



Contents lists available at ScienceDirect

## Asian Pacific Journal of Tropical Biomedicine

journal homepage: www.elsevier.com/locate/apjtb



Document heading doi:10.1016/S2221-1691(13)60186-0 © 2013 by the Asian Pacific Journal of Tropical Biomedicine. All rights reserved.

## Analysis of phytochemical profile of *Terminalia arjuna* bark extract with antioxidative and antimicrobial properties

Shreya Mandal, Arpita Patra, Animesh Samanta, Suchismita Roy, Arpita Mandal, Tapasi Das Mahapatra, Shrabani Pradhan, Koushik Das, Dilip Kumar Nandi\*

Department of Nutrition, Microbiology and Human Physiology, Raja N L Khan Women's College, Midnapore, Pin-721102, West Bengal, India

## PEER REVIEW

**Peer reviewer**

Dr. R. N. S. Yadav, Centre For Studies In Biotechnology, Dibrugarh University, Dibrugarh, Assam-786004, India.

Tel: +91-9435032590

E-mail: yadavrns1956@yahoo.co.in

**Comments**

The aim of the study is to investigate phytochemical screening, antimicrobial activity and qualitative thin layer chromatographic separation of flavonoid components, antioxidant activity and total flavonoid compound of *T. arjuna*. The *T. arjuna* bark extract revealed the presence of bio-active constituents which are known to exhibit medicinal as well as physiological activities.

Details on Page 965

## ABSTRACT

**Objective:** To investigate phytochemical screening, antimicrobial activity and qualitative thin layer chromatographic separation of flavonoid components, antioxidant activity and total flavonoid compound of *Terminalia arjuna*.

**Methods:** For phytochemical screening, some common and available standard tests were done. Antimicrobial bioassay was done through agar well diffusion method. Detection of antioxidant activity and flavonoid compounds were done through thin layer chromatography. Total antioxidant activity was measured by 2, 2-diphenyl-1-picrylhydrazyl (DPPH) in colorimetric method. Aluminum chloride colorimetric method was used for total flavonoid determination.

**Results:** Phytochemical screening showed the active compounds presence in high concentration, such as phytosterol, lactones, flavonoids, phenolic compounds and tannins and glycosides. The antimicrobial activity of extract showed that greater inhibition zone against Gram negative bacteria than Gram positive bacteria. This methanolic extract showed a promising antioxidant activity, as absorption of DPPH redicles decreased in DPPH free radical scavenging assay. Flavonoids components having antioxidant property present in the methanol extract at a level of 199.00 mg quercetin equivalent/g of dried methanol extract in colorimetric method.

**Conclusions:** The *Terminalia arjuna* bark extract revealed the presence of bio-active constituents which are known to exhibit medicinal as well as physiological activities.

## KEYWORDS

*Terminalia arjuna*, Phytocompound, TLC, Antioxidant, Total flavonoid, Antimicrobial activity, DPPH

### 1. Introduction

In the ancient India, medicinal plants were used to prevent various critical diseases. The plant kingdom is an important source of herbal drugs. Even in recent years, there has been an increasing awareness about the importance of medicinal plants. Generally, herbal drugs are easily available, safe, less expensive, efficient, and rarely have side effects. According to World Health Organization, medicinal plants would be the best source to obtain

variety of drugs[1]. Medicinal plants contain some organic compounds which provide definite physiological action on the human body and these bioactive substances include tannins, alkaloids, carbohydrates, terpenoids, steroids, flavonoids and phenols.

The bio-active phytocompounds are synthesized by primary or rather secondary metabolism of living organisms. Secondary metabolites are chemically and taxonomically extremely diverse compounds with obscure function. They are widely used in the human therapy,

\*Corresponding author: Dilip Kumar Nandi, Department of Human Physiology, Nutrition, and Microbiology, Raja N L Khan Women's College, Midnapore, Pin-721102, West Bengal, India.

