

Seroepidemiologies of Human Metapneumovirus and Respiratory Syncytial Virus in Young Children, Determined with a New Recombinant Fusion Protein Enzyme-Linked Immunosorbent Assay

Sarah R. Dunn,^a Alex B. Ryder,^a Sharon J. Tollefson,^a Meng Xu,^c Benjamin R. Saville,^c John V. Williams^{a,b}

Departments of Pediatrics,^a Pathology, Microbiology, and Immunology,^b and Biostatistics,^c School of Medicine, Vanderbilt University Medical Center, Nashville, Tennessee, USA

We compared antibodies against human metapneumovirus (HMPV) and respiratory syncytial virus (RSV) in children. The antibody nadirs for both viruses were at 3 to 5 months, and the majority of children were seropositive for both by 2 years. There was no significant difference in the kinetics of maternal antibody decline or seroconversion relative to the two viruses.

Human metapneumovirus (HMPV) is a leading cause of lower respiratory tract illness (LRI) in children and adults (1–9). HMPV and respiratory syncytial virus (RSV) share a number of clinical and genetic similarities (1, 4, 10, 11). HMPV infection leads to significant morbidity in infants and other populations, including immunocompromised, high-risk, and elderly patients (7, 12–19). Interestingly, several studies have reported that the mean age of infants with LRI due to HMPV is higher than that of infants with RSV-associated LRI (1, 8, 10, 20–23). To test the hypothesis that this age discrepancy was due to a difference in maternally derived antibody titers, we determined seropositivity rates by age in a prospective collection of sera from young children.

Serum specimens were obtained from the Vanderbilt Vaccine Clinic, a clinic established for the purpose of evaluating investigational vaccines in young children and conducting viral surveillance (24, 25). Term infants were enrolled at birth and were followed until 5 years of age, although specimens were collected occasionally from older subjects. Serum was obtained and stored at -20°C until testing. Specimens used in the present study were each collected from a unique patient between December 1989 and August 2001 and were selected randomly to yield roughly similar numbers of specimens per 1-year age group. The Vanderbilt Institutional Review Board approved the study.

Human sera were tested for the presence of HMPV or RSV F protein-specific antibodies by enzyme-linked immunosorbent assay (ELISA). Soluble HMPV and RSV F proteins were expressed in Freestyle 293 cells (Invitrogen) and purified as described previously (26). ELISA methods were developed and conditions optimized using known negative and positive sera (not shown). Briefly, 100 ng/well of purified HMPV or RSV F protein was adsorbed onto 384-well polystyrene plates (Nunc) overnight in carbonate buffer (pH 9.8) at 4°C . Plates were blocked with 5% nonfat dried milk in phosphate-buffered saline (PBS) with 0.5% Tween 20 (PBS-T) for 2 h at room temperature. After the plates were washed with PBS-T, serial 4-fold dilutions of serum in duplicate were added, and the plates were incubated for 1 h at room temperature. Plates were washed with PBS-T, alkaline phosphatase-conjugated anti-human IgG (Southern Biotech) was added, and the plates were incubated for 1 h. Finally, plates were washed with PBS-T, and *p*-nitrophenyl phosphate substrate (Sigma) was added. The absorbance at 405 nm was read at 30 min and the mean of duplicate wells taken. Based on the baseline signal of the refer-

ence sera to each antigen, a specimen was designated positive if the ratio of the specimen absorbance compared to negative-control serum value was >2 at a dilution of $\geq 1:20$ for HMPV F or $\geq 1:80$ for RSV F. The ELISA endpoint titer assigned to each specimen was the reciprocal dilution that yielded a positive result. We used logistic regression to model the binary outcome of seropositivity as a function of age and virus, and we used linear regression to model the natural logarithm of titer as a function of age and virus. Restricted cubic splines were used to provide a smoothed estimate of the group means by age.

Two hundred eighty-two sera were included, with 144 (51%) collected from males. The racial distribution of the subjects was 145 (51%) white, 125 (44%) black, and 12 other. The age distribution of the children is shown in Table 1. Of children that were 60 months old or older, the mean age was 78 months (range, 60 to 122 months).

