A Susceptibility Locus for Migraine with Aura, on Chromosome 4q24

Maija Wessman,^{1,3,4} Mikko Kallela,⁵ Mari A. Kaunisto,^{3,4} Pia Marttila,² Eric Sobel,² Jaana Hartiala,² Greg Oswell,^{1,2} Suzanne M. Leal,⁹ Jeanette C. Papp,² Eija Hämäläinen,^{1,2,3} Petra Broas,² Geoffrey Joslyn,² Iiris Hovatta,⁷ Tero Hiekkalinna,² Jaakko Kaprio,^{6,8,10} Jürg Ott,⁹ Rita M. Cantor,² John-Anker Zwart,^{2,11} Matti Ilmavirta,¹² Hannele Havanka,¹³ Markus Färkkilä,⁵ Leena Peltonen,^{2,7} and Aarno Palotie^{1,2,3}

¹Department of Pathology and Laboratory Medicine and ²Department of Human Genetics, University of California, Los Angeles; Departments of ³Clinical Chemistry, ⁴Biosciences, ⁵Neurology, and ⁶Public Health, University of Helsinki, and Departments of ⁷Human Molecular Genetics and ⁸Mental Health, National Public Health Institute, Helsinki; ⁹Laboratory of Statistical Genetics, the Rockefeller University, New York; ¹⁰Department of Public Health and General Practice, University of Oulu, Oulu, Finland; ¹¹Department of Clinical Neuroscience, Faculty of Medicine, Norwegian University of Science and Technology, Trondheim, Norway; ¹²Department of Neurology, Central Hospital of Central Finland, Jyväskylä, Finland; and ¹³Department of Neurology, Länsi-Pohja Central Hospital, Kemi, Finland

Migraine is a complex neurovascular disorder with substantial evidence supporting a genetic contribution. Prior attempts to localize susceptibility loci for common forms of migraine have not produced conclusive evidence of linkage or association. To date, no genomewide screen for migraine has been published. We report results from a genomewide screen of 50 multigenerational, clinically well-defined Finnish families showing intergenerational transmission of migraine with aura (MA). The families were screened using 350 polymorphic microsatellite markers, with an average intermarker distance of 11 cM. Significant evidence of linkage was found between the MA phenotype and marker D4S1647 on 4q24. Using parametric two-point linkage analysis and assuming a dominant mode of inheritance, we found for this marker a maximum LOD score of 4.20 under locus homogeneity (P = .000006) or locus heterogeneity (P = .000011). Multipoint parametric (HLOD = 4.45; P = .0000058) and nonparametric (NPL_{all} = 3.43; P = .0007) analyses support linkage in this region. Statistically significant linkage was not observed in any other chromosomal region.

Introduction

Migraine (MIM 157300) is a highly prevalent primary headache disorder, which affects ~10%-12% of the white population (Henry et al. 1992; Rasmussen and Olesen 1992; Stewart et al. 1992; O'Brien et al. 1994; Hagen et al. 2000). It is more prevalent among women, and, for both sexes, its prevalence peaks during middle age and declines thereafter (Lipton et al. 1994, 2001). The most common form of migraine is migraine without aura (MO), which, according to the International Headache Society's (IHS) standardized guidelines, is characterized by unilateral pulsating pain of moderate to severe intensity, aggravated by physical activity and lasting 4 -72 h. The attacks are associated with nausea, vomiting, photophobia, and phonophobia. Migraine with aura (MA) shares the same headache qualities, but the headache is usually preceded by aura—attacks of focal neurological symptoms, usually visual, that develop grad-

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Address for correspondence and reprints: Dr. Aarno Palotie, Department of Pathology, UCLA, School of Medicine 695 Charles E. Young Drive South, Box 95-7088 Los Angeles, CA 90095-7088. Email: apalotie@mednet.ucla.edu

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ually within 5–20 min and that persist for <60 min (Headache Classification Committee of the International Headache Society 1988). Approximately one-third of individuals with migraine experience both types of migraine during their lifetimes.

Twin studies indicate that migraine has a significant genetic component, with heritability estimates of 40%–65% (Honkasalo et al. 1995; Larsson et al. 1995; Gervil et al. 1999b; Ulrich et al. 1999b). Approximately 50% of migraine sufferers have an affected first-degree relative (Bille 1997). However, the mode of transmission of the disorder is not clear (Mochi et al. 1993; Russell and Olesen 1993; Russell et al. 1995; Ulrich et al. 1999a). Several population-based family studies have suggested that both genetic and environmental factors are involved in migraine but that genetic factors are more influential in MA than in MO (Russell and Olesen 1995; Ziegler et al. 1998; Gervil et al. 1999a; Ulrich et al. 1999a, 1999b).

