amyloid-beta peptide fragments: Formation of two-coordinate bis(histidine) complexes. *Angew Chem Int Ed Engl* 47(47):9084–9087.
35. Himes RA, Park GY, Barry AN, Blackburn NJ, Karlin KD (2007) Synthesis and X-ray absorption spectroscopy structural studies of Cu(I) complexes of histidylhistidine peptides: The predominance of linear 2-coordinate geometry. *J Am Chem Soc* 129(17):5352–5353.
36. Gray HB, Malmström BG, Williams RJP (2000) Copper coordination in blue proteins. *J Biol Inorg Chem* 5(5):551–559.
37. Dunitz JD (1994) The entropic cost of bound water in crystals and biomolecules. *Science* 264(5159):670.
38. Wirstam M, Blomberg MRA, Siegbahn PER (1999) Reaction mechanism of compound I formation in heme peroxidases: A density functional theory study. *J Am Chem Soc* 121(43):10178–10185.
39. Heimdal J, Rydberg P, Ryde U (2008) Protonation of the proximal histidine ligand in heme peroxidases. *J Phys Chem B* 112(8):2501–2510.
40. Kaukonen M (2007) Calculated reaction cycle of cytochrome c oxidase. *J Phys Chem B* 111(43):12543–12550.
41. Baik M, Freisner RA (2002) Computing redox potentials in solution: Density functional theory as a tool for rational design of redox agents. *J Phys Chem A* 106(32):7407–7412.
42. Roy LE, Jakubikova E, Guthrie MG, Batista ER (2009) Calculation of one-electron redox potentials revisited. Is it possible to calculate accurate potentials with density functional methods? *J Phys Chem A* 113(24):6745–6750.
43. Martens CF, et al. (1993) Redox behaviour of novel copper(II) crown ether-pyrazole complexes. *J Chem Soc Chem Commun* 1:88–90.
44. Martens CF, et al. (1995) X-ray structures and redox properties of copper(II) bis(pyrazole) complexes. *Inorg Chem* 34(19):4735–4744.
45. Chen P, Bell J, Eipper BA, Solomon EI (2004) Oxygen activation by the noncoupled binuclear copper site in peptidylglycine α -hydroxylating monooxygenase. Spectroscopic definition of the resting sites and the putative Cu(II)-OOH intermediate. *Biochemistry* 43(19):5735–5747.
46. Pettingill TM, Strange RW, Blackburn NJ (1991) Carbonmonoxy dopamine beta-hydroxylase. Structural characterization by Fourier transform infrared, fluorescence, and x-ray absorption spectroscopy. *J Biol Chem* 266(26):16996–17003.
47. Fox S, Nanthakumar A, Wikström M, Karlin KD, Blackburn NJ (1996) XAS structural comparisons of reversible inter-convertible oxo- and hydroxo-bridged heme-copper oxidase model compounds. *J Am Chem Soc* 118(1):24–34.
48. Obias HV, et al. (1998) Heterobinuclear ligand-induced structural and chemical variations in [(L)Fe^{III}-O-Cu^{II}]⁺ μ -oxo complexes. *J Am Chem Soc* 120(37):9696–9697.
49. Baron R, McCammon JA (2007) Dynamics, hydration, and motional averaging of a loop-gated artificial protein cavity: The W191G mutant of cytochrome c peroxidase in water as revealed by molecular dynamics simulations. *Biochemistry* 46(37):10629–10642.
50. Tegoni M, Yu F, Bersellini M, Penner-Hahn JE, Pecoraro VL (2012) Designing a functional type 2 copper center that has nitrite reductase activity within α -helical coiled coils. *Proc Natl Acad Sci USA* 109(52):21234–21239.
51. Qin L, et al. (2009) Redox-dependent conformational changes in cytochrome C oxidase suggest a gating mechanism for proton uptake. *Biochemistry* 48(23):5121–5130.
52. Brand SE, et al. (2007) A new ruthenium complex to study single-electron reduction of the pulsed O(H) state of detergent-solubilized cytochrome oxidase. *Biochemistry* 46(50):14610–14618.
53. Rousseau DL, Ching YC, Brunori M, Giacometti GM (1989) Axial coordination of ferric Aplysia myoglobin. *J Biol Chem* 264(14):7878–7881.
54. Sharma V, Wikström M, Kaila VR (2011) Stabilization of the peroxy intermediate in the oxygen splitting reaction of cytochrome cbb₃. *Biochim Biophys Acta* 1807(7):813–818.