Of the 282 specimens, 219 (78%) were positive for HMPV and 219 (78%) were positive for RSV. Rates of seropositivity for both viruses were high for subjects <6 months old, and titers against both viruses initially decreased with age, reaching a nadir at 5 to 6 months. Seropositivity for HMPV and RSV increased with age after 6 months, reaching $\geq 80\%$ in all subjects that were >2 years old. The probability of seropositivity varied by age for each virus (Fig. 1), but the probabilities of HMPV seropositivity and RSV seropositivity did not differ ($P = 0.12$). Similarly, the mean endpoint titers against both viruses were higher in subjects that were <6 months old and reached a nadir between 3 and 5 months. Serum titers against HMPV remained low until 13 months of age, when the mean log titer began to increase (Fig. 2). In contrast, antibody titers against RSV rose at an earlier age.

We compared titers of serum antibody against HMPV and RSV in a prospectively collected cohort of children. The nadirs for both viruses were between 3 and 5 months of age, consistent with the rate of decline for maternally derived antibodies. Thus, the

Received 17 December 2012 Returned for modification 13 January 2013

Accepted 9 July 2013

Published ahead of print 14 August 2013

Address correspondence to John V. Williams, john.williams@vanderbilt.edu.

Copyright © 2013, American Society for Microbiology. All Rights Reserved.

doi:10.1128/CVI.00750-12

TABLE 1 Number of serum samples from each age group

Child age group (mo)	No. of specimens
<3	8
3–5	9
6–8	16
9–11	9
12–23	48
24–35	52
36–47	48
48–59	54
>60	38
Total	282

waning titer of maternally derived antibody does not correlate with the observed higher mean age of infants with HMPV-associated LRI and does not explain the susceptibility of older infants to HMPV-associated LRI. Consistent with an older age for HMPV infection, the mean endpoint titer against HMPV remained low until 1 year of age. The delayed rise in seropositivity against HMPV compared to seropositivity against RSV reflects the observed older age for HMPV infection but fails to provide a biological explanation. Possible mechanisms include the protective serum antibody threshold against HMPV being lower than that against RSV or age-related differences in the contribution of immune response to disease. The serological data may also reflect differing transmission rates for HMPV and RSV or discordant ability of young infants to mount anti-F antibody responses to HMPV and RSV.

Our study has some limitations. Neutralizing antibodies might provide a better indication of protection against disease, but there was insufficient specimen remaining for this testing. Further, the

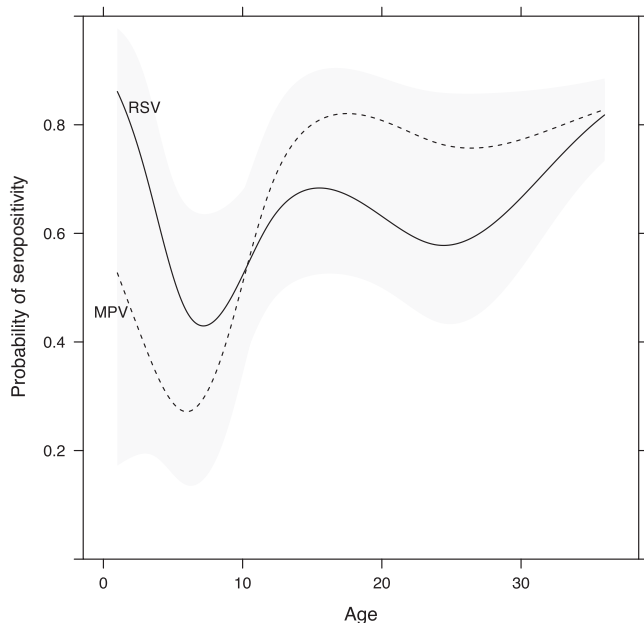


FIG 1 Estimated probabilities of seropositivity by virus and age (in months). Logistic regression was used to model seropositivity to HMPV (dotted) and RSV (solid) by age in months. The 95% confidence intervals (CIs) are indicated as shaded regions.