Tel: +91-9434229882

E-mail: dilipnandi2004@yahoo.co.in

Article history:

Received 20 Sep 2013

Received in revised form 27 Sep, 2nd revised form 4 Oct, 3rd revised form 10 Oct 2013

Accepted 15 Nov 2013

Available online 28 Dec 2013

veterinary, agriculture, scientific research and countless other areas[2]. Medicinal plants containing active chemical constituents with high antioxidant property play an important role in the prevention of various degenerative diseases[3] and have possible benefits to the humanity. A large number of phytochemicals belonging to several chemical classes have been shown to have inhibitory effects on all types of microorganisms *in vitro*. Botanical medicines or phytomedicines refer to the use of seeds, berries, leaves, bark, root or flowers of any plant for medicinal purposes by significant number of people. Knowledge of the chemical constituents of plants is desirable because such information will be value for synthesis of complex chemical substances[4–6].

*Terminalia arjuna* (*T. arjuna*) is a deciduous large-sized fluted tree to 30 m tall and 2–2.5 m diameter at breast height, with an often buttressed trunk. *T. arjuna* (family Combretaceae), a large tree, is found throughout the South Asian region. This tree is usually an evergreen tree with new leaves appearing in the hot season (February to April) before leaf fall. This tree is an exotic tree in India. It is one of the most versatile medicinal plants having a wide spectrum of biological activity. The bark of *T. arjuna* is anti-dysentric, antipyretic, astringent, cardiogenic, lithotriptic, anticoagulant, hypolipidemic, antimicrobial[7] and antiuremic[8] agent. Many useful phytoconstituents have been isolated from *T. arjuna* which included triterpenoids for cardiovascular properties, tannins and flavonoids for its anticancer, antimicrobial properties and so on[9]. The powder of the bark acts as a diuretic in cirrhosis of liver and gives relief in symptomatic hypertension[10]. In studies in mice, its leaves have been shown to have analgesic and anti-inflammatory properties[11]. The purpose of the study was to find out the preliminary phytochemical screening of the extract and to determine the antioxidant and antimicrobial activity of extract of the bark of *T. arjuna*.

## 2. Methods and materials

### 2.1. Plant material collection

The bark of *T. arjuna* was collected from University Road, Vidyasagar University, Midnapore, Paschim Medinipur, district of West Bengal. The material was identified by the taxonomist of the Botany Department at the Raja N. L. Khan Women's College, Midnapore. The voucher specimens were deposited in the Department of Botany, Raja N. L. Khan Women's College.

### 2.2. Bacterial strain and culture conditions

Two Gram negative and two Gram positive indicator bacteria used for antimicrobial assay respectively, *Escherichia coli* (*E. coli*) (MTCC 443), *Klebsiella pneumoniae* (*K. pneumoniae*) (MTCC 109), *Staphylococcus aureus* (*S. aureus*) (MTCC 3160) and *Streptococcus mutans* (*S. mutans*) (MTCC 890) were provided by microbiological laboratory and clinical detection center Midnapur (West Medinipur, India). They were cultured in tryptone soy broth or agar (TSB or TSA) in aerobic condition at 37 °C.

### 2.3. Preparation of methanolic extract of bark of *T. arjuna*

The collected *T. arjuna* barks were cut into small pieces. The plant parts were dried in an incubator for 7 d at 40 °C, crushed in an electrical grinder and then the powder was separated. A total of 100 g of bark powder of said plant material was washed in 400 mL of petroleum ether for 24 h to remove the greasy pigmented non polar materials. Then the petroleum ether was discarded and residue was dissolved in 500 mL diethyl ether for 2 h in a soxhlet apparatus. The extract was filtered through Whatman No. 1 filter paper and the resulting filtrate was dried in the air. The ether solid extract was dissolved in 300 mL acetone for 1 h in a soxhlet apparatus. Then the extract was filtered through Whatman No. 1 filter paper and the resulting filtrate was dried under reduced pressure at 40 °C on a rotary evaporator. The acetone solid extract was dissolved in 200 mL methanol and was dried in the air. The methanol extract was stored in refrigerator for phytochemical screening, antioxidant and antimicrobial activity study. Percent of yield[12] was calculated as follows:

$$\text{Extract yield \%} = (W_1/W_2) \times 100$$

Where,  $W_1$  is net weight of powder in grams after extraction and  $W_2$  is total weight of wood powder in grams taken for extraction.

