Cellular/Molecular

# $D_1$ Dopamine Receptor Coupling to PLC $\beta$ Regulates Forward Locomotion in Mice

Ivan O. Medvedev,<sup>1\*</sup> Amy J. Ramsey,<sup>2\*</sup> Shababa T. Masoud,<sup>2</sup> Marie Kristel Bermejo,<sup>2</sup> Nikhil Urs,<sup>1</sup> Tatyana D. Sotnikova,<sup>3</sup> Jean-Martin Beaulieu,<sup>6</sup> Raul R. Gainetdinov,<sup>3,4,5</sup> and Ali Salahpour<sup>2</sup>

<sup>1</sup>Department of Cell Biology, Duke University Medical Center, Durham, North Carolina 27710, <sup>2</sup>Department of Pharmacology and Toxicology, Faculty of Medicine, University of Toronto, Toronto, Ontario M5S 1A8, Canada, <sup>3</sup>Department of Neuroscience and Brain Technologies, Istituto Italiano di Tecnologia, 16163 Genova, Italy, <sup>4</sup>Skolkovo Institute of Science and Technology, Skolkovo, Moscow Region, 143025, Russia, <sup>5</sup>Faculty of Biology and Soil Science, St. Petersburg State University, St. Petersburg 199034, Russia, and <sup>6</sup>Department of Psychiatry and Neuroscience, Faculty of Medicine, Laval University and Institut Universitaire en Santé Mentale de Québec, Québec City, Québec G1J 2G3, Canada

Several studies have reported the coupling of dopamine signaling to phospholipase C  $\beta$  (PLC $\beta$ ) both *in vitro* and *in vivo*. However, the precise physiological relevance of this signaling pathway in mediating dopamine behaviors is still unclear. Here we report that stimulation of dopamine receptor signaling *in vivo* with systemic administration of apomorphine, amphetamine, and cocaine leads to increased production of inositol triphosphate (IP $_3$ ) in the mouse striatum. Using selective antagonists and dopamine D $_1$  and D $_2$  receptor knock-out animals, we show that the production of IP $_3$  is mediated by the D $_1$  receptor, but not the D $_2$  receptor. A selective blocker of PLC $\beta$ , U73122, was used to assess the physiological relevance of D $_1$ -mediated IP $_3$  production. We show that U73122 inhibits the locomotor-stimulating effects of apomorphine, amphetamine, cocaine, and SKF81297. Furthermore, U73122 also suppresses the spontaneous hyperactivity exhibited by dopamine transporter knock-out mice. Importantly, the effects of U73122 are selective to dopamine-mediated hyperactivity, as this compound does not affect hyperactivity induced by the glutamate NMDA receptor antagonist MK801. Finally, we present evidence showing that an imbalance of D $_1$ - and D $_2$ -mediated signaling following U73122 treatment modifies the locomotor output of animals from horizontal locomotor activity to vertical activity, further highlighting the importance of the PLC $\beta$  pathway in the regulation of forward locomotion via dopamine receptors.

### Introduction

Dopamine neurotransmission regulates both motor and nonmotor behaviors (Carlsson, 2001; Greengard, 2001; Beaulieu and Gainetdinov, 2011). The role of dopamine in facilitating voluntary movement is evidenced by the locomotor-stimulant effects of dopaminergic drugs like amphetamine, and conversely by the inhibitory effects of dopamine antagonists like haloperidol (Carlsson, 2001).

Dopamine acts on the following two subclasses of receptors: the  $D_1$  class, composed of  $D_1$  and  $D_5$  receptors; and the  $D_2$  class, composed of the  $D_2$ ,  $D_3$ , and  $D_4$  receptors (Missale et al., 1998; Beaulieu and Gainetdinov, 2011). Classically, the  $D_1$  class has been shown to be positively coupled to cAMP production through a  $G_{\text{s/olf}}$  heterotrimeric G-protein, while activation of the

Received June 4, 2013; revised Sept. 4, 2013; accepted Oct. 3, 2013.

Author contributions: I.O.M. and A.S. designed research; I.O.M., A.J.R., S.T.M., M.K.B., N.U., T.D.S., J.-M.B., R.R.G., and A.S. performed research; I.O.M., A.J.R., J.-M.B., R.R.G., and A.S. analyzed data; I.O.M., A.J.R., and A.S. wrote the naner

This work was partially supported by National Institutes of Health K99 Grant 1K99ES016816-01, and Canadian Institutes of Health Research Operating Grants 210296 (to A.S.) and 258294 (to A.J.R.). We thank Marc G. Caron for support and critical reading of the manuscript.

\*I.O.M. and A.J.R. contributed equally to this work.

The authors declare no competing financial interests.

Correspondence should be addressed to Ali Salahpour, Department of Pharmacology and Toxicology, University of Toronto, 1 Kings College Circle, Toronto, ON MSS 1A8, Canada. E-mail: ali.salahpour@utoronto.ca.

DOI:10.1523/JNEUROSCI.2382-13.2013

Copyright © 2013 the authors 0270-6474/13/3318125-09\$15.00/0

 $D_2$  class leads to reduction in cAMP levels through  $G_{i/o}$  proteins (Missale et al., 1998; Neve et al., 2004; Beaulieu and Gainetdinov, 2011)

Traditionally, the cAMP pathway has been thought to mediate the locomotor behaviors ascribed to dopamine. However, several published studies raise the possibility that cAMP is not the only pathway for dopamine-mediated locomotion. Indeed, some dopamine-mediated locomotor behaviors are preserved when the G<sub>s</sub>/adenylyl cyclase/protein kinase A (PKA) pathway is perturbed by knocking out key signaling molecules. For example, Golf knock-out mice are hyperactive (Zhuang et al., 2000), and basal activity is essentially normal in mice lacking the catalytic subunit of PKA or dopamine- and cAMP-regulated phosphoprotein of 32 kDa (DARPP-32; Brandon et al., 1998; Nally et al., 2003). Alterations in the cAMP pathway are most evident in psychostimulant response, where mice deficient in DARPP-32 and G<sub>olf</sub> are less responsive to cocaine and amphetamine (Fienberg et al., 1998; Zachariou et al., 2006; Corvol et al., 2007). Furthermore, selective deletion of DARPP-32 in D<sub>1</sub> or D<sub>2</sub> neurons bidirectionally affects basal activity (Bateup et al., 2010). These observations highlight the complexity of D<sub>1</sub> signaling and raise the possibility that alternate signaling pathways converge or function in parallel with cAMP signaling to elicit dopamine-mediated locomotion (Neve, 2010).

