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## Regulation of cortical and hippocampal 5-HT<sub>1A</sub> receptor function by corticosterone in GR+/- mice

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### SUMMARY

Our objective in the present study was to examine 5-HT<sub>1A</sub> receptor function in prefrontal cortex and hippocampus of GR+/- mice, which appear to be an appropriate murine model of depression. 5-HT<sub>1A</sub> receptor function was determined by measuring [<sup>35</sup>S]GTP S binding stimulated by the 5-HT<sub>1A</sub> receptor agonist 8-OH-DPAT (1 μM), an indication of the capacity of the receptor to activate G proteins. 5-HT<sub>1A</sub> receptor expression was determined by measuring the binding of [<sup>3</sup>H]8-OH-DPAT (2 nM). We observed no effect of the constitutive reduction in GR on 5-HT<sub>1A</sub> receptor-stimulated [<sup>35</sup>S]GTP S binding or 5-HT<sub>1A</sub> receptor binding sites. Corticosterone treatment (10 mg/kg, sc once daily for 21 days) of wild-type mice resulted in a decrease in 5-HT<sub>1A</sub> receptor function in prefrontal cortex [8-OH-DPAT-stimulated [<sup>35</sup>S]GTP S binding (% above basal), vehicle-treated: 39±4.9; corticosterone-treated: 17±2.8], but not in hippocampus. The constitutive reduction in GR expression prevented the down-regulation of 5-HT<sub>1A</sub> receptor function in frontal cortex by chronic corticosterone administration. In contrast, corticosterone treatment of GR+/- mice resulted in an increase in 5-HT<sub>1A</sub> receptor function in hippocampus which reached statistical significance in CA2/3 region [8-OH-DPAT-stimulated [<sup>35</sup>S]GTP S binding (% above basal), vehicle-treated: 41±9.7; corticosterone-treated: 94±23]. These changes seem to be evoked by a combined effect of high corticosterone levels and GR deficiency. Although GR+/- mice do not exhibit changes in baseline corticosterone, the constitutive deficiency in GR appears to have unmasked regulatory effects of elevated corticosterone in the maintenance of 5-HT<sub>1A</sub> receptor function in prefrontal cortex and hippocampus.

### Keywords

glucocorticoid receptor; quantitative autoradiography; serotonin; corticosterone; major depression; [<sup>35</sup>S]GTP S binding

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### CONTRIBUTORS

PG designed the study and wrote the protocol. MAV performed the animal experiments. JGH supervised the collection of data and undertook the statistical analyses. JGH wrote the first draft of the manuscript. All authors managed literature searches, and contributed to and have approved the final manuscript.

### CONFLICT OF INTEREST

None

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## INTRODUCTION

Neurotransmitter and hormonal responses associated with acute stress are adaptive in the short-term in that they are necessary to promote homeostasis. However, repeated exposure to stress over time may lead to pathophysiological changes due to chronic elevations in glucocorticoid levels. The deleterious effects of prolonged exposure to elevated glucocorticoid levels, which include neurochemical and morphological changes in forebrain structures, are associated with cognitive deficits and affective disorders, specifically major depression (Herman et al., 2003; Sapolsky, 2003; McEwen, 2007).

Hyperactivity of the hypothalamic-pituitary-adrenal (HPA) system and elevated plasma cortisol levels associated with major depressive disorder may be due to diminished glucocorticoid receptor (GR) function or expression, and as a consequence, deficient feedback regulation of cortisol (Holsboer, 2000; Pariante, 2006). GR heterozygous (GR<sup>+/-</sup>) mice, with a 50% reduction in GR expression, are indistinguishable from wild-type control mice at baseline, but exhibit increased sensitivity to stress and dysregulation of the HPA axis. GR<sup>+/-</sup> mice display increased helplessness after stress exposure, a behavioral correlate of depression in mice, and increased stress-induced plasma corticosterone levels (Ridder et al., 2005). GR<sup>+/-</sup> mice also exhibit abnormal responses in the dexamethasone suppression test and dexamethasone/corticotropin-releasing hormone test (DEX/CRH test), similar to what is observed clinically in severely depressed patients (Ridder et al., 2005). Consistent with the neurotrophin hypothesis of depression, GR<sup>+/-</sup> mice exhibit a down regulation of brain-derived neurotrophic factor (BDNF) protein in hippocampus (Ridder et al., 2005).