### 2.4. Phytochemicals analysis

Phytochemical analysis of the test sample was carried out according to standard methods[13–15].

#### 2.4.1. Salkowski reaction test for phytosterols

To 0.5 mL chloroform extract in a test tube add 1 mL of concentrated (conc.)  $H_2SO_4$  from the sides of the test tube. Appearance of reddish brown colour in chloroform layer indicates presence of phytosterols.

#### 2.4.2. Liebermann–Burchard's test for triterpenoids

Extract was treated with few drops of acetic anhydride, boil and cool. Conc. sulfuric acid was added from the sides of the test tube which showed a brown ring at the junction of two layers, and formation of deep red color indicated the presence of triterpenoids.

#### 2.4.3. Foam test for saponins

Small amount of extract was taken in a test tube with little quantity of water and shake vigorously. Appearance of foam persisting for 10 min indicated presence of saponins.

#### 2.4.4. Dragendroff's test for alkaloids

Various extracts of the herbal drug were dissolved in chloroform. Chloroform was evaporated and the residue was acidified by adding few drops of Dragendroff's reagent (Potassium bismuth iodide). Appearance of orange red precipitate indicated presence of alkaloids.

#### 2.4.5. Molisch's test for carbohydrates

The extract was mixed with Molisch reagent, and then added conc. H<sub>2</sub>SO<sub>4</sub> along the sides of the test tube to form layers. Appearance of reddish violet ring the interference indicated the presence of carbohydrates.

#### 2.4.6. Lead acetate test for flavanoids

To the alcoholic solution of the extract add few drops of 10% lead acetate solution. Appearance of yellow precipitate indicated presence of flavanoids.

#### 2.4.7. Legal's test for lactones

To the extract mixtures add sodium nitroprusside and pyridine. Then the mixture was treated with NaOH. Appearance of deep red colour indicated the presence of lactones.

#### 2.4.8. Ferric chloride test for phenolic compounds and tannins

Take 2 mL of extract in a test tube and add ferric chloride solution drop by drop. Appearance of bluish black precipitate indicated presence of phenolic compounds and tannins.

#### 2.4.9. Ninhydrin test for proteins

Few drops of ninhydrin added to the extract. Appearance of blue colour indicated presence of amino acid where as proteins may rarely give positive result.

#### 2.4.10. Keller–Killiani test for glycosides

A total of 1 mL of glacial acetic acid, few drops of ferric

chloride solution and conc. H<sub>2</sub>SO<sub>4</sub> (Slowly through the sides of the test tube) were add to the extract. Appearance of reddish brown ring at the junction of the liquids indicated the presence of de-oxy sugars.

#### 2.5. Thin layer chromatography (TLC) analysis for antioxidant constituents

About 2 µg of extracts of *T. arjuna* was loaded on TLC plates (Merck, 20 cm×20 cm). The plates were developed in methanol: chloroform: hexane (7:2:1, v/v/v) to separate various constituents of the extracts. The developed plates were air dried. Then the antioxidant constituents were analyzed by DPPH technique<sup>[16,17]</sup>. For this 0.05% of DPPH solution in methanol was sprayed on the surface of developed TLC plates and incubated for 10 min at room temperature. The active antioxidant constituents of the *T. arjuna* extract was detected as yellow spots produced via reduction of DPPH by resolved bands against purple back ground on the TLC plates. Ascorbic acid was used as standard antioxidant<sup>[18]</sup>.

#### 2.6. TLC analysis for flavonoid constituents

About 2 µg of extracts of *T. arjuna* was loaded on TLC plates (Merck, 20 cm×20 cm). The plates were developed in toluene: chloroform: methanol (4:4:1, v/v/v) to separate flavonoid compounds of the extracts. The developed plate was air dried. Then anisaldehyde sulfuric acid was sprayed on the surface of the plate and incubated for 20 min at 100 °C. The present flavonoid compound of this extracts was detected as blue spot on developed TLC plate. The R<sub>f</sub> value of the bands were also determined.

