# **Mass Resolution and Mass Accuracy: How Much Is Enough?**

Alan G. Marshall,\*,1,2 Greg T. Blakney,<sup>2</sup> Tong Chen,<sup>1</sup> Nathan K. Kaiser,<sup>2</sup> Amy M. McKenna,<sup>2</sup> Ryan P. Rodgers,<sup>1,2</sup> Brian M. Ruddy,<sup>1</sup> and Feng Xian<sup>2</sup>

<sup>1</sup>*Department of Chemistry & Biochemistry, Florida State University, 95 Chieain Way, Tallahassee, FL 32306, U.S.A.* <sup>2</sup> *Ion Cyclotron Resonance Program, National High Magnetic Field Laboratory, Florida State University, 1800 East Paul Dirac Drive, Tallahassee, FL 32310, U.S.A.*

Accurate mass measurement requires the highest possible mass resolution, to ensure that only a single elemental composition contributes to the mass spectral peak in question. Although mass resolution is conventionally defined as the closest distinguishable separation between two peaks of equal height and width, the required mass resolving power can be ∼10× higher for equal width peaks whose peak height ratio is 100:1. Ergo, minimum resolving power requires specification of maximum dynamic range, and is thus 10–100× higher than the conventional definition. Mass resolving power also depends on massto-charge ratio. Mass accuracy depends on mass spectral signal-to-noise ratio and digital resolution. Finally, the reliability of elemental composition assignment can be improved by resolution of isotopic fine structure. Thus, the answer to the question of "how much is enough mass resolving power" requires that one first specify *S*/*N* ratio, dynamic range, digital resolution, mass-to-charge ratio, and (if available) isotopic fine structure. The highest available broadband mass resolving power and mass accuracy is from Fourier transform ion cyclotron resonance mass spectrometry. Over the past five years, FT-ICR MS mass accuracy has improved by about an order of magnitude, based on higher magnetic field strength, conditional averaging of time-domain transients, better mass calibration (spectral segmentation; inclusion of a space charge term); radially dispersed excitation; phase correction to yield absorption-mode display; and new ICR cell segmentation designs.

*Keywords*: Fourier transform, ion cyclotron resonance, FTMS, isotopic fine structure

(Received September 1, 2012; Accepted December 28, 2012)

# **INTRODUCTION**

A primary goal in mass spectrometry is to identify an analyte, particularly in the presence of other analytes, based on measurement of ion mass. This paper addresses the question of the minimum mass resolving power required to provide mass accuracy sufficient to assign a unique elemental composition,  $C_cH_hN_nO_oS_s...$ , from measurement of mass spectral peak position. Our treatment focuses on Fourier transform ion cyclotron resonance mass spectrometry,<sup>1)</sup> because it offers the highest broadband resolution of any mass analyzer. Recent examples include resolution and identification of tens of thousands of monoisotopic elemental compositions from a single mass spectrum of petroleum crude oil<sup>2)</sup> and unit mass resolution of an intact 147 kDa antibody.<sup>3)</sup>

Recent improvements in FT-ICR mass resolution and accuracy are conveniently separated into instrumental and data reduction categories. Optimization of the experiment has until now been based on single-ion simulations,<sup>1)</sup> but may in future be guided by large-scale simulations of the trajectories of up to a million or more ions<sup>4-6)</sup> as they are injected into an ICR ion trap for subsequent excitation and detection. We begin with experimental advances, and then proceed to software developments.

### **EXPERIMENTAL METHODS**

#### **ICR trapped ion cell conguration**

FT-ICR excitation and detection were originally performed with a cubic cell, with dc "trapping" voltage applied to a pair of opposed axial end cap electrodes, rf excitation voltage applied to one pair of opposed transverse electrodes, and induced rf signal detected on the other pair of opposed transverse electrodes. 7) Cubic and tetragonal cells were then replaced by a cylindrical cell,<sup>8)</sup> with a variety of segmentation configurations designed to approach a threedimensional axial quadrupolar dc trapping potential, and more spatially uniform excitation and detection electric fields.<sup>9,10)</sup> The best current performance has been obtained with the cells shown schematically in Fig.  $1$ .<sup>11-14</sup>) The Tolmachev cell extends an original Gabrielse idea of multiple axial segments to which different dc voltages are applied to produce a nearly quadrupolar trapping potential.<sup>15)</sup> The Nikolaev "dynamically harmonized" cell relies on ion cyclotron motion itself to average out imperfections in the trapping electric field produced by segmented "leaf" electrodes. The Nikolaev cell has achieved mass resolving power greater than 1,000,000 for bovine serum albumin and 20,000,000 for narrow-band detection of reserpine (*m*/*z* 609). 16)