None of the numerous studies performed to date have led to identification of a gene responsible for the more common forms of migraine—that is, MO or MA—or even to specification of their mode of transmission. Only familial hemiplegic migraine (FHM1 [MIM 141500]), a rare autosomal dominant subtype of migraine, has had a gene identified for it, on 19p13 (Joutel et al. 1993; Ophoff et al. 1996). Mutations in this gene, *CACNA1A*

Table 1
Families with Migraine Who Were Used for Genomewide Screen

Category	No. (Range)			
Pedigree size:				
One generation	5			
Two generations	19			
Three generations	24			
Four generations	2			
Total	50			
Subjects:				
Total	646			
Total genotyped	430			
Per-pedigree mean:				
Total	13.0 (3-34)			
Genotyped	8.6 (3–20)			
Affected (with either MA or MO)	5.9 (3–17)			

(MIM 601011), a brain-specific P/Q-type calcium-channel gene, account for only a small fraction of all patients with migraine and for 50% of families with the FHM subtype (Ducros et al. 2001). Interestingly, mutations in this same gene also can cause either episodic ataxia type 2 (EA-2 [MIM 108500]) or spinocerebellar ataxia type 6 (SCA6 [MIM 183086]) (Ophoff et al. 1996; Jodice et al. 1997); on the other hand, 20% of families with FHM show linkage to 1q21-31 (FHM2 [MIM 602481), providing evidence that FHM is heterogeneous (Ducros et al. 1997; Gardner et al. 1997). Several studies have suggested that the 19p13 CACNA1A locus may be involved in nonhemiplegic migraine as well (May et al. 1995; Ophoff et al. 1997; Nyholt et al. 1998*b*; Terwindt et al. 2001), although contradictory data also have been reported (Hovatta et al. 1994; White et al. 2001).

Thus far, 1q has not been linked to either MA or MO; on the other hand, Xq has been implicated in typical familial migraine (MIM 300125) (Nyholt et al. 1998a, 2000; Oterino et al. 2001). In addition, a number of association studies have shown linkage between various gene loci and either MA or MO (Pardo et al. 1995; Peroutka et al. 1997; Del Zompo et al. 1998; Ogilvie et al. 1998; Kowa et al. 2000; Lea et al. 2000; Tzourio et al. 2001). Unfortunately, most of these studies have remained single reports and await confirmatory replication studies. No genomewide screens for susceptibility loci for migraine have been published thus far.

Identification of genes and allelic variants predisposing to common traits such as migraine has been complicated by inconsistencies in clinical diagnosis, a variable age at onset, an unknown mode of inheritance, locus heterogeneity, and the poorly understood role of environmental factors. However, the small number of founders, along with environmental and cultural homogeneity, can potentially make the Finnish population useful in genetic studies of complex diseases (de la Chapelle 1993; Lander and Schork 1994; Peltonen et al.

2000). This population, with well-established genealogical registries and well-kept population records, has been used successfully in past genetic studies, and most identified loci have been replicated also in mixed populations, suggesting their general significance (Mahtani et al. 1996; Kuokkanen et al. 1997; Hovatta et al. 1999; Leppävuori et al. 1999; Pajukanta et al. 1999; Ekelund et al. 2000; for a review, see Peltonen et al. 2000; Laitinen et al. 2001). We report here the results of a genomewide screen of 50 Finnish families with MA, implicating a susceptibility locus on 4q24.

Subjects and Methods

Study Design

The present study focuses on families ascertained for MA. This restriction was imposed because data suggest that the genetic component of MA may be stronger than that of MO (Honkasalo et al. 1995; Russell et al. 1995; Ulrich et al. 1999a). Also, by mandating that an aura be one of the symptoms, we diminish the risk that other types of headache will be included. Several studies also have indicated that MA and MO might be distinct disorders (Russell et al. 1996, 2001). Genetic analysis was performed only on individuals who gave their informed consent to the study. The ethics committees of the Helsinki University Central Hospital and the University of California, Los Angeles, approved the study protocol.

Consecutively identified families with at least three first-degree family members affected with MA were considered for the study. The families were recruited on the basis of patients attending headache clinics in three Finnish cities (Helsinki, Kemi, and Jyväskylä). Three neurologists (M.K., H.H., and M.I.) were in charge of the recruitment. Once a member of the family, the index case, had been clinically diagnosed, by a neurologist, as suffering from migraine, the index case was asked to contact all other members of the family who were believed to suffer from migraine and to ask if they would be willing to participate in the study. If at least three possible migraine sufferers were willing to participate, the validated Finnish Migraine Specific Questionnaire for Family Studies (FMSQ_{ES}) (Kallela et al. 2001) was mailed to each of them, to their parents, and to their siblings. The FMSQ_{FS} has been shown to be both specific and sensitive in both diagnosing MA and MO in a family setting (Kallela et al. 2001) and differentiating MA from MO. From among the first 300 participating families, 50 independent, multigenerational families including a total 646 family members were selected for genotyping. These families were chosen because they displayed a seemingly autosomal dominant mode of inheritance, with the MA phenotype passed from one generation to the next. In rare cases, an unaffected individual (i.e., a subject without MA) had affected offspring; this suggests