#### 2.7. Antioxidant activity determination by DPPH free radical scavenging assay

DPPH radical scavenging activity of the extract was measured by the method described by Barros *et al*<sup>[19]</sup>. For this, different concentrations of extract and ascorbic acid (standard) were prepared with methanol (Sigma–Aldrich) as the test solutions. About 1 mL of each prepared concentrations were placed into test tubes and 0.5 mL of 1 mmol/L DPPH solution in methanol was added. The test tubes were incubated for 15 min and the absorbance was read at 517 nm. A blank solution consisted of DPPH dissolved in same amount of methanol. The DPPH radical scavenging activity percentage was calculated by using the following formula:

$$\text{DPPH radical scavenging activity (\%)} = \frac{A_{\text{control}} - A_{\text{extract}}}{A_{\text{control}}} \times 100$$

Where  $A_{\text{control}}$  is the absorbance of a DPPH solution without extract;  $A_{\text{extract}}$  is the absorbance of the tested extract. All measurements were performed in triplicate.

### 2.8. Determination of total flavonoid content

Aluminum chloride colorimetric method was used for flavonoids determination<sup>[20]</sup>. About 1 mL of the plant extracts/standard of different concentration solution was mixed with 3 mL of methanol, 0.2 mL of aluminum chloride, 0.2 mL of 1 mol/L potassium acetate and 5.6 mL of distilled water. It remained at room temperature for 30 min. The absorbance of the reaction mixture was measured at 415 nm with spectrophotometer against blank. Methanol served as blank. The total content of flavonoid compounds in plant methanol extracts in quercetin equivalents was calculated by the following equation:

$$C=(c \times V)/m$$

Where C is total content of flavonoid compounds, mg/g plant extract, in quercetin equivalent; c is the concentration of quercetin established from the calibration curve in mg/mL, V is the volume of extract in mL, and m is the weight of crude plant extract in g.

### 2.9. Antimicrobial analysis

The antimicrobial activity was determined in the methanolic *T. arjuna* bark extract using agar well diffusion method. The antibacterial activities of *T. arjuna* bark extract (concentration of compound 50%, 100 %) were tested against two Gram-positive *S. aureus*, *S. mutans* and two Gram-negative *E. coli* and *K. pneumoniae*, human pathogenic bacteria. Zone of inhibition of *T. arjuna* bark extract were compared with standards like chloramphenicol for antibacterial activity. The results showed that the remarkable inhibition of the bacterial growth was against the tested organisms<sup>[9]</sup>.

## 3. Result

### 3.1. Percent of yield determination

The obtained yield (%) of the *T. arjuna* bark methanolic extract was 0.3%.

### 3.2. Preliminary phytochemical screening

The preliminary phytochemical analysis in *T. arjuna*

bark methanolic extract showed the active compounds presence in high concentration, such as phytosterol, lactones, flavonoids, phenolic compounds and tannins and glycosides. Also, the active compounds presence in low concentration, such as triterpenoids, saponins, alkaloids, carbohydrates and proteins are shown in Table 1.

**Table 1**

Preliminary phytochemical analysis of *T. arjuna* bark extract.

Phytoconstituents	Tests	Conclusion
Phytosterols	Salkowski reaction	++
Triterpenoids	Liebermann–Burchard's test	+
Saponins	Foam test	+
Alkaloids	Dragendroff's test	+
Carbohydrates	Molisch's test	+
Flavanoids	Lead Acetate test	++
Lactones	Legal's test	++
Phenolic Compounds and Tannins	5% FeCl <sub>3</sub> Test	++
Proteins	Ninhydrin test	+
Glycosides	Keller–Killiani test	++

+: Present in low concentration; ++: Present in high concentration.

### 3.3. TLC analysis for antioxidant constituents

The plates TLC were developed in methanol: chloroform: hexane (7:2:1, v/v/v) and sprayed with 0.05% DPPH reagent. Purple colour of DPPH reagent was bleached by yellow spots which was the indication of positive antioxidant activity. The bark extract of *T. arjuna* in terms of DPPH free radical scavenging activity showed one resolved TLC band with strong antioxidant activity and another spot with weak antioxidant activity as compared to standard antioxidant ascorbic acid (Figure 1).

