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cMET and phospho-cMET protein levels in breast cancers and survival outcomes

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Abstract

Purpose—To evaluate cMET and phospho-cMET (p-cMET) levels in breast cancer subtypes and its impact on survival outcomes.

Experimental Design—We measured protein levels of cMET and p-cMET in 257 breast cancers using reverse phase protein array. Regression tree method and Martingale residual plots were applied to find best cutoff point for high and low levels. Kaplan-Meier survival curves were used to estimate relapse-free (RFS) and overall (OS) survival. Cox proportional hazards models were fit to determine associations of cMET/p-cMET with outcomes after adjustment for other characteristics.

Results—Median age was 51years. There were 140 (54.5%) hormone receptor (HR)-positive, 53 (20.6%) HER2-positive and 64 (24.9%) triple-negative tumors. Using selected cutoffs, 181 (70.4%) and 123 (47.9%) cancers had high levels of cMET and p-cMET, respectively. There were no significant differences in mean expression of cMET (P<0.128) and p-cMET (P<0.088) by breast cancer subtype. Dichotomized cMET and p-cMET level was a significant prognostic factor for RFS (HR:2.44,95% CI:1.34-4.44,P=0.003 and HR:1.64,95% CI:1.04-2.60,P=0.033) and OS (HR:3.18,95% CI:1.43-7.11,P=0.003 and HR:1.92,95% CI:1.08-3.44,P=0.025). Within breast cancer subtypes, high cMET levels were associated with worse RFS (P=0.014) and OS (P=0.006) in HR-positive tumors, and high p-cMET levels were associated with worse RFS (P=0.019) and OS (P=0.014) in HER2-positive breast cancers. In multivariable analysis patients with high cMET had a significantly higher risk of recurrence (HR:2.06; 95% CI:1.08-3.94,P=0.028) and death (HR:

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2.81; 95%CI:1.19-6.64,P=0.019). High p-cMET level was associated with higher risk of recurrence (HR:1.79,95%CI 1.08-2.95.77,P=0.020).

Conclusions—High levels of cMET and p-cMET were seen in all breast cancer subtypes and correlated with poor prognosis.

Keywords

cMET; phospho-cMET; breast cancer prognosis; breast cancer subtype

Introduction

The proto-oncogene *cMET* (mesenchymal-epithelial transition factor gene) on chromosome 7q31 encodes a receptor tyrosine kinase (RTK) which acts as the receptor for hepatocyte growth factor (HGF) or scatter factor (SF) (1-3). This RTK is a single-pass type I transmembrane heterodimer protein made of a disulfide bridge that links an -chain (50 kDa) and a -chain (145 kDa) (4). The -chain in cMET traverses the cell membrane and contains the cytoplasmic kinase domain and a docking site, comprised of tyrosine residues Y1349 and Y1356 (5). HGF binding to the extracellular domain activates the intrinsic kinase activity which phosphorylates the tyrosines at the carboxy-terminal docking site. Phosphorylated cMET (phospho-cMET or p-cMET) binds Grb2 (Growth factor receptorbound protein 2) and Gab1 (GRB2-associated-binding protein 1) and activates downstream signaling molecules such as Phosphatidylinositol 3-kinase (PI3K/AKT) and Mitogenactivated protein kinase (ERK/MAPK) pathways (6, 7). Through these cellular signaling pathways HGF/cMET interaction plays a key role in cellular proliferation, survival, migration and invasion (8-10). The HGF/cMET axis contributes a critical physiological function in embryogenesis, angiogenesis and wound healing (11, 12).