FT-ICR mass resolving power is directly proportional to the duration of an undamped time-domain ICR signal, which in turn requires maintenance of tight spatial coherence of an orbiting packet of ions of a given *m*/*z* ratio. Largescale simulations have shown that such spatial coherence

<sup>\*</sup>Correspondence to: Alan G. Marshall, *Ion Cyclotron Resonance Program, National High Magnetic Field Laboratory, Florida State University, 1800 East Paul Dirac Drive, Tallahassee, FL 32310, U.S.A.*, e-mail: marshall@ magnet.fsu.edu



Fig. 1. Seven-section<sup>15)</sup> (top) and dynamically harmonized<sup>16)</sup> (bottom) ICR cells.

requires a minimum number of trapped ions. 4) Those same simulations also predict coalescence of the trajectories of ions of closely spaced *m*/*z*, as a function of the number of trapped ions. 17) Thus, for compositionally complex mixtures containing thousands of different elemental compositions, it may be necessary to average 100 or more FT-ICR mass spectra to achieve the desired signal-to-noise ratio and dynamic range. Finally, ion trajectory simulations are also helping to predict (and rectify) degradation in performance due to imperfect construction/alignment of electrodes, 6) magnetic field inhomogeneity, 18) choice of excitation waveform, *etc.*

### **Sample preparation**

Russian bitumen was dissolved in toluene to yield a 1 mg/mL stock solution. The sample was further diluted to 0.25 mg/mL with 49:49 (v/v) toluene-methanol and 2% formic acid for protonation during positive ion electrospray ionization (ESI).

### **FT-ICR mass spectrometry**

A custom-built FT-ICR mass spectrometer<sup>13)</sup> equipped with a passively shielded 9.4 tesla superconducting horizontal solenoid magnet and an electrically compensated cell, 12,13) was used for all experiments. Samples were directly pumped through a fused silica capillary at  $0.5 \mu L$ / min through a 50  $\mu$ m i.d. fused silica microESI needle under typical ESI conditions (needle, 2.3 kV; tube lens, 350V; heated metal capillary current, ∼5.0A). External quadrupole isolation was performed with an Extrel QC 150 quadrupole mass filter (Pittsburgh, PA) and a custom-built accumulation octopole.<sup>19-21)</sup> Ions in the accumulation octopole were cooled with helium gas at ∼2mTorr before transfer to the ICR cell. Initial broadband analysis guided the construction of a 4Mword, 10Msample/s time-domain excitation waveform. 22,23) The SWIFT waveform was stored in the hard drive of a PC data system prior to loading of both the isolation and broadband excitation waveforms to an arbitrary waveform generator (National Instruments (NI) model PXI 5421) and later amplified for output to the ICR cell. After quadrupole and SWIFT isolation of the initial ion population, the resultant spectrum was acquired after broadband excitation of remaining ions at 96 kHz to 961 kHz at 50Hz/  $\mu$ s and 378  $V_{p-p}$ . Time-domain data (8 Mword, low *m/z* cutoff =211) were routed to a differential preamplifier and



Fig. 2. Discrete mass spectrum. A: peak amplitude. ∆*m*50%: peak full width at half-maximum peak height.  $\sigma$ : standard deviation of baseline noise. Predicted mass measurement precision is given by Eq. (1) (see the text). (Reproduced, with permission, from ref. 24.)