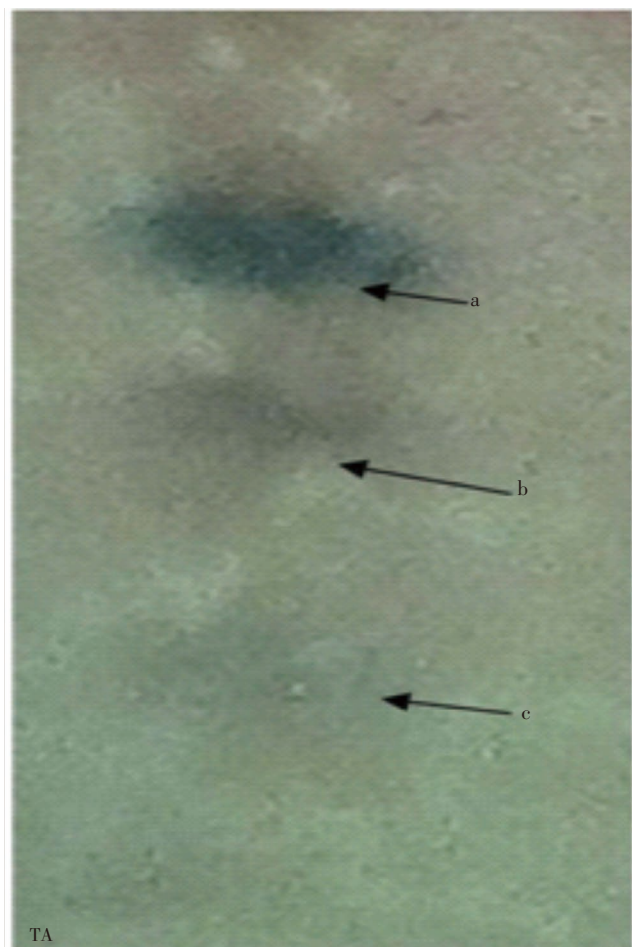


**Figure 1.** TLC antioxidant activity analysis of *T. arjuna* constituents. Standard: Ascorbic acid; TA: *T. arjuna*.

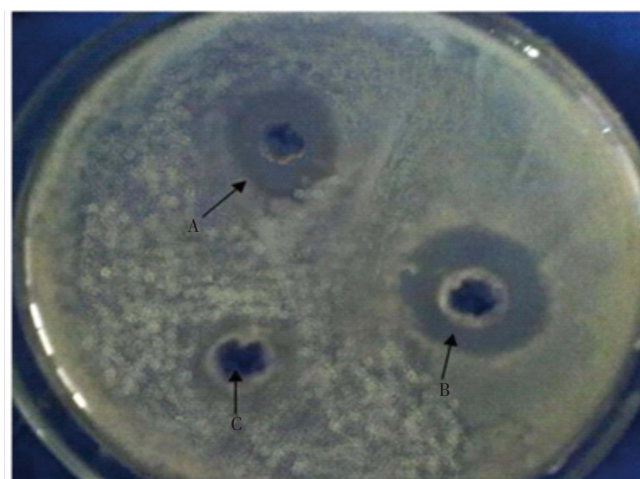


### 3.4. TLC analysis for flavonoid constituents

The plates TLC were developed in chloroform: toluene: methanol (4:4:1) and sprayed with anisaldehyde sulfuric acid reagent. It gives three flavonoid constituents in the TA bark extract with  $R_f$  value of 0.25, 0.32, and 0.38. The eluted compounds showed blue color corresponding with flavonoid behavior (Figure 2 and Table 2).



**Figure 2.** TLC analysis for flavonoid compounds in *T. arjuna* bark methanolic extract.



**Figure 3.** Inhibition zone against *E. coli* (Indicator microbes of Gram negative bacteria). A: Chloramphenicol (Std), B: TA methanolic extract, C: Control (methanol).

**Table 2**

Detection of flavonoids through TLC.

