Perspective

The VEGF signaling pathway in cancer: the road ahead

Steven A. Stacker^{1,2} and Marc G. Achen^{1,2}

Abstract

The vascular endothelial growth factor (VEGF) family of soluble protein growth factors consists of key mediators of angiogenesis and lymphangiogenesis in the context of tumor biology. The members of the family, VEGF-A (also known as VEGF), VEGF-B, VEGF-C, VEGF-D, and placenta growth factor (PIGF), play important roles in vascular biology in both normal physiology and pathology. The generation of a humanized neutralizing antibody to VEGF-A (bevacizumab, also known as Avastin) and the demonstration of its benefit in numerous human cancers have confirmed the merit of an anti-angiogenesis approach to cancer treatment and have validated the VEGF-A signaling pathway as a therapeutic target. Other members of the VEGF family are now being targeted, and their relevance to human cancer and the development of resistance to anti-VEGF-A treatment are being evaluated in the clinic. Here, we discuss the potential of targeting VEGF family members in the diagnosis and treatment of cancer.

Key words Angiogenesis, vascular endothelial growth factor (VEGF), VEGF receptors, monoclonal antibody, cancer metastasis

The theory that blood vessels might be critical for the growth of tumors has its roots throughout the early and mid-20th century[1,2]. Initially, observations by pathologists and cancer biologists suggested an association between blood vessels and tumors^[3,4]. Using experimental models. cancer biologists observed that blood vessels were essential to support tumor growth beyond the size allowed by oxygen diffusion alone [5]. The concept of blood vessels supporting tumor growth gained critical support from the theory and work of Judah Folkman, who proposed the relationship between neo-vessels and tumor growth [6]. While Folkman's work highlighted the importance of angiogenic factors in tumors, it was not until the discovery and characterization of vascular endothelial growth factor-A (VEGF-A; also known as VEGF), and the development of inhibitory monoclonal antibodies that blocked the binding of VEGF-A to key receptors, that an in vivo proof-of-principle experiment targeting a known tumor angiogenic factor was performed[2,7].

Authors' Affiliations: 'Tumour Angiogenesis Program, Peter MacCallum Cancer Centre, St. Andrews Place, East Melbourne, Victoria 3002, Australia. 2Sir Peter MacCallum Department of Oncology, University of Melbourne, Parkville, Victoria 3050, Australia.

Corresponding Authors: Steven A. Stacker and Marc G. Achen, Peter MacCallum Cancer Centre, Locked Bag 1, A'Beckett Street, Victoria 8006, Australia. Tel: +61-3-96565263; Fax: +61-3-96561411; Email: steven. stacker@petermac.org, marc.achen@petermac.org.

doi: 10.5732/cjc.012.10319

The VEGF family of proteins first came to the attention of cancer biologists in the early 1980s when vascular permeability factor (VPF) was discovered in the ascites fluid of cancer patients[8]. It was another 6 years before this molecule was discovered to be identical to VEGF-A, a mitogen in endothelial cells capable of promoting angiogenesis in vivo [9,10]. With the isolation of the VEGF-A cDNA, the entire predicted amino acid sequence for this multifunctional cytokine was defined. Structural similarity to the transforming growth factor-B (TGF-β) family and a highly conserved cysteine-rich motif quickly assigned VEGF-A to the burgeoning family of cystine-knot growth factors[11]. Since then, related genes encoding other members of the VEGF family have been isolated, and their protein products have been characterized in developmental, vascular, and cancer biology [12-18]. As is commonly observed with ligands of a growth factor family, the related growth factors share a common series of receptors for signal transduction. The VEGF family signals predominantly through the receptor tyrosine kinases VEGF receptor (VEGFR)-1, VEGFR-2, and VEGFR-3, in combination with the co-receptors neuropilin (NP)-1 and NP-2, and, in some cases, can use other receptors such as integrins[19].