As GR<sup>+/-</sup> mice appear to be an appropriate murine model of depression (Ridder et al., 2005), our objective in the present study was to examine serotonin (5-hydroxytryptamine, 5-HT) 5-HT<sub>1A</sub> receptor function in GR<sup>+/-</sup> mice. 5-HT<sub>1A</sub> receptors are present in high density in cortical and limbic areas, where they are post-synaptic to serotonergic neurons. The distribution of 5-HT<sub>1A</sub> receptors in brain is consistent with a role for this 5-HT receptor in cognitive or integrative functions, as well as in emotional states. We have found that 5-HT<sub>1A</sub> receptor function in hippocampus, but not in frontal cortical areas, is attenuated in BDNF<sup>+/-</sup> mice and mice with a forebrain-specific reduction in BDNF (Hensler et al., 2003, 2007). As hippocampal BDNF protein levels are also diminished in GR<sup>+/-</sup> mice, we hypothesized that hippocampal 5-HT<sub>1A</sub> receptor function would be attenuated in GR<sup>+/-</sup> mice. Moreover, we were interested in examining the effect of chronic corticosterone treatment, a means to mimic chronic stress, on 5-HT<sub>1A</sub> receptor function and expression. 5-HT<sub>1A</sub> receptor function was determined by measuring 5-HT<sub>1A</sub> receptor agonist 8-OH-DPAT stimulated [<sup>35</sup>S]GTP S binding to G proteins, an indication of the capacity of the receptor to activate G proteins. 5-HT<sub>1A</sub> receptor expression was determined by measuring the binding of the agonist radioligand [<sup>3</sup>H]8-OH-DPAT to 5-HT<sub>1A</sub> receptor sites. As cortical and limbic structures play a key role in the integration and association of stressful stimuli with previous experiences, and therefore are brain regions of particular interest when examining the long-term effects of repeated exposure to stress, we focused our attention in the current study on prefrontal cortex and dorsal hippocampus.

## MATERIALS AND METHODS

### Mice

GR-heterozygous mice (GR<sup>+/-</sup>) were generated as previously described to obtain F1 hybrid mice with the same background as used in behavioral and neurochemical studies (Ridder et al., 2005; Schulte-Herbrüggen et al., 2007; Trajkovska et al., 2009). Male GR<sup>+/-</sup> mice and wildtype littermate controls were 4–5 months old at the beginning of experiments. Mice

were housed individually under a reverse phase 12:12 hour light/dark cycle, with *ad libitum* access to food and water. German animal welfare authorities approved all experiments.

### Corticosterone Treatment

Mice were injected subcutaneously with either sesame oil vehicle or corticosterone (10 mg/kg) once daily for 21 days. This corticosterone treatment of mice results in attenuation of 5-HT<sub>1A</sub> receptor function in the dorsal raphe, consistent with the effects of chronic stress on somatodendritic autoreceptor function (Hensler et al., 2007).

### Tissue Preparation

Mice were sacrificed 24 hours after the last injection of vehicle or corticosterone. Brains were rapidly removed and stored at  $-80^{\circ}\text{C}$ . Coronal sections of 20  $\mu\text{m}$  thickness were cut in a cryostat microtome at the level of the frontal cortex (plates 13–14) and dorsal hippocampus (plates 46–47) according to the atlas of the mouse brain (Paxinos and Franklin, 1997), and thaw-mounted onto gelatin-coated glass slides. After desiccation at  $4^{\circ}\text{C}$ , sections were stored at  $-80^{\circ}\text{C}$ .

