Corrections

BIOCHEMISTRY. For the article "Redox regulation of surface protein thiols: Identification of integrin α -4 as a molecular target by using redox proteomics," by Teresa Laragione, Valentina Bonetto, Filippo Casoni, Tania Massignan, Giancarlo Bianchi, Elisabetta Gianazza, and Pietro Ghezzi, which appeared in issue 25, December 9, 2003, of *Proc. Natl. Acad. Sci. USA* (100, 14737–14741; first published December 1, 2003; 10.1073/pnas.2434516100), the authors note that the *x*-axis label for the NAC concentration in Fig. 1*B* should be micromolar. In addition, the indications for the black and white bars were switched in the legend for Fig. 6. The legend should have read: "No antibody, black bars; anti-VLA-4 (10 ng/ml), gray bars; and anti-VLA-4 (10 μ g/ml), white bars." The corrected figures and their legends appear below.

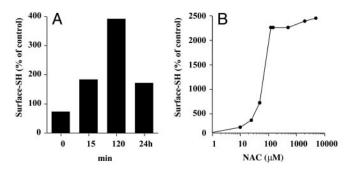


Fig. 1. Time course (*A*) and dose response (*B*) of NAC-induced increase of surface SH expression. PBMCs (10^6 cells per ml) were cultured with 5 mM NAC for the indicated time (*A*), after which surface thiols were quantified with DTNB. In the experiment shown in *B*, cells were incubated for 2 h with the indicated concentration of NAC. Data are the mean of duplicate experiments.

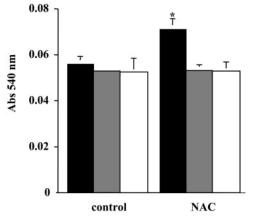


Fig. 6. Quantitation of the effect of NAC on cell adhesion. Experiments were performed exactly as in Fig. 5. Cells were solubilized and eosin Y and absorbance was quantitated at 540 nm. No antibody, black bars; anti-VLA-4 (10 ng/ml), gray bars; and anti-VLA-4 (10 μ g/ml), white bars. Data are means ± SE (n = 3). *, P < 0.01 vs. control.

www.pnas.org/cgi/doi/10.1073/pnas.0401128101

BIOCHEMISTRY. For the article "Myc-interacting protein 1 target gene profile: A link to microtubules, extracellular signal-regulated kinase, and cell growth," by Joseph Ziegelbauer, Joyce Wei, and Robert Tjian, which appeared in issue 2, January 13, 2004, of *Proc. Natl. Acad. Sci. USA* (101, 458–463; first published January 2, 2004; 10.1073/pnas.0307562100), the authors note that the supporting information was omitted from the article and that the following statement should be added in *Methods* at the end of *High-Density Oligonucleotide Microarray*: "Tables with lists of individual transcripts discussed in the microarray experiments are published as supporting information on the PNAS web site." The supporting information has been added online.

www.pnas.org/cgi/doi/10.1073/pnas.0400811101

CHEMISTRY, BIOCHEMISTRY. For the article "DNA-mediated charge transport for DNA repair," by Elizabeth M. Boon, Alison L. Livingston, Nikolas H. Chmiel, Sheila S. David, and Jacqueline K. Barton, which appeared in issue 22, October 28, 2003, of *Proc. Natl. Acad. Sci. USA* (100, 12543–12547; first published October 14, 2003; 10.1073/pnas.2035257100), the authors note that the calibrations to potentials versus NHE using Ag wire were not accurate. Measurements using an Ag/AgCl reference electrode rather than Ag wire show the midpoint potentials for DNA-bound MutY to be -110 mV or 90 mV versus NHE. Similarly, for the DNA-bound MutY mutant, C199H, the midpoint potential is -135 mV or 65 mV versus NHE. This correction does not affect the conclusions of the article.

www.pnas.org/cgi/doi/10.1073/pnas.0401134101

GENETICS. For the article "A strabismus susceptibility locus on chromosome 7p," by Vaishali Parikh, Yin Yao Shugart, Kimberly F. Doheny, Jie Zhang, Lan Li, John Williams, David Hayden, Brian Craig, Hilda Capo, Denise Chamblee, Cathy Chen, Mary Collins, Stuart Dankner, Dean Fiergang, David Guyton, David Hunter, Marcia Hutcheon, Marshall Keys, Nancy Morrison, Michelle Munoz, Marshall Parks, David Plotsky, Eugene Protzko, Michael X. Repka, Maria Sarubbi, Bruce Schnall, R. Michael Siatkowski, Elias Traboulsi, Joanne Waeltermann, and Jeremy Nathans, which appeared in issue 21, October 14, 2003, of *Proc. Natl. Acad. Sci. USA* (100, 12283–12288; first published September 30, 2003; 10.1073/pnas.2035118100), the authors note that in Fig. 14 the color coding of the list of numbers under subject 412 was incorrect. The corrected figure and its legend appear on the opposite page.

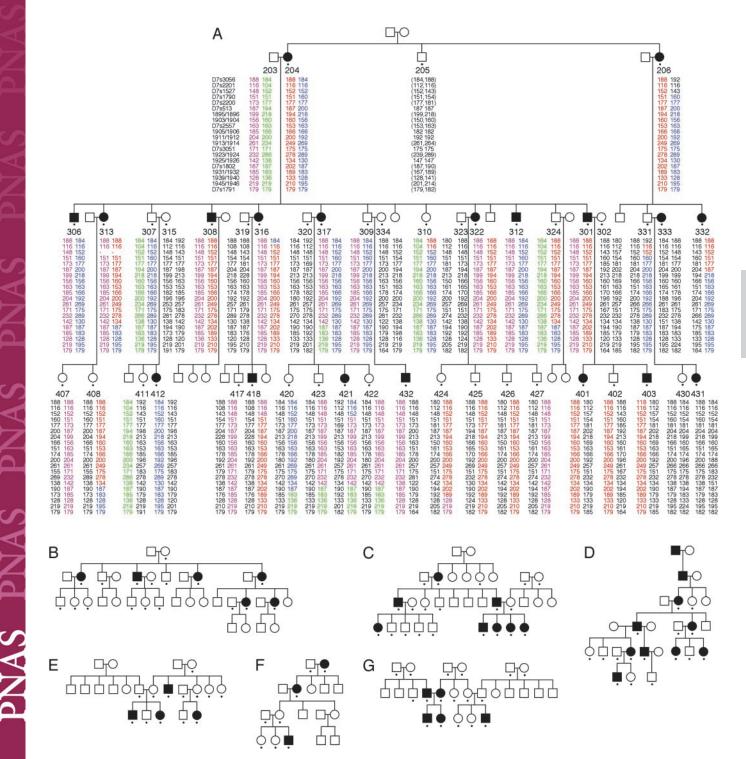


Fig. 1. Seven pedigrees analyzed by whole genome scanning. The assignment of affected individuals (filled symbols) is described in *Materials and Methods*. Dots indicate those individuals for whom genotypes were obtained. Haplotypes are shown for family A with the 20 high-resolution chromosome 7p markers (Fig. 2*B* and Table 3) arranged in order with the most telomere-proximal marker (D7s3056) at the top and the most centromere-proximal marker (D7s1791) at the bottom, as listed to the left of subject 203. For each individual, the paternal haplotype is on the left and maternal haplotype is on the right. Numbers indicate the size of the PCR products in bp. The haplotype for individual 203 was reconstructed from the haplotypes of his children; the phase of the markers for individual 205 cannot be determined with the available data. The four chromosomes in individuals 203 and 204 are color coded.

www.pnas.org/cgi/doi/10.1073/pnas.0400841101