

Arginine Cools the Inflamed Gut

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Maintenance of immunophysiological homeostasis and regulation of gut barrier function are essential for the defense of the host. Severe alterations, including infections and chronic inflammation, have been associated with increased intestinal permeability, leading to deregulation of gut function and homeostasis. To establish and reinforce this crucial balance, the intestinal metabolism plays a key role and has to be tightly regulated since in the human intestinal mucosa, the protein fractional synthesis rate is approximately 50% per day. This value is higher than that of other major metabolically active tissues, such as liver and muscle, and depends on the accessibility of the metabolic precursor pool (1). The amino acid L-arginine (L-Arg) is a central intestinal metabolite, both as a constituent of protein synthesis and as a regulatory molecule limiting intestinal alterations and maintaining immunophysiological functions (1, 2). Infection-associated L-Arg deficiency has been shown to contribute to immunopathology, and clinical trials involving L-Arg administration have shown substantial decreases in inflammation and infectious complications (3). In this issue, [Chau and colleagues](#) demonstrate that malaria-associated hypoargininemia impairs intestinal barrier function and predisposes the host to coinfection with *Salmonella*. Increasing bioavailability of L-Arg through oral supplementation ameliorates intestinal inflammation and pathology, demonstrating that pharmacological intervention at the metabolic-precursor level can be utilized to regulate mucosal immunohomeostasis (4).

METABOLISM AND CATABOLISM OF L-ARGININE

L-Arg is derived from the diet, turnover of proteins, and endogenous production through synthesis from L-citrulline (L-Cit) and successive actions of argininosuccinate synthetase (AS) and argininosuccinate lyase (AL), the third and fourth enzymes of the urea cycle. The major site of L-Arg metabolism is the liver, where L-Arg generated in the urea cycle is rapidly converted to urea and ornithine by arginases, however, with no net synthesis of L-Arg. Although synthesis of L-Arg from L-Cit can occur in many cell types, a major part of endogenous synthesis occurs via “the intestinal-renal axis,” a postnatally established collaboration between epithelial cells of the small intestine and proximal tubule cells of the kidney. In adult animals, L-Cit is produced primarily by intestinal epithelial cells from NH₃, CO₂, and ornithine by carbamyl-phosphate synthetase I and ornithine transcarbamylase, the first two enzymes of the urea cycle, and is supplied to the kidney and probably to other tissues for synthesis of L-Arg (2, 5, 6).

L-Arg is a crucial amino acid that serves to modulate immune responses through conversion by several intracellular classes of enzymes, with isoforms of arginase and nitric oxide synthase (NOS) being the two major enzyme families exerting key immunological functions (7). However, catabolism of extracellular L-Arg requires active and regulated uptake via specific cationic amino acid transporters (CAT) or heteromeric amino acid transporters (HAT) that act as H⁺-coupled symporters or antiporters (8). There are two isoforms of arginase, cytosolic arginase I (Arg1)

and mitochondrial arginase II. Arg1 is abundant in liver as part of the urea cycle, and only low levels are found in extrahepatic tissue. In contrast, Arg2 is abundant mostly in extrahepatic tissues and cells such as kidney, brain, gut, and hematopoietic cells, with a main role in the production of L-ornithine (L-Orn), L-proline (L-Pro), and L-glutamate (L-Glu). In addition, three isozymes of NOS catalyze the production of NO and L-Cit from L-Arg. While neuronal NOS (nNOS or NOS1) and endothelial NOS (eNOS or NOS3) are mostly constitutively expressed, inducible NOS (iNOS or NOS2) expression is positively regulated by proinflammatory cytokines and microbial constituents activating pattern recognition molecules (7).

While, in healthy adults, the level of endogenous L-Arg synthesis is sufficiently great that L-Arg is not an essential amino acid, catabolic stress as well as dysfunction of the kidneys or small intestine can lead to hypoargininemia, a condition where levels of endogenous L-Arg may not suffice to meet metabolic demands. As such, L-Arg homeostasis is maintained primarily by modulation of L-Arg catabolism rather than L-Arg synthesis. Accordingly, L-Arg is classified as a semiessential or conditionally essential amino acid (9). Although it is known that regulation of L-Arg catabolism is influenced by (i) the expression levels of L-Arg transporters and L-Arg-converting enzymes, (ii) their cofactor availability, and (iii) substrate competition between the enzymatic systems (10), the molecular mechanisms and consequences of hypoargininemia on the host's immune response are poorly understood.