Fig. 3. Barely resolved pairs of Lorentzian magnitude-mode spectral peaks of equal width. Peak height ratio is 1 : 1 (top) and 100 : 1 (bottom). Note that the resolving power required to distinguish the peaks is ∼10 times higher for the 100 : 1 pair.

ultimately digitized (NI Model PXI 5122) by a Predator data station. 23)

## **RESULTS AND DISCUSSION**

### **Signal-to-noise ratio and mass measurement precision**

For a discrete mass spectrum (see Fig. 2), mass measurement precision may be defined as *m*/σ(*m*), in which *m* is measured mass and  $\sigma(m)$  is the standard deviation for a large number of measurements. Fortunately, under very general conditions, it is possible to predict the precision for a large number of measurements from the signal-to-noise ratio, *S*/*N,* of a *single* measurement, according to Eq. (1):

$$
m/\sigma(m) = \text{constant}
$$
  
×SNR $\sqrt{\text{(Number of Data Points per Peak Width)}}$  (1)

in which the constant depends on mass spectral peak shape and is of order one in magnitude. $24$  In the absence of systematic error (as recently demonstrated for FT-ICR MS, 25) mass accuracy is the same as mass precision. The importance of Eq. (1) is that it shows that mass imprecision (which ultimately limits mass accuracy) depends directly on SNR.

Thus, SNR must be specified in any report of mass measurement accuracy.

#### **Dynamic range and mass accuracy**

Mass resolution is conventionally defined as the minimum separation between two mass spectral peaks of equal height and width, such that there is a detectable "valley" between them. Mass resolving power is then defined as  $(m_1/$  $(m_2-m_1)$ ) (Fig. 3, top). Under that definition, mass resolving power (even for a single peak) is *m*/∆*m*50%, in which ∆*m*50% is mass spectral peak full width at half-maximum peak height. However, if (as is almost always the case experimentally) the two peaks are of unequal height, then the minimum mass resolving power to produce a valley between them can be much higher (*e.g.*, ten times higher for a peak height ratio of 100 : 1 (Fig. 3, bottom). The dynamic range for FT-ICR MS peaks can exceed 10,000 : 1, and may be increased by radially dispersed excitation. 26) Thus, dynamic range must also be specified in determining the minimal mass resolving power for a particular application.

#### **Elemental composition from mass defect**

Every isotope of every element has a different mass defect (*i.e.*, the difference between the exact mass of an atom (in Da) and the nearest integer), as shown in Fig. 4 for isotopes of some common elements. Thus, a unique elemental composition may be determined for any molecule from a

#### **Conditional averaging to reduce rms mass error**

As noted above, when direct infusion is available, it is desirable to sum many FT-ICR mass spectra to improve SNR and dynamic range. However, an ionization source may not



Fig. 4. Atomic mass defects for selected isotopes of some common chemical elements. (Reproduced, with permission, from ref. 35.)



Fig. 5. (+) ESI 9.4 T FT-ICR mass spectra of a Russian bitumen. Bottom: Broadband (+) ESI 9.4 T FT-ICR mass spectrum (black) with a 4Mword 10MHz SWIFT excitation waveform overlaid (red). Middle: Mass scale-expanded segment for SWIFT-isolated ions, including assignment of two monoisotopic molecular formulas. Top: Mass scale-expanded segments for ions 1 or 2Da higher in mass than the assigned monoisotopic ions. Appearance of the <sup>18</sup>O and <sup>34</sup>S isotopologues serves to validate the monoisotopic compositional assignments. Also note the detection of isotopologues containing  ${}^{2}H, {}^{13}C,$  or  ${}^{15}N.$ 



Fig. 6. Magnitude (top) and absorption (bottom) (+) ESI 9.4 T FT-ICR mass spectral segments for a North American petroleum crude oil. Note the absorption-mode resolution of three pairs of peaks (compositions differing by  $C_3$  *vs.* SH<sub>4</sub>, 0.0034Da and H<sub>4</sub>N<sub>2</sub>S<sub>2</sub> *vs.*  $C_4O_3$ , 0.0032Da) that are unresolved in magnitude-mode display.

