Bacteriocin Typing of Serratia marcescens Isolates of Known Serotype/-Group

WALTER H. TRAUB

Department of Pathology, Bowman Gray School of Medicine, Winston-Salem, North Carolina 27103

Received for publication 14 February 1972

The number of isolates of Serratia marcescens that could be typed by sensitivity to bacteriocins was compared with the nature of the serotype/-group of each of the isolates. Ninety-four of 101 isolates (93.1%) could be bacteriocintyped; this compares with 80 of the isolates (79.2%) that had been serotyped, and with 91 of the isolates (90.1%) that carried determinable O antigens. It is recommended that bacteriocin typing of S. marcescens be adopted by reference laboratories, because this technique is simple, inexpensive, and appears to be of somewhat higher epidemiological resolution than classic serological procedures.

Recently we developed a simple, inexpensive, apparently reproducible technique for typing clinical isolates of Serratia marcescens based on bacteriocin sensitivity (4, 5). Unfortunately, specific antisera were not available to us; therefore, it was not possible to determine whether given bacteriocin types of this organism correspond to certain serotypes/-groups, or whether certain serotypes can be further divided into different bacteriocin types analogous to colicin typing of Shigella sonnei isolates (1). To date, 15 somatic (O) antigens and 13 flagellar (H) antigens have been defined among isolates of S. marcescens (2). For purposes of this study, B. Davis and G. J. Hermann of the Center for Disease Control (CDC), Atlanta, Ga., kindly provided us with isolates of S. marcescens of known serotype/-group from their collection. Here we wish to report comparative results obtained with our technique of bacteriocin typing.

MATERIALS AND METHODS

Bacteria. A total of 101 isolates of *S. marcescens* were received from B. Davis and G. J. Hermann, Enterobacteriology Unit, CDC, Atlanta, Ga., in August 1971. The isolates had been submitted to the CDC from various states during the period of 1969 to 1971. The isolates had been coded. After receipt, the isolates were checked for viability and purity by subculture to MacConkey agar (Fisher Scientific Co., Raleigh, N.C.). The isolates were screened for deoxyribonuclease production, employing deoxyribonuclease test agar (Difco Laboratories, Detroit, (Mich.); the plates were flooded with 1 N HCl after incubation at 33 C for 18 hr (3). All isolates proved

¹Present address: Institut fur Hygiene and Medizinische Mikrobiologie, 8520 Erlangen, Wasserturmstr. 3, West Germany. deoxyribonuclease-positive. The isolates were maintained on Brain Heart Infusion agar slants (Difco) at 4 C.

Bacteriocin typing. The isolates were typed according to their sensitivity to bacteriocins as previously described (5). After completion of the experiments, the results obtained were sent to the CDC; in turn, we were informed as to the nature of the serotype/-group of each of the isolates.

RESULTS

All but 7 of the 101 isolates (93.1%) could be typed with our routine bacteriocin typing procedure. Shown in Table 1 are the distributions of the various serotypes/-groups of *S. marcescens* into bacteriocin types. Of the 101 isolates, one isolate carried indeterminate O and H antigens, and this isolate proved refractory to the killing action of all 10 bacteriocins employed. Nine additional isolates were O-nontypable; of these, six isolates were bacteriocin-typable, and three were tolerant to all bacteriocins. Twenty of the isolates were H-nontypable, but 19 of these proved susceptible to one or more bacteriocins.

The majority of the isolates were of bacteriocin types 1, 4, 9, 14, 15, 16, 18, and 21, a finding in agreement with the data for isolates of S. marcescens from our clinical material (4), except that we had encountered only one previous isolate of S. marcescens of bacteriocin type 15.

The majority of the isolates examined were of serogroups O2, 3, 4, 5, 6, and 14. Yet isolates of any one of these six serogroups could be further subdivided into at least four or more bacteriocin types, a rate comparing favorably with that achieved through the determination of

 TABLE 1. Distribution of serotypes/-groups of
 Serratia marcescens into bacteriocin types

Bacteriocin type	Serotypes/-groups	Total
1	O2:H1 (9 isolates); O15:H5	10
4	O5:H1; O undet.:H1 (2 isolates);	9
	rel. 05:H13; 05:H11; 05:	
	H13; rel. O5:H1 (2 isolates);	
	O14:H12	
7	O4:H1 (2 isolates)	2
9	O14:H10 (3 isolates); O14:H4 (2	10
	isolates); O14:H12 (2 isolates);	
	014:H undet.; rel. 014:H8;	
10	06:H1	
10	04:H1	1
12	014:H5	1 2
13	03:H12; 0 undet.:H12	10
14	rel. O2:H undet.; O3:H5; O3: nonmotile; O6:H undet.; O	10
	undet.: $H1$; O14: H5 (3 isolates);	
	O14: H undet. (2 isolates)	
15	O undet.: $H5$; O2: H undet.; O2:	17
10	nonmotile; rel. O2:H4; rel.	
	O2:nonmotile; O2:H5 (2 iso-	
	lates); O3:H5 (3 isolates); O3:	
	H undet.; O3:nonmotile; O6:	
	nonmotile; O14: H5 (2 isolates);	
	O14:H undet.; O14:nonmotile	
16	O1:H5 (2 isolates); O5:H6; O5:	4
	H undet.	
17	O3:H1 (3 isolates)	3
18	O14:H4 (2 isolates); O14:H12 (4	6
	isolates)	
21	O5:H1; O5:nonmotile; O6:H2 (2	6
	isolates); O6:H undet.; O14:	
00	H11	1
23 24	O14:H3 O4:H4: O13:H4	$\frac{1}{2}$
24 25	$O_4: H_4; O_{13}: H_4$ O undet.: H10	
20 33	O undet. 1110 O1:H undet.	1
37	01:H4; rel. 02:H1	2
39	014:H11	1
43	O14:H undet.	1
44	O4:H12; rel. O4 + O15:H12	2
45	O5:H2 (2 isolates)	2
Nontyp-	012:H undet.; rel. 013:H10;	7
able	O14:H5; O undet.:H6 (2 iso-	
	lates); O undet.:H undet.; O	
	undet.:H11	
Total		101