Multiple germline and somatic mutations in tumors result in anomalous signaling via diverse mechanisms such as activating mutations of *cMET* gene and over-expression of HGF/SF or cMET (13-15). The HGF/cMET signaling cascade has been repeatedly shown to be dysregulated in a variety of tumors such as lung, kidney, head & neck and colorectal cancers (16-17). Increased HGF/cMET signaling in these tumors correlates with poor outcomes (18-20). Additionally, p-cMET has also been shown to be an important predictor of tumor aggressiveness, metastatic potential and poor survival (21, 22). In summary, aberrant HGF/ cMET activation results in an aggressive phenotype and is associated with tumor progression and poor outcomes. The key role as a mitogenic, motogenic and angiogenic molecule makes HGF/cMET inhibition an attractive therapeutic strategy in cancer (23, 24).

Overexpression of cMET has been shown to contribute to the development of invasive phenotype during progression of breast cancer *in vivo and in vitro* (25). In addition, Lindemann et al. showed that differential expression of cMET between tumor and adjacent normal tissue was associated with aggressive ductal carcinoma in situ (DCIS) phenotype (26). Prior studies have shown that protein tyrosine phosphorylation profiling of basal-like breast carcinomas is characterized by elevated tyrosine phosphorylation of cMET (27, 28). Hence, the HGF/cMET axis seems to play a significant role in tumor progression in breast cancer (29).

We performed an analysis of primary breast cancer specimens to evaluate the protein levels of total cMET and p-cMET by breast cancer subtype using reverse phase protein arrays (RPPA), and their correlation with patient outcome.

Materials and Methods

Patients and tumor samples

Fine needle aspirates from 257 primary invasive breast cancers were obtained and snap frozen. All specimens were collected under Institutional Review Board (IRB)-approved protocols. The breast tumors were classified into three clinically relevant subtypes defined by immunohistochemistry (IHC) for estrogen receptor (ER) and progesterone receptor (PR) status and by IHC or fluorescent in situ hybridization (FISH) for HER2 status as per American Society of Clinical Oncology and College of American Pathologists (ASCO/CAP) guidelines (30). These subtypes were defined by the dominant traditional prognostic molecular marker (ER, PR and HER2). Hormone receptor-positive (HR-positive) tumors were ER-positive and/or PR-positive and HER2-negative. Similarly, HER2-positive group included all HER2 positive tumors irrespective of hormone receptor status. Triple negative subtype included all cases that were ER/PR and HER2 negative.

The samples used for the study were archived samples from previously collected tumor specimens from patient treated at M D Anderson Cancer Center between 1986 and 2007. The corresponding clinical data was obtained from the Breast Cancer Management System database at MD Anderson Cancer Center. Missing information from the database was collected by chart review.

Reverse phase protein lysate microarray (RPPA)

Protein was extracted from the human tumors and RPPA was performed in our laboratory as described previously (31). Briefly, lysis buffer was used to lyse frozen tumors by homogenization. Tumor lysates were normalized to 1 µg/µL concentration using bicinchoninic acid assay and boiled with 1% SDS, and the supernatants were manually diluted in six or eight 2-fold serial dilutions with lysis buffer. An Aushon Biosystems (Burlington, MA) 2470 arrayer created 1,056 sample arrays on nitrocellulose-coated FAST slides (Schleicher & Schuell BioScience, Inc.) from the serial dilutions. A slide was then probed with validated primary cMET and p-cMET antibodies (Cell Signaling Technology, Danvers, MA) and the signal was amplified using a DakoCytomation-catalyzed system. The antibodies for cMET (Mouse) and p-cMET (Rabbit, Y1235) were used at a dilution of 1:250 for RPPA. A secondary antibody was used as a starting point for amplification. The slides were scanned, analyzed, and quantitated using Microvigene software (VigeneTech Inc.) to generate serial dilution-signal intensity curves for each sample with the logistic fit model: $\ln(y) = a + (b - a) / (1 + \exp \{c^*[d - \ln(x)]\})$. A representative natural logarithmic value of each sample curve on the slide (curve average) was then used as a relative quantification of the amount of each protein in each sample. The level of cMET and p-cMET in each sample was expressed as a log mean centered value after correction for protein loading using the average expression levels of over 150 proteins as previously described (31).