VEGF-A

VEGF-A has become the most widely studied tumor

angiogenic factor. The generation of a humanized monoclonal antibody (mAb) that inhibits the interaction of VEGF-A with its receptors VEGFR-1 and VEGFR-2 [20] and the development of small-molecule protein tyrosine kinase (PTK) inhibitors to the cytoplasmic regions of the VEGF receptors^[21] have provided avenues to explore the efficacy of targeting the VEGF-A signaling pathway in cancer. Extensive mouse tumor models developed through the 1990s [7] led to the success of the VEGF-A mAb known as Avastin (or bevacizumab) in treating a range of human cancers including colorectal cancer, lung cancer, and renal cell carcinoma. However, the recent withdrawal of Avastin for treating breast cancer in the United States has highlighted the limitations of anti-angiogenic treatments [22], as well as the need for a careful cost-benefit analysis to ensure that scarce healthcare resources are well spent in the treatment of cancer. While Avastin is clearly beneficial in a subset of cancer patients, the development of resistance to Avastin and the pre-existing refractoriness of some tumors suggest the role of other growth factors in sustaining tumor angiogenesis. Alternative ligands (other than VEGF-A) for the angiogenic receptor VEGFR-2 (i.e., VEGF-C and VEGF-D; see below) and other angiogenic factors from distinct growth factor families [e.g., fibroblast growth factor (FGF)] are logical candidates for promoting resistance to Avastin. In addition, other mechanisms, such as vascular mimicry, the involvement of endothelial precursor cells (EPCs) or altered tumor metabolism leading to reduced oxygen requirement, may also contribute to the development of resistance^[21].

Placenta Growth Factor (PIGF)

PIGF is a ligand for VEGFR-1, which has been reported to play a dual role in cancer as a stimulator of angiogenesis and as an autocrine or paracrine factor promoting proliferation of tumor cells expressing VEGFR-1^[23,24]. It has also been proposed that PIGF may modulate the effect of VEGF-A by displacing VEGF-A from VEGFR-1 expressed on endothelial cells. A range of animal tumor models have provided supportive or contradictory data on the role of PIGF in driving tumor angiogenesis and tumor growth[25,26]. These findings suggest a context-dependent activity for PIGF in cancer. either due to the presence of other VEGF family members or the diverse expression of VEGF receptors on various cell types including endothelial cells, tumor cells, or associated immune cells[27,28].

VEGF-B

While VEGF-B, a ligand for VEGFR-1, has been shown to be expressed in human cancer^[29], its functions

in arteriogenesis [30] and in fatty acid uptake by cells [31] have highlighted its role in other pathologies, predominantly cardiovascular disease. Recently, VEGF-B was shown to regulate endothelial cell uptake and transport of fatty acids in muscle, leading to the hypothesis that VEGF-B antagonists could be used as novel pharmacologic agents in the treatment of type 2 diabetes[31]. Although inhibitory antibodies against VEGF-B have been developed[32], so far there is little data on the role of this growth factor in tumor biology; it was reported, however, that overexpression of VEGF-B in a mouse model of pancreatic cancer suppressed tumor growth, highlighting the differences in biological functions between the VEGF family members[33].

VEGF-C

VEGF-C and VEGF-D form a subfamily within the VEGF family, consisting of highly related polypeptides that require post-translational cleavage of the N- and Cterminal propeptides to generate mature forms with enhanced binding affinities for VEGFR-2 and VEGFR-3^[16,34-36]. Notably, VEGFR-3 is expressed on lymphatic endothelium, where it can promote lymphangiogenesis[37], and it is also expressed on angiogenic blood vessels [38]. VEGF-C is expressed in a range of human cancers, including solid tumors such as nasopharyngeal, liver and gastric cancers, with a preponderance in Asian populations [39]. Mouse tumor studies have displayed the ability of VEGF-C to promote tumor angiogenesis and lymphangiogenesis in vivo and to drive tumor growth and metastatic spread [40-42]. Given that the mature form of VEGF-C is a high affinity ligand for VEGFR-2 and that VEGF-C is expressed in many human cancers, VEGF-C is likely to be an alternative ligand to VEGF-A for VEGFR-2 - binding, which could in turn promote tumor angiogenesis. Therefore, VEGF-C, in combination with anti-angiogenic drugs such as Avastin, may be a viable target for anti-cancer therapy.