### [<sup>35</sup>S]GTP $\gamma$ S Autoradiography

Autoradiography of 8-OH-DPAT-stimulated [<sup>35</sup>S]GTP  $\gamma$  S binding in brain sections was performed as previously described (Hensler et al., 2007). Sections were incubated in assay buffer containing 40 pM [<sup>35</sup>S]GTP  $\gamma$  S (1250 Ci/mmol, Dupont/NEN) either in the absence or in the presence of ( $\pm$ )8-OH-DPAT (1  $\mu\text{M}$ ) (Tocris). Basal [<sup>35</sup>S]GTP  $\gamma$  S binding was defined in the absence of 8-OH-DPAT. Nonspecific [<sup>35</sup>S]GTP  $\gamma$  S binding was defined in the absence of 8-OH-DPAT and in the presence of 10  $\mu\text{M}$  GTP  $\gamma$  S (Roche/Boehringer-Manheim). The incubation was stopped by two washes for 5 minutes each in ice-cold HEPES buffer, followed by a brief immersion in ice-cold de-ionized water. Dried sections were exposed to Kodak Biomax MR film (Amersham) for 48 hours.

### [<sup>3</sup>H]8-OH-DPAT Autoradiography

Autoradiography of the binding of [<sup>3</sup>H]8-OH-DPAT to 5-HT<sub>1A</sub> receptors in brain sections was performed as described with slight modification (Hensler et al., 1991). Sections incubated in assay buffer containing 2 nM [<sup>3</sup>H]8-OH-DPAT (210 Ci/mmol, Amersham). Nonspecific binding was defined by incubating adjacent sections in the presence of 10  $\mu\text{M}$  WAY-100635 (Sigma/RBI, St. Louis, MO). Incubation was terminated by two washes for 5 minutes each in ice-cold buffer, followed by a dip in ice-cold de-ionized water. Dried sections exposed to Kodak BioMax MR Film (Amersham, Piscataway, NJ) for 9 weeks.

### Image Analysis

Analysis of the digitized autoradiograms was performed using the image analysis program NIH Image, version 1.47 (NIH, Bethesda, MD) as previously described (see Hensler et al., 1991; Hensler et al., 2007).

### Data Analysis

Statistical comparisons were made by two-way ANOVA with genotype and treatment as factors. F values reaching significance ( $p < 0.05$ ) were evaluated further by post hoc analysis using the Newman-Keuls test. Statistical tests were performed using Statistica software (version 4.1, Statsoft, Tulsa, OK).

## RESULTS

### Differences in 5-HT<sub>1A</sub> Receptor Function: wild-type versus GR+/- Mice

[<sup>35</sup>S]GTP S binding stimulated by the 5-HT<sub>1A</sub> receptor agonist 8-OH-DPAT was assessed in the dorsal hippocampus (Figure 1, panels A, B, C). In CA1, dentate gyrus and CA2/3 regions we found no significant main effect of genotype (CA1:  $F_{1,22}=1.71$ ,  $p=0.20$ ; CA2/3:  $F_{1,22}=1.35$ ,  $p=0.26$ ; dentate gyrus:  $F_{1,22}=1.83$ ,  $p=0.19$ ). There was also no difference between wild-type and GR+/- mice in the number of 5-HT<sub>1A</sub> receptor binding sites as measured by the binding of [<sup>3</sup>H]8-OH-DPAT [main effect of genotype, CA1:  $F_{1,22}=0.134$ ,  $p=0.72$ ; CA2/3:  $F_{1,22}=1.74$ ,  $p=0.20$ ; dentate gyrus:  $F_{1,22}=0.033$ ,  $p=0.86$  (Figure 1, panels D, E, F). Thus the deficiency in GR did not alter 5-HT<sub>1A</sub> receptor binding or function, specifically the capacity of 5-HT<sub>1A</sub> receptors to activate G proteins, in dorsal hippocampus.

We also assessed 8-OH-DPAT-stimulated [<sup>35</sup>S]GTP S binding in the prefrontal cortex (Figure 2A). Here we found a significant main effect of genotype ( $F_{1,22}=5.86$ ,  $p=0.02$ ). Post hoc analysis, however, revealed no significant difference between vehicle-treated wild-type and GR+/- mice in this brain region ( $p=0.88$ ). There was also no difference between genotypes in the number of 5-HT<sub>1A</sub> receptors. As observed for the hippocampus, the deficiency in GR did not alter 5-HT<sub>1A</sub> receptor binding or function in the prefrontal cortex.