Statistical Methods

Boxplots were generated for original and log2 transformed expressions of total cMET and pcMET by breast cancer subtypes. The original expressions were right-skewed but the log2 transformation data was normally distributed. Hence, all following statistical analyses were based on the log2 transformation of the original expression values. P values less than 0.05 were considered statistically significant and all tests were two-sided. Statistical analyses were done with R statistical software version 2.12.0 (R Development Core Team, 2010, Vienna, Austria).

Mean and standard deviations were generated for total cMET and p-cMET by tumor subtypes. Linear regression models were used to determine if the mean total cMET and p-

cMET expression was different by tumor subtypes. Martingale residual plots with lowess smooth for Cox's model for total cMET and p-cMET separately as covariate by tumor subtypes suggested a non-linear effect of total and p-cMET. A regression tree method was applied to find the best cutoff point for total cMET and p-cMET expression. Combining the results from martingale residual plots and regression trees, total cMET expression was divided into high level (>0) expression and low level (0) expression. Similarly, p-cMET was divided as high level (>0.35) and low level (0.35). Patient and tumor characteristics including age, stage, grade and subtype were tabulated between high and low level expressions of total cMET and p-cMET individually. Groups were then compared with the Chi-square tests (32).

Overall survival (OS) and corresponding censoring were computed in months from diagnosis to death for each patient. Relapse-free survival (RFS) was regarded as the time to first relapse after diagnosis. Median RFS and OS were estimated nonparametrically with the use of Kaplan-Meier curves by patient characteristics and levels of total cMET and p-cMET expression and compared by the log-rank statistic. Log-rank tests were used to evaluate the hazard ratio by total cMET and p-cMET expression levels among all patients and patients within each subtype. Cox proportional hazards models were fit to determine the association of cMET and p-cMET levels with the risk of recurrence and death after adjustment for other patient and disease characteristics.

Results

Median patient age was 51 years (range 23-85 years). There were a total of 140 (54.5%) hormone receptor (HR)-positive tumors, 53 (20.6%) HER2-positive tumors and 64 (24.9%) triple receptor-negative (TN) tumors. Using the selected cutoffs, a total of 181 (70.4%) and 123 (47.9%) patients had high expression of cMET and p-cMET, respectively.

Patient and clinic characteristics by levels of total cMET and p-cMET are summarized in table 1. There was no statistically significant difference in clinical or pathologic parameters in patients with high or low level of total cMET. Patients with high p-cMET expression tended to be older (Age > 50: 60.2% vs. 41.0%, P = .003) and had fewer high grade tumors (Grade III: 60% vs. 72.2% P = .046).

No significant differences in mean levels of total cMET expressions (P = 0.128) and pcMET expressions (P = 0.088) were seen between different tumor subtypes, as seen in table 2. At a median follow up of 42.23 months (5.17-277.77 months), there were 76 (30%) relapses and 50 (20%) deaths.

Table 3 summarizes the median RFS estimates by c-MET and p-cMET expression and by tumor subtypes. Dichotomized total cMET expression (cutoff point 0) was a significant prognostic factor for RFS (HR: 2.44, 95% CI: 1.34-4.44, P = 0.003). Estimated 5-year RFS rates were 61.3% (95% CI: 53.2%-70.7%) and 78.9% (95% CI: 68.6%-90.8%) for patients with high cMET and low cMET level, respectively (P = 0.003). Likewise, dichotomized p-cMET expression (cutoff point 0.35) was also a significant prognostic factor for RFS (HR: 1.64, 95% CI: 1.04-2.60, P = 0.033) and estimated 5-year RFS rates for patients with high p-cMET and low p-cMET levels were 58.9% (95% CI: 49.1%-70.7%) and 73.8% (95% CI: 65.6%-83.1%), respectively (P = 0.033). Total cMET was also a significant predictor of RFS within the HR-positive subtype (HR: 3.44, 95% CI: 1.21-9.81, P = 0.014). In contrast, p-cMET was a significant predictor of RFS within the HER2-positive subtype (HR: 3.02, 95% CI: 1.15-7.96, P = 0.019). The Kaplan-Meier survival curves for RFS for all patients and by breast tumor subtypes are as shown in figure 1. Although, there was a trend towards worse

RFS with high cMET levels (HR 2.36; 95% CI: 0.86-6.51) in triple-negative subtype, this did not reach statistical significance (P = 0.086).