VEGF-D

The growth factor VEGF-D is closely related in structure to VEGF-C, and it includes a central VEGF homology domain (VHD) related to other VEGF family members, with N- and C-terminal propeptides^{16]} that can be proteolytically cleaved by enzymes such as proprotein convertases and plasmin[36,43,44]. VEGF-D is expressed in a range of human cancers [39,45,46] and has been associated with poor patient outcome in some tumor types [47,48]. Importantly, animal models of cancer have demonstrated that VEGF-D can promote tumor angiogenesis and lymphangiogenesis, solid tumor growth, dilation of collecting lymphatic vessels, and lymphatic and distant

organ metastasis [49-53]. The proteolytic processing of VEGF-D is required for promoting tumor growth and spread^[54]. Opportunities for targeting VEGF-D signaling in cancer could involve mAbs specific to the VHD of VEGF-D that are capable of inhibiting binding to VEGFR-2 and VEGFR-3^[49,55,56]. Alternatively, PTK inhibitors that block VEGFR-2 and VEGFR-3 signaling would interfere with VEGF-D-mediated signal transduction [21]. Further, mAbs to VEGFR-2 and VEGFR-3[57,58] that would prevent the binding of VEGF-D, or a soluble form of VEGFR-3 that could sequester both VEGF-C and VEGF-D [59], could be employed. Targeting the VEGF-D signaling pathway would likely have the merit of inhibiting both tumor angiogenesis and lymphangiogenesis^{60]}, which could, in turn, restrict both solid tumor growth and metastatic spread.

Future

What have the past 20 years taught us about targeting VEGF-A signaling? Clearly, this period has provided biochemists, biologists, and clinicians the time to design and evaluate a variety of agents that modify or inhibit these signaling pathways. The agents that were developed have included a broad range of molecules targeting different components of the pathways, including VEGF-A itself, VEGF receptors, VEGF co-receptors, PTKs and signaling intermediates, and transcription factors. During this period, other VEGF family members were also identified, enhancing the diversity of signaling induced by the VEGF family of ligands.

Avastin, a humanized mAb to VEGF-A, has been widely used in a range of prevalent human cancers over the past 8 years, typically in combination with cytotoxic chemotherapy. Although this agent has provided significant benefit to cancer patients, there is a need for other drugs that could be combined with Avastin to deliver improved clinical outcomes. Use of agents targeting other VEGF family members, in combination with Avastin, may be a potential approach. Further, small-molecule PTK inhibitors of VEGF receptors (that are not highly specific) have been employed although dose-limiting toxicity in combination with cytotoxic chemotherapy has restricted their widespread use. Further insights into the structure and function of the PTK domains of VEGF receptors may allow development of more specific small-molecule PTK inhibitors.

Further studies delving into the complex network of signaling cascades that drive angiogenesis, lymphangiogenesis, and resistance to anti-angiogenic drugs such as Avastin are clearly required. Genome-wide functional approaches such as those using small interfering RNA (siRNA) technology, supported by bioinformatics, could help strategize effective targeting of growth factors, receptors, and PTKs.

Agents specifically targeting VEGF family members and their receptors are currently in various stages of development; mAbs to VEGF-B, VEGF-C, VEGF-D, PIGF, VEGFR-1, VEGFR-2, and VEGFR-3 are being evaluated by the biotechnology and pharmaceutic industries for their efficacy as anti-cancer agents. For these mAbs, a key challenge will be identifying specific cancer indications in which clinical benefit can be achieved. Given that many cancers are resistant to Avastin or develop resistance over the course of treatment, these mAbs may be tested in patients whose cancers are Avastin-resistant and/or whose cancers express the appropriate target growth factor. Hence, assays that quantitate the levels of these growth factors or other relevant biomarkers will be required to identify the appropriate patients, although assessing these biomarkers may not be straightforward. VEGF family members are secreted proteins, which may easily allow testing in blood-drawn samples; however, some family members (e.g., VEGF-C and VEGF-D) are proteolytically processed to generate forms with a different bioactivity. Hence, it will be important to fully appreciate which forms are most bioactive and clinically relevant.

It is important to note that inhibiting signaling by VEGF family members could potentially have pro-tumor effects. Recent studies have demonstrated that antiangiogenic treatments could cause increased tumor metastasis through mechanisms such as the induction of hypoxia (caused by VEGF-A inhibition), resulting in increased tumor cell motility [61,62]. Nonetheless, these issues have been addressed in various tumor models [63] and by comparing the effect of antibodies and smallmolecule inhibitors to show that anti-VEGF-A treatment does not promote metastasis[64].