necessarily produce the same number of ions each time. Because space charge shifts FT-ICR MS peak positions, any variation in ion number will degrade mass measurement accuracy and mass resolving power. We therefore introduced "conditional averaging." Basically, the idea is to average only those mass spectra whose summed peak heights are the same to within a specified range (say, 10%). It is possible to reduce rms mass error by a factor of 2–3 by such conditional averaging. 26)

Mass measurement error may be further reduced by adding a third term to the ICR frequency-to-*m*/*z* conversion (mass "calibration") equation, and (for compositionally complex mass spectra) by separate calibration of multiple mass spectral segments.<sup>29)</sup> The combination of all of the above data treatments has been shown to reduce rms mass measurement random error to ∼25ppb, and corresponding systematic error to ∼5ppb (*i.e.*, essentially negligible). 29)

## **Isotopic ne structure for validation of elemental composition assignment**

It is widely appreciated that the confidence in assigning a monoisotopic elemental composition (mass, M) can be improved by accurate determination of the combined heights of unresolved isotopologue signals at  $(M+1)$  and  $(M+2)$ .<sup>30)</sup> It is less widely understood that even higher confidence can follow from ultrahigh-resolution mass measurement of individual isotopologues, as shown in Fig. 5. For example, identification of the <sup>34</sup>S isotopologue can unequivocally confirm the presence and number of <sup>32</sup>S in the monoisotopic

species. 31)

**Phase correction to yield an absorption-mode spectrum** We have recently described broadband phase correction<sup>32,33)</sup> to yield an absorption-mode FT-ICR mass spectrum, whose resolving power can be up to twice that for conventional magnitude-mode display.<sup>34)</sup> Moreover, close mass doublets partially or unresolved in magnitude mode may become resolved in absorption mode (see Fig. 6), thereby increasing the number of correctly assigned elemental compositions. Thus, the improvement in resolution based on absorption-mode display is not only quantitative; it can be qualitative (*i.e.*, being able to assign a peak or not). Finally, the improvement due to phase correction is much greater near the baseline of the peak, where (*e.g.*) a Lorentzian magnitude-mode peak can be ten-fold wider than for absorption mode.

### **Acknowledgments**

This work was supported by the National Science Foundation (DMR-06-54118; CHE-1016942; CHE-1019193), the Florida State University Future Fuels Institute, the BP/Gulf of Mexico Research Initiative, and the State of Florida.

#### **REFERENCES**

1) A. G. Marshall, C. L. [Hendrickson,](http://dx.doi.org/10.1002/(SICI)1098-2787(1998)17:1%3C1::AID-MAS1%3E3.0.CO;;2-K) G. S. Jackson. Fourier transform ion cyclotron resonance mass [spectrometry:](http://dx.doi.org/10.1002/(SICI)1098-2787(1998)17:1%3C1::AID-MAS1%3E3.0.CO;;2-K) A primer. *Mass [Spectrom.](http://dx.doi.org/10.1002/(SICI)1098-2787(1998)17:1%3C1::AID-MAS1%3E3.0.CO;;2-K) Rev.* 17: 1–35, 1998.