Extract	Solvent system	Revealing reagent	No. of Spot	$R_f$ value
Methanol	Chloroform: Toluene: Methanol (4:4:1,v/v/v)	Anisaldehyde sulfuric acid	3	a. 0.25 b. 0.32 c. 0.38

### 3.5. Determination of total flavonoid content

The total flavonoid content of the *T. arjuna* bark was estimated by using aluminium chloride colorimetric technique and found to be 199.00 mg quercetin equivalent/g of dried methanol extract.

### 3.6. Antimicrobial activity

Antimicrobial activity of *T. arjuna* bark extract showed greater result against gram negative bacteria than Gram positive bacteria (Figure 3 and Table 3).

**Table 3**

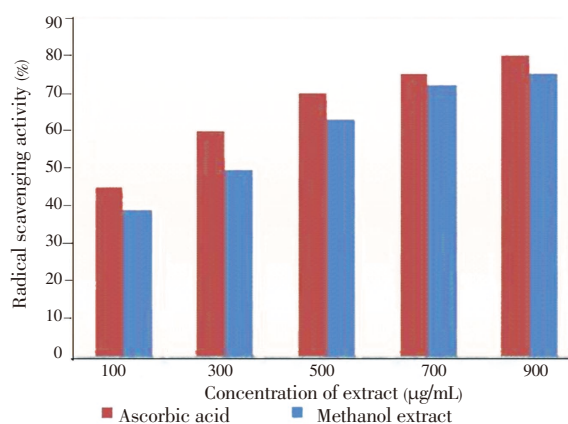
Antimicrobial activities of *T. arjuna* bark methanolic extract and zone of inhibition.

Extract	Diameter of zone of inhibition (mm)* against							
	<i>S. aureus</i>		<i>S. mutans</i>		<i>K. pneumoniae</i>		<i>E. coli</i>	
	100%	50%	100%	50%	100%	50%	100%	50%
Methanolic	11	5.2	16	5.8	21	4.5	32	20
Chloramphenicol (std)	18	10	20	12	28	15	45	30

\*The zone of inhibition (mm) taken as average. Std: Standard Test Dose.

### 3.7. Antioxidant activity determination by DPPH free radical scavenging assay

DPPH is a free radical and it gives strong absorption band at 517 nm in the visible region of electromagnetic radiation. As antioxidant compounds donate protons to these radicals, the absorption decreased. The decrease in absorption was



**Figure 4.** DPPH radical scavenging activity of methanolic extract of *T. arjuna* bark.

taken as a measure of the extent of radical scavenging activity. The results were compared with that of ascorbic acid and the methanolic extract of *T. arjuna* (Figure 4).

#### 4. Discussion

Medicinal plants were of great importance to the health of individuals and communities[21]. Phytochemical analysis conducted on the plant extracts revealed the presence of constituents which are known to exhibit medicinal as well as physiological activities[22]. Analysis of the plant extracts revealed the presence of phytochemicals, such as proteins, carbohydrates, phenols, tannins, flavonoids, saponins, glycosides, steroids, terpenoids and alkaloids. Several studies have described the antioxidant properties of different parts of various medicinal plants which are rich in phenolic compounds[23,24]. *T. arjuna* is a widespread medicinal plant used in the pharmacological system of medicine to care for various degenerative diseases[9]. In this present study, preliminary phytochemical analysis revealed a large amount of phytosterol, lactones, flavonoids, phenolic compounds and tannins and glycosides present in methanol extract of *T. arjuna* bark. Natural antioxidants mainly come from plants in the form of phenolic compounds, such as flavonoids, phenolic acids, tocopherols *etc*[25]. The antioxidative properties of flavonoids are due to several different mechanisms, such as scavenging of free radicals, chelation of metal ions, such as iron and copper and inhibition of enzymes responsible for free radical generation[26]. This methanolic extract has great free radical scavenging property and also contains liberal amount of flavonoid components. Flavonoids are hydroxylated phenolic substances known to be synthesized by plants in response to microbial infection and they have been found to be antimicrobial substances against wide array of microorganisms *in vitro*. Their activity is probably due to their ability to complex with extracellular and soluble proteins and to complex with bacterial cell wall[27]. More than 2000 flavonoids have been reported among woody and non-woody plants[28]. Activity of methanolic extract of *T. arjuna* was comparable to that of reference standard drug chloramphenicol disc. *T. arjuna* bark extract exhibited good antimicrobial activity. The maximum inhibition zone of methanolic extract shows against two gram negative bacteria. Thus, the methanolic extract of *T. arjuna* has great antioxidant and antimicrobial activities.