Clearly, other signaling pathways (i.e., not directly involving VEGF family members) such as those driven by members of the platelet-derived growth factor (PDGF), insulin-like growth factor (IGF), and FGF growth factor families, exert control on tumor angiogenesis and may be important in promoting tumor growth. Other alternative molecular targets, such as c-MET, may also be able to affect multiple tumor properties such as metastasis, angiogenesis, tumor growth, and cellular motility upon hypoxia^[65-67]. Although not the subject of this review, such pathways may provide valid therapeutic targets for inhibiting angiogenesis and/or lymphangiogenesis in cancer, thereby restricting tumor growth and spread. Furthermore, recent studies from our own laboratory have indicated the role of known inflammatory pathways in enhancing the spread of cancer^[51], opening avenues for the use of existing anti-inflammatory drugs in cancer. This possibility is further exemplified by the recent studies published by Rothwell et al. [68,69] demonstrating the anti-cancer effect of non-steroidal anti- inflam matory drugs (NSAIDs) such as aspirin. While expensive drugs such as Avastin allow the treatment of specific cancers in the Western world, the capacity to employ relatively inexpensive "off-patent" small molecules (such as aspirin) with a defined chemistry and known biological could provide a cost-effective population-based preventative approach to cancer in Asian countries.

Acknowledgments

This work was funded partly by a program grant from

the National Health and Medical Research Council of Australia (NH&MRC). SAS and MGA are supported by Senior Research Fellowships from the NH&MRC. SAS would like to acknowledge the support of the Pfizer Australia Fellowship, SAS and MGA are consultants for Vegenics Ltd, a company which develops inhibitors of angiogenesis.

Received: 2012-12-27; revised: 2013-01-21;

accepted: 2013-01-22.

References

- [1] Ferrara N, Hillan KJ, Gerber HP, et al. Discovery and development of bevacizumab, an anti-VEGF antibody for treating cancer. Nat Rev Drug Discov, 2004,3:391-400.
- Ferrara N. VEGF and the quest for tumour angiogenesis factors. Nat Rev Cancer, 2002,2:795-803.
- Lewis WH. The vascular pattern of tumors. Johns Hopkins Hosp Bull, 1927,41:156-162.
- [4] Algire GH, Chalkley HW. Vascular reactions of normal and malignant tissues in vivo. I. Vascular reactions of mice to wounds and to normal and neoplastic transplants. J Natl Cancer Inst, 1945,6:73-85.
- Folkman J, Long DM, Becker FF. Growth and metastasis of tumor in organ culture. Cancer, 1963, 16:453-467.
- [6] Folkman J. Tumor angiogenesis: therapeutic implications. N Engl J Med, 1971,285:1182-1186.
- Kim KJ, Li B, Winer J, et al. Inhibition of vascular endothelial growth factor-induced angiogenesis suppresses tumour growth in vivo. Nature, 1993,362:841-844.
- [8] Senger DR, Galli SJ, Dvorak AM, et al. Tumour cells secrete a vascular permeability factor that promotes accumulation of ascities fluid. Science, 1983,219:983-985.
- Keck PJ, Hauser SD, Krivi G, et al. Vascular permeability factor, an endothelial cell mitogen related to PDGF. Science, 1989,246:1309-1312.
- [10] Leung DW, Cachianes G, Kuang WJ, et al. Vascular endothelial growth factor is a secreted angiogenic mitogen. Science, 1989,246:1306-1309.
- [11] McDonald NQ, Hendrickson WA. A structural superfamily of growth factors containing a cystine knot motif. Cell, 1993,73: 421 - 424
- [12] Olofsson B, Pajusola K, von Euler G, et al. Genomic organization of the mouse and human genes for vascular endothelial growth factor B (VEGF-B) and characterization of a second splice isoform. J Biol Chem, 1996,271:19310-19317.
- [13] Paavonen K, Horelli-Kuitunen N, Chilov D, et al. Novel human vascular endothelial growth factor genes VEGF-B and VEGF-C localize to chromosomes 11g13 and 4g34, respectively. Circulation, 1996,93:1079-1082.
- [14] Joukov V, Pajusola K, Kaipainen A, et al. A novel vascular endothelial growth factor, VEGF-C, is a ligand for the Flt4 (VEGFR-3) and KDR (VEGFR-2) receptor tyrosine kinases. EMBO J, 1996,15:290-298.
- [15] Yamada Y, Nezu JI, Shimane M, et al. Molecular cloning of a novel vascular endothelial growth factor, VEGF-D. Genomics, 1997,42:483-488.