Several studies have described the coupling of a  $D_1$ -like receptor to phospholipase C  $\beta$  (PLC $\beta$ ). (Undie and Friedman, 1990;

Friedman et al., 1997; Undieh, 2010). In addition, it was shown that concomitant activation of  $D_1$  and  $D_2$  receptors within a heterodimeric complex is linked to an increase in intracellular calcium levels (Lee et al., 2004; Rashid et al., 2007; Hasbi et al., 2010; Verma et al., 2010; Perreault et al., 2011), likely via multiple mechanisms (Chun et al., 2013).

Despite these studies that have convincingly demonstrated that PLC $\beta$  is a downstream effector of dopamine signaling, the precise physiological role of this dopamine-signaling pathway remains unclear. We sought to investigate in vivo the role of dopamine receptor signaling through PLC $\beta$  and inositol triphosphate (IP<sub>3</sub>) by evaluating the contribution of this pathway to the modulation of locomotor activity. Our study shows that PLC $\beta$  is a critical modulator of dopaminemediated forward locomotor activity, and that, in vivo, direct and indirect dopamine agonists lead to stimulation of IP<sub>3</sub> production via the D<sub>1</sub> dopamine receptor. Furthermore, the inhibition of PLCB signaling shifts the L-DOPAmediated locomotor activity of animals from horizontal activity to vertical activity, stressing the importance of the PLC $\beta$  pathway for forward locomotion.

### **Materials and Methods**

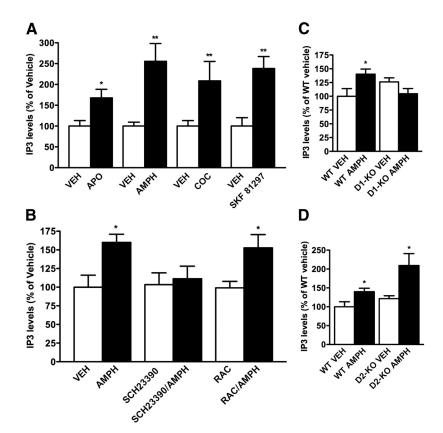
Animals. All behavioral experiments were performed during the light cycle and used 3- to 5-month-old C57BL/6J age- and sex-matched

animals of either sex. Dopamine transporter knock-out (DAT-KO) mice and wild-type littermates (WT) (Giros et al., 1996) maintained on a C57BL/6J genetic background of the same age were also used. Animals were housed four to five per cage in a humidity- and temperature-controlled room with 12 h light/dark cycle (lights on at 8:00 A.M.). Mice were provided food and water *ad libitum*. All mice were experimentally naïve, and a separate group of mice was used for each testing paradigm. Experiments were conducted in accordance with the National Institutes of Health or Canadian Council for Animal Care guidelines for the care and use of animals and an approved animal protocol from either the Duke University Animal Care and Use Committee or the Faculties of Medicine and Pharmacy Animal Care Committee at the University of Toronto

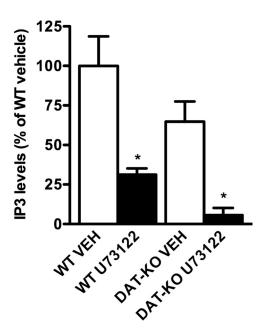
Reagents. Apomorphine, α-methyl-para-tyrosine (αMPT), raclopride, SKF81297, SCH 23390, MK801, L-DOPA, carbidopa, U73122, and U73343 were purchased from Sigma-Aldrich. Anti-phospho-GluR1-Ser-845 antibody and anti-total-GluR1 antibody were purchased from Millipore. Anti-GAPDH antibody was purchased from Sigma. Species-specific Alexa Fluor 680 antibody was purchased from Invitrogen. Species-specific IRDye 800CW antibody was purchased from LI-COR Biosciences.

*Drugs.* Compounds or saline (0.9% NaCl) were administered intraperitoneally or subcutaneously in a volume of 10 ml/kg. For U73122 and U73343, 5 mg of compound was first dissolved in 20  $\mu$ l of Tween 20 and then resuspended in saline solution for intraperitoneal injection. Apomorphine was dissolved in distilled water containing 0.1% ascorbate and injected subcutaneously.

Locomotor analysis. Locomotor activity was measured in an automated infrared beam-break apparatus (AccuScan Instruments) during the light phase of the light/dark cycle. Animals were placed in an activity monitor



**Figure 1.** Direct and indirect dopamine agonists lead to accumulation of IP $_3$  in the striatum via D $_1$  receptors. **A**, C57BL/6J mice were injected with 1 mg/kg apomorphine (APO), 3 mg/kg amphetamine (AMPH), 20 mg/kg cocaine (COC), or 10 mg/kg SKF81297, and IP $_3$  levels were quantified using radioimmunoassay. **B**, C57BL/6J mice were pretreated with vehicle (VEH), 0.1 mg/kg SCH23390 (D $_1$  antagonist), or 2 mg/kg raclopride (RAC; D $_2$  antagonist) before amphetamine administration (3 mg/kg). **C**, **D**, IP $_3$  levels were assessed after amphetamine (3 mg/kg) or vehicle injection in D $_1$ -K0 mice (**C**) or D $_2$ -K0 mice (**D**) and their respective WT littermates. All data are presented as the means  $\pm$  SEM. N = 5. \*p < 0.05; \*\*p < 0.01 as determined by Student's t test.



**Figure 2.** Selective PLC $\beta$  inhibition reduces IP $_3$  levels in the striatum of WT and DAT-KO mice. WT and DAT-KO mice were treated with vehicle (VEH) or a selective PLC $\beta$  inhibitor, U73122 (10 mg/kg). IP $_3$  levels were assessed using radioimmunoassay and were reported as a percentage of WT vehicle-treated mice. Data are presented as the mean  $\pm$  SEM. N=5. \*p<0.05, Student's t test.