HYPARGININEMIA CAUSES ALTERED INTESTINAL IMMUNOHOMEOSTASIS

L-Arg availability in the alimentary tract has previously been shown to play a key role in intestinal immunohomeostasis upon catabolic stress induced by inflammation and infection (7). L-Arg supplementation attenuated the degree of tissue damage in intestinal ischemia and promoted healing of intestinal mucosa (11, 12). Similarly, in a murine model of dextran sulfate sodium (DSS)-induced colitis, oral L-Arg treatment improved clinical parameters by dampening proinflammatory responses and inflammatory cell infiltration, leading to improved mucosal integrity and enhanced epithelial cell migration in an iNOS-dependent manner (13). In addition, infection by the intestinal pathogen *Citrobacter rodentium* was shown to cause a significant decrease in the serum L-Arg concentration, with associated infection-induced immunopa-

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thology being partially reversed after L-Arg supplementation (14). In this issue, Chau and colleagues demonstrate now that L-Arg availability is also critical for maintaining intestinal barrier function during malaria parasite infection, as malaria-associated hypoargininemia impairs intestinal barrier function and predisposes to coinfection with *Salmonella* (4).

It is known that during malaria parasite infection, L-Arg bioavailability drops due to enhanced destruction by host- and parasite-encoded arginases, utilization by host nitric oxide synthase (NOS), and increased scavenging of NO by cell-free hemoglobin from lysed red blood cells (RBCs) (15–17). Importantly, impaired bioavailability of L-Arg and NO has been shown to alter endothelial dysfunction in infected adults and has been suggested to be a major contributing factor to pathogenesis during infection (18). In a rodent model of experimental malaria, exogenous NO provision restored NO-mediated signaling in the brain, decreased systemic inflammation, reduced vascular leakage and hemorrhage in the brain, and provided marked protection (19). In addition, L-Arg infusion into adults infected with falciparum malaria parasites reversed infection-induced endothelial dysfunction (20), highlighting the importance of L-Arg bioavailability for the host's homeostasis during infection.

Immunopathology during malaria infection is also observed in the gastrointestinal (GI) tract. During severe *Plasmodium falciparum* infection, sequestration of parasitized RBCs and capillary blockage are prominent in intestinal villi and are associated with ischemia, malabsorption, and increased GI permeability (21–23). In addition, 50% of individuals with uncomplicated malaria have GI disturbances (24). Several features of malaria pathology suggest that the increased risk of developing bacteremia during malaria parasite and nontyphoidal *Salmonella* serotype (NTS) coinfection results from malaria-induced damage to the intestinal epithelium (25–27). Chau and colleagues demonstrate that malaria-associated hypoargininemia causes increased circulating and tissue histamine levels, mast cell activation, and ileal mastocytosis, which collectively alter intestinal epithelial integrity, predisposing the host to secondary bacterial infections (4).

REGULATION OF INTESTINAL BARRIER FUNCTION AND INFLAMMATION BY L-ARGININE

Malaria-induced hypoargininemia enhances parasite sequestration and basophil transmigration, as well as mast cell activation, which collectively result in an immunopathology that resembles allergic inflammation, leading to increased intestinal permeability (28). Although, their distinct contribution remains to be explored, it is likely that altered activities of L-Arg-catabolizing enzyme families, arginase(s), and NOS contribute to the observed immunopathology. It can be postulated that the activity of arginase through conversion of L-Arg into L-Orn enhances epithelial barrier function (29), while NOS activity confers an anti-inflammatory state through regulation of energy metabolism (30).

L-Orn produced by arginase is used by ornithine decarboxylase (ODC) to produce the polyamine putrescine, which is then converted into the polyamines spermidine and spermine by constitutively expressed spermidine and spermine synthases, respectively. Notably, polyamines are associated with mucosal protection in the GI tract and with intestinal epithelial cell migration. L-Orn can also be acted upon by ornithine aminotransferase (OAT) to produce L-proline (L-Pro), an important precursor in collagen syn-

thesis, and is involved in wound healing and cell migration in fibroblasts and epithelial cells (10).