- 2) R. P. Rodgers, A. G. Marshall. Petroleomics: Chemistry of the underworld. *Proc. Natl. Acad. Sci. U.S.A.* 105: 1–6, 2008.
- 3) S. G. Valeja, N. K. Kaiser, F. Xian, C. L. [Hendrickson,](http://dx.doi.org/10.1021/ac202429c) J. C. Rouse, A. G. Marshall. Unit mass [resolution](http://dx.doi.org/10.1021/ac202429c) for an intact 148 kDa therapeutic [monoclonal](http://dx.doi.org/10.1021/ac202429c) antibody by FT-ICR mass spectrometry. *Anal. Chem.* 83: [8391–8395,](http://dx.doi.org/10.1021/ac202429c) 2011.
- 4) E. N. Nikolaev, R. M. A. Heeren, A. M. Popov, A. V. [Pozdneev,](http://dx.doi.org/10.1002/rcm.3234) K. S. Chingin. Realistic [modeling](http://dx.doi.org/10.1002/rcm.3234) of ion cloud motion in a Fourier [transform](http://dx.doi.org/10.1002/rcm.3234) ion cyclotron resonance cell by use of a particlein-cell approach. *Rapid Commun. Mass Spectrom.* 21: [3527–3546,](http://dx.doi.org/10.1002/rcm.3234) [2007.](http://dx.doi.org/10.1002/rcm.3234)
- 5) F. E. Leach 3rd, A. [Kharchenko,](http://dx.doi.org/10.1016/j.jasms.2009.10.001) R. M. A. Heeren, E. N. Nikolaev, I. J. Amster. Comparison of [particle-in-cell](http://dx.doi.org/10.1016/j.jasms.2009.10.001) simulations with [experimentally](http://dx.doi.org/10.1016/j.jasms.2009.10.001) observed frequency shifts between ions of the same [mass-to-charge](http://dx.doi.org/10.1016/j.jasms.2009.10.001) in Fourier transform ion cyclotron resonance mass [spectrometry.](http://dx.doi.org/10.1016/j.jasms.2009.10.001) *J. Am. Soc. Mass Spectrom.* 21: [203–208,](http://dx.doi.org/10.1016/j.jasms.2009.10.001) 2010.
- 6) G. Vladimirov, C. L. [Hendrickson,](http://dx.doi.org/10.1007/s13361-011-0268-8) G. T. Blakney, A. G. Marshall, R. M. A. Heeren, E. N. Nikolaev. Fourier [transform](http://dx.doi.org/10.1007/s13361-011-0268-8) ion cyclotron resonance mass [resolution](http://dx.doi.org/10.1007/s13361-011-0268-8) and dynamic range limits [calculated](http://dx.doi.org/10.1007/s13361-011-0268-8) by computer modeling of ion cloud motion. *J. Am. Soc. Mass [Spectrom.](http://dx.doi.org/10.1007/s13361-011-0268-8)* 23: 375–384, 2012.
- 7) M. B. [Comisarow,](http://dx.doi.org/10.1016/0009-2614(74)89137-2) A. G. Marshall. Fourier transform ion cyclotron resonance [spectroscopy.](http://dx.doi.org/10.1016/0009-2614(74)89137-2) *Chem. Phys. Lett.* 25: 282–283, [1974.](http://dx.doi.org/10.1016/0009-2614(74)89137-2)
- 8) S. C. Beu, D. A. Laude, Jr. Open trapped ion cell [geometries](http://dx.doi.org/10.1016/0168-1176(92)80006-M) for [FT/ICR/MS.](http://dx.doi.org/10.1016/0168-1176(92)80006-M) *Int. J. Mass Spectrom. Ion Process.* 112: 215–230, [1992.](http://dx.doi.org/10.1016/0168-1176(92)80006-M)
- 9) A. G. Marshall. [Milestones](http://dx.doi.org/10.1016/S1387-3806(00)00324-9) in Fourier transform ion cyclotron resonance mass [spectrometry](http://dx.doi.org/10.1016/S1387-3806(00)00324-9) technique development. *Int. J. Mass [Spectrom.](http://dx.doi.org/10.1016/S1387-3806(00)00324-9)* 200: 331–336, 2000.
- 10) A. G. Marshall, C. L. [Hendrickson.](http://dx.doi.org/10.1016/S1387-3806(01)00588-7) Fourier transform ion cyclotron resonance detection: Principles and [experimental](http://dx.doi.org/10.1016/S1387-3806(01)00588-7) configurations. *Int. J. Mass [Spectrom.](http://dx.doi.org/10.1016/S1387-3806(01)00588-7)* 215: 59–75, 2002.