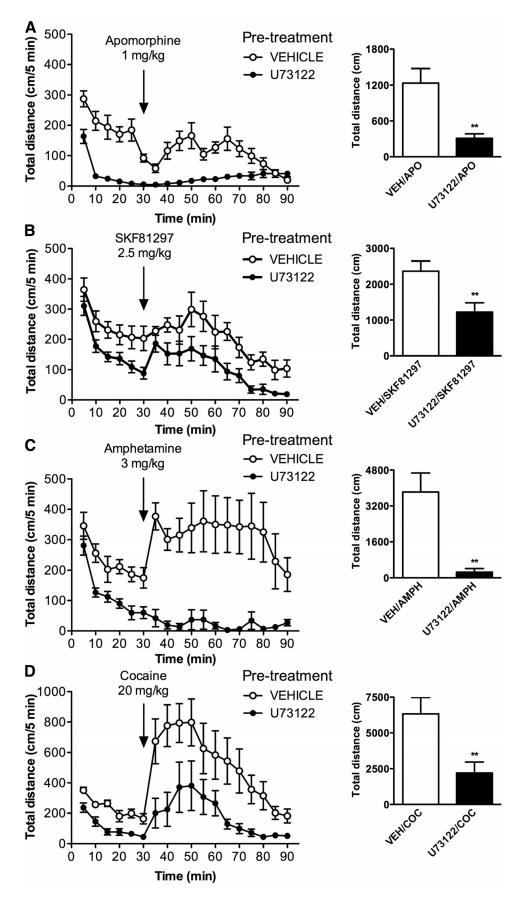


Figure 3. Inhibition of PLCβ reduces the locomotor response of mice to direct and indirect dopamine agonists. Mice were pretreated with vehicle (VEH) or U73122 (10 mg/kg) and placed in locomotor activity chambers. *A–D*, After 30 min, mice were injected with 1 mg/kg apomorphine (APO; *A*), 2.5 mg/kg SKF81297 (*B*), 3 mg/kg amphetamine (AMPH; *C*), or 20 mg/kg coaine (COC; *D*), and locomotor activity was monitored for another 60 min. Distance traveled (in centimeters) was measured over 5 min intervals, and total distance is shown as a sum of 60 min. Data are presented as the mean ± SEM. *N* = 8.\*\**p* < 0.01, Student's *t* test.

chamber ( $20 \times 20$  cm), and individual activity data were collected at 5 min intervals. Forward locomotion was measured as the total distance traveled unless otherwise indicated. Vertical activity was measured as the time spent in the vertical rearing position.

 $IP_3$  level measurement.  $IP_3$  measurement was performed using the GE Healthcare  $IP_3$  [  $^3\mathrm{H}$ ] Biotrak Assay System (TRK1000) according to manufacturer notes. Briefly, mice were killed by cervical dislocation, and striatal tissue was dissected rapidly within 1 min. The tissue was immediately submerged in 1 ml of 10% perchloric acid and incubated for 10 min, after which the tissue was sonicated three times for 10 s. Following sonication, the sample was centrifuged for 15 min at 10,000  $\times$  g at 4°C. The supernatant was then transferred to a 15 ml conical tube and titrated to pH 7.5 using 1.5 m KOH containing 60 mm HEPES. The pH of each sample was verified to be 7.5  $\pm$  0.1. The samples were then centrifuged at 2000  $\times$  g to precipitate KClO $_4$ . The supernatant protein concentration was measured using the Bradford assay. A protein sample in the supernatant of 100  $\mu$ l was then assayed using the TRK1000 kit following the procedure outlined by the manufacturer.

Western blot. Mice were pretreated with either vehicle or U73122 (10 mg/kg, i.p.) 30 min before administration of either vehicle or SKF81297 (3 mg/kg, i.p.). Fifteen minutes after the second injection, mice were killed by cervical dislocation, and striatum was rapidly dissected and snap frozen in liquid nitrogen. As described by Ghisi et al. (2009), striatum was homogenized in 1% SDS and 2 µM okadaic acid with a hand-held homogenizer, and the protein concentration determined using a BCA assay (Pierce). Fifty micrograms of striatal protein was resolved on a 10% SDS polyacrylamide gel and transferred to a PVDF membrane. The immunostaining of blots was performed overnight at 4°C with the following primary antibodies: anti-phospho-GluR1-Ser-845 (1:500) and antitotal-GluR1 (1:1500). Appropriate secondary antibodies (1:5000, Alexa Fluor 680 or IRDye 800CW) were used, and blots were developed using the LI-COR Biosciences Odyssey Imaging System. Densitometric analysis was performed with ImageJ software. Total GluR1 protein signal was used as the loading control for phospho-GluR1 protein levels.

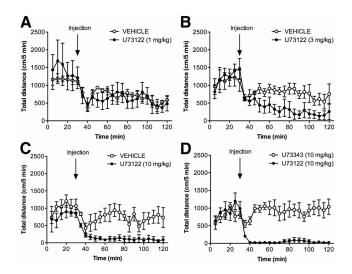
### **Results**

## Direct and indirect dopamine agonists increase IP<sub>3</sub> levels in vivo

To test whether activation of dopamine receptors can lead to an activation of PLC $\beta$  *in vivo*, we measured levels of IP<sub>3</sub> accumulation in the striatal tissue following systemic administration of direct and indirect dopamine receptor agonists. As shown in Figure 1A, the injection of mice with 1 mg/kg apomorphine, 3 mg/kg amphetamine, 20 mg/kg cocaine, or 10 mg/kg SKF81297 leads respectively to a 50, 150, 100, and 120% increase in IP<sub>3</sub> levels compared with saline treatment.

Based on the observation that SKF81297, a  $D_1$ -selective agonist, increases striatal IP $_3$  levels, we performed a series of studies to determine whether the increase of IP $_3$  is due to the activation of a dopamine  $D_1$  or  $D_2$  receptor. Mice were injected with either SCH23390 ( $D_1$  antagonist) or raclopride ( $D_2$  antagonist) before injections with amphetamine (3 mg/kg). As shown in Figure 1B, injection of SCH23390 (0.1 mg/kg) completely abolishes production of IP $_3$  by amphetamine, while pretreatment with raclopride (2 mg/kg) has no effect on amphetamine-induced IP $_3$  accumulation. This indicates that stimulation of IP $_3$  production in the mouse striatum *in vivo* is mediated by  $D_1$  and not  $D_2$  class receptors.