It has been shown that *T. arjuna* bark consists of many useful compounds, such as flavonoids, tannins, phenols, phytosterols, saponins and alkaloids. Its antioxidant

activity is largely due to flavonoids. The antioxidant and anti-microbial properties of *T. arjuna* are responsible on presence of large amount of flavonoid components. So the results further supported the view that the bark of *T. arjuna* is promising source of natural useful therapeutic agents. The traditional medicine practice is recommended strongly for this plant as well as it is suggested that further work should be carried out to isolate, purify, and characterize the active constituents responsible for the bioactivity study.

#### Conflict of interest statement

We declare that we have no conflict of interest.

#### Acknowledgements

We are thankful to Raja N. L. Khan Women's College, Paschim Medinipur for the grand of UGC Major Project [F. No. 41–68/2012(SR)] and Dr. Keshab Chandra Mandal, Head of Department Microbiology, Vidyasagar University, Paschim Medinipur, West Bengal, India for providing facilities for this research work.

---

#### Comments

##### Background

Medicinal plants were of great importance to the health of individuals and communities. Medicinal plants would be the best source to obtained variety of drugs. Medicinal plants contain some organic compounds which provide definite physiological action on the human body and these bioactive substances include tannins, alkaloids, carbohydrates, terpenoids, steroids, flavonoids and phenols.

##### Research frontiers

The present work shows the antioxidant and antimicrobial activity measures of *T. arjuna* of bark in methanolic extract and also analysis of phytochemical profile.

##### Related reports

Some lifestyle diseases such as cancer, diabetes, cardiovascular diseases, kidney disease are very common nowadays. *T. arjuna* has bio-effective phytochemicals which are responsible to cure in those diseases.

##### Innovations and breakthroughs

*T. arjuna* is an exotic tree in India. It is one of the most

versatile medicinal plants having a wide spectrum of biological activity. In this present study, authors have demonstrated antioxidant, antimicrobial study along with total flavonoid compound measure respectively.

### Applications

The purpose of the study was to find out the preliminary phytochemical screening of the extract and to determine the antioxidant and antimicrobial activity of extract of the bark of *T. arjuna* and in future it may be an agent for treating oxidative stress related disease along with microbial infection.

### Peer review

Aim of the study is to investigate phytochemical screening, antimicrobial activity, and qualitative thin layer chromatographic separation of flavonoid components, antioxidant activity and total flavonoid compound of *T. arjuna*. The *T. arjuna* bark extract revealed the presence of bio-active constituents which are known to exhibit medicinal as well as physiological activities.