- [16] Achen MG, Jeltsch M, Kukk E, et al. Vascular endothelial growth factor D (VEGF-D) is a ligand for the tyrosine kinases VEGF receptor 2 (Flk1) and VEGF receptor 3 (Flt4). Proc Natl Acad Sci USA, 1998,95:548-553.
- [17] Orlandini M, Marconcini L, Ferruzzi R, et al. Identification of a c-fos-induced gene that is related to the platelet-derived growth factor/vascular endothelial growth factor family. Proc Natl Acad Sci USA 1996 93:11675-11680
- [18] Maglione D, Guerriero V, Viglietto G, et al. Isolation of a human placenta cDNA coding for a protein related to the vascular permeability factor. Proc Natl Acad Sci USA, 1991,88: 9267-9271
- [19] Koch S, Tugues S, Li X, et al. Signal transduction by vascular endothelial growth factor receptors. Biochem J, 2011,437:169-
- [20] Hurwitz H, Fehrenbacher L, Novotny W, et al. Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. N Engl J Med, 2004,350:2335-2342.
- [21] Halford MM, Tebbutt NC, Desai J, et al. Towards the biomarker-guided rational use of anti-angiogenic agents in the treatment of metastatic colorectal cancer. Colorectal Cancer, 2012.1:149-161.
- [22] Rossari JR, Metzger-Filho O, Paesmans M, et al. Bevacizumab and breast cancer: a meta-analysis of first-line phase III studies and a critical reappraisal of available evidence. J Oncol. 2012.2012:417673
- [23] Fischer C, Mazzone M, Jonckx B, et al. Flt1 and its ligands VEGFB and PLGF: drug targets for anti-angiogenic therapy? Nat Rev Cancer, 2008,8:942-956.
- [24] Yao J, Wu X, Zhuang G, et al. Expression of a functional VEGFR-1 in tumor cells is a major determinant of anti-PLGF antibodies efficacy. Proc Natl Acad Sci U S A, 2011,108: 11590-11595
- [25] Bais C, Wu X, Yao J, et al. PLGF blockade does not inhibit angiogenesis during primary tumor growth. Cell, 2010,141: 166-177
- [26] Van de Veire S, Stalmans I, Heindryckx F, et al. Further pharmacological and genetic evidence for the efficacy of PIGF inhibition in cancer and eye disease. Cell, 2010,141:178-190.
- [27] Fischer C, Jonckx B, Mazzone M, et al. Anti-PLGF inhibits growth of VEGF(R)-inhibitor-resistant tumors without affecting healthy vessels. Cell, 2007,131:463-475.
- [28] Ribatti D. Novel angiogenesis inhibitors: addressing the issue of redundancy in the angiogenic signaling pathway. Cancer Treat Rev, 2011,37:344-352.