To further validate the results in Figure 1*B*, we measured the stimulation of  $IP_3$  accumulation after amphetamine administration (3 mg/kg) in  $D_1$  and  $D_2$  receptor knock-out animals ( $D_1$ -KO and  $D_2$ -KO). As shown in Figure 1, *C* and *D*, the accumulation of  $IP_3$  levels after amphetamine injection is completely abolished in  $D_1$ -KO animals, while  $IP_3$  accumulation is still observed in  $D_2$ -KO mice. These results, using both pharmacological and genetic approaches, clearly demonstrate that the accumulation of  $IP_3$  after



**Figure 4.** Selective inhibition of PLC $\beta$  dose-dependently reduces hyperactivity in DAT-KO mice. *A–C*, DAT-KO mice were habituated to the activity monitor chambers for 30 min before injection with vehicle or increasing doses of U73122 (1 mg/kg, A; 3 mg/kg, B; 10 mg/kg, C; N = 4). The hyperactivity of DAT-KO animals was dose-dependently reduced by U73122 treatment. D, Mice were treated with either U73122 (10 mg/kg) or U73343 (10 mg/kg), the inactive analog of U73122 (N = 8). Distance traveled (in centimeters) was measured in 5 min intervals for 30 min before injection and for 90 min after injection. Data are presented as the mean  $\pm$  SEM.

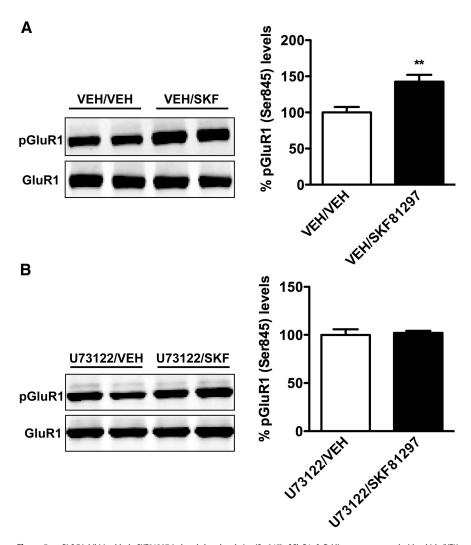
amphetamine injection in vivo is mediated by the  $\mathrm{D}_1$  and not the  $\mathrm{D}_2$  dopamine receptor.

# The PLC $\beta$ pathway is critical for dopamine-mediated locomotor activity

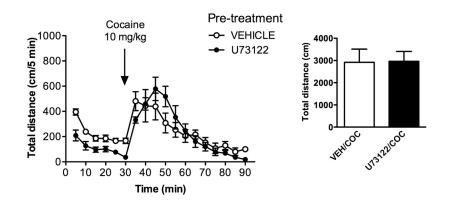
Having shown that stimulation of D<sub>1</sub> receptors leads to an increase in striatal IP3 levels, we next investigated the role of this pathway in dopamine-mediated locomotor activity. For this, the selective PLC $\beta$  inhibitor U73122 was used. We first determined that U73122 effectively reduces IP<sub>3</sub> levels in the brain. Peripheral injection of U73122 (10 mg/kg) reduces the measureable levels of IP<sub>3</sub> in the striatum of both WT and DAT-KO mice, which have a fivefold increase in extracellular dopamine (Giros et al., 1996; Fig. 2). Interestingly, DAT-KO mice show a trend decrease in their basal levels of IP<sub>3</sub>, and U73122 has a greater effect in reducing IP<sub>3</sub> levels in DAT-KO mice (Fig. 2). These results may be explained by the 50% reduction in D<sub>1</sub> receptor levels in the DAT-KO mice (Giros et al., 1996; Ghisi et al., 2009), a compensatory adaptation to sustained hyperdopaminergia. Thus, although DAT-KO mice have high extracellular dopamine levels, signaling through the PLC $\beta$  pathway is likely desensitized through compensatory adaptations.

We next tested whether the PLCβ/IP<sub>3</sub> pathway plays a role in dopamine-mediated locomotor activity. In the first set of experiments, C57BL/6J mice were pretreated with U73122 before administration of direct or indirect dopamine receptor agonists. Pretreatment with U73122 (10 mg/kg) reduces basal activity, and also reduces the locomotor responses of mice to the direct dopamine agonists apomorphine (1 mg/kg) and SKF81297 (2.5 mg/kg), as well as to indirect dopamine agonists amphetamine (3 mg/kg) and cocaine (20 mg/kg; Fig. 3). These results suggest a prominent role of the PLCβ/IP<sub>3</sub> pathway in mediating dopamine-stimulated locomotor activity.

To corroborate these pharmacological results, we used a genetic model of enhanced dopamine transmission. For this, U73122 was used to reduce IP<sub>3</sub> levels in DAT-KO animals. DAT-KO mice have increased extracellular dopamine levels, and display hyperactivity and impaired habituation in a novel envi-



**Figure 5.** PLC $\beta$  inhibition blocks SKF81297-induced phosphorylation (Ser845) of GluR1. **A, B,** Mice were pretreated with vehicle (VEH; **A**) or U73122 (10 mg/kg; **B**) and then injected with vehicle or SKF81297 (3 mg/kg). Phosphorylated GluR1 (Ser845) and total GluR1 protein levels in the striatum were assessed by Western blot 15 min after SKF injection. Phosphorylated GluR1 levels were corrected to total GluR1 levels and relative to vehicle in each pretreatment group. Data are presented as the mean  $\pm$  SEM. N = 4-5. \*\*p < 0.01, Student's t test.



**Figure 6.** Inhibition of PLC $\beta$  does not affect hyperactivity induced by a low dose of cocaine (COC; 10 mg/kg). Mice were pretreated with vehicle (VEH) or U73122 (10 mg/kg). After 30 min, mice were injected with cocaine (10 mg/kg), and locomotor activity was monitored for another 60 min. Distance traveled (in centimeters) was measured over 5 min intervals, and the total distance is shown as a sum of 60 min. Data are the mean  $\pm$  SEM. N=8.

ronment (Giros et al., 1996; Jones et al., 1998). The administration of U73122 to DAT-KO mice dose dependently reduces their hyperlocomotor activity (Fig. 4). Importantly, the injection of DAT-KO animals with U73343 (10 mg/kg), the inactive analog of

U73122, has no major effect on locomotor hyperactivity, indicating that the effects of U73122 (10 mg/kg) are mediated through its inhibition of PLC $\beta$  (Fig. 4D).