### References

- [1] Yadav RN, Agrawala M. Phytochemical analysis of some medicinal plants. *J Physiol* 2011; **3**(12): 10–14.
- [2] Vasu K, Goud JV, Suryam A, Singara Chary MA. Biomolecular and phytochemical analyses of three aquatic angiosperms. *Afr J Microbiol Res* 2009; **3**(8): 418–421.
- [3] Lukmanul H, Giriya A, Boopathy R. Antioxidant property of selected *Ocimum* species and their secondary metabolite content. *J Med Plants Res* 2008; **2**(9): 250–257.
- [4] Mojab F, Kamalinejad M, Ghaderi N, Vanidipour HR. Phytochemicals screening of some species of Iranian plants. *Iran J Pharm Res* 2003; **3**: 77–82.
- [5] Parekh J, Chanda S. Antibacterial and phytochemical studies on twelve species of Indian medicinal plants. *Afr J Biomed Res* 2007; **10**: 175–181.
- [6] Parekh J, Chanda S. Phytochemicals screening of some plants from western region of India. *Plant Arch* 2008; **8**: 657–662.
- [7] Mandal A, Das K, Nandi D K. *In vitro* bioactivity study of bark extract of *Terminalia arjuna* on probiotics, commercially available probiotic formulation. *Int J Phytopharmacol* 2010; **1**(2): 109–113.
- [8] Das K, Chakraborty PP, Ghosh D, Nandi DK. Protective effect of aqueous extract of *Terminalia arjuna* against dehydrating induced oxidative stress and uremia in male rat. *Iran J Pharma Res* 2010; **9**(2): 153–161.
- [9] Nema R, Jain P, Khare S, Pradhan A, Gupta A, Singh D. Antibacterial and antifungal activity of *Terminalia arjuna* leaves extract with special reference to flavanoids. *Basic Res J Med Clin Sci* 2012; **1**(5): 63–65.
- [10] Chatterjee AS. *The treatise on indian medicinal plants: Council of scientific and industrial research*. New Delhi: Publication and Information Directorate; 1994.
- [11] Biswas M, Biswas K, Karan TK, Bhattacharya S, Ghosh AK, Haldar PK. Evaluation of analgesic and anti-inflammatory activities of *Terminalia arjuna* leaf. *J Phytol* 2011; **3**(1): 33–38.
- [12] Patil UH, Gaikwad DK. Phytochemical evaluation and bactericidal potential of *Terminalia arjuna* stem bark. *Int J Pharm Sci Res* 2010; **2**(3): 614–619.
- [13] Harbone JB. *Phytochemical methods*. London: Chapman and Hall; 1998, p. 117–119.
- [14] Fransworth NR. Biological and phytochemical screening of plants. *J Pharm Sci* 1996; **55**: 225–227.
- [15] Rangari VD. *Pharmacognosy and phytochemistry*. Nasik: Carrier Publication; 2002, p.132.
- [16] Kannan R, Arumugam R, Meenakshi S. Thin layer chromatography analysis of antioxidant constituents from seagrasses of Gulf of Mannar Biosphere Reserve, South India. *Int J ChemTech Res* 2010; **2**: 1526–1530.
- [17] Raj RS, Radhamany PM. Preliminary phytochemical and *in vitro* antioxidant properties of *Brunfelsia americana* L. *J Pharm Res* 2010; **3**: 2712–2713.
- [18] Dzomba P, Mupa M. Wild *Cucumis* leaves: phytochemical profile and antioxidant capacity. *Asian Pac J Trop Biomed* 2013; forthcoming.
- [19] Barros L, Falcao S, Baptista P, Freire C, Vilas-Boas M, Ferreira IC. Antioxidant activity of *Agaricus* sp. mushrooms by chemical, biochemical and electrochemical assays. *Food Chem* 2008; **111**: 61–66.
- [20] Wang SY, Jiao H. Correlation of antioxidant capacities to oxygen radical scavenging enzyme activities in blackberry. *J Agric Food Chem* 2000; **48**(11): 5672–5676.
- [21] Pascaline J, Charles M, Lukhoba C, George O. Phytochemical constituents of some medicinal plants used by the Nandis of South Nandi district, Kenya. *J Anim Plant Sci* 2011; **9**: 1201–1210.
- [22] Sofowora A. *Medicinal plants and traditional medicine in Africa*. New York: John Wiley and Sons; 1993, p. 191–289.
- [23] Brown JE, Rice-Evans CA. Luteolin rich artichoke extract protects low density lipoprotein from oxidation *in vitro*. *Free Radic Res* 1998; **29**: 247–255.
- [24] Krings U, Berger RG. Antioxidant activity of roasted foods. *Food Chem* 2001; **72**: 223–229.
- [25] Ali SS, Kasoju N, Luthra A, Singh A, Sharanabasava H, Sahuand A, et al. Indian medicinal herbs as source of antioxidants. *Food Res Int* 2008; **41**: 1–15.
- [26] Benavente-Garcia O, Castillo J, Marin FR, Ortuno A, Del-Rio JA. Uses and properties of *Citrus* flavonoids. *J Agric Food Chem* 1997; **45**(12): 4505–4515.
- [27] Marjorie C. Plant products as antimicrobial agents. *Clin Microbiol Rev* 1999; **12**: 564–582.
- [28] Harborne JB. Plant phenolics. In: Bell EA, Charlewood BV, editors. *Secondary plant products*. Berlin: Verlag Springer; 1980, p. 320.