NOS activity has recently been shown to dampen inflammation through regulation of myeloid and lymphoid cell activation. Nitric oxide produced by iNOS in inflammatory monocytes and dendritic cells regulates inflammatory cytokine production, cell differentiation, and survival (31–33). In addition, the regulated release of NO by nonhematopoietic stromal cells controls the expansion of activated T cells (34–36), and T cell-intrinsic iNOS activity regulates polarization of antigen-specific CD4⁺ T helper cells (37–39). Moreover, expression of iNOS by intestinal B cells is critical for immunoglobulin A class switch recombination and intestinal immunohomeostasis at steady state and upon intestinal infection (40). Future studies will have to detail the tissue-, cell-, and context-dependent role of iNOS and the molecular mechanisms that regulate nonhematopoietic, myeloid, and lymphoid immune cell function at steady state and during inflammation and infection. Modulating arginase- and NOS-mediated pathways through regulation of the bioavailability of L-Arg or its precursor L-Cit by oral supplementation provides an efficient and practical strategy to dampen intestinal inflammation and pathology, demonstrating that pharmacological intervention at the metabolic-precursor level can be utilized to regulate mucosal immunohomeostasis.

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REFERENCES

- Bertrand J, Goichon A, Dechelotte P, Coeffier M. 2012. Regulation of intestinal protein metabolism by amino acids. *Amino Acids* doi:10.1007/s00726-012-1325-8.
- Morris SM, Jr. 2007. Arginine metabolism: boundaries of our knowledge. *J. Nutr.* 137:1602S–1609S.
- Drover JW, Dhaliwal R, Weitzel L, Wischmeyer PE, Ochoa JB, Heyland DK. 2011. Perioperative use of arginine-supplemented diets: a systematic review of the evidence. *J. Am. Coll. Surg.* 212:385–399.
- Chau JY, Tiffany CM, Nimishakavi S, Lawrence JA, Pakpour N, Mooney JP, Lokken KL, Caughey GH, Tsolis RM, Luckhart S. 2013. Malaria-associated L-arginine deficiency induces mast cell-associated disruption to intestinal barrier defenses against nontyphoidal *Salmonella* bacteremia. *Infect. Immun.* 81:3515–3526.
- Morris SM, Jr. 2002. Regulation of enzymes of the urea cycle and arginine metabolism. *Annu. Rev. Nutr.* 22:87–105.
- Mori M, Gotoh T. 2004. Arginine metabolic enzymes, nitric oxide and infection. *J. Nutr.* 134:2820S–2825S, 2853S.
- Das P, Lahiri A, Chakravorty D. 2010. Modulation of the arginase pathway in the context of microbial pathogenesis: a metabolic enzyme moonlighting as an immune modulator. *PLoS Pathog.* 6:e1000899. doi:10.1371/journal.ppat.1000899.
- Closs EI, Simon A, Vekony N, Rotmann A. 2004. Plasma membrane transporters for arginine. *J. Nutr.* 134:2752S–2759S, 2765S–2767S.
- Wu G, Meininger CJ, Knabe DA, Bazer FW, Rhoads JM. 2000. Arginine nutrition in development, health and disease. *Curr. Opin. Clin. Nutr. Metab. Care* 3:59–66.
- Luiking YC, Ten Have GA, Wolfe RR, Deutz NE. 2012. Arginine de novo and nitric oxide production in disease states. *Am. J. Physiol. Endocrinol. Metab.* 303:E1177–E1189.
- Fotiadis C, Adamis S, Misiakos EP, Genetzakis M, Antonakis PT, Tsekouras DK, Gorgoulis VG, Zografos GC, Papalois A, Fotinou M, Perrea D. 2007. The prophylactic effect of L-arginine in acute ischaemic