Because D<sub>1</sub> receptors are known to signal through G<sub>s/olf</sub> coupling to elevate cAMP, we next performed studies to examine potential cross talk between cAMP and IP3 pathways. One of the established indicators of D<sub>1</sub>-G<sub>s/olf</sub> signaling is the dopamine-mediated phosphorylation of the AMPA GluR1 subunit. Residue Ser845 is selectively phosphorylated by PKA in a dopamine-dependent manner (Roche et al., 1996; Corvol et al., 2007). We examined whether PLCB inhibition would alter phosphorylation of GluR1 by comparing the levels of phospho-GluR1 after SKF81297 injection in the presence or absence of U73122. Interestingly, we discovered that U73122 pretreatment prevented the SKFinduced phosphorylation of GluR1 (Fig. 5), suggesting the existence of some level of cross talk between the two pathways.

While U73122 administration fully inhibited locomotor responses to amphetamine, apomorphine, and SKF81297, it did not fully inhibit the effect of cocaine on locomotion at a 20 mg/kg dose (Fig. 3D). To examine this discrepancy further, we evaluated the effect of U73122 on the locomotor activity elicited by a lower (10 mg/kg) dose of cocaine. To our surprise, while the U73122 pretreatment had a clear effect on basal activity, there was no effect on the cocaine-stimulated locomotor activity at 10 mg/kg (Fig. 6). Furthermore, the level of activity induced by cocaine at this dose was within the same range as the response observed following the administration of 20 mg/kg cocaine along with a U73122 pretreatment. This suggests that at least two complementary mechanisms may underlie locomotor responsiveness to cocaine. It is noteworthy that previous findings have shown that DARPP-32 knock-out mice are insensitive to 10 mg/kg doses of cocaine, but respond normally to higher doses (20 mg/ kg; Fienberg et al., 1998). Together, these observations hint that perhaps at low doses, it is the cAMP/DARPP32 pathway that mediates the locomotor-stimulating effects of cocaine (Corvol et al., 2007), while activation of the PLC pathway would contribute to enhanced responses at higher cocaine doses.

The question of whether PLC $\beta$  inhibition selectively affects dopamine-mediated activity was addressed using MK-801,

an NMDA receptor antagonist that increases locomotor activity independent of dopamine (Gainetdinov et al., 2001; Chartoff et al., 2005). As shown in Figure 7A, the injection of WT mice with MK-801 (0.3 mg/kg) results in a robust increase in locomotor

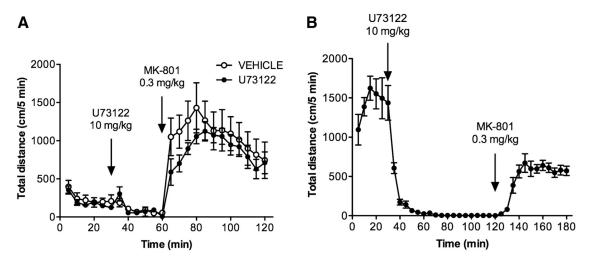


Figure 7. The PLC $\beta$  inhibitor U73122 does not suppress locomotor hyperactivity induced by MK801, an NMDA receptor antagonist. *A, B,* C57BL/6J (*A*) and DAT-K0 (*B*) mice were habituated to the locomotor activity chamber for 30 min and then treated with vehicle (only C57BL/6J mice) or U73122 (10 mg/kg, i.p.; both C57BL/6J and DAT-K0 mice). After 30 min (for C57BL/6J mice) or 90 min (for DAT-K0 mice), animals were injected with MK-801 (0.3 mg/kg, i.p.), and locomotor activity was assessed for 60 min. Distance traveled (in centimeters) was measured over 5 min intervals. Data are presented as the mean  $\pm$  SEM. N = 8.

activity that is unaffected by U73122 pretreatment (10 mg/kg). MK-801 treatment also stimulates locomotor activity in DAT-KO mice pretreated with U73122 (Fig. 7*B*). These studies demonstrate that U73122 selectively inhibits dopaminemediated activity, and that the locomotor effects of the inhibitor are not due to general sedation, neuromuscular block, or muscular weakness.

Last, we evaluated the effect of the PLC $\beta$ /IP<sub>3</sub> pathway on the behavior elicited by dopamine itself. The previous experiments used direct or indirect agonists of the dopamine system or a genetic model of hyperdopaminergia. However, a unique feature of DAT-KO mice allows the assessment of the effect of dopamine depletion and dopamine restoration on motor activity. In DAT-KO animals, all available dopamine results from de novo synthesis by tyrosine hydroxylase (TH). Inhibition of TH by  $\alpha$ MPT leads to complete depletion of the brain dopamine content in these animals, resulting in complete akinesia and immobility (Sotnikova et al., 2005; Costa et al., 2006; Dzirasa et al., 2006; Managò et al., 2012). These animals are termed dopaminedeficient DAT-KO mice (DDD mice). Importantly, dopamine can be restored in DDD mice by injecting them with L-DOPA, which bypasses the inhibition of TH and restores locomotor activity. This system thus allows the assessment of dopamine signaling directly, without the use of direct or indirect ligands. We therefore investigated the functional role of the PLCβ/IP<sub>3</sub> pathway on dopamine motor activity using DDD mice.

DAT-KO mice were first injected with  $\alpha$ MPT (250 mg/kg), and their locomotor activity was monitored for 30 min. This produced complete immobility, as expected. Mice were subsequently injected with U73122 (10 mg/kg) or vehicle. Thirty minutes after the U73122 injection, animals received an injection of L-DOPA/carbidopa (50/50 mg/kg), which normally restores locomotor activity (Sotnikova et al., 2005; Costa et al., 2006; Dzirasa et al., 2006; Managò et al., 2012).

The administration of U73122 prevents the restoration of locomotor activity by L-DOPA as measured by the total distance traveled (Fig. 8A). Interestingly, however, there is a striking switch in the locomotor behavior of the mice injected with U73122. As shown in Figure 8B, the animals that were injected with U73122 display a vertical activity phenotype after L-DOPA

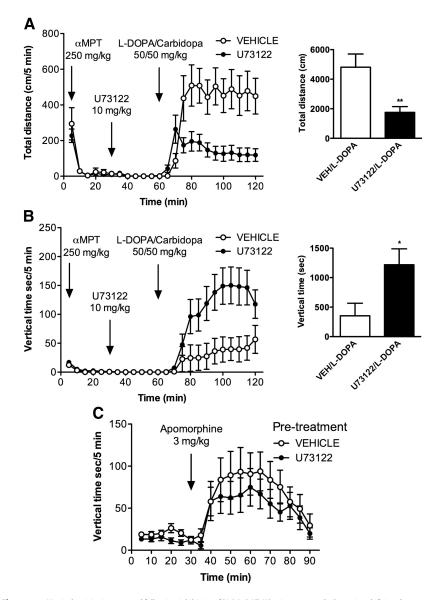
injection instead of horizontal activity. This result suggests that the  $PLC\beta/IP_3$  pathway is primarily important for regulating forward locomotor activity in mice, and that the inhibition of this pathway can switch the behavioral locomotor output of the dopamine system from horizontal to vertical (climbing) activity.

