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PALB2 mutations in familial breast and pancreatic cancer

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Abstract

PALB2 (Partner And Localizer of *BRCA2*) binds to and colocalizes with *BRCA2* in DNA repair. Germline mutations in *PALB2* have been identified in approximately 1–2% of familial breast cancer and 3–4% of familial pancreatic cancer cases. The goal of this study was to evaluate the prevalence of *PALB2* mutations in women with breast cancer without *BRCA1/2* mutations who also had a personal or family history of pancreatic cancer. *PALB2* mutation analysis was performed in 94 non-*BRCA1/2* breast cancer patients with a personal or family history of pancreatic cancer. Two truncating *PALB2* mutations, c.3549C>CA and c.2962C>CT, were identified resulting in a mutation prevalence of 2.1%. The proband found to carry the c.3549C>CA *PALB2* mutation had a mother diagnosed with both breast and pancreatic cancer; this

relative was subsequently confirmed to carry the identical mutation. The proband with the c. 2962C>CT mutation had a father and paternal aunt diagnosed with pancreatic cancer; neither relative was available for testing. Two novel *PALB2* missense variants were also found, one of which was deemed potentially deleterious. The prevalence rate of *PALB2* mutations in a non-*BRCA1/2* breast cancer population specifically selected for a family history of pancreatic cancer does not appear to be significantly increased compared to that observed in other breast cancer populations studied thus far. Further evaluation is needed to determine the prevalence of *PALB2* mutations and the clinical utility of such testing in those individuals affected with both breast and pancreatic cancers.

Keywords

BRCA2; Breast cancer; PALB2; Pancreatic cancer

Introduction

Breast cancer is a common disease, affecting over one million women worldwide every year [1]. Though the vast majority of breast cancers are sporadic in nature, approximately 5–10% of breast cancers are due to autosomal dominant inheritance of a specific genetic mutation [2, 3]. *BRCA1* and *BRCA2* are the most widely known and best understood genes among the current panel of breast cancer susceptibility genes. However, mutations in *BRCA1* and *BRCA2* account for no more than half of all hereditary breast cancers [4, 5]. Thus, the need to identify the contribution of other breast cancer susceptibility genes is crucial.

PALB2 (Partner And Localizer of BRCA2) is a recently discovered, moderate-risk breast cancer susceptibility gene [6, 7]. *PALB2* (*FANCN*) and *BRCA2* (*FANCD1*) are Fanconi anemia (FA) genes that function in the FA-Breast Cancer (BRCA) DNA repair pathway. Biallelic mutations in *PALB2* or *BRCA2* result in the development of Fanconi anemia [8]. The *PALB2* gene product functions as a tumor suppressor and interacts closely with both *BRCA1* and *BRCA2* during double-strand DNA repair. *PALB2* acts as a physical link between *BRCA1* and *BRCA2* to form a “BRCA complex” which then binds with *RAD51* to initiate homologous recombination [9–12]. It is this close functional interaction with known high-risk breast cancer genes, particularly *BRCA2*, that ultimately helped to identify *PALB2* as a candidate breast cancer susceptibility gene [13].

Though *PALB2* gene mutations are rare among the general population, studies estimate a prevalence of 1–2% in patients with familial breast cancer [6, 14]. *PALB2* gene mutation carriers appear to have a two- to four-fold relative risk of developing breast cancer; however, these risk estimates may be even higher in those patients with a family history of breast cancer [6, 15]. There is additional data to suggest that some *PALB2*-associated breast cancers display a more aggressive tumor phenotype, including triple-negative disease, higher tumor grade and higher *Ki67* expression [16]. In fact, nearly 40% of the *PALB2*-associated breast cancers identified to date display a triple-negative phenotype, regardless of the specific *PALB2* mutation [12].

In addition, *PALB2* mutations have recently been shown to be associated with an increased risk of familial pancreatic cancer. Tischkowitz et al. [17] screened a cohort of 254 pancreatic cancer patients unselected for family history and found a *PALB2* mutation prevalence of <1%. However, Jones et al. [18] sequenced *PALB2* in 96 patients with familial pancreatic cancer and found an overall prevalence of 3.1%. Slater et al. [19] found a similar prevalence of *PALB2* mutations (3.7%) in a panel of 81 European patients with familial pancreatic

cancer. Thus, *PALB2* gene mutations appear to be more prevalent in those patients with a family history of pancreatic cancer.