### Effect of Corticosterone Treatment

There was no significant main effect of corticosterone treatment on 8-OH-DPAT-stimulated [<sup>35</sup>S]GTP S binding in subregions of dorsal hippocampus (CA1:  $F_{1,22}=3.73$ ,  $p=0.07$ ; CA2/3:  $F_{1,22}=3.87$ ,  $p=0.06$ ; dentate gyrus:  $F_{1,22}=1.63$ ,  $p=0.21$ ) (see Figure 1). The interaction between factors (genotype and treatment), although not significant for CA1 ( $F_{1,22}=2.47$ ,  $p=0.13$ ) or dentate gyrus ( $F_{1,22}=1.63$ ,  $p=0.18$ ), was significant for the CA2/3 region ( $F_{1,22}=4.59$ ,  $p=0.04$ ). Post hoc analysis indicated that repeated corticosterone treatment of GR+/- mice resulted in a significant increase in 8-OH-DPAT-stimulated [<sup>35</sup>S]GTP S binding in CA2/3 region (Figure 1, panel B). The effect of corticosterone treatment on the binding of [<sup>3</sup>H]8-OH-DPAT to 5-HT<sub>1A</sub> receptors sites was also measured (Figure 1, panel D, E, F). There was no significant main effect of treatment (CA1:  $F_{1,22}=0.018$ ,  $p=0.89$ ; CA2/3:  $F_{1,22}=3.29$ ,  $p=0.08$ ; dentate gyrus:  $F_{1,22}=0.09$ ,  $p=0.76$ ) or interaction between factors (genotype and treatment) (CA1:  $F_{1,22}=0.129$ ,  $p=0.72$ ; CA2/3:  $F_{1,22}=0.11$ ,  $p=0.74$ ; dentate gyrus:  $F_{1,22}=0.33$ ,  $p=0.57$ ). Thus although chronic corticosterone treatment of GR+/- mice resulted in an increase in the capacity of 5-HT<sub>1A</sub> receptors in dorsal hippocampus to activate G proteins, this was not accompanied by an increase in 5-HT<sub>1A</sub> receptor binding sites.

In the prefrontal cortex, we found a significant main effect of corticosterone treatment on 8-OH-DPAT stimulated [<sup>35</sup>S]GTP S binding ( $F_{1,22}=8.28$ ,  $p=0.01$ ) and a significant interaction between factors (genotype and treatment) ( $F_{1,22}=6.92$ ,  $p=0.02$ ). Post hoc analysis indicated that corticosterone treatment of wild-type, but not of GR+/- mice resulted in a significant attenuation in 5-HT<sub>1A</sub> receptor-stimulated [<sup>35</sup>S]GTP S binding in this brain region (Figure 2A). We observed no significant main effect of corticosterone treatment on [<sup>3</sup>H]8-OH-DPAT binding to 5-HT<sub>1A</sub> receptors sites ( $F_{1,22}=0.018$ ,  $p=0.89$ ). Although the interaction between factors (genotype and treatment) was significant ( $F_{1,22}=9.58$ ,  $p=0.01$ ), post hoc analysis indicated no significant difference between groups ( $p>0.05$ ) (Figure 2B). Thus although chronic corticosterone administration resulted in a significant attenuation in 5-HT<sub>1A</sub> receptor function in prefrontal cortex of wild-type mice, this was not accompanied by a down-regulation of 5-HT<sub>1A</sub> receptor binding sites.