Table 3 summarizes the median OS estimates by c-MET and p-cMET expression and by tumor subtypes. At the time of analysis, 207 of the 257 patients (80.5%) were still alive. As was seen with RFS analysis, dichotomized cMET level was a significant prognostic factor of OS (HR: 3.18, 95% CI: 1.43-7.11, P = 0.003). Estimated 5-year OS rates were 72.4% (95% CI: 64.7%-81.0%) and 93.3% (95% CI: 87.8%-99.2%) for patients with high cMET and low cMET levels, respectively (P = 0.003). Dichotomized p-cMET level was a significant prognostic factor of OS (HR: 1.92, 95% CI: 1.08-3.44, P = 0.025). The estimated 5-year OS rates for patients with high p-cMET and low p-cMET levels were 72.4% (95% CI: 63.7%-82.3%) and 85.8% (95% CI: 79.4%-92.7%), respectively (P = 0.006) and p-cMET (HR: 5.49, 95% CI: 1.20-25.10, P = 0.014) were significant predictor of OS within HR-positive tumors and HER2-positive tumors, respectively. The Kaplan-Meier survival curves for OS for all patients and by subtypes are as shown in figure 2. Although, there was a trend towards worse OS with high p-cMET levels (HR 2.02; 95% CI: 0.80-5.13) in triple-negative subtype, this did not reach statistical significance (P = 0.128).

Multivariable models for RFS and OS are summarized on table 4. After adjustment for patient factors, tumor characteristics and treatment, patients with tumors expressing high levels of cMET had a significant higher risk of recurrence (HR 2.06; 95% CI: 1.08-3.94; P = 0.028) and death (HR 2.81; 95% CI: 1.19-6.64; P = 0.019) compared to patients with low cMET levels. Also, patients with tumors expressing high levels of p-cMET had a significant higher risk of recurrence (HR 1.79; 95% C: 1.08-2.95; P = 0.020) compared to patients with high p-cMET levels.

To evaluate whether cMET confers radioresistance, we performed an exploratory sub-group analysis among 164 patients who received radiation therapy. Dichotomized total cMET level was a significant prognostic factor for both RFS (HR 3.37; 95% CI: 1.50-7.57, P = 0.002) and OS (HR 4.03; 95% CI: 1.39-11.67, P = 0.006) for patients who received radiation therapy. Similarly, dichotomized p-cMET was a significant prognostic factor for RFS (HR 2.07; 95% CI: 1.12-3.84, P = 0.017) and OS (HR 2.25; 95% CI: 1.05-4.85, P = 0.033) in this group. In contrast, among 93 patients who did not receive radiation therapy, total cMET and p-cMET were not significant prognostic factors for either RFS or OS.

Discussion

The cMET proto-oncogene encodes a RTK which binds with high affinity to its ligand HGF/ SF (1). This receptor-ligand interaction leads to formation of phosphorylated cMET which activates downstream effectors such as PI3K (Phosphatidylinositol 3 kinase)-AKT, PLC (Phospholipase C), RAS-MAPK (mitogen-activated protein kinase), c-Src and STATs (signal transducer and activator of transcription) (4, 5). Together this cascade contributes to carcinogenesis and angiogenesis in a wide variety of human malignancies including cervical, endometrial, gastric, kidney, liver, sarcoma, lung, colorectal and breast (4).