Taken together, these studies raise the question as to whether *PALB2* gene mutations are more prevalent in families with *both* breast and pancreatic cancers. We hypothesized that a higher prevalence of *PALB2* mutations might exist in this population given: (1) the close functional relationship that *PALB2* shares with *BRCA2*; (2) the known association of *PALB2* mutations with both breast and pancreatic cancers; and (3) the well-recognized *BRCA2* phenotype of familial breast and pancreatic cancers [20, 21]. Thus, the goal of this study was to determine the prevalence of *PALB2* mutations in a population of *BRCA1/2*-negative breast cancer patients specifically selected for a personal and/or family history of pancreatic cancer.

Methods

Subject selection

Eligible subjects were identified through three university-affiliated cancer risk programs. All subjects were required to have a personal history of breast cancer (invasive or DCIS) and negative testing for both *BRCA1* and *2* mutations (Table 1). Other specific eligibility criteria included: (1) Personal history of breast cancer at age ≥ 50 with family history of one or more cases of pancreatic cancer in first or second degree relatives in the same lineage of the family; or (2) Personal history of breast cancer at age ≥ 65 with family history of one or more cases of breast cancer diagnosed at age ≥ 65 and one or more cases of pancreatic cancer in first or second degree relatives in the same lineage of the family; or (3) Personal history of breast cancer at age ≥ 65 with family history of two or more cases of pancreatic cancer in first or second degree relatives in the same lineage of the family; or (4) Personal history of breast cancer at age ≥ 65 with personal history of pancreatic cancer; or (5) Personal history of bilateral breast cancer with family history of one or more cases of pancreatic cancer in first or second degree relatives in the same lineage of the family. Cancer diagnoses for probands were confirmed by pathology reports; cancer diagnoses for relatives were determined only through proband interviews, though pathologic confirmation was occasionally possible.

Data collection

Clinical information relevant to eligible patients was abstracted from medical records and included: age at breast cancer diagnosis, breast cancer treatment and outcome. Information regarding breast cancer pathology was abstracted from pathology reports and included histology, estrogen receptor (ER), progesterone receptor (PR) and HER2 status, grade and nodal status. Information regarding family history of cancers was also collected from both lineages and included: types of cancers, age at cancer diagnoses, and family ethnicity.

Laboratory methods

Blood or saliva was collected from each study subject for DNA analysis. DNA was extracted from blood using a Qiagen DNA extraction kit and from saliva using the Oragene DNA extraction procedure. Patients were tested for *PALB2* alterations using Capillary Exon Grouping Analysis (cEGAN). Exon grouping analysis is based on Conformation-Specific Capillary Electrophoresis [22, 23]. All coding exons and surrounding intronic sequences were amplified with 19 primer pairs and analyzed on ABI-3730XL instruments (Applied Biosystems, Foster City, California). Polymerase chain reaction fragments with aberrant mobility were sequenced for the first 12 exons. Exon 13 was directly sequenced in all patients. This method permits detection of mutations and polymorphisms in *PALB2* coding

regions as well as splice-site mutations. *BRCA1* and *2* gene sequencing for all cases was performed through Myriad Genetics Inc.

Statistical methods

The 90% confidence interval (CI) of the prevalence of germline *PALB2* mutations was computed using the method of exact binomial confidence interval [24].

All patients provided informed consent for tissue analysis and medical record review. This study was approved by the institutional review boards of Dana Farber/Harvard Cancer Center and University of Pennsylvania Cancer Center.

Results

A total of 94 cases from independent families were selected for *PALB2* sequence analysis. None of the subjects had a *BRCA* mutation or variant of uncertain significance. Characteristics of the 94 study subjects are summarized in Table 1. Median age at breast cancer diagnosis was 41 years, with 29.8% of subjects being diagnosed with multiple breast cancers. Three subjects had a personal history of both breast and pancreatic cancers. All subjects except one had at least one first- or second-degree family member with pancreatic cancer, and 11.7% of subjects had two relatives with pancreatic cancer. (One patient had a personal history of breast and pancreatic cancer, but no family history of pancreatic cancer). The majority of the subjects in our study were of Northern European descent (62/94, 65.9%).

Truncating *PALB2* mutations were identified in 2/94 subjects (2.1%; 90% CI = 0.40%, 6.5%). Both subjects were of Northern European/Caucasian descent. In the first subject, *PALB2* sequence analysis revealed a nonsense mutation in exon 9, c.2962C>CT, which results in a stop codon at amino acid 988. This subject was diagnosed with pre-menopausal breast cancer at age 49. Pathology review showed a 1.5 cm, poorly differentiated, invasive ductal breast carcinoma that was negative for both estrogen receptor (ER) and Her2 over-expression. The subject also had a basal cell carcinoma of the skin at age 47. The family history was remarkable for a father with pancreatic cancer at age 62 and a paternal aunt with pancreatic cancer at age 60 (Fig. 1A). Of note, the subject's mother and two maternal aunts were diagnosed with post-menopausal breast cancer. The subject's mother was also affected with uterine cancer at age 75. Tissue from family members was unavailable for analysis.