If so, the induction of vertical (climbing) activity in mice should be relatively insensitive to U73122 inhibition. It is well described that, at doses of 2–4 mg/kg, apomorphine induces vertical activity (climbing) in mice (Chow and Beck, 1984). We therefore determined whether U73122 would affect apomorphine-induced climbing. As shown in Figure 8C, injection of U73122 (10 mg/kg) had no significant effect on apomorphine (3 mg/kg)-induced vertical activity, highlighting the importance of the PLC $\beta$ /IP $_3$  pathway in the selective modulation of horizontal locomotor activity.

#### Discussion

Here we report *in vivo* evidence that dopamine stimulation of the  $PLC\beta/IP_3$  pathway is an important contributor to locomotor activity in mice. Over the years, there have been numerous studies that have reported the ability of the dopamine system to modulate the  $PLC\beta/IP_3$  pathway; however, to date, a clear physiological role for this pathway has not been established (Undie and Friedman, 1990; Hasbi et al., 2010). We show that blockade of this pathway leads to the inhibition of both basal and psychostimulant-mediated locomotor activity. Importantly, the  $PLC\beta/IP_3$  pathway is selective for dopamine-mediated behavior, as the stimulant effects of MK-801 were not blocked by U73122 treatment.

Additionally, using a high dose of apomorphine, we demonstrate that the  $PLC\beta/IP_3$  pathway selectively modulates "forward" locomotor activity and does not inhibit vertical activity (climbing). Furthermore, in DDD mice treated with U73122, restoring locomotor activity with L-DOPA led to vertical rather than the horizontal activity. These observations clearly indicate that the  $PLC\beta/IP_3$  pathway is more important for forward locomotion than vertical activity. Intriguingly, vertical activity induced by dopamine agonists has been linked by some to L-DOPA-induced dyskinesas (Johnston et al., 2005). Although the signaling mechanisms underlying these diverse stereotypic behaviors are poorly understood (Undie et al., 2000), our study



**Figure 8.** Vertical activity is preserved following inhibition of PLC $\beta$ . DAT-KO mice were made dopamine deficient by treatment with  $\alpha$ MPT (250 mg/kg, i.p.) and were placed in locomotor activity chambers. After 30 min, mice were treated with vehicle (VEH; N=13) or U73122 (10 mg/kg, i.p.; N=16). After another 30 min, all mice were injected with a combination of ι-DOPA/carbidopa (50/50 mg/kg, i.p.). **A**, Locomotor activity was measured as the distance traveled (in centimeters) over 5 min intervals (left). Total distance traveled in the 60 min after ι-DOPA/carbidopa injection is shown in the right panel. **B**, During the same treatment regimen, time (in seconds) spent in vertical activity was quantified in 5 min intervals (left). The total time spent in vertical activity over the 60 min after ι-DOPA/carbidopa injection is shown in the right panel. **C**, C57BL/6J mice were pretreated with vehicle or U73122 (10 mg/kg, i.p.) and placed in locomotor activity chambers. After 30 min, mice were injected with apomorphine (3 mg/kg, s.c.), and locomotor activity was recorded for another 60 min. The time (in seconds) spent in vertical activity is shown in 5 min intervals. N=8. All data are presented as the mean  $\pm$  SEM. \*p<0.05, \*\*p<0.01 as determined by Student's t test.

may provide evidence for the role of  $D_2$  receptors in vertical activity. Alternately, our studies could be interpreted to indicate that  $D_1$  signaling through other pathways besides  $PLC\beta/IP_3$  is responsible for dopamine-mediated vertical activity.

Evidence that G-protein-coupled dopamine receptors signal through multiple pathways has been demonstrated in several systems (Kotecha et al., 2002; Beaulieu et al., 2005; Masri et al., 2008; Rajagopal et al., 2010; Chun et al., 2013). Our studies lend support to the notion that  $D_1$  receptors can signal through both PKA and PLC $\beta$ , and that these two pathways can exhibit cross talk in modulating Ser845, a PKA-dependent AMPA receptor phosphorylation site. In line with this, it has previously been shown that the activation of PKC, the downstream effector

of PLC $\beta$ , can modulate PKA activity through a CDK5/DARPP32 signaling pathway (Bibb et al., 1999; Sahin et al., 2008). This raises the possibility that activation of PLC $\beta$  signaling by D<sub>1</sub> receptor may also contribute to the maintenance of PKA activity following its activation by this same receptor.

It has long been known that a subpopulation of D<sub>1</sub>-like receptors are able to stimulate IP<sub>3</sub> production through a PLCβ mechanism (Undieh, 2010). These studies have been performed mainly on brain slices ex vivo. Sahu et al. (2009) showed that dopamine-induced IP<sub>3</sub> accumulation was attenuated in striatal slices from D5-KO animals. In another study, Friedman et al. (1997) were still able to detect IP3 production following dopamine stimulation in cortical slices from D<sub>1</sub>-KO mice. However, it is important to note that Friedman et al. (1997) conducted IP3 measurements on cortical slices, while our studies were performed in the striatum. Therefore, it is possible that different receptor subtypes may be mediating dopamine-induced IP3 production in different brain structures.

In addition to reports that D<sub>1</sub>-like receptors mediate PLCβ/IP<sub>3</sub> production, a series of studies have also highlighted the ability of the D<sub>1</sub>/D<sub>2</sub> receptor heteromer to mobilize calcium through a  $G_{\alpha}/PLC\beta$ mediated pathway (Hasbi et al., 2009, 2010; Perreault et al., 2011). However, a recent study reports that dopamine receptor-mediated calcium signaling can occur through multiple pathways, including those that are independent of D<sub>1</sub>/D<sub>2</sub> heteromers or G<sub>q</sub> (Chun et al., 2013). It should also be noted that the population of medium spiny neurons coexpressing D<sub>1</sub> and D<sub>2</sub> receptors are mainly located in the nucleus accumbens (Perreault et al., 2010), while locomotor activity is mainly regulated by the dorsal striatum (Hnasko et al., 2006). Consequently, the modulation of the PLCβ/IP<sub>3</sub> pathway by dopamine receptors may be different in ventral and dorsal striatum. Thus, while activa-

tion of the  $D_1/PLC\beta/IP_3$  pathway in the dorsal striatum may be required to modulate locomotor activity, stimulation of the  $PLC\beta/IP_3$  pathway in the ventral striatum may be required to mediate limbic responses.