In this analysis of 257 breast cancer samples we demonstrated that increased levels of total cMET and p-cMET using RPPA are seen in nearly 70% and 50% of breast cancers, respectively. We also show that there is no significant difference in the levels of total cMET and p-cMET among different breast cancer subtypes. Survival analysis reveals that total cMET and p-cMET levels are significant prognostic factors for both RFS and OS. Analysis of survival outcomes among various tumor subtypes show that high cMET levels is a poor

prognostic factor for hormone receptor-positive breast cancer and high p-cMET levels is a poor prognostic factor for HER2-positive breast cancers.

A number of studies have shown correlation of cMET expression with progression, aggressive behavior and poor survival outcomes in breast cancers (33-36). However, no study, to the best of our knowledge, has yet investigated the significance of differential expression of cMET and p-cMET in the different breast cancer subtypes (HR-positive, HER2-positive and TNBC. Also, there are no studies reporting p-cMET levels as a prognostic factor in breast cancer. Previous work studying this receptor has been performed using enzyme-linked immunosorbent assay (ELISA), immunoperoxidase (IPO), immunohistochemistry (IHC) and Immunofluorescence (IF) techniques; however, cMET expression has not been evaluated to date using RPPA. Recent data from our laboratory has demonstrated significant correlations between RPPA and IHC in snap frozen primary breast tumors and has established reliability of RPPA in functional proteomic "fingerprinting" (31). Compared to IHC or ELISA, RPPA is more sensitive, reduces variability and avoids observer dependency (37). Applying RPPA analysis for molecular targets promotes practical development for clinical application since it allows for a cost-effective, quick, precise, reliable and reproducible quantification of protein expression and phosphorylation states in multiple samples simultaneously and requires only limited clinical material (38).

Both HGF expression and cMET expression have been shown to correlate with poor prognosis in breast cancer. These clinical outcomes are a consequence of multiple phenotypic properties that are imparted to tumor cells by HGF/cMET activation. HGF/ cMET signaling enhances the transition from pre-invasive DCIS to invasive carcinoma (39). The pathway also promotes cell motility and angiogenesis (11, 40). HGF-dependent – catenin stabilization leads to the establishment of bone metastasis in breast cancers (41). HGF/cMET receptor signals synergize with HER2 and promote breakdown of cell-cell junctions and enhance cell invasiveness (42). This cross talk is possibly responsible for poor prognosis seen in HER2-positive breast cancers with high levels of p-cMET (RFS: P = 0.019and OS: P = 0.014).

Development of therapy resistance is a major cause of treatment failure in breast cancer. In vitro studies have shown that HGF/SF/cMET signaling pathway can confer resistance against induction of apoptosis by various DNA damaging-agents (radiation and cytotoxic agents such as anthracyclines and taxanes) (43). Moreover, the pathway is also involved in promoting cell survival by enhancing DNA repair (44). Studies also suggest overexpression of the HGF/SF and cMET contributes to resistance, both inherent and treatment-acquired, to endocrine therapy and to trastuzumab treatment (45, 46). Radiotherapy, anthracyclines, taxanes, endocrine therapy and trastuzumab form the backbone of breast cancer therapy. The anti-apoptotic prosurvival effect of the HGF/SF/cMET signaling pathway makes MET inhibition a potential therapeutic target for breast cancer that are resistant and refractory to conventional therapies. Since cMET participates in acquisition of resistance, a subset of breast cancers may benefit from combination of MET inhibitors as first line therapy with traditional treatments.

Recent literature has shown that MET expression confers radioresistance in cancer cells (47). De Bacco et al. demonstrated that human breast cancer cell lines (MDA-MB-231 and MDA-MB-435S) subjected to therapeutic doses of ionizing radiation (IR) showed increased MET expression, ligand-independent MET phosphorylation/signal transduction and promoted cell invasion and survival (47). Further, MET silencing by siRNA or treatment with kinase inhibitors counteracted these effects. Expression of cMET in oropharyngeal squamous cell carcinomas treated with radiotherapy has been shown to correlate with decreased rates of complete remission, shorter disease-free survival and overall survival

(47). This data suggests that targeting MET may increase radiosensitivity of tumor cells and is an attractive target for radiosensitization.