- 11) I. A. Boldin, E. N. Nikolaev. Fourier [transform](http://dx.doi.org/10.1002/rcm.4838) ion cyclotron resonance cell with dynamic [harmonization](http://dx.doi.org/10.1002/rcm.4838) of the electric field in the whole volume by shaping of the [excitation](http://dx.doi.org/10.1002/rcm.4838) and detection electrode assembly. *Rapid [Commun.](http://dx.doi.org/10.1002/rcm.4838) Mass Spectrom.* 25: [122–126,](http://dx.doi.org/10.1002/rcm.4838) 2011.
- 12) A. V. [Tolmachev,](http://dx.doi.org/10.1016/j.jasms.2008.01.006) E. W. Robinson, S. Wu, H. Kang, N. M. Lourette, L. Paša-Tolić, R. D. Smith. [Trapped-ion](http://dx.doi.org/10.1016/j.jasms.2008.01.006) cell with improved DC potential [harmonicity](http://dx.doi.org/10.1016/j.jasms.2008.01.006) for FT-ICR MS. *J. Am. Soc. Mass [Spectrom.](http://dx.doi.org/10.1016/j.jasms.2008.01.006)* 19: 586–597, 2008.
- 13) N. K. Kaiser, J. J. Savory, A. M. [McKenna,](http://dx.doi.org/10.1021/ac201546d) J. P. Quinn, C. L. [Hendrickson,](http://dx.doi.org/10.1021/ac201546d) A. G. Marshall. Electrically compensated FT-ICR cell for complex mixture analysis. *Anal. Chem.* 83: [6907–6910,](http://dx.doi.org/10.1021/ac201546d) [2011.](http://dx.doi.org/10.1021/ac201546d)
- 14) A. M. Brustkern, D. L. Rempel, M. L. Gross. An [electrically](http://dx.doi.org/10.1016/j.jasms.2008.05.016) [compensated](http://dx.doi.org/10.1016/j.jasms.2008.05.016) trap designed to eighth order for FT-ICR mass [spectrometry.](http://dx.doi.org/10.1016/j.jasms.2008.05.016) *J. Am. Soc. Mass Spectrom.* 19: 1281–1285, 2008.
- 15) G. Gabrielse, L. Haarsma, S. L. Rolston. [Open-endcap](http://dx.doi.org/10.1016/0168-1176(89)85027-X) Penning traps for high precision [experiments.](http://dx.doi.org/10.1016/0168-1176(89)85027-X) *Int. J. Mass Spectrom. Ion Process.* 88: [319–332,](http://dx.doi.org/10.1016/0168-1176(89)85027-X) 1989.
- 16) E. N. [Nikolaev,](http://dx.doi.org/10.1007/s13361-011-0125-9) I. A. Boldin, R. Jertz, G. Baykut. Initial experimental [characterization](http://dx.doi.org/10.1007/s13361-011-0125-9) of a new ultra-high resolution FTICR cell with dynamic [harmonization.](http://dx.doi.org/10.1007/s13361-011-0125-9) *J. Am. Soc. Mass Spectrom.* 22: [1125–1133,](http://dx.doi.org/10.1007/s13361-011-0125-9) 2011.
- 17) I. A. Boldin, E. N. Nikolaev. Theory of peak [coalescence](http://dx.doi.org/10.1002/rcm.4231) in Fourier transform ion cyclotron resonance mass [spectrometry.](http://dx.doi.org/10.1002/rcm.4231) *Rapid Commun. Mass Spectrom.* 23: [3213–3219,](http://dx.doi.org/10.1002/rcm.4231) 2009.
- 18) G. Vladimirov, Y. Kostyukevich, A. G. Marshall, C. L. Hendrickson, G. T. Blakney, E. N. Nikolaev. In influence of different components of magnetic field inhomogeneity on cyclotron motion coherence at very high magnetic field. Proc. 58th Amer. Soc. Mass Spectrom. Annual Conf. on Mass Spectrometry & Allied Topics, Salt Lake City, UT, 23–27 May, 2010.
- 19) N. K. Kaiser, J. P. Quinn, G. T. Blakney, C. L. [Hendrickson,](http://dx.doi.org/10.1007/s13361-011-0141-9) A.