In the second subject found to have a mutation, *PALB2* sequence analysis revealed a nonsense mutation in exon 13, c.3549C>CA, which results in a stop codon at amino acid 1183. This subject was initially diagnosed with pre-menopausal breast cancer at age 43. Pathology review showed a 2.5 cm invasive ductal carcinoma, ER-positive, Her2 status unknown. Despite treatment with mastectomy, chemotherapy and tamoxifen, the breast cancer recurred with distant metastases at age 47. The family history was remarkable for a mother with breast cancer at age 50 and pancreatic cancer at age 61 (Fig. 1B). Other notable cancers within the pedigree included a maternal uncle with colon cancer at age 48, a maternal grandfather with pharyngeal cancer at age 71, and a paternal aunt with colon cancer at age 30. *PALB2* sequence analysis of the mother's blood revealed the same *PALB2* mutation identified in the proband. Tissue from other family members was unavailable for analysis.

In addition, sequence analysis revealed thirteen different *PALB2* variants and polymorphisms (Table 2). Among these, two novel missense variants were identified: 1222T>TC (p.Y408YH) and 3128G>GC(p.G1043GA). Both of the subjects found to have a novel missense variant were of Northern European descent. Pathogenicity of the novel

missense variants was analyzed using three in silico tests (PolyPhen, SIFT and PMut). Based on these results, the variant 1222T>TC was predicted to be a neutral change by two of three tests (SIFT, PMut). However, the variant 3128G>GC was predicted to have potential deleterious effects. This variant codes for 1043G>AG, which is located in WD repeat 4 (1010–1052). Interaction with *BRCA2* occurs through a hydrophobic pocket at the crossover between WD repeats 4 and 5 (1053–1057). Thus, it is quite possible that this alteration may abrogate the interaction of *PALB2* and *BRCA2* [25]. The subject with the 3128G>GC variant was diagnosed with pre-menopausal breast cancer at age 40. Pathology review showed a 2.3 cm, grade 3, invasive ductal carcinoma which was ER-positive and Her 2-negative. Extensive lymphatic invasion was noted in addition to three positive axillary lymph nodes. The family history was remarkable for a paternal grandfather with pancreatic cancer. No family members were available for genetic testing. In addition a novel synonymous variant was also found, 1881G>GT (p. V627VV). Consistent with previously reported synonymous variants, this variant is predicted to be non-deleterious and pathogenicity was therefore not assessed.

Discussion

We found *PALB2* mutations in two of 94 subjects, a prevalence of 2.1% in this specific breast cancer population. This rate is consistent with the 1–2% prevalence of mutations reported in other familial breast cancer populations [6, 14]. A recent study by Adank et al. [26] evaluated the prevalence of *PALB2* mutations in 110 *BRCA2*-like families, including 45 breast cancer patients with a first or second degree relative with pancreatic cancer. While one truncating *PALB2* mutation was found in a male breast cancer patient, none of the 45 subjects with breast cancer and a family history of pancreatic cancer was found to carry a pathogenic mutation. The difference in *PALB2* mutation prevalence between our study and that of Adank et al. is likely not significant and may be due to the small sample sizes. While the *PALB2* mutation prevalence rate of 2.1% in our population is higher than that reported by Adank et al., it is lower than that found in studies of familial pancreatic cancer (3.1–3.7%) [18, 19].

The c.3549C>CA and c.2962C>CT *PALB2* mutations identified in our sample have been previously described in breast cancer and Fanconi anemia patients, respectively [8, 18]. However, our study is the first to specifically identify the c.3549C>CA *PALB2* mutation in association with pancreatic cancer. This mutation was identified in the mother of a proband within our sample; the mother was affected with both breast and pancreatic cancer. It is likely that the c.2962C>CT *PALB2* mutation is specifically associated with the pancreatic cancer in our proband's family as well, however tissue from family members affected with pancreatic cancer was unavailable for analysis. With the addition of our data on the family with the c.3549C>CA *PALB2* mutation, there are now nine *PALB2*-related pancreas families identified to date [17–19]. Interestingly, eight of these families include one or more cases of breast cancer, and three families contain individuals with both breast and pancreatic cancer. Within the familial pancreatic cancer population described in the report of Tischkowitz et al. [17] one of nine subjects (11.1%; 90% CI = 0.57%, 42.9%) with both breast and pancreatic cancer carried a *PALB2* mutation. We performed *PALB2* sequence analysis on four individuals with concomitant breast and pancreatic cancer, namely three probands and one relative (mother) of a proband with breast cancer. Of these, one mutation carrier was found. Though numbers are small, these rates of *PALB2* mutations within individuals with both breast and pancreatic cancer is noteworthy and warrants further study of this patient population.