In conclusion, we report the first demonstration of the involvement of a  $PLC\beta/IP_3$  pathway in selectively regulating dopamine-mediated locomotor activity. Our results show that this pathway is crucial for horizontal activity and does not influence stereotypical vertical activity displayed by mice after apomorphine injection. Interestingly, our *in vivo* studies indicate that direct or indirect dopamine agonists stimulate  $IP_3$  production exclusively though  $D_1$  dopamine receptor. This signaling cascade can potentially be targeted for the develop-

ment of novel therapies for movement disorders including Parkinson's disease.

#### References

- Bateup HS, Santini E, Shen W, Birnbaum S, Valjent E, Surmeier DJ, Fisone G, Nestler EJ, Greengard P (2010) Distinct subclasses of medium spiny neurons differentially regulate striatal motor behaviors. Proc Natl Acad Sci U S A 107:14845–14850. CrossRef Medline
- Beaulieu JM, Gainetdinov RR (2011) The physiology, signaling, and pharmacology of dopamine receptors. Pharmacol Rev 63:182–217. CrossRef Medline
- Beaulieu JM, Sotnikova TD, Marion S, Lefkowitz RJ, Gainetdinov RR, Caron MG (2005) An Akt/beta-arrestin 2/PP2A signaling complex mediates dopaminergic neurotransmission and behavior. Cell 122:261–273. CrossRef Medline
- Bibb JA, Snyder GL, Nishi A, Yan Z, Meijer L, Fienberg AA, Tsai LH, Kwon YT, Girault JA, Czernik AJ, Huganir RL, Hemmings HC Jr, Nairn AC, Greengard P (1999) Phosphorylation of DARPP-32 by Cdk5 modulates dopamine signalling in neurons. Nature 402:669–671. CrossRef Medline
- Brandon EP, Logue SF, Adams MR, Qi M, Sullivan SP, Matsumoto AM, Dorsa DM, Wehner JM, McKnight GS, Idzerda RL (1998) Defective motor behavior and neural gene expression in RIIβ-protein kinase A mutant mice. J Neurosci 18:3639–3649. Medline
- Carlsson A (2001) A paradigm shift in brain research. Science 294:1021–1024. CrossRef Medline
- Chartoff EH, Heusner CL, Palmiter RD (2005) Dopamine is not required for the hyperlocomotor response to NMDA receptor antagonists. Neuropsychopharmacology 30:1324–1333. CrossRef Medline
- Chow HL, Beck HM (1984) The effect of apomorphine on the open-field behavior of rats: alone and in pairs. Pharmacol Biochem Behav 21:85–88. CrossRef Medline
- Chun LS, Free RB, Doyle TB, Huang XP, Rankin ML, Sibley DR (2013) D1–D2 dopamine receptor synergy promotes calcium signaling via multiple mechanisms. Mol Pharmacol 84:190–200. CrossRef Medline
- Corvol JC, Valjent E, Pascoli V, Robin A, Stipanovich A, Luedtke RR, Belluscio L, Girault JA, Hervé D (2007) Quantitative changes in Galphaolf protein levels, but not D1 receptor, alter specifically acute responses to psychostimulants. Neuropsychopharmacology 32:1109–1121. CrossRef Medline
- Costa RM, Lin SC, Sotnikova TD, Cyr M, Gainetdinov RR, Caron MG, Nicolelis MA (2006) Rapid alterations in corticostriatal ensemble coordination during acute dopamine-dependent motor dysfunction. Neuron 52:359–369. CrossRef Medline
- Dzirasa K, Ribeiro S, Costa R, Santos LM, Lin SC, Grosmark A, Sotnikova TD, Gainetdinov RR, Caron MG, Nicolelis MA (2006) Dopaminergic control of sleep-wake states. J Neurosci 26:10577–10589. CrossRef Medline
- Fienberg AA, Hiroi N, Mermelstein PG, Song W, Snyder GL, Nishi A, Cheramy A, O'Callaghan JP, Miller DB, Cole DG, Corbett R, Haile CN, Cooper DC, Onn SP, Grace AA, Ouimet CC, White FJ, Hyman SE, Surmeier DJ, Girault J, et al. (1998) DARPP-32: regulator of the efficacy of dopaminergic neurotransmission. Science 281:838–842. CrossRef Medline
- Friedman E, Jin LQ, Cai GP, Hollon TR, Drago J, Sibley DR, Wang HY (1997) D1-like dopaminergic activation of phosphoinositide hydrolysis is independent of D1A dopamine receptors: evidence from D1A knockout mice. Mol Pharmacol 51:6–11. Medline
- Gainetdinov RR, Mohn AR, Bohn LM, Caron MG (2001) Glutamatergic modulation of hyperactivity in mice lacking the dopamine transporter. Proc Natl Acad Sci U S A 98:11047–11054. CrossRef Medline
- Ghisi V, Ramsey AJ, Masri B, Gainetdinov RR, Caron MG, Salahpour A (2009) Reduced D2-mediated signaling activity and trans-synaptic upregulation of D1 and D2 dopamine receptors in mice overexpressing the dopamine transporter. Cell Signal 21:87–94. CrossRef Medline
- Giros B, Jaber M, Jones SR, Wightman RM, Caron MG (1996) Hyperlocomotion and indifference to cocaine and amphetamine in mice lacking the dopamine transporter. Nature 379:606–612. CrossRef Medline
- Greengard P (2001) The neurobiology of slow synaptic transmission. Science 294:1024–1030. CrossRef Medline
- Hasbi A, Fan T, Alijaniaram M, Nguyen T, Perreault ML, O'Dowd BF, George SR (2009) Calcium signaling cascade links dopamine D1–D2 receptor