Table 2
Diagnosis of 430 Genotyped Subjects with Migraine

Diagnosis (IHS Classification)	No. of Subject		
MA (1.2 ^a)	252		
MO (1.1)	53		
HA	10		
NoHA	91		
DU	24		

^a Includes EQV (IHS classification 1.2.5), in 6 subjects.

that there is reduced penetrance for the disease gene or that phenocopies may exist. In several instances, there appeared to have been assorted mating within the pedigrees—that is, an offspring was the result of mating between an individual with the common MA phenotype and an individual who either (a) displayed either the MA or MO phenotype or (b) came from a family with a history of either MA or MO. In these instances, the spouse and his or her offspring were excluded from the genetic analysis. In addition to completing the FMSQ_{FS}, the participants were asked to provide a sample of their blood, for genotyping; blood samples were received from 430 individuals.

Diagnoses

Clinical data were available on 646 individuals (368 women and 278 men), including 246 individuals (174 women and 72 men) with MA and 53 individuals (36 women and 17 men) with MO. Table 1 describes the family sample used in our genomewide screen. Diagnoses of all index cases and family members were made according to the IHS criteria (Headache Classification Committee of the International Headache Society 1988), on the basis of the FMSQ_{FS} (Kallela et al. 2001). Table 2 describes the diagnosis of the 430 individuals from whom a blood sample was available for genotyping. Table 3 describes the clinical characteristics of headache, and table 4 describes the aura symptoms in the 246 subjects with MA. Families with FHM (IHS classification 1.2.3) were not included in our study.

Genotyping

Linkage analysis was conducted for the 22 autosomes and the X chromosome. In all, 350 polymorphic microsatellite markers were genotyped in the 430 subjects from the 50 families that provided blood. The markers were from The Human MapPairs Genome-Wide Screening Set (LI-COR). This compilation is the ninth version of the Weber-lab screening set (Broman et al. 1998), with a few modifications. The order and distances between the markers were determined on the basis of the genome database

of the Center for Medical Genetics, Marshfield Medical Research Foundation. The markers are approximately evenly spaced throughout the genome, with an average of 11 cM between loci. The markers that failed to genotype well were replaced by microsatellite markers from The Genome Database and from the Marshfield genome database. Genomic DNA was extracted from peripheral blood samples, by standard techniques. DNA was amplified by multiplex PCR assays with fluorescent primers designed to detect the microsatellite loci. Pipetting of the reactions was performed by a Hydra Microdispenser (Robbins Scientific). The amplification reactions were run in microtiter 96-well plates, by Tetrad thermal cyclers (MJ Research). The resulting PCR fragments, along with a size-standard ladder and fragments from known control individuals from Fondation Jean Dausset CEPH, were separated on 6% acrylamide gels by electrophoresis using a LI-COR DNA 4200 Genetic Analyzer (LI-COR). The bands on the gels were automatically interpreted into genotypes by the Saga1.0 software package (University of Washington and LI-COR). Saga takes digital gel images, tracks the lanes, and calls the alleles on the basis of a combination of size, peak shape, and marker characteristics (e.g., di-, tri-, or tetranucleotide repeat). Allele sizes were standardized to those of CEPH control individuals. All genotypes were verified by human inspection. The PedCheck1.1 (O'Connell and Weeks 1998) and Sim-Walk2 2.82 (Sobel and Lange 1996) computer programs were used to detect genotyping errors. If the mistyping was not resolved by review of the gel images, the suspected genotypes were set to be unknown.

Linkage Analyses

All analyses were performed with the disease-gene frequency set at 0.001, under the assumption of autosomal dominant inheritance and a phenocopy rate of 2.4%

 Table 3

 Clinical Characteristics of Headache

Symptom	Proportion of Subjects (%)
Unilateral headache	76
Pulsating headache	75
Headache intensity:	
Unbearable	39
Severe	42
Moderate	17
Mild	2
Aggravated by physical activity	76
Associated symptoms:	
Nausea	85
Vomiting	57
Photophobia	87
Phonophobia	79

Table 4
Aura Symptoms in 246 Subjects with MA

Type of Aura	Proportion of Subjects (%)
Any visual aura	94
Hemianopia	40
Scintillating scotoma	56
Photopsia	49
Blurring of vision	43
Any sensory aura	36
Any motor aura	14
Speech disturbance	29