- [29] Kanda M, Nomoto S, Nishikawa Y, et al. Correlations of the expression of vascular endothelial growth factor B and its isoforms in hepatocellular carcinoma with clinico-pathological parameters. J Surg Oncol, 2008,98:190-196.
- [30] Lahteenvuo JE, Lahteenvuo MT, Kivela A, et al. Vascular endothelial growth factor-B induces myocardium-specific angiogenesis and arteriogenesis via vascular endothelial growth factor receptor-1- and neuropilin receptor-1-dependent mechanisms. Circulation, 2009, 119:845-856.
- [31] Hagberg CE, Falkevall A, Wang X, et al. Vascular endothelial growth factor B controls endothelial fatty acid uptake. Nature, 2010 464:917-921
- [32] Scotney PD, MacKenzie A, Maccarone P, et al. Human vascular endothelial growth factor B: characterization of recombinant isoforms and generation of neutralizing monoclonal antibodies. Clin Exp Pharmacol Physiol, 2002,29:1024-1029.
- [33] Albrecht I, Kopfstein L, Strittmatter K, et al. Suppressive effects of vascular endothelial growth factor-B on tumor growth in a mouse model of pancreatic neuroendocrine tumorigenesis. PLoS One 2010 5:e14109
- [34] Joukov V, Pajusola K, Kaipainen A, et al. A novel vascular endothelial growth factor, VEGF-C, is a ligand for the Flt-4 (VEGFR-3) and KDR (VEGFR-2) receptor tyrosine kinases. EMBO J, 1996,15:290-298
- [35] Joukov V, Sorsa T, Kumar V, et al. Proteolytic processing regulates receptor specificity and activity of VEGF-C. EMBO J, 1997,16:3898-3911.
- [36] Stacker SA, Stenvers K, Caesar C, et al. Biosynthesis of vascular endothelial growth factor-D involves proteolytic processing which generates non-covalent homodimers. J Biol Chem. 1999.274:32127-32136.
- [37] Veikkola T, Jussila L, Makinen T, et al. Signalling via vascular endothelial growth factor receptor-3 is sufficient for lymphangiogenesis in transgenic mice. EMBO J, 2001,20:1223-1231.
- [38] Tammela T, Zarkada G, Nurmi H, et al. VEGFR-3 controls tip to stalk conversion at vessel fusion sites by reinforcing notch signalling, Nature Cell Biol. 2011.13:1202-1213.
- [39] Stacker SA, Williams RA, Achen MG. Lymphangiogenic growth factors as markers of tumor metastasis. APMIS, 2004,112:539-
- [40] Skobe M, Hawighorst T, Jackson DG, et al. Induction of tumor lymphangiogenesis by VEGF-C promotes breast cancer metastasis. Nature Med, 2001,7:192-198.
- [41] Mandriota SJ, Jussila L, Jeltsch M, et al. Vascular endothelial growth factor-C-mediated lymphangiogenesis promotes tumour metastasis. EMBO J, 2001,20:672-682.
- [42] Karpanen T, Alitalo K. Lymphatic vessels as targets of tumor therapy. J Exp Med, 2001,194:F37-F42.
- [43] McColl BK, Baldwin ME, Roufail S, et al. Plasmin activates the lymphangiogenic growth factors VEGF-C and VEGF-D. J Exp Med 2003 198:863-868
- [44] McColl BK, Paavonen K, Karnezis T, et al. Proprotein convertases promote processing of VEGF-D, a critical step for binding the angiogenic receptor VEGFR-2. FASEB J, 2007,21: 1088-1098
- [45] Debinski W, Slagle-Webb B, Achen MG, et al. VEGF-D is an X-linked/AP-1 regulated putative onco-angiogen in human glioblastoma multiforme. Mol Med, 2001,7:598-608.
- [46] Achen MG, Williams RA, Minekus MP, et al. Localization of vascular endothelial growth factor-D in malignant melanoma suggests a role in tumour angiogenesis. J Pathol, 2001,193: 147-154
- [47] White JD, Hewett PW, Kosuge D, et al. Vascular endothelial growth factor-D expression is an independent prognostic