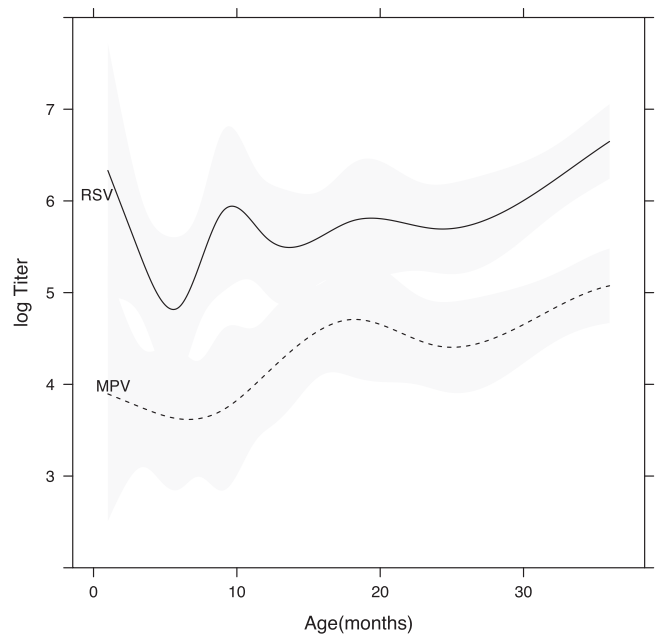


FIG 2 Log titer by virus and age. Linear regression was used to model the natural logarithm of endpoint titer for HMPV (dotted line) and RSV (solid line) by age. The 95% CIs are indicated as shaded regions.

clinical and demographic histories of these specimens were not available.

Others have reported seroepidemiologic studies of HMPV using different methods and age groups (27–33). Only one report compared the seroepidemiologies of HMPV and RSV, but the results were grouped into age groups of 6 months to 5 years and 6 to 10 years (34). Our findings demonstrate that the kinetics of antibody titers against HMPV and RSV are similar during the first year of life. Further studies with humans and using animal models are needed to understand the reason for the reported difference between ages of LRI hospitalization of HMPV and RSV patients, which has implications for vaccine development.

ACKNOWLEDGMENTS

This study was supported by an Infectious Diseases Society of America Summer Scholarship for Medical Students Award (for A.B.R. and S.R.D.). J.V.W. was supported by NIH grant AI085062. The work was supported in part by CTSA award no. UL1 TR000445 from the National Center for Advancing Translational Sciences.

J.V.W. serves on the Scientific Advisory Board of Quidel.

REFERENCES

- van den Hoogen BG, van Doornum GJ, Fockens JC, Cornelissen JJ, Beyer WE, de Groot R, Osterhaus AD, Fouchier RA. 2003. Prevalence and clinical symptoms of human metapneumovirus infection in hospitalized patients. *J. Infect. Dis.* 188:1571–1577.
- van den Hoogen BG, de Jong JC, Groen J, Kuiken T, de Groot R, Fouchier RA, Osterhaus AD. 2001. A newly discovered human pneumovirus isolated from young children with respiratory tract disease. *Nat. Med.* 7:719–724.
- Sloots TP, Mackay IM, Bialasiewicz S, Jacob KC, McQueen E, Harnett GB, Siebert DJ, Masters BI, Young PR, Nissen MD. 2006. Human metapneumovirus, Australia, 2001–2004. *Emerg. Infect. Dis.* 12:1263–1266.
- Peiris JS, Tang WH, Chan KH, Khong PL, Guan Y, Lau YL, Chiu SS. 2003. Children with respiratory disease associated with metapneumovirus in Hong Kong. *Emerg. Infect. Dis.* 9:628–633.