Pre clinical data suggests that cMET inhibition in tumor cells impairs cell proliferation, survival, motility, invasion and angiogenesis (48, 49). Several small molecule cMET kinase inhibitors and antibodies against HGF and against cMET are in various stages of development as potential cancer therapies (24, 25, 50). Targeted therapy for breast cancers with pre-selection based on overexpression of cMET and p-cMET with MET inhibition needs further exploration after adequate optimization of predictive markers. Levels of total cMET and p-cMET are uniformly elevated in all breast cancers subtypes and were found to be significant prognostic factors for both RFS and OS. The predictive potential of cMET however cannot be reliably assessed from this retrospective analysis and needs evaluation in clinical trials of targeted therapy against cMET. cMET inhibition is a promising new strategy for treatment of breast cancers and deserves further assessment.

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Statement of Translational Relevance

The proto-oncogene cMET (mesenchymal-epithelial transition factor gene) encodes a tyrosine-kinase membrane receptor, which acts as a ligand for hepatocyte growth factor (HGF). Dysregulation of cMET and its phosphorylated form, phospho-cMET (p-cMET) have been implicated in tumor proliferation, survival, angiogenesis and migration. High expression of cMET has been shown to correlate with adverse survival outcomes in various tumor types and confers chemo-resistance and radio-resistance. The HGF/cMET axis also appears to play a key role in tumor progression in breast cancers. Clarifying the prognostic implication and differential impact of cMET expression on survival in breast cancer subtypes is a necessary first step to application of targeted therapy against cMET in breast cancers. We found that high levels of cMET and p-cMET were seen in all breast cancer subtypes and correlated with poor prognosis. Inhibition of the cMET axis is a promising new therapeutic strategy and needs further investigation.

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Figure 1.

Kaplan-Meier estimates illustrating the Relapse free survival (RFS) of patients by (A) total cMET and (B) p-cMET expression levels and Overall survival (OS) of patients by (C) total cMET and (D) p-cMET expression levels. High: total cMET > 0, Low: total cMET -0. High: p-cMET > 0.35, Low: p-cMET -0.35.

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Figure 2.

Kaplan-Meier estimates illustrating (A) Overall survival (OS) and (B) Relapse free survival (RFS) of patients by total cMET in Hormone receptor positive breast cancer. Kaplan-Meier estimates illustrating (C) Overall survival (OS) and (D) Relapse free survival (RFS) of patients by p-cMET in HER2 positive breast cancer. High: total cMET > 0, Low: total cMET -0. High: p-cMET > 0.35, Low: p-cMET -0.35.

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Table 1	and phospho-cMET levels
	y total cMET :
	Characteristics b
	Clinical (
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					cMEJ					p-cMET		
	Ŏ Ľ	erall 257)	High () =N	cMET -0) 181)	Low (N	cMET 0) =76)		High I (>(S=	-cMET .35) =123)	Low p (0 (N=	-cMET .35) 134)	
	z	%	z	%	z	%	Ρ	z	%	z	%	Р
Age												
50	128	49.8	83	45.9	45	59.2		49	39.8	79	59.0	
>50	129	50.2	98	54.1	31	40.8	0.069	74	60.2	55	41.0	0.003
Stage												
I	8	3.1	4	2.2	4	5.3		2	1.6	9	4.5	
П	141	55.3	43	56.4	40	52.6		67	54.9	74	55.6	
Ш	106	41.6	39	41.3	32	42.1	0.426	53	43.4	53	39.8	0.391
Grade												
1	12	4.7	6	5.1	3	4.0		6	7.5	3	2.3	
2	73	28.9	55	30.9	18	24.0	_	39	32.5	34	25.6	
3	168	66.4	114	64.0	54	72.0	0.473	72	60.0	96	72.2	0.046
Chemotherapy												
Anthracycline based	17	6.7	11	6.2	6	7.9		10	8.3	7	5.3	
Taxane based	21	8.3	16	16.9	5	6.6		10	8.3	11	8.3	
Anthracycline & Taxane	180	<i>4</i> .0 <i>7</i>	121	68.1	59	77.6		62	65.3	101	75.9	
No Chemotherapy	36	14.1	30	16.8	9	7.9	0.228	22	18.2	14	10.5	0.214
Hormone Therapy												
Yes	107	40.5	70	38.7	37	48.7		59	48.0	48	35.8	
No	150	59.5	111	61.3	39	51.3	0.178	64	52.0	86	64.2	0.065
Radiation Therapy												
Yes	164	63.8	108	59.7	56	73.7		79	64.2	85	63.4	
No	93	36.2	73	40.3	20	26.3	0.046	44	35.8	49	36.6	0.998
Subtype												
HER2-positive	53	20.6	40	22.1	13	17.1	0.079	25	20.3	28	20.9	0.064