- colitis in a rat model of ischaemia/reperfusion injury. *Acta Chir. Belg.* 107:192–200.
12. Cross RK, Wilson KT. 2003. Nitric oxide in inflammatory bowel disease. *Inflamm. Bowel Dis.* 9:179–189.
 13. Coburn LA, Gong X, Singh K, Asim M, Scull BP, Allaman MM, Williams CS, Rosen MJ, Washington MK, Barry DP, Piazuelo MB, Casero RA, Jr, Chaturvedi R, Zhao Z, Wilson KT. 2012. L-Arginine supplementation improves responses to injury and inflammation in dextran sulfate sodium colitis. *PLoS One* 7:e33546. doi:10.1371/journal.pone.0033546.
 14. Gobert AP, Cheng Y, Akhtar M, Mersey BD, Blumberg DR, Cross RK, Chaturvedi R, Drachenberg CB, Boucher JL, Hacker A, Casero RA, Jr, Wilson KT. 2004. Protective role of arginase in a mouse model of colitis. *J. Immunol.* 173:2109–2117.
 15. Lopansri BK, Anstey NM, Weinberg JB, Stoddard GJ, Hobbs MR, Levesque MC, Mwaikambo ED, Granger DL. 2003. Low plasma arginine concentrations in children with cerebral malaria and decreased nitric oxide production. *Lancet* 361:676–678.
 16. Olszewski KL, Morrissey JM, Wilinski D, Burns JM, Vaidya AB, Rabinowitz JD, Llinas M. 2009. Host-parasite interactions revealed by Plasmodium falciparum metabolomics. *Cell Host Microbe* 5:191–199.
 17. Omodeo-Sale F, Cortelezzi L, Vommario Z, Scaccabarozzi D, Dondorp AM. 2010. Dysregulation of L-arginine metabolism and bioavailability associated to free plasma heme. *Am. J. Physiol. Cell Physiol.* 299:C148–C154.
 18. Weinberg JB, Lopansri BK, Mwaikambo E, Granger DL. 2008. Arginine, nitric oxide, carbon monoxide, and endothelial function in severe malaria. *Curr. Opin. Infect. Dis.* 21:468–475.
 19. Gramaglia I, Sobolewski P, Meays D, Contreras R, Nolan JP, Frangos JA, Intaglietta M, van der Heyde HC. 2006. Low nitric oxide bioavailability contributes to the genesis of experimental cerebral malaria. *Nat. Med.* 12:1417–1422.
 20. Yeo TW, Lampah DA, Gitawati R, Tjitra E, Kenangalem E, McNeil YR, Darcy CJ, Granger DL, Weinberg JB, Lopansri BK, Price RN, Duffull SB, Celermajer DS, Anstey NM. 2007. Impaired nitric oxide bioavailability and L-arginine reversible endothelial dysfunction in adults with falciparum malaria. *J. Exp. Med.* 204:2693–2704.
 21. Seydel KB, Milner DA, Jr, Kamiza SB, Molyneux ME, Taylor TE. 2006. The distribution and intensity of parasite sequestration in comatose Malawian children. *J. Infect. Dis.* 194:208–215.
 22. Molyneux ME, Looareesuwan S, Menzies IS, Grainger SL, Phillips RE, Wattanagoon Y, Thompson RP, Warrell DA. 1989. Reduced hepatic blood flow and intestinal malabsorption in severe falciparum malaria. *Am. J. Trop. Med. Hyg.* 40:470–476.
 23. Wilairatana P, Meddings JB, Ho M, Vannaphan S, Looareesuwan S. 1997. Increased gastrointestinal permeability in patients with Plasmodium falciparum malaria. *Clin. Infect. Dis.* 24:430–435.
 24. Sowunmi A, Ogundahunsi OA, Falade CO, Gbotosho GO, Oduola AM. 2000. Gastrointestinal manifestations of acute falciparum malaria in children. *Acta Trop.* 74:73–76.
 25. Cunnington AJ, de Souza JB, Walther M, Riley EM. 2012. Malaria impairs resistance to Salmonella through heme- and heme oxygenase-dependent dysfunctional granulocyte mobilization. *Nat. Med.* 18:120–127.
 26. Tabu C, Breiman RF, Ochieng B, Aura B, Cosmas L, Audi A, Olack B, Bigogo G, Ongus JR, Fields P, Mintz E, Burton D, Oundo J, Feikin DR. 2012. Differing burden and epidemiology of non-Typhi Salmonella bacteremia in rural and urban Kenya, 2006–2009. *PLoS One* 7:e31237. doi:10.1371/journal.pone.0031237.