5. Mullins JA, Erdman DD, Weinberg GA, Edwards K, Hall CB, Walker FJ, Iwane M, Anderson LJ. 2004. Human metapneumovirus infection among children hospitalized with acute respiratory illness. *Emerg. Infect. Dis.* 10:700–705.
6. Martinello RA, Esper F, Weibel C, Ferguson D, Landry ML, Kahn JS. 2006. Human metapneumovirus and exacerbations of chronic obstructive pulmonary disease. *J. Infect.* 53:248–254.
7. Falsey AR, Erdman D, Anderson LJ, Walsh EE. 2003. Human metapneumovirus infections in young and elderly adults. *J. Infect. Dis.* 187:785–790.
8. Esper F, Martinello RA, Boucher D, Weibel C, Ferguson D, Landry ML, Kahn JS. 2004. A 1-year experience with human metapneumovirus in children aged <5 years. *J. Infect. Dis.* 189:1388–1396.
9. Boivin G, De Serres G, Cote S, Gilca R, Abed Y, Rochette L, Bergeron MG, Dery P. 2003. Human metapneumovirus infections in hospitalized children. *Emerg. Infect. Dis.* 9:634–640.
10. McAdam AJ, Hasenbein ME, Feldman HA, Cole SE, Offermann JT, Riley AM, Lieu TA. 2004. Human metapneumovirus in children tested at a tertiary-care hospital. *J. Infect. Dis.* 190:20–26.
11. van den Hoogen BG, Bestebroer TM, Osterhaus AD, Fouchier RA. 2002. Analysis of the genomic sequence of a human metapneumovirus. *Virology* 295:119–132.
12. Englund JA, Boeckh M, Kuypers J, Nichols WG, Hackman RC, Morrow RA, Fredricks DN, Corey L. 2006. Brief communication: fatal human metapneumovirus infection in stem-cell transplant recipients. *Ann. Intern. Med.* 144:344–349.
13. Larcher C, Geltner C, Fischer H, Nachbaur D, Muller LC, Huemer HP. 2005. Human metapneumovirus infection in lung transplant recipients: clinical presentation and epidemiology. *J. Heart Lung Transplant.* 24:1891–1901.
14. Boivin G, De Serres G, Hamelin ME, Cote S, Argouin M, Tremblay G, Maranda-Aubut R, Sauvageau C, Ouakki M, Boulianne N, Couture C. 2007. An outbreak of severe respiratory tract infection due to human metapneumovirus in a long-term care facility. *Clin. Infect. Dis.* 44:1152–1158.
15. Hamelin ME, Cote S, Laforge J, Lampron N, Bourbeau J, Weiss K, Gilca R, DeSerres G, Boivin G. 2005. Human metapneumovirus infection in adults with community-acquired pneumonia and exacerbation of chronic obstructive pulmonary disease. *Clin. Infect. Dis.* 41:498–502.
16. van den Hoogen BG. 2007. Respiratory tract infection due to human metapneumovirus among elderly patients. *Clin. Infect. Dis.* 44:1159–1160.
17. Vicente D, Montes M, Cilla G, Perez-Trallero E. 2004. Human metapneumovirus and chronic obstructive pulmonary disease. *Emerg. Infect. Dis.* 10:1338–1339.
18. Williams JV, Crowe JE, Jr, Enriquez R, Minton P, Peebles RS, Jr, Hamilton RG, Higgins S, Griffin M, Hartert TV. 2005. Human metapneumovirus infection plays an etiologic role in acute asthma exacerbations requiring hospitalization in adults. *J. Infect. Dis.* 192:1149–1153.
19. Williams JV, Martino R, Rabella N, Otegui M, Parody R, Heck JM, Crowe JE, Jr. 2005. A prospective study comparing human metapneumovirus with other respiratory viruses in adults with hematologic malignancies and respiratory tract infections. *J. Infect. Dis.* 192:1061–1065.
20. Williams JV, Edwards KM, Weinberg GA, Griffin MR, Hall CB, Zhu Y, Szilagyi PG, Wang CK, Yang CF, Silva D, Ye D, Spaete RR, Crowe JE, Jr. 2010. Population-based incidence of human metapneumovirus infection among hospitalized children. *J. Infect. Dis.* 201:1890–1898.