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					cMET					p-cMET		
	ove N≕	trall 257)	High <) (N=	cMET -0) 181)	Low ((N=	cMET 0) =76)		High I (>(N=	-cMET .35) :123)	Low p- (0) (N=	cMET 35) 134)	
	N	%	Z	₀%₀	z	%	Ρ	Z	%	N	%	Ρ
HR-positive	140	54.5	103	56.9	37	48.7		75	61.0	65	48.5	
Triple Negative	64	24.9	38	31.0	26	34.2		23	18.7	41	30.6	

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cMET: Total cMET, p-cMET: Phospho-cMET, HR-positive: Hormone receptor positive

	Table 2
Total cMET and phospho-cMET of	expressions by tumor subtype

			Total cMI	ET	P	hospho-cN	1ET
Subtype	N	Mean	SD	F-test P Value	Mean	SD	F-test P Value
HER2-positive	53	0.355	0.0799		0.180	0.1027	
Hormone receptor-positive	140	0.391	0.0541	0.128	0.257	0.0651	0.088
Triple receptor-negative	64	0.192	0.0904		0.001	0.0990	

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				R	elapse-Free Su	rvival (RFS)			Overall Surv	rival (OS)	
Tumor Subtypes	Protein	Level	Patients	Events	5-year RFS (%)	95% CI	Ρ	Events	5-year OS (%)	13 %S6	Ρ
	LEIN	High	181	63	61.3	53.2-70.7	200 U	43	72.4	64.7-81.0	0000
IlourerO	CIMET	Low	76	13	78.9	68.6-90.8	cnn.n	L	93.3	87.8-99.2	c00.0
Overall	TTM.	High	123	45	58.9	49.1-70.7	<i></i>	32	72.4	63.7-82.3	2000
	p-civie i	Low	134	31	73.8	65.6-83.1	ccu.u	18	85.8	79.4-92.7	CZU.U
		High	103	31	65.4	54.7-78.2	100	17	82.8	75.0-91.3	200.0
Hormone	CIMET	Low	37	4	85.3	78.2-100	0.014	1	100	NA-NA	000.0
receptor-positive	TTM.	High	75	21	67.6	55.6-82.2	0 510	11	85.2	77.0-94.2	0.510
	p-civile 1	Low	65	14	75.1	63.3-89.2	610.0	L	90.4	82.6-99.0	61C.U
	LEIN	High	40	17	55.5	40.8-75.4	0 6 3 0	13	59.4	41.4-85.2	0.120
	CIMET	Low	13	4	0.00	NA-NA	ncc.n	1	100	NA-NA	0C1.U
avuisod-2721		High	25	15	36.5	19.5-68.3	0100	12	52.3	33.3-82.2	100
	p-civit-1	Low	28	6	74.1	57.4-95.5	610.0	2	91.7	81.3-100	0.014
	TIME	High	38	15	59.5	45.5-77.7	2000	13	56.1	38.0-82.7	0 107
Triple	CIMIDI	Low	26	5	80.4	66.4-97.4	0.000	5	66.2	66.2-97.4	0.10/
receptor-negative		High	23	6	6.09	43.9-84.5	1200	6	58.8	41.1-84.1	0170
	p-civit-1	Low	41	11	72.3	59.6-87.7	107.0	6	74.2	60.5-91.0	0.120
CI: Confidence Interv	val, cMET: 1	Total cME	ET, p-cMET	: Phospho-	cMET						