Previous reports have suggested that breast tumors associated with some *PALB2* mutations are aggressive in nature [12, 16]. Our data further supports this concept, as both mutation

carriers in our sample proved to have aggressive disease. The breast cancer in the first mutation carrier was notable for poor differentiation and triple-negative status, lacking ER, PR and HER2 over-expression. In the second mutation carrier, the breast tumor recurred with distant metastases 4 years after diagnosis despite initial aggressive therapy for ER-positive, node-negative disease. These observations must be interpreted with caution given the small sample size, but are nevertheless intriguing.

In conclusion, we found that *PALB2* mutations occur with a prevalence of 2.1% in a population of *BRCA1/2*-negative breast cancer patients specifically selected for a personal and/or family history of pancreatic cancer. This prevalence rate appears comparable to the rate of *PALB2* mutations published in other breast cancer populations. Further study is needed to determine the prevalence of *PALB2* mutations and the clinical utility of such testing in those individuals affected with both breast and pancreatic cancers. The benefit of pancreatic cancer screening in patients at increased risk of developing pancreatic cancer, such as those who carry a *BRCA2* or *PALB2* mutation, is uncertain. Recently, Verna et al. [27] reported that endoscopic ultrasound (EUS) and MRI were able to detect early neoplastic pancreatic lesions in 12% of 51 individuals with a family history of pancreatic cancer. Other studies evaluating the benefit of such screening are ongoing. If pancreatic cancer screening is found to be beneficial in high risk populations, identifying those individuals with *PALB2* mutations will likely have even greater clinical significance. Currently, however, routine diagnostic screening for *PALB2* mutations in any breast cancer patient is not indicated.

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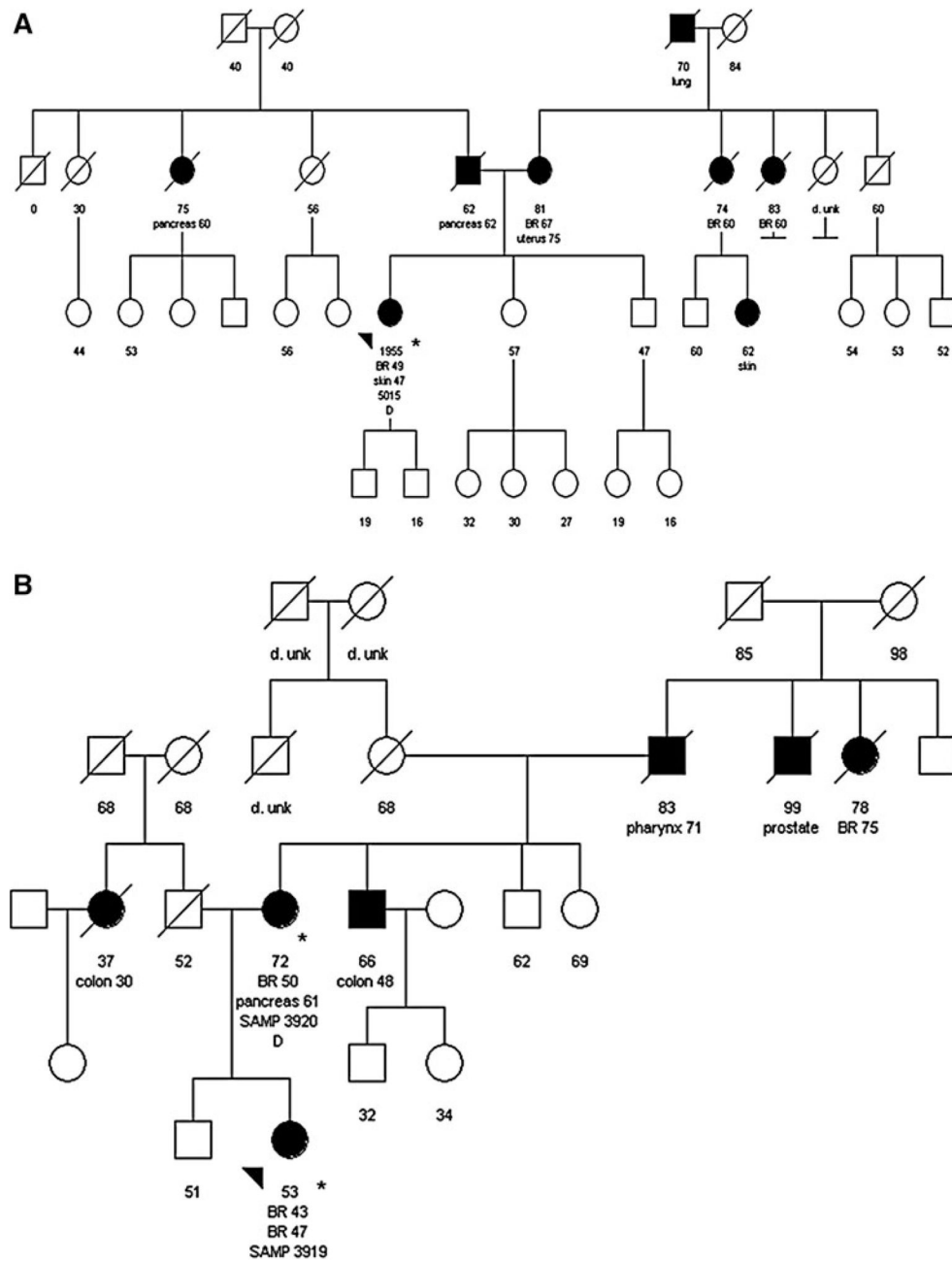