- marker for survival in colorectal carcinoma. Cancer Res, 2002.62:1669-1675.
- [48] Nakamura Y. Yasuoka H. Tsuiimoto M. et al. Prognostic significance of vascular endothelial growth factor-D in breast carcinoma with long-term follow-up. Clin Cancer Res, 2003,9: 716-721
- [49] Stacker SA, Caesar C, Baldwin ME, et al. VEGF-D promotes the metastatic spread of tumor cells via the lymphatics. Nature Med, 2001,7:186-191.
- [50] Kopfstein L, Veikkola T, Djonov VG, et al. Distinct roles of vascular endothelial growth factor-D in lymphangiogenesis and metastasis. Am J Pathol, 2007,170:1348-1361.
- [51] Karnezis T, Shayan R, Caesar C, et al. VEGF-D promotes tumor metastasis by regulating prostaglandins produced by the collecting lymphatic endothelium. Cancer Cell, 2012,21:181 -195
- [52] Achen MG, McColl BK, Stacker SA. Focus on lymphangiogenesis in tumor metastasis. Cancer Cell, 2005,7:121-127.
- [53] Achen MG, Stacker SA. Molecular control of lymphatic metastasis Ann NY Acad Sci 2008 1131:225-234
- [54] Harris NC, Paavonen K, Davydova N, et al. Proteolytic processing of vascular endothelial growth factor-D is essential for its capacity to promote the growth and spread of cancer. FASEB J, 2011,25:2615-2625.
- [55] Achen MG, Roufail S, Domagala T, et al. Monoclonal antibodies to vascular endothelial growth factor-D block interactions with both VEGF receptor-2 and VEGF receptor-3. Eur J Biochem, 2000,267:2505-2515.
- [56] Davydova N, Roufail S, Streltsov VA, et al. The VD1 neutralizing antibody to vascular endothelial growth factor-D: binding epitope and relationship to receptor binding. J Mol Biol, 2011,407:581-593.
- [57] Youssoufian H, Hicklin DJ, Rowinsky EK. Review: monoclonal antibodies to the vascular endothelial growth factor receptor-2 in cancer therapy. Clin Cancer Res, 2007,13:5544s-5548s.
- [58] Pytowski B, Goldman J, Persaud K, et al. Complete and specific inhibition of adult lymphatic regeneration by a novel VEGFR-3 neutralizing antibody. J Natl Cancer Inst, 2005,97: 14-21.
- [59] He Y, Kozaki K, Karpanen T, et al. Suppression of tumor lymphangiogenesis and lymph node metastasis by blocking vascular endothelial growth factor receptor 3 signaling. J Natl Cancer Inst, 2002,94:819-825.
- [60] Achen MG, Mann GB, Stacker SA. Targeting lymphangiogenesis to prevent tumour metastasis. Br J Cancer, 2006,94: 1355-1360.
- [61] Ebos JM, Lee CR, Cruz-Munoz W, et al. Accelerated metastasis after short-term treatment with a potent inhibitor of tumor angiogenesis. Cancer Cell, 2009, 15:232-239.
- [62] Paez-Ribes M, Allen E, Hudock J, et al. Antiangiogenic therapy elicits malignant progression of tumors to increased local invasion and distant metastasis. Cancer Cell, 2009, 15: 220-231.
- [63] Singh M, Couto SS, Forrest WF, et al. Anti-VEGF antibody therapy does not promote metastasis in genetically engineered mouse tumour models. J Pathol, 2012,227:417-430.
- [64] Chung AS, Kowanetz M, Wu X, et al. Differential drug classspecific metastatic effects following treatment with a panel of angiogenesis inhibitors. J Pathol, 2012,227:404-416.
- [65] Yakes FM, Chen J, Tan J, et al. Cabozantinib (xl184), a novel MET and VEGFR2 inhibitor, simultaneously suppresses metastasis, angiogenesis, and tumor growth. Mol Cancer Ther, 2011.10:2298-2308.
- [66] You WK, Sennino B, Williamson CW, et al. VEGF and c-Met

- blockade amplify angiogenesis inhibition in pancreatic islet cancer. Cancer Res, 2011,71:4758-4768.
- [67] Qin L, Bromberg-White JL, Qian CN. Opportunities and challenges in tumor angiogenesis research: back and forth between bench and bed. Adv Cancer Res, 2012,113:191-239.
- [68] Rothwell PM, Wilson M, Price JF, et al. Effect of daily aspirin
- on risk of cancer metastasis: a study of incident cancers during randomised controlled trials. Lancet, 2012,379:1591 -1601.
- [69] Rothwell PM, Fowkes FG, Belch JF, et al. Effect of daily aspirin on long-term risk of death due to cancer: analysis of individual patient data from randomised trials. Lancet, 2011,377:31-41.

Submit your next manuscript to Chinese Journal of Cancer and take full advantage of:

- Open access
- No charge to authors
- Quickly published
- Thorough peer review
- Professionally edited
- No space constraints
- · Indexed by PubMed, CA, and Google Scholar

Submit your manuscript at www.cjcsysu.com