(Hovatta et al. 1994). Allele frequencies were calculated from the genotypes of all individuals genotyped in the analysis. Initially, subjects with MA (IHS classification 1.2), with the exception of the subjects with migraine aura without headache (EQV [IHS classification 1.2.5]), were classified as affected. All other subjects, with MO (IHS classification 1.1), headache other than migraine (HA), no headache (NoHA), or diagnosis unavailable (DU), were classified as unknown. Their genotypes are useful in the reconstruction of missing parental genotypes and phase information. The initial linkage analysis was performed by means of a two-point approach—that is, by using a single marker and the trait. Additionally, two-point linkage analyses were also performed with a broader phenotype, in which both individuals with MA and individuals with MO were considered to be affected. Two-point analysis is known to be less sensitive to map and genotyping errors than are multipoint methods (Risch and Giuffra 1992). Two-point parametric linkage analysis was performed both under locus homogeneity and under locus heterogeneity, by the LINKAGE (Lathrop and Lalouel 1984) and HOMOG (Ott 1983) computer programs. To investigate the possibility that a putative migraine gene may act in a recessive fashion, we employed an affected-sib-pair (ASP) analysis. The identity-by-descent (IBD) status of each ASP was estimated by the SIBPAIR program. Within the ASP analysis, we anticipate that some power is lost because pedigrees have been split into nuclear families, which have been analyzed as though they had been ascertained independently; however, this is not expected to bias the linkage results in any manner (Göring and Terwilliger 2000b). The program ANALYZE (Göring and Terwilliger 2000a) was used to conduct these analyses.

For regions showing evidence of linkage in the parametric two-point analysis, multipoint parametric and nonparametric analyses were performed by the GENE-HUNTER program, version 2.1_r2beta (Kruglyak et al. 1996). Parametric linkage analysis was performed using the model presented above, while allowing for locus het-

erogeneity. The nonparametric statistic, NPL $_{\rm all}$, which estimates the statistical significance of alleles shared IBD between all affected family members, was calculated also, together with an estimated P value.

P values for LOD scores detected under homogeneity were estimated according to the method proposed by Nyholt (2000); P values for LOD scores detected under heterogeneity were estimated according to the method proposed by Chiano and Yates (1995). To aid in the interpretation of linkage results, the thresholds were set according to the recommendation made by Lander and Kruglyak (1995). For LOD scores detected under homogeneity, the thresholds are as follows: a pointwise threshold of P = .000049 (which is equivalent to a LOD score of 3.3), for significant linkage; a pointwise threshold of P = .0017 (LOD score 1.9), for suggestive linkage, and a pointwise threshold of P = .05 (LOD score 0.59), for nominal linkage.

Results

Parametric and Nonparametric Two-Point Linkage Analyses

Parametric and nonparametric LOD scores were calculated using an affecteds-only strategy, for 350 markers. Initially, family members with MA (IHS classification 1.2.1) were classified as affected, and all others were classified as unknown (see the "Subjects and Methods" section); all results based on this phenotype classification are presented first. For parametric analyses, a dominant mode of inheritance and locus heterogeneity were employed. Plots of the two-point parametric LOD scores for each chromosome in the genomewide screen are shown in figure 1. In the genome screen, marker D4S1647 gave the high-

est LOD score, 4.20 (under homogeneity, P = .000006; under heterogeneity, P = .000011), at a recombination fraction (θ) of 0.18. This marker also gave an ASP LOD score of 2.25 (P = .0064) (table 5). Marker D4S2380, ~4 cM proximal to marker D4S1647, had a two-point maximum LOD score of 2.26 (under heterogeneity; $\theta = 0.08$ and P = .0011), and an ASP LOD score of 1.30 (P = .0073). Marker D4S3240, ~9 cM distal to marker D4S1647, had a two-point LOD score of 1.01 (under heterogeneity; $\theta = 0.16$ and P = .023) and an ASP LOD score of 0.63 (P = .041). Five other markers adjacent to marker D4S1647, spanning a region of 59 cM (D4S1517–D4S1520), showed two-point maximum LOD scores from 1.01 (P = .023) to 1.89 (P = .0026). Detailed results are presented in table 5.