Fig. 1. Pedigrees of the families with *PALB2* mutations
A depicts the family carrying the c.2962C>CT mutation. **B** depicts the family carrying the c.3549C>CA mutation. Individuals with cancer are shown in *black*. The *arrow* indicates the proband carrying the *PALB2* mutation. Cancer type (BR breast cancer), age at diagnosis, and age of death are shown below each individual. *Asterisks* indicate those patients for whom cancer diagnoses were confirmed by pathology report

Table 1

Characteristics of study subjects

	<i>N</i> (%)
Total number of subjects	94
Median age at first breast cancer diagnosis (range)	41 (31–66)
Subjects with multiple breast cancers	28 (29.8%)
Subjects with both breast and pancreatic cancers	3 (3.2%)
Family history of breast cancer in eligible lineage ^a	
1 family member	38 (40.4%)
2 family members	16 (17.0%)
Family member with bilateral breast cancer	4 (4.3%)
Family history of pancreatic cancer in eligible lineage ^b	
1 family member	82 (87.2%)
2 family members	11 (11.7%)

^aFamily members include both 1st and 2nd degree relatives

^bOne subject had only a personal history of pancreatic cancer (and no family history)

Table 2

Identified *PALB2* variants

Variant type	<i>PALB2</i> cDNA change	<i>PALB2</i> protein change	Number observed	Prediction programs		
				PolyPhen	SIFT	PMut
<i>Novel variants</i>						
Mis	c.3128G > GC	p.L1043G > AG	1	Possibly damaging	Affect protein function	Pathological
Mis	c.1222T > TC	p.Y408Y > YH	1	Possibly damaging	Tolerated	Neutral
Syn	c.1881G > GT	p.V627VV	1	N/A	N/A	N/A
<i>Known variants</i>						
Mis	c.1010T > TC ^a	p.L337LS	5	Possibly damaging	Affect protein function	–
Mis	c.1676A > AG ^a	p.Q559QR	16	Benign	Tolerated	–
Mis	c.2014G > GC ^a	p.E672EQ	2	Benign	Affect protein function	–
Mis	c.2590C > CT ^a	p.P864PS	3	Probably Damaging	Tolerated	–
Mis	c.2794G > GA ^b	p.V932VM	1	Benign	Tolerated	–
Mis	c.2816T > TG ^a	p.L939LW	1	Possibly damaging	Affect protein function	–
Mis	c.2993G > GA ^a	p.G998GE	1	Probably damaging	Affect protein function	–
Syn	c.1194G > GA ^b	p.V398VV	1	N/A	Tolerated	N/A
Syn	c.1572A > AG ^b	p.S524SS	1	N/A	Tolerated	N/A
Syn	c.3300T > TG ^b	p.T1100TT	2	N/A	Tolerated	N/A

Mis: Missense variant, *Syn*: synonymous variant, *PolyPhen*: Polymorphism Phenotyping prediction (<http://genetics.bwh.harvard.edu/pph>), *SIFT*: Sorting Intolerant From Tolerant (http://blocks.fhcrc.org/sift/SIFT_BLink_submit.html), *PMut* (<http://mmb2.pcb.ub.es:8080/PMut>)

^aPreviously reported [26]

^bPreviously reported [6]

N/A Not applicable

– Information not available