## DISCUSSION

In the present study we found no effect of the constitutive reduction in GR on 5-HT<sub>1A</sub> receptor function or binding sites in hippocampus or in prefrontal cortex. The lack of effect of genotype on 5-HT<sub>1A</sub> receptor-stimulated [<sup>35</sup>S]GTP S binding in hippocampus was somewhat unexpected given the reduction in hippocampal BDNF in GR+/- mice (Ridder et al., 2005) and our previous findings of reduced hippocampal 5-HT<sub>1A</sub> receptor-stimulated [<sup>35</sup>S]GTP S binding in mice deficient in BDNF (Hensler et al., 2003, 2007). Our previous observations in BDNF deficient mice have led us to speculate that BDNF may support or promote 5-HT<sub>1A</sub> receptor function in the hippocampus. However, in GR+/- mice the hypothesized effect of reduced BDNF to decrease 5-HT<sub>1A</sub> receptor function appears to be countered or opposed by the constitutive reduction in GR.

Chronic administration of corticosterone to wild-type mice resulted in an attenuation of 5-HT<sub>1A</sub> receptor-stimulated [<sup>35</sup>S]GTP S binding in prefrontal cortex. Interestingly, the constitutive reduction in GR expression prevented the down-regulation of 5-HT<sub>1A</sub> receptor function in frontal cortex by chronic corticosterone administration. Thus, the negative regulatory effect of corticosterone administration on 5-HT<sub>1A</sub> function in frontal cortex appears to be lost in GR+/- mice. In hippocampus, corticosterone treatment of GR+/- mice resulted in an increase in 5-HT<sub>1A</sub> receptor-stimulated [<sup>35</sup>S]GTP S binding in hippocampus. These functional changes in 5-HT<sub>1A</sub> receptors seem to be evoked by a combined effect of high corticosterone levels and GR deficiency. The constitutive deficiency in GR may have unmasked regulatory effects of corticosterone in the maintenance of 5-HT<sub>1A</sub> receptor function in prefrontal cortex and hippocampus.

GR, with low affinity for glucocorticoids, have been implicated in the negative feedback regulation of the HPA axis when glucocorticoid levels are high (de Kloet 2007). Mineralocorticoid receptors (MR), with high affinity for glucocorticoids, are believed to play a role in homeostasis as they are activated when glucocorticoid levels are low (de Kloet 2007). In the hippocampus, 5-HT<sub>1A</sub> receptor number and function are under stringent regulation by MR and GR. Occupancy of MR and GR by high levels of corticosterone results in the potentiation of 5-HT<sub>1A</sub> receptor-mediated responses in CA1 pyramidal neurons. By contrast, the strict inhibitory regulation of hippocampal 5-HT<sub>1A</sub> receptor expression and function by corticosterone is predominantly mediated by MR (Kuroda et al., 1994; Meijer and de Kloet, 1995; Hesen and Joëls, 1996). Human imaging and postmortem studies indicate that hippocampal and cortical 5-HT<sub>1A</sub> receptors are down regulated in major depressive disorder (see Drevets et al., 2007, and references therein), an effect believed to be mediated predominantly through MR as a result of elevated basal corticosteroid levels (Meijer et al., 1997). Hippocampal MR expression in cytosol and nuclear fractions, as determined by Western blot analysis, is not altered in GR+/- mice (Riva and Gass, unpublished observations). That 5-HT<sub>1A</sub> receptors are unchanged in GR+/- mice suggests that MR function in these mice, at least with regard to the regulation of 5-HT<sub>1A</sub> receptors, is not altered.

MRs and GRs function together in a complementary manner to promote adaptive responses to stress, eg. facilitation of memory storage, behavioral adaptation, and preparation for future events. 5-HT<sub>1A</sub> receptors in cortical and limbic structures may play an important role in these adaptive processes. Perturbation of the MR/GR balance, specifically decreased GR function, is implicated in major depressive disorder, and appears to be involved in increased vulnerability to stress (Ridder et al., 2005; de Kloet et al., 2007). However, in the present study, the constitutive reduction in GR appears to have unmasked a positive regulatory effect of corticosterone, most likely through activation of both MR and GR, in the

maintenance of 5-HT<sub>1A</sub> receptor function in hippocampus and prefrontal cortex. Future experiments have to clarify by which molecular mechanisms this regulation is mediated.