Statistically significant or suggestive linkage was not observed in any other chromosomal region. However, nominal linkage with parametric two-point LOD scores giving P values $\leq .05$ were found at 21 additional loci,

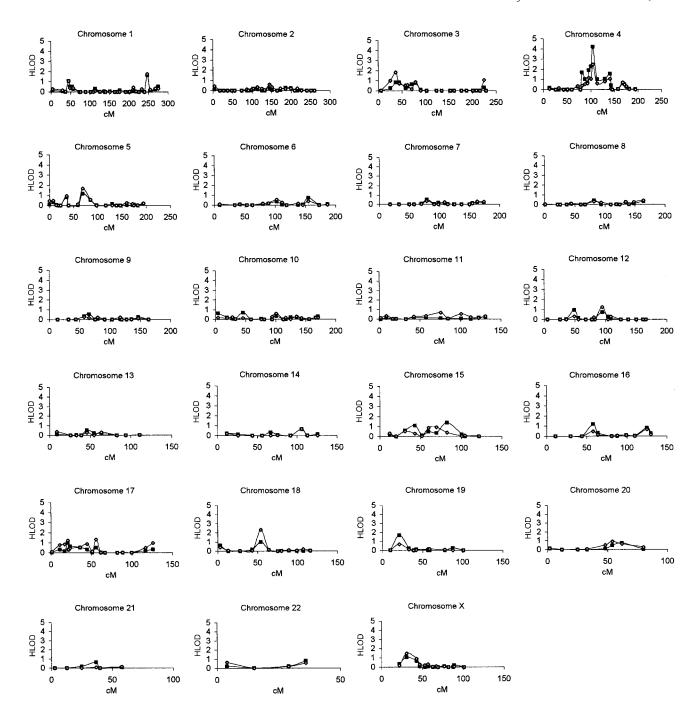


Figure 1 Two-point parametric linkage analyses (dominant model allowing for heterogeneity) for the 350-marker genomewide screen of 50 families with migraine. Blackened squares denote the two-point LOD scores when only individuals suffering from MA were considered to be affected; unblackened diamonds denote the two-point LOD scores for a broader phenotype, in which both individuals with MA and individuals with MO were considered to be affected. Analyses were performed by the LINKAGE program, as described in the text.

in 1p, 1q, 3p, 5p, 5q, 6q, 12p, 12q, 15q, 16p, 16q, 17p, 18q, 19p, 22q, and Xp. These results are presented in detail in table 5. Of these regions, two—1q and 19p—are of specific interest because they previously have been linked to either FHM (Joutel et al. 1993; Ducros et al. 1997; Gardner et al. 1997) or MA and MO (May et al.

1995; Ophoff et al. 1997; Nyholt et al. 1998b; Terwindt et al. 2001). Therefore, results regarding these regions are presented here in more detail. Also, results regarding the X chromosome will be presented in more detail because several previous and recent studies have provided evidence of linkage of the X chromosome to common

Table 5

Locations and Two-Point LOD Scores for Markers with P < .05 in the Genomewide Screen

Marker (Location)		Results under Locus Homogeneity		Results under Locus Heterogeneity			RESULTS OF ASP ANALYSIS		
	Position ^a (cM)	LOD Score	P	$ heta^{ m b}$	HLOD Score	P	$ heta^{ ext{b}}$	LOD Score	P
D1S552 (1p36.13)	45	(.36)	(.099)	.28	1.08	.020	.00	(.00)	(.50)
D1S3462 (1q42.2)	247	.70	.036	.30	1.66	.0046	.34	(.00)	(.50)
D3S1259 (3p25.2)	37	.80	.026	.30	.82	.038	.26	.71	.036
D3S3038 (3p24.3)	45	.82	.026	.30	.82	.038	.30	(.52)	(.062)
D3S2409 (3p21.31)	71	.64	.043	.36	(.64)	(.061)	.36	(.13)	(.22)
D3S1766 (3p14.2)	79	.77	.030	.30	.77	.045	.00	(.12)	(.23)
D4S1517 (4q13.3)	82	1.66	.0029	.26	1.66	.0046	.26	1.02	.015
D4S3243 (4q)	88	1.01	.016	.26	1.01	.023	.26	(.17)	(.19)
D4S2361 (4q21.23)	93	.61	.047	.30	(.68)	(.055)	.22	(.12)	(.23)
D4S2409 (4q22.1)	96	1.85	.0018	.20	1.89	.0026	.16	1.00	.016
D4S2380	101	1.75	.0023	.22	2.26	.0011	.08	1.30	.0073
D4S1647 (4q24)	105	4.20	.000006	.18	4.20	.000011	.18	2.25	.0064
D4S3240 (4q25)	114	.76	.031	.30	1.01	.023	.16	.63	.041
D4S2394 (4q28.2)	130	.98	.017	.28	1.05	.021	.22	(.02)	(.38)
D4S1520 (4q31.1)	141	1.55	.0038	.20	1.55	.0061	.20	(.47)	(.071)
D5S2845 (5p14.3)	36	.82	.026	.24	.81	.039	.24	(.36)	(.10)
D5S2500 (5q12.1)	69	1.11	.012	.26	1.16	.016	.20	(.36)	(.099)
D6S2436 (6q25.2)	155	.75	.032	.34	.75	.046	.34	.61	.047
D12S1042 (12p11.23)	49	.94	.019	.32	.94	.028	.32	(.35)	(.10)
D12S1064 (12q21.33)	95	(.42)	(.082)	.36	.74	.047	.00	(.00)	(.50)
D15S659 (15q21.1)	43	(.05)	(.32)	.42	1.09	.019	.06	(.00)	(.50)
D15S655 (15q25.3)	83	.70	.036	.24	1.38	.0092	.04	(.31)	(.12)
D16S753 (16p12.3)	58	(.004)	(.45)	.40	1.22	.014	.00	(.07)	(.29)
D16S539 (16q24.1)	125	.84	.025	.34	.84	.036	.34	(.00)	(.5)
D17S945 (17p13.1)	21	.96	.018	.30	1.01	.023	.24	.61	.047
D18S877 (18q12.1)	54	(.18)	(.18)	.40	1.00	.024	.00	(.08)	(.27)
D19S427 (19p13.2)	21	1.70	.0026	.22	1.70	.0042	.22	(.08)	(.27)
D22S683 (22q12.3)	36	(.41)	(.085)	.34	.82	.038	.08	(.44)	(.077)
DXS9896 (Xp)	31	1.08	.013	.30	1.08	.020	.30	(.13)	(.22)
DXS6810 (Xp11.4)	43	.65	.042	.34	(.65)	(.060)	.34	(.31)	(.12)