 27. Roux CM, Butler BP, Chau JY, Paixao TA, Cheung KW, Santos RL, Luckhart S, Solis RM. 2010. Both hemolytic anemia and malaria parasite-specific factors increase susceptibility to nontyphoidal Salmonella enterica serovar Typhimurium infection in mice. *Infect. Immun.* 78:1520–1527.
 28. Maarsingh H, Zaagsma J, Meurs H. 2008. Arginine homeostasis in allergic asthma. *Eur. J. Pharmacol.* 585:375–384.
 29. Wang WW, Qiao SY, Li DF. 2009. Amino acids and gut function. *Amino Acids* 37:105–110.
 30. Dai Z, Wu Z, Yang Y, Wang J, Satterfield MC, Meininger CJ, Bazer FW, Wu G. 2013. Nitric oxide and energy metabolism in mammals. *Biofactors.* doi:10.1002/biof.1099.
 31. Everts B, Amiel E, van der Windt GJ, Freitas TC, Chott R, Yarasheski KE, Pearce EL, Pearce EJ. 2012. Commitment to glycolysis sustains survival of NO-producing inflammatory dendritic cells. *Blood* 120:1422–1431.
 32. Giordano D, Li C, Suthar MS, Draves KE, Ma DY, Gale M, Jr, Clark EA. 2011. Nitric oxide controls an inflammatory-like Ly6C(hi)PDCA1+ DC subset that regulates Th1 immune responses. *J. Leukoc. Biol.* 89:443–455.
 33. Mishra BB, Rathinam VA, Martens GW, Martinot AJ, Kornfeld H, Fitzgerald KA, Sasseti CM. 2013. Nitric oxide controls the immunopathology of tuberculosis by inhibiting NLRP3 inflammasome-dependent processing of IL-1beta. *Nat. Immunol.* 14:52–60.
 34. Lukacs-Kornek V, Malhotra D, Fletcher AL, Acton SE, Elpek KG, Tayalia P, Collier AR, Turley SJ. 2011. Regulated release of nitric oxide by nonhematopoietic stroma controls expansion of the activated T cell pool in lymph nodes. *Nat. Immunol.* 12:1096–1104.
 35. Khan O, Headley M, Gerard A, Wei W, Liu L, Krummel MF. 2011. Regulation of T cell priming by lymphoid stroma. *PLoS One* 6:e26138. doi:10.1371/journal.pone.0026138.
 36. Siegert S, Huang HY, Yang CY, Scarpellino L, Carrie L, Essex S, Nelson PJ, Heikenwalder M, Acha-Orbea H, Buckley CD, Marsland BJ, Zehn D, Luther SA. 2011. Fibroblastic reticular cells from lymph nodes attenuate T cell expansion by producing nitric oxide. *PLoS One* 6:e27618. doi:10.1371/journal.pone.0027618.
 37. Niedbala W, Besnard AG, Jiang HR, Alves-Filho JC, Fukada SY, Nascimento D, Mitani A, Pushparaj P, Alqahtani MH, Liew FY. 2013. Nitric oxide-induced regulatory T cells inhibit th17 but not Th1 cell differentiation and function. *J. Immunol.* 191:164–170.
 38. Obermajer N, Wong JL, Edwards RP, Chen K, Scott M, Khader S, Kolls JK, Odunsi K, Billiar TR, Kalinski P. 2013. Induction and stability of human Th17 cells require endogenous NOS2 and cGMP-dependent NO signaling. *J. Exp. Med.* 210:1433–1445.
 39. Yang J, Zhang R, Lu G, Shen Y, Peng L, Zhu C, Cui M, Wang W, Arnaboldi P, Tang M, Gupta M, Qi CF, Jayaraman P, Zhu H, Jiang B, Chen SH, He JC, Ting AT, Zhou MM, Kuchroo VK, Morse HC, III, Ozato K, Sikora AG, Xiong H. 2013. T cell-derived inducible nitric oxide synthase switches off TH17 cell differentiation. *J. Exp. Med.* 210:1447–1462.
 40. Fritz JH, Rojas OL, Simard N, McCarthy DD, Hapfelmeier S, Rubino S, Robertson SJ, Larjani M, Gosselin J, Ivanov II, Martin A, Casellas R, Philpott DJ, Girardin SE, McCoy KD, Macpherson AJ, Paige CJ, Gommerman JL. 2012. Acquisition of a multifunctional IgA+ plasma cell phenotype in the gut. *Nature* 481:199–203.