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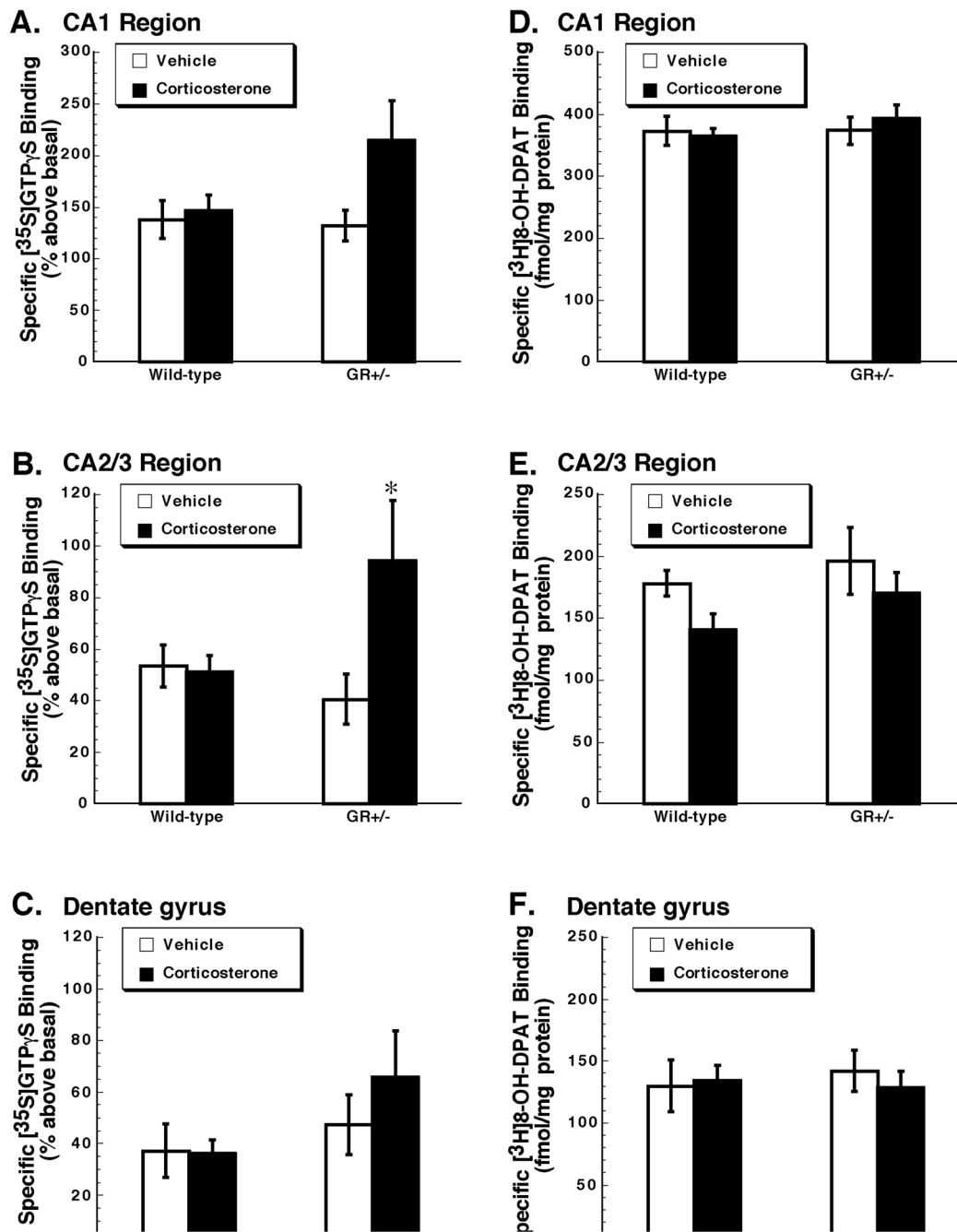
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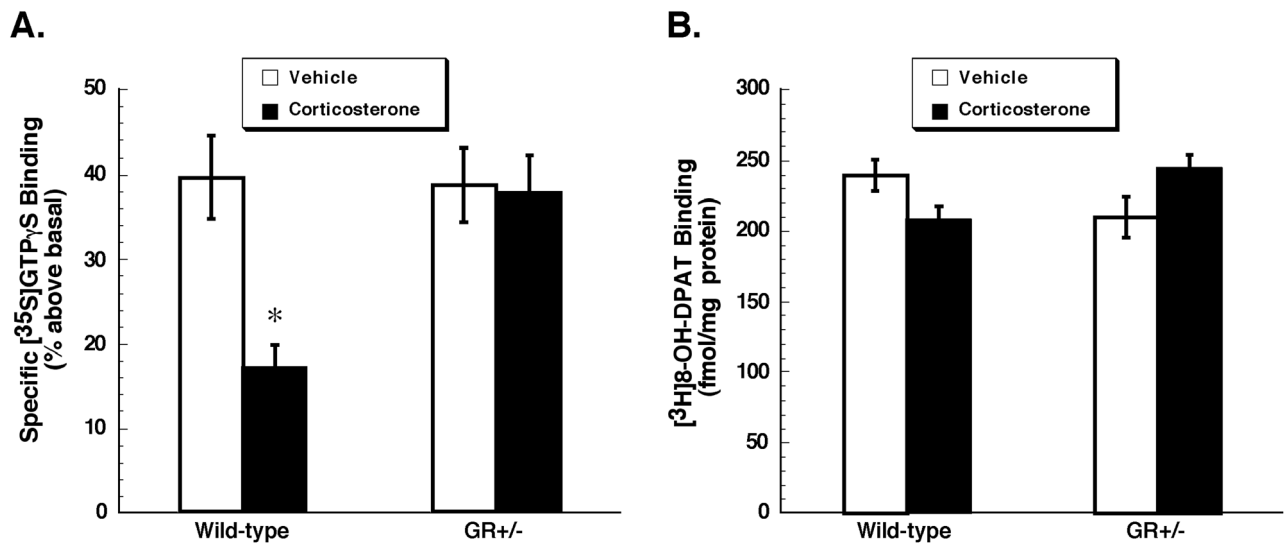
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**Figure 1. Chronic corticosterone treatment increases 5-HT<sub>1A</sub> receptor function in the hippocampus of GR+/- but not wild-type mice**