NOTE.—Results are based on analyses of individuals with MA; parentheses denote entries for which P > .05; the significant P value (and the marker for it) is underlined, and the suggestive P value (and the marker for it) is in boldface italic.

forms of migraine (Nyholt et al. 1998a, 2000; Oterino et al. 2001; Wieser et al. 2001).

Ten markers were genotyped on chromosome 19, from 10 to 101 cM (where pter is at 0 cM and qter is at 105 cM). Marker D19S1150, at a distance of ~38 cM, is an intragenic marker of CACNA1A and was included in the screen. This marker showed no evidence of linkage in our families (HLOD 0.00). However, marker D19S427, located ~20 cM distal to marker D19S1150, showed nominal evidence of linkage, with a maximum LOD score of 1.70 ($\theta = 0.22$; under homogeneity, P = .0026; under heterogeneity, P = .0042).

Thirty-one markers were genotyped on chromosome 1, from 4 to 275 cM (where pter is at 0 cM and qter is at 290 cM). Marker D1S3462, on 1q42 (at 247 cM),

and marker D1S522, on 1p36 (at 45 cM), showed nominal evidence of linkage, with HLOD values of 1.66 ($\theta = 0.34$; P = .0046) and 1.08 ($\theta = 0.00$; P = .02), respectively.

Fourteen markers were genotyped on the X chromosome from 22 to 101 cM (where pter is at 0 cM and qter is at 103 cM); of these, marker DXS9896, at 31 cM, showed a two-point maximum LOD score of 1.08 ($\theta = 0.30$; under homogeneity, P = .013; under heterogeneity, P = .02), with nominal evidence of linkage. No evidence of linkage was observed on Xq.

In addition, markers D14S1426 (at 126 cM), D17S1830 (at 117 cM), and D18S535 (at 64 cM) gave LOD scores of 1.13 (P=.011), 1.44 (P=.005), and 1.41 (P=.005), respectively, in the nonparametric ASP analysis.

^a Distance from pter, according to the map published by the Center for Medical Genetics, Marshfield Medical Research Foundation.

^b Values are those at which the maximum LOD score was found.

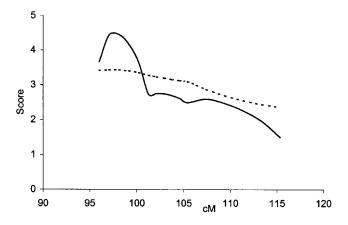


Figure 2 Multipoint parametric and nonparametric linkage analyses of 4q, with markers D4S2409 (96 cM), D4S2380 (101 cM), D4S1647 (105 cM), and D4S3240 (114 cM). Analyses were performed by the GENEHUNTER program, as described in the text. The vertical axis presents both the parametric (i.e., HLOD [*unbroken line*]) and nonparametric (i.e., NPL_{all} [*dotted line*]) values along a 20-cM region of chromosome 4.

Parametric Three-Point Linkage Analysis

Parametric three-point linkage analysis—that is, analysis using two markers and the trait, on all pairs of adjacent markers—was performed in the region from marker D4S2409 (at 96 cM) to marker D4S3240 (at 114 cM). The maximum three-point HLOD, 3.68 (P = .0004), was observed with the D4S2380-D4S1647 pair, suggesting that the best evidence of linkage is obtained in this 4-cM region.

