

NIH Public Access

Author Manuscript

J Med Chem. Author manuscript; available in PMC 2013 December 04.

Published in final edited form as:

J Med Chem. 2013 September 12; 56(17): . doi:10.1021/jm400720h.

Substituted Tetrahydroisoquinolines as Selective Antagonists for the Orexin 1 Receptor

David A. Perrey†, **Nadezhda A. German**†, **Brian P. Gilmour**†, **Jun-Xu Li**‡, **Danni L. Harris**†, **Brian F. Thomas**†, and **Yanan Zhang**†,*

† Research Triangle Institute, Research Triangle Park, North Carolina 27709

‡ Department of Pharmacology and Toxicology, University at Buffalo, the State University of New York, Buffalo, NY 14214

Abstract

Increasing evidence implicates the orexin 1 (OX₁) receptor in reward processes, suggesting OX₁ antagonism could be therapeutic in drug addiction. In a program to develop an $OX₁$ selective antagonist, we designed and synthesized a series of substituted tetrahydroisoquinolines and determined their potency in OX_1 and OX_2 calcium mobilization assays. Structure-activity relationship (SAR) studies revealed limited steric tolerance and preference for electron deficiency at the 7-position. Pyridylmethyl groups were shown to be optimal for activity at the acetamide position. Computational studies resulted in a pharmacophore model and confirmed the SAR results. Compound **72** significantly attenuated the development of place preference for cocaine in rats.

Keywords

Orexin; antagonist; selective; tetrahydroisoquinoline

Introduction

Orexin A and B, also known as hypocretin 1 and 2, are hypothalamic neuropeptides independently discovered by two groups in 1998 .^{1, 2} They are the endogenous ligands for two G protein-coupled receptors (GPCRs), orexin 1 (OX₁) and orexin 2 (OX₂).^{3, 4} Orexinexpressing neurons are located predominantly in a small area of the hypothalamus.^{2, 5-7} However, the nerve fibers of orexin neurons project throughout the central nervous system (CNS).1, 7-9 Interestingly, orexin receptors have different patterns of expression: Some brain regions express predominantly OX₁, others predominantly OX₂,^{2, 10, 11} suggesting that these receptors might modulate many unrelated functions as also suggested by antagonist and knock-out mouse studies.^{3, 4, 12} The orexin system has been shown to play a role in a variety of important biological processes, including sleep/wake cycle, $^{13, 14}$ feeding, 2 and energy homeostasis.²

Author Contributions

^{*}**Corresponding Author** Yanan Zhang, Ph.D. yzhang@rti.org; Tel: 919-541-1235; Fax: 919-541-6499.

The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript.

Supporting Information. HPLC analysis of target compounds. This material is available free of charge via the Internet at [http://](http://pubs.acs.org) [pubs.acs.org.](http://pubs.acs.org)

Recently, the orexin system, particularly the $OX₁$ receptor, was implicated in drug reward, reinstatement of drug seeking and psychomotor sensitization.15-18 This is consistent with the findings that orexin neurons have a prominent input to basal ganglia and forebrain structures (central amygdala, ventral bed nucleus of the stria terminalis, nucleus accumbens shell, ventral pallidum and ventral tegmental area) that underlie motivation, reward, stress and addiction-related behaviors.19 Orexinergic signaling seems to be involved in the rewarding effects of natural rewards and some drugs. Mice lacking orexins not only have reduced appetite but also show much less addiction-like sequelae associated with exposure to morphine and amphetamines.^{20, 21} Chemical activation of lateral hypothalamus (LH) neurons reinstates extinguished morphine seeking in rats, and the effect is blocked by the selective OX_1 antagonist SB334867 (1).¹⁶ Blockade of orexin A transmission also decreases alcohol self-administration and cue-induced reinstatement of extinguished alcohol and cocaine seeking, 2^{2} , 2^{3} and attenuates stress-induced reinstatement of extinguished cocaine and alcohol seeking.^{19, 24} Moreover, antagonism of orexin A transmission at the OX_1 receptor decreases nicotine self-administration in rats and the motivation to obtain the d rug.²⁵

Early efforts to modulate the orexin system have been focused on the blockade of both receptors, or $OX₂$ selectively, because of their well-recognized implications in the pathophysiology of sleep disorders. As a result, several structural classes of dual OX_1/OX_2 antagonists and OX_2 selective antagonists were developed.^{12, 26-29} The dual antagonists almorexant (**2**), SB-649868 (**3**) and suvorexant (**4**) entered clinical trials for insomnia.27, 28 However, relatively few OX₁ selective antagonists have been reported to date. Compound 1, developed by GlaxoSmithKline, was the first selective OX_1 antagonist described.^{30, 31} It has a potency of $~40$ nM at OX₁ and is at least 50-fold selective for OX₁ over OX₂. Structureactivity relationship (SAR) studies have resulted in analogs with improved $OX₁$ selectivity,³² but 1 remains an important tool to probe the pharmacology and function of OX1. Despite its wide application, **1** has limitations including a less than desirable pharmacokinetic profile and the recently discovered hydrolytic instability where the 2 methylbenzoxazole undergoes ring opening under acidic conditions.31, 33 Other antagonists with OX_1 selectivity or preference are the pyrrolidine-based SB674042 and disubstituted piperidines based on the structure of **3**. 34, 35 Two compounds of the latter series showed OX_1 selectivity or preference, though significant OX_2 activity remains for most.³⁵ A similar approach by Rottapharm resulted in a number of spiro-piperidines and spiro-pyrrolidines with reasonable OX_1 potency and selectivity.³⁶ During the preparation of this manuscript, a phenylglycine-amide substituted tetrahydroisoquinoline derivative (ACT-335827) was reported by Actelion that was potent and OX_1 selective; however some OX_2 activity remained (IC₅₀ OX₁ = 6 nM, OX₂ = 417 nM).³⁷

Here we report our efforts in developing OX_1 selective antagonists based on a tetrahydroisoquinoline scaffold as present in both **2** and TCS-OX2-29 (5), a selective OX₂ antagonist.38 Compound **6**, identified in a high throughput screening campaign by Actelion which led to the discovery of **2**, showed reasonable OX_1 activity and selectivity (IC₅₀ OX₁ = 119 nM, $OX_2 = 8100$ nM).³⁹ Initial SAR studies by the same group identified several positions that could be modified to improve OX_1 activity. For example, replacement of the methoxy group at the 7-position with larger groups such as ethoxy or propoxy further increased OX_1 potency while little OX_2 activity was observed. The present work further examines SARs within the tetrahydroisoquinoline series, with the aim to improve potency and selectivity over OX_2 and identify a viable tool compound for pharmacological studies. Our investigation looked specifically at the structural requirement at the 7-position of the tetrahydroisoquinoline, as well as the substitution pattern at the acetamide.

Results and Discussion

Chemistry

7-Alkoxytetrahydroisoquinolines