Table 4

Multivariable Cox Proportional Hazards Model for Relapse Free Survival (RFS) and Overall Survival (OS) relative to total cMET and phospho-cMET expression

			cME	L						p-cM	Π		
		RFS			SO				RFS			SO	
Model	HR	95% CI	Ч	HR	95% CI	Ь	Model	HR	95% CI	4	HR	95% CI	Ч
cMET: High vs. Low	2.06	1.08-3.94	0.028	2.81	1.19-6.64	0.019	p-cMET: High vs. Low	1.79	1.08-2.95	0.020	1.84	0.97-3.48	0.062
Subtype: HER2+ vs. HR+	1.03	0.54-1.98	0.925	1.24	0.53-2.90	0.621	Subtype: HER2+ vs. HR+	1.00	0.52-1.92	0.999	1.11	0.48-2.60	0.803
Subtype: TN vs. HR+	0.65	0.34-1.25	0.201	2.00	0.87-4.58	0.103	Subtype: TN vs. HR+	0.59	0.31-1.14	0.115	1.62	0.71-3.73	0.249
Age: >50 vs. 50	0.79	0.48-1.28	0.339	0.97	0.53-1.77	0.924	Age: >50 vs. 50	0.83	0.51-1.34	0.441	1.06	0.58-1.93	0.848
Chemotherapy: A vs. N	0.06	0.01-0.25	0.0001	0.20	0.03-1.07	0.061	Chemotherapy: A vs. N	0.05	0.01-0.23	0.0001	0.15	0.03-0.78	0.024
Chemotherapy: T vs. N	0.13	0.05-0.33	<0.0001	0.26	0.08-0.82	0.025	Chemotherapy: T vs. N	0.13	0.53-0.34	<0.0001	0.28	0.08-0.89	0.032
Chemotherapy: B vs. N	0.10	0.05-0.18	<0.0001	0.28	0.13-0.63	0.002	Chemotherapy: B vs. N	0.10	0.56-0.19	<0.0001	0.26	0.13-0.54	0.0003
Hormonal Therapy: Y vs. N	0.21	0.10-0.43	0.0001	0.31	0.11-0.84	0.022	Hormonal Therapy: Y vs. N	0.16	0.08-0.33	<0.0001	0.23	0.08-0.63	0.004
Radiation Therapy: Y vs. N	0.91	0.53-1.56	0.740	0.80	0.41-1.57	0.523	Radiation Therapy: Y vs. N	0.84	0.49-1.45	0.528	0.81	0.41-1.59	0.545
Grade: I, II vs. III	0.84	0.48-1.50	0.419	0.72	0.33-1.53	0.390	Grade: I, II vs. III	0.94	0.52-1.67	0.819	0.77	0.36-1.68	0.514
Clinical Stage: I, II vs. III	0.37	0.22-0.62	0.0002	0.67	0.33-1.34	0.243	Clinical Stage: I, II vs. III	0.40	0.24-0.69	0.000	0.77	0.38-1.55	0.458
RFS: Relapse Free Survival, Of	: Overa	ll Survival, cN	AET: total c	MET. p-	-cMET: phos	sho-cME	T, HR: Hazard ratio, HR+: Horr	none rec	eptor positiv	e, TN: Tripl	le Negati	ive, A: Anthr.	acycline b

B: Both Anthracycline & Taxane, T: Taxane based