[<sup>35</sup>S]GTP S binding (A, B, C) was stimulated by the 5-HT<sub>1A</sub> receptor agonist 8-OH-DPAT (1 μM), and is expressed as % above basal. Specific binding of [<sup>3</sup>H]8-OH-DPAT (2 nM) (D, E, F) is expressed as fmol/mg protein. Panels A and B show data for the CA1 region. Panels C and D show corresponding data in the CA2/3 region. Panels E and F show data for the dentate gyrus. Presented are the mean ± S.E.M. n = 6, wild-type mice treated with vehicle; n = 8, wild-type mice treated with corticosterone; n = 6, GR+/- mice treated with vehicle; n = 6, GR+/- mice treated with corticosterone. Data were analyzed by two-way ANOVA. \*p<0.05 when compared with vehicle-treated GR+/- mice, post hoc Newman-Keuls test.





**Figure 2. Chronic corticosterone treatment decreases 5-HT<sub>1A</sub> receptor function in prefrontal cortex of wild-type but not GR<sup>+/-</sup> mice**

[<sup>35</sup>S]GTP S binding (A) was stimulated by the 5-HT<sub>1A</sub> receptor agonist 8-OH-DPAT (1 μM), and is expressed as % above basal. Specific binding of [<sup>3</sup>H]8-OH-DPAT (2 nM) (B) is expressed as fmol/mg protein. Presented are mean ± S.E.M., n= 6 wild-type mice treated with vehicle; n = 8, wild-type mice treated with corticosterone; n = 6, GR<sup>+/-</sup> mice treated with vehicle; n = 6, GR<sup>+/-</sup> mice treated with vehicle. Data were analyzed by two-way ANOVA. \*p<0.05 when compared with vehicle-treated wild-type mice, post hoc Newman-Keuls test.