flagellar antigens, i.e., serotyping. Serogroup O14 consisted of 31 isolates; with the exception of one nontypable isolate, the isolates could be subdivided into 10 different bacteriocin types, as compared with 7 serotypes (H antigens 3, 4, 5, 8, 10, 11, and 12). Comparison of the bacteriocin sensitivity/tolerance patterns (4) of bacteriocin types 4, 9, and 21, as well as 14 and 15, disclosed that isolates of the respective former and latter bacteriocin types appeared to be related, since the bacteriocin patterns differed only with regard to sensitivity or tolerance to one of the ten bacteriocins. Furthermore, it was noted that isolates of serogroup O3 that comprised bacteriocin types 14 and 15 differed in sensitivity to one bacteriocin, as was true for isolates of serogroups 6 and 14 that were of bacteriocin type 14 and 15, respectively. Likewise, isolates of serogroup 1 and bacteriocin types 33 and 37, as well as isolates of serogroup O5 and bacteriocin type 4 and 21, appeared to be closely related as judged by the similar bacteriocin sensitivity/ tolerance patterns.

It was of interest to note that 9 of 10 isolates comprising bacteriocin type 1 were of identical serotype (O2:H1). Similarly, the two isolates of serotypes O4:H1 and O5:H2 were found to belong to bacteriocin types 7 and 45, respectively. However, in most instances the number of isolates of specific serotype/-group was too small to allow any definitive conclusions.

DISCUSSION

The purpose of this study was to determine how bacteriocin typing of isolates of S. marcescens compared with serologic procedures and whether isolates of S. marcescens of known serotype/-group could be further divided into different bacteriocin types. The number of isolates of known serotype/-group at hand was limited; we were not able to examine isolates of S. marcescens carrying O antigens 7, 8, 9, 10, or 11. For instance, isolates of serotype O11:H4 and O11:H13 had been found to be among the more prevalent at Boston City Hospital (6). Similarly, flagellar antigens H7 and 9 were not represented among the isolates studied. Nevertheless, certain conclusions could be drawn.

With the aid of the bacteriocin typing procedure, 94 of the 101 isolates (93.1%) could be typed. Serological tests permitted typing of 80 of the isolates (79.2%) and grouping of 91 (90.1%). It appears that a larger number of isolates of *S. marcescens* can be typed according to their sensitivity/tolerance to 10 selected bacteriocins than through serological methods. In some instances there appears to exist a correlation between certain serotypes and bacteriocin types. Also, certain serogroups appeared to be related to given bacteriocin types. However, in most cases the number of isolates of known serotype/-group received was too small to permit any further conclusions.

It is recommended that reference laboratories adopt the bacteriocin technique for typing isolates of *S. marcescens*, because this procedure is simple, inexpensive, and apparently of higher "epidemic resolution" than hitherto employed classic serological procedures.

ACKNOWLEDGMENTS

I sincerely thank B. Davis and G. J. Hermann for their gracious and kind cooperation; without their help this study would not have been feasible.

This study was supported by a grant from the Schering Corporation, Kenilworth, N.J., whom we thank for their generous financial support.

LITERATURE CITED

 Abbot, J. D., and R. Shannon. 1958. A method for typing Shigella sonnei, using colicine production as a marker. J. Clin. Pathol. 11:71-77.

- Edwards, P. R., and W. H. Ewing. 1962. Identification of Enterobacteriaceae, 2nd ed., p. 223. Burgess Publishing Co., Minneapolis, Minn.
 Jeffries, C. D., D. F. Holtman, and D. G. Guse. 1957.
- Jeffries, C. D., D. F. Holtman, and D. G. Guse. 1957. Rapid method for determining the activity of microorganisms on nucleic acids. J. Bacteriol. 73:590-591.
- Traub, W. H., and E. A. Raymond. 1971. Epidemiological surveillance of Serratia marcescens infections by bacteriocin typing. Appl. Microbiol. 22:1058-1063.
- Traub, W. H., E. A. Raymond, and T. S. Startsman. 1971. Bacteriocin (marcescin) typing of clinical isolates of Serratia marcescens. Appl. Microbiol. 21:837-840.
- Wilfert, J. N., F. F. Barrett, W. H. Ewing, M. Finland, and E. H. Kass. 1970. Serratia marcescens: biochemical, serological, and epidemiological characteristics and antibiotic susceptibility of strains isolated at Boston City Hospital. Appl. Microbiol. 19:345-352.