G. Marshall. A novel 9.4 tesla FTICR mass [spectrometer](http://dx.doi.org/10.1007/s13361-011-0141-9) with improved sensitivity, mass [resolution,](http://dx.doi.org/10.1007/s13361-011-0141-9) and mass range. *J. Am. Soc. Mass Spectrom.* 22: [1343–1351,](http://dx.doi.org/10.1007/s13361-011-0141-9) 2011.

- 20) M. W. Senko, C. L. [Hendrickson,](http://dx.doi.org/10.1016/S1044-0305(97)00126-8) M. R. Emmett, S. D.-H. Shi, A. G. Marshall. External [accumulation](http://dx.doi.org/10.1016/S1044-0305(97)00126-8) of ions for enhanced electrospray [ionization](http://dx.doi.org/10.1016/S1044-0305(97)00126-8) Fourier transform ion cyclotron resonance mass [spectrometry.](http://dx.doi.org/10.1016/S1044-0305(97)00126-8) *J. Am. Soc. Mass Spectrom.* 8: 970–976, 1997.
- 21) B. E. Wilcox, C. L. [Hendrickson,](http://dx.doi.org/10.1016/S1044-0305(02)00622-0) A. G. Marshall. Improved ion [extraction](http://dx.doi.org/10.1016/S1044-0305(02)00622-0) from a linear octopole ion trap: SIMION analysis and experimental [demonstration.](http://dx.doi.org/10.1016/S1044-0305(02)00622-0) *J. Am. Soc. Mass Spectrom.* 13: [1304–1312,](http://dx.doi.org/10.1016/S1044-0305(02)00622-0) 2002.
- 22) A. G. Marshall, T.-C. L. Wang, T. L. Ricca. Tailored [excitation](http://dx.doi.org/10.1021/ja00312a015) for Fourier transform ion cyclotron resonance mass [spectrom](http://dx.doi.org/10.1021/ja00312a015)etry. *J. Am. Chem. Soc.* 107: [7893–7897,](http://dx.doi.org/10.1021/ja00312a015) 1985.
- 23) G. T. Blakney, C. L. [Hendrickson,](http://dx.doi.org/10.1016/j.ijms.2011.03.009) A. G. Marshall. Predator data station: A fast data [acquisition](http://dx.doi.org/10.1016/j.ijms.2011.03.009) system for advanced FT-ICR MS [experiments.](http://dx.doi.org/10.1016/j.ijms.2011.03.009) *Int. J. Mass Spectrom.* 306: 246–252, 2011.
- 24) L. Chen, C. E. Cottrell, A. G. Marshall. Effect of [signal-to-noise](http://dx.doi.org/10.1016/0169-7439(86)80025-9) ratio and number of data points upon precision in [measurement](http://dx.doi.org/10.1016/0169-7439(86)80025-9) of peak [amplitude,](http://dx.doi.org/10.1016/0169-7439(86)80025-9) position, and width in Fourier transform [spectrometry.](http://dx.doi.org/10.1016/0169-7439(86)80025-9) *Chemom. Intell. Lab. Syst.* 1: 51–58, 1986.
- 25) C. M. [Yanofsky,](http://dx.doi.org/10.1021/ac050640q) A. W. Bell, S. Lesimple, F. Morales, T. T. Lam, G. T. Blakney, A. G. [Marshall,](http://dx.doi.org/10.1021/ac050640q) B. Carrillo, K. Lekpor, D. Boismenu, R. E. Kearney. [Multicomponent](http://dx.doi.org/10.1021/ac050640q) internal recalibration of an [LC-FTICR-MS](http://dx.doi.org/10.1021/ac050640q) analysis employing a partially characterized complex peptide mixture: [Systematic](http://dx.doi.org/10.1021/ac050640q) and random errors. *Anal. Chem.* 77: [7246–7254,](http://dx.doi.org/10.1021/ac050640q) 2005.
- 26) A. G. Marshall, G. T. Blakney, C. L. Hendrickson, A. M. Mc-Kenna, R. P. Rodgers. Fourier transform ion cyclotron resonance: The mass spectrometry zenith. New Alternatives in High-Resolution Mass Spectrometry Symp., Pittcon 2012, Orlando, FL, 11–15 March, 2012.