- heteromer to striatal BDNF production and neuronal growth. Proc Natl Acad Sci U S A 106:21377–21382. CrossRef Medline
- Hasbi A, O'Dowd BF, George SR (2010) Heteromerization of dopamine D2 receptors with dopamine D1 or D5 receptors generates intracellular calcium signaling by different mechanisms. Curr Opin Pharmacol 10:93–99. CrossRef Medline
- Hnasko TS, Perez FA, Scouras AD, Stoll EA, Gale SD, Luquet S, Phillips PE, Kremer EJ, Palmiter RD (2006) Cre recombinase-mediated restoration of nigrostriatal dopamine in dopamine-deficient mice reverses hypophagia and bradykinesia. Proc Natl Acad Sci U S A 103:8858–8863. CrossRef Medline
- Johnston TH, Lee J, Gomez-Ramirez J, Fox SH, Brotchie JM (2005) A simple rodent assay for the in vivo identification of agents with potential to reduce levodopa-induced dyskinesia in Parkinson's disease. Exp Neurol 191:243–250. CrossRef Medline
- Jones SR, Gainetdinov RR, Jaber M, Giros B, Wightman RM, Caron MG (1998) Profound neuronal plasticity in response to inactivation of the dopamine transporter. Proc Natl Acad Sci U S A 95:4029–4034. CrossRef Medline
- Kotecha SA, Oak JN, Jackson MF, Perez Y, Orser BA, Van Tol HH, MacDonald JF (2002) A D2 class dopamine receptor transactivates a receptor tyrosine kinase to inhibit NMDA receptor transmission. Neuron 35: 1111–1122. CrossRef Medline
- Lee SP, So CH, Rashid AJ, Varghese G, Cheng R, Lança AJ, O'Dowd BF, George SR (2004) Dopamine D1 and D2 receptor Co-activation generates a novel phospholipase C-mediated calcium signal. J Biol Chem 279: 35671–35678. CrossRef Medline
- Managò F, Espinoza S, Salahpour A, Sotnikova TD, Caron MG, Premont RT, Gainetdinov RR (2012) The role of GRK6 in animal models of Parkinson's disease and L-DOPA treatment. Sci Rep 2:301. CrossRef Medline
- Masri B, Salahpour A, Didriksen M, Ghisi V, Beaulieu JM, Gainetdinov RR, Caron MG (2008) Antagonism of dopamine D2 receptor/beta-arrestin 2 interaction is a common property of clinically effective antipsychotics. Proc Natl Acad Sci U S A 105:13656–13661. CrossRef Medline
- Missale C, Nash SR, Robinson SW, Jaber M, Caron MG (1998) Dopamine receptors: from structure to function. Physiol Rev 78:189–225. Medline
- Nally RE, McNamara FN, Clifford JJ, Kinsella A, Tighe O, Croke DT, Fienberg AA, Greengard P, Waddington JL (2003) Topographical assessment of ethological and dopamine receptor agonist-induced behavioral phenotype in mutants with congenic DARPP-32 "knockout." Neuropsychopharmacology 28:2055–2063. CrossRef Medline
- Neve KA (2010) The dopamine receptors. New York: Humana.
- Neve KA, Seamans JK, Trantham-Davidson H (2004) Dopamine receptor signaling. J Recept Signal Transduct Res 24:165–205. CrossRef Medline
- Perreault ML, Hasbi A, Alijaniaram M, Fan T, Varghese G, Fletcher PJ, Seeman P, O'Dowd BF, George SR (2010) The dopamine D1–D2 receptor heteromer localizes in dynorphin/enkephalin neurons: increased high affinity state following amphetamine and in schizophrenia. J Biol Chem 285:36625–36634. CrossRef Medline
- Perreault ML, Hasbi A, O'Dowd BF, George SR (2011) The dopamine d1–d2 receptor heteromer in striatal medium spiny neurons: evidence for a third distinct neuronal pathway in basal ganglia. Front Neuroanat 5:31. CrossRef Medline
- Rajagopal S, Rajagopal K, Lefkowitz RJ (2010) Teaching old receptors new tricks: biasing seven-transmembrane receptors. Nat Rev Drug Discov 9:373–386. CrossRef Medline
- Rashid AJ, So CH, Kong MM, Furtak T, El-Ghundi M, Cheng R, O'Dowd BF, George SR (2007) D1–D2 dopamine receptor heterooligomers with unique pharmacology are coupled to rapid activation of Gq/11 in the striatum. Proc Natl Acad Sci USA 104:654–659. CrossRef Medline
- Roche KW, O'Brien RJ, Mammen AL, Bernhardt J, Huganir RL (1996) Characterization of multiple phosphorylation sites on the AMPA receptor GluR1 subunit. Neuron 16:1179–1188. CrossRef Medline
- Sahin B, Hawasli AH, Greene RW, Molkentin JD, Bibb JA (2008) Negative regulation of cyclin-dependent kinase 5 targets by protein kinase C. Eur J Pharmacol 581:270–275. CrossRef Medline

- Sahu A, Tyeryar KR, Vongtau HO, Sibley DR, Undieh AS (2009) D5 dopamine receptors are required for dopaminergic activation of phospholipase C. Mol Pharmacol 75:447–453. CrossRef Medline
- Sotnikova TD, Beaulieu JM, Barak LS, Wetsel WC, Caron MG, Gainetdinov RR (2005) Dopamine-independent locomotor actions of amphetamines in a novel acute mouse model of Parkinson disease. PLoS Biol 3:e271. CrossRef Medline
- Undie AS, Friedman E (1990) Stimulation of a dopamine D1 receptor enhances inositol phosphates formation in rat brain. J Pharmacol Exp Ther 253:987–992. Medline
- Undie AS, Berki AC, Beardsley K (2000) Dopaminergic behaviors and signal transduction mediated through adenylate cyclase and phospholipase C pathways. Neuropharmacology 39:75–87. CrossRef Medline
- Undieh AS (2010) Pharmacology of signaling induced by dopamine D(1)-like receptor activation. Pharmacol Ther 128:37–60. CrossRef Medline
- Verma V, Hasbi A, O'Dowd BF, George SR (2010) Dopamine D1–D2 receptor Heteromer-mediated calcium release is desensitized by D1 receptor occupancy with or without signal activation: dual functional regulation by G protein-coupled receptor kinase 2. J Biol Chem 285:35092–35103. CrossRef Medline
- Zachariou V, Sgambato-Faure V, Sasaki T, Svenningsson P, Berton O, Fienberg AA, Nairn AC, Greengard P, Nestler EJ (2006) Phosphorylation of DARPP-32 at Threonine-34 is required for cocaine action. Neuropsychopharmacology 31:555–562. CrossRef Medline
- Zhuang X, Belluscio L, Hen R (2000)  $G_{OLF\alpha}$  mediates dopamine  $D_1$  receptor signaling. J Neurosci 20:RC91. Medline