Multipoint Analyses

To pool information from multiple markers and to position the linked locus on 4q24, we performed parametric and nonparametric multipoint analyses, using the GENEHUNTER program. The parametric test gave the highest HLOD, 4.45 (P = .0000058), between markers D4S2409 and D4S2380 (fig. 2). The estimated fraction of linked pedigrees, α , at HLOD was 0.23–0.5. The results of the nonparametric analysis are consistent with those of the parametric result, giving a maximum NPL_{all} of 3.43 (P = .0007) at the same position (fig. 2).

Parametric Two-Point Linkage Analysis for a Broader Phenotype

Two-point linkage analyses were also performed with use of a broader phenotype, in which both individuals with MA and individuals with MO were considered to be affected. A plot of the two-point parametric LOD scores (under heterogeneity) for each chromosome is shown in figure 1. No significant linkage to any chromosomal region was detected. Marker D4S1647 showed

only suggestive evidence of linkage, with a LOD score of 2.48 ($\theta = 0.22$), both under homogeneity (P = .00037) and under heterogeneity (P = .00064). Marker D18S877, at 31 cM, also showed suggestive evidence of linkage, with a two-point maximum LOD score of 2.32 ($\theta = 0.00$; under heterogeneity, P = .00093).

Discussion

Our results, based on the genotyping of 50 Finnish families with MA, provide strong evidence of a novel migraine-susceptibility locus on 4q24. Statistically significant evidence of linkage, both under locus homogeneity and under locus heterogeneity (LOD score 4.20; P = .000006 and P = .000011, respectively), between the common MA phenotype and marker D4S1647 was obtained with a parametric two-point linkage analysis, under a dominant mode of inheritance. Parametric and nonparametric multipoint analyses supported linkage to this chromosomal region. The multipoint analyses found the highest HLOD scores, 4.45 (P = .0000058), and NPL_{all} = 3.43 (P = .0007), in the region between markers D4S2409 and D4S2380, 7 cM proximal of the D4S1647 locus.

Our successful identification of linkage to 4q24 was most probably due to the selection strategy used in our study sample, which emphasized clustering in families with a specific phenotype, MA. Additionally, to reduce heterogeneity, a single neurologist carefully analyzed all the clinical data. Only families with MA throughout the pedigree were considered for further analysis. On the basis of these carefully diagnosed families with predominant MA, we were able to select reasonably large, multigenerational families for our linkage study. The importance of careful family selection in linkage studies of complex traits has recently, elegantly been demonstrated by Brzustowicz et al. (2000), who were able to locate, on 1q21-q22, a major susceptibility locus for familial schizophrenia in 22 extended families with high rates of schizophrenia. In genetically homogeneous populations, such as the Finns, in which there are potentially only a few affected founders, linkage analysis of complex traits can be especially powerful, provided that the clinical phenotype is properly dissected. Finland is not the only country in which there is a genetically isolated population appropriate for genetic studies, but the additional features of well-kept population records and a high-quality, state-run health-care system make it a very reliable resource for the genetic studies of complex traits (Peltonen et al. 2000).

The strength of analysis of a more stringently defined migraine phenotype was reflected in the drop in the twopoint LOD scores for 4q when a broader disease classification was included, in which both individuals with MA and individuals with MO were considered to be affected. Additionally, no other statistically significant locus was detected for this combined phenotype. It is likely that the increased heterogeneity and possible phenocopies introduced into the etiology by inclusion of the MO phenotypes are responsible for these differences. However, the current sample is not large enough to allow us to investigate whether MA and MO are genetically distinct entities.

To the best of our knowledge, no other report of a statistically significant linkage of common MA phenotypes to any autosomal region has been published. The present study's finding therefore suggests the existence of a novel MA locus and must be tested in independent studies. However, results of two other candidate-gene studies have indicated that two possible loci on chromosome 4 are associated with migraine. An association study by Pardo et al. (1995), of 112 unrelated patients with migraine (62 patients with MO and 50 patients with MA), provided evidence that the group-component locus (GC [MIM 139200]) on 4q12-13.3 is involved in migraine. GC codes for a multifunctional protein found in plasma, for example, where it carries the vitamin-D sterols and prevents polymerization of actin by binding its monomers. Tzourio et al. (2001) found an association between migraine and a polymorphism at the endothelin type A-receptor gene (ENDRA [MIM 131243]) on 4q31. Endothelin receptors mediate the biological effects of endothelin (ET-1 [MIM 131240]), a potential vasoconstrictor, which has been implicated in migraine, by recent studies showing increased plasma ET-1 levels during and between migraine attacks (Färkkilä et al. 1992; Kallela et al. 1998; Hasselblatt et al. 1999). GC and ENDRA are good candidate genes for our further studies, although they are located some distance from marker D4S1647 (GC is 30 Mb proximal, and ENDRA is 50 Mb distal). The ~60-cM region around our most informative marker, D4S1647, contains ≥170 known genes, including several obvious candidate genes (UCSC Human Genome Project Working Draft).