- 27) J. H. Beynon. Qualitative analysis of organic [compounds](http://dx.doi.org/10.1038/174735a0) by mass [spectrometry.](http://dx.doi.org/10.1038/174735a0) *Nature* 174: 735–737, 1954.
- 28) S. Kim, R. P. Rodgers, A. G. [Marshall.](http://dx.doi.org/10.1016/j.ijms.2006.02.001) Truly 'exact' mass: Elemental [composition](http://dx.doi.org/10.1016/j.ijms.2006.02.001) can be determined uniquely from molecular mass [measurement](http://dx.doi.org/10.1016/j.ijms.2006.02.001) at ∼0.1mDa accuracy for molecules up to ∼500Da. *Int. J. Mass [Spectrom.](http://dx.doi.org/10.1016/j.ijms.2006.02.001)* 251: 260–265, 2006.
- 29) J. J. Savory, N. K. Kaiser, A. M. [McKenna,](http://dx.doi.org/10.1021/ac102943z) F. Xian, G. T. Blakney, R. P. Rodgers, C. L. [Hendrickson,](http://dx.doi.org/10.1021/ac102943z) A. G. Marshall. Parts-perbillion Fourier [transform](http://dx.doi.org/10.1021/ac102943z) ion cyclotron resonance mass measurement accuracy with a "walking" [calibration](http://dx.doi.org/10.1021/ac102943z) equation. *Anal. Chem.* 83: [1732–1736,](http://dx.doi.org/10.1021/ac102943z) 2011.
- 30) T. Kind, O. Fiehn. [Metabolomic](http://dx.doi.org/10.1186/1471-2105-7-234) database annotations *via* query of elemental [compositions:](http://dx.doi.org/10.1186/1471-2105-7-234) Mass accuracy is insufficient even at less than 1ppm. *BMC [Bioinformatics](http://dx.doi.org/10.1186/1471-2105-7-234)* 7: 234, 2006.
- 31) S. D.-H. Shi, C. L. [Hendrickson,](http://dx.doi.org/10.1073/pnas.95.20.11532) A. G. Marshall. Counting individual sulfur atoms in a protein by [ultrahigh-resolution](http://dx.doi.org/10.1073/pnas.95.20.11532) Fourier transform ion cyclotron resonance mass [spectrometry:](http://dx.doi.org/10.1073/pnas.95.20.11532) Experimental [resolution](http://dx.doi.org/10.1073/pnas.95.20.11532) of isotopic fine structure in proteins. *Proc. Natl. Acad. Sci. U.S.A.* 95: [11532–11537,](http://dx.doi.org/10.1073/pnas.95.20.11532) 1998.
- 32) S. C. Beu, G. T. Blakney, J. P. Quinn, C. L. [Hendrickson,](http://dx.doi.org/10.1021/ac049733i) A. G. Marshall. [Broadband](http://dx.doi.org/10.1021/ac049733i) phase correction of FT-ICR mass spectra *via* [simultaneous](http://dx.doi.org/10.1021/ac049733i) excitation and detection. *Anal. Chem.* 76: [5756–5761,](http://dx.doi.org/10.1021/ac049733i) 2004.
- 33) F. Xian, C. L. [Hendrickson,](http://dx.doi.org/10.1021/ac101091w) G. T. Blakney, S. C. Beu, A. G. Marshall. [Automated](http://dx.doi.org/10.1021/ac101091w) broadband phase correction of Fourier transform ion cyclotron [resonance](http://dx.doi.org/10.1021/ac101091w) mass spectra. *Anal. Chem.* 82: [8807–8812,](http://dx.doi.org/10.1021/ac101091w) 2010.
- 34) A. G. Marshall, M. B. [Comisarow,](http://dx.doi.org/10.1063/1.438196) G. Parisod. Relaxation and spectral line shape in Fourier [transform](http://dx.doi.org/10.1063/1.438196) ion cyclotron resonance [spectroscopy.](http://dx.doi.org/10.1063/1.438196) *J. Chem. Phys.* 71: 4434–4444, 1979.
- 35) V. V. Lobodin, R. P. Rodgers, A. G. Marshall. Analysis of complex organic mixtures by FT-ICR mass spectrometry. Petroleomics. In Comprehensive Environmental Mass Spectrometry (Ed.: A. Lebedev), ILM Publications, St. Albans, Hertfordshire, UK, pp. 415–442, 2012.