21. Bosis S, Esposito S, Niesters HG, Crovari P, Osterhaus AD, Principi N. 2005. Impact of human metapneumovirus in childhood: comparison with respiratory syncytial virus and influenza viruses. *J. Med. Virol.* 75:101–104.
22. Døllner H, Risnes K, Radtke A, Nordbo SA. 2004. Outbreak of human metapneumovirus infection in Norwegian children. *Pediatr. Infect. Dis. J.* 23:436–440.
23. Klein MI, Coviello S, Bauer G, Benitez A, Serra ME, Schiatti MP, Delgado MF, Melendi GA, Novalli L, Pena HG, Karron RA, Kleeberger SR, Polack FP. 2006. The impact of infection with human metapneumovirus and other respiratory viruses in young infants and children at high risk for severe pulmonary disease. *J. Infect. Dis.* 193:1544–1551.
24. Williams JV, Harris PA, Tollefson SJ, Halburnt-Rush LL, Pingsterhaus JM, Edwards KM, Wright PF, Crowe JE, Jr. 2004. Human metapneumovirus and lower respiratory tract disease in otherwise healthy infants and children. *N. Engl. J. Med.* 350:443–450.
25. Williams JV, Wang CK, Yang CF, Tollefson SJ, House FS, Heck JM, Chu M, Brown JB, Lintao LD, Quinto JD, Chu D, Spaete RR, Edwards KM, Wright PF, Crowe JE, Jr. 2006. The role of human metapneumovirus in upper respiratory tract infections in children: a 20-year experience. *J. Infect. Dis.* 193:387–395.
26. Cseke G, Wright DW, Tollefson SJ, Johnson JE, Crowe JE, Jr, Williams JV. 2007. Human metapneumovirus fusion protein vaccines that are immunogenic and protective in cotton rats. *J. Virol.* 81:698–707.
27. Ebihara T, Endo R, Kikuta H, Ishiguro N, Yoshioka M, Ma X, Kobayashi K. 2003. Seroprevalence of human metapneumovirus in Japan. *J. Med. Virol.* 70:281–283.
28. Wolf DG, Zakay-Rones Z, Fadeela A, Greenberg D, Dagan R. 2003. High seroprevalence of human metapneumovirus among young children in Israel. *J. Infect. Dis.* 188:1865–1867.
29. Hamelin ME, Boivin G. 2005. Development and validation of an enzyme-linked immunosorbent assay for human metapneumovirus serology based on a recombinant viral protein. *Clin. Diagn. Lab. Immunol.* 12:249–253.
30. Leung J, Esper F, Weibel C, Kahn JS. 2005. Seroepidemiology of human metapneumovirus (hMPV) on the basis of a novel enzyme-linked immunosorbent assay utilizing hMPV fusion protein expressed in recombinant vesicular stomatitis virus. *J. Clin. Microbiol.* 43:1213–1219.
31. Pavlin JA, Hickey AC, Ulbrandt N, Chan YP, Endy TP, Boukhvalova MS, Chunsuttiwat S, Nisalak A, Libraty DH, Green S, Rothman AL, Ennis FA, Jarman R, Gibbons RV, Broder CC. 2008. Human metapneumovirus reinfection among children in Thailand determined by ELISA using purified soluble fusion protein. *J. Infect. Dis.* 198:836–842.
32. Lüsebrink J, Wiese C, Thiel A, Tillmann RL, Ditt V, Muller A, Schildgen O, Schildgen V. 2010. High seroprevalence of neutralizing capacity against human metapneumovirus in all age groups studied in Bonn, Germany. *Clin. Vaccine Immunol.* 17:481–484.
33. Banerjee S, Sullender WM, Ahuja RK, Broor S. 2011. Seroepidemiological study of human metapneumovirus in New Delhi, India. *Indian J. Med. Microbiol.* 29:363–367.
34. Cusi MG, Terrosi C, Kleines M, Schildgen O. 2011. RSV and HMPV seroprevalence in Tuscany (Italy) and North-Rhine Westfalia (Germany) in the winter season 2009/2010. *Influenza Other Respir. Viruses* 5:380–381.