In addition to the significant finding with regard to 4q, the present study has revealed 21 markers, on 12 different chromosomes, that provide nominal evidence of linkage. However, in light of the LOD scores for them, the risk of false-positive signals is considerable. Nonetheless, of special interest are regions on chromosomes 1q, 19p, and X, where several earlier studies found evidence of linkage or association with migraine. The *CACNA1A* locus on 19p13, which Joutel et al. (1993) reported as being linked to FHM, has also been indicated as being involved in nonhemiplegic migraine. May et al. (1995) studied how *CACNA1A* markers are involved with migraine in 28 German families, and Terwindt et al. (2001) do so later in 36 extended Dutch families. Both groups concluded that the increased allele sharing around the *CACNA1A*

gene on 19p13 is consistent with an important involvement of this region in migraine, especially MA. Nyholt et al. (1998b) reported significant cosegregation and allele sharing for markers situated within or adjacent to the CACNA1A locus in one of the four large Australian families that they analyzed. On the other hand, our twopoint linkage analysis of Finnish families showed no evidence of linkage of the MA phenotype to the intragenic marker D19S1150 of CACNA1A. This finding is in agreement with our previous results excluding this region as a site for a migraine locus in Finnish families with migraine (Hovatta et al. 1994); however, two of the four families in that previous study (Hovatta et al. 1994) also were included in the present study. It should be stressed that our data are from families collected from a genetically homogeneous population and that, when other ethnic groups are analyzed, locus heterogeneity is likely to exist; however, we have found nominal linkage to marker D19S427 (LOD score 1.70; P = .0042), located in the proximity of the insulin-receptor gene (INSR [MIM 147670]) that White et al. (2001) recently have associated with migraine. Both the findings by White et al. (2001) and those of our group support the notion that genes other than CACNA1A might contribute to the linkage and association findings on 19p13. These regions obviously require further study with other samples and, possibly, with further phenotypic stratification.

We have found nominal evidence of linkage at marker D1S3462 (LOD score 1.70; P = .0046) on 1q42. There is no previous linkage evidence that would indicate that this region is involved in migraine. One locus—or, possibly, two loci—for FHM have been mapped to 1q21-31 (Ducros et al. 1997; Gardner et al. 1997). Interestingly, the 1q21-42 region harbors several ion-channel genes that are possible candidates for both FHM and MA.

Our finding of nominal linkage to the X chromosome, at marker DXS9896 (LOD score 1.08; P = .02) is noteworthy, since several other studies have provided evidence of linkage of the X chromosome to migraine. Studies by Nyholt et al. (1998a, 2000) have implicated a locus on Xq24-28 in two Australian families with migraine. Interestingly, Oterino et al. (2001) have suggested the connexin 32 gene (GJB1 [MIM 304040]) on Xq13 as a possible candidate for MA in a family with both Chargot-Marie-Tooth disease (MIM 302800) and MA. Furthermore, Wieser et al. (2001) have reported 80 families (from Spain, Germany, and the United States) with migraine and possible X-linked dominant inheritance, providing evidence of allele sharing and linkage to the X chromosome. Further studies are necessary to confirm the role of the X-chromosomal loci in genetic predisposition to migraine.

In conclusion, the present study provides strong evidence of an MA-susceptibility locus on 4q24. Further studies are needed to delineate the role of the other

potential loci involved in MA in the families analyzed in the present study. However, the statistically significant evidence of the linkage to 4q24 should facilitate efforts to identify an underlying migraine-susceptibility gene. The detection of underlying mutation(s) will provide clues to the further elucidation of the complex molecular pathways of migraine and, finally, will help in the development of rational treatment strategies.

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Electronic Data-Base Information

Accession numbers and URLs for data in this article are as follows:

- Center for Medical Genetics, Marshfield Medical Research Foundation, http://research.marshfieldclinic.org/genetics/ (for genetic-linkage distances)
- Fondation Jean Dausset CEPH, http://www.cephb.fr (for heterozygosity and allele sizes)
- Genome Database, The, http://www.gdb.org/ (for primers used for genotyping, heterozygosity, and allele sizes)
- Online Mendelian Inheritance in Man (OMIM), http://www .ncbi.nlm.nih.gov/Omim/ (for migraine [MIM 157300], FHM1 [MIM 141500], CACNA1A [MIM 601011], EA-2 [MIM 108500], SCA6 [MIM 183086], FHM2 [MIM 602481], typical familial migraine [MIM 300125], GC [MIM 139200], ENDRA [MIM 131243], ET-1 [MIM 131240], INSR [MIM 147670], GJB1 [MIM 304040], and Chargot-Marie-Tooth disease [MIM 302800])
- UCSC Human Genome Project Working Draft, http://genome .ucsc.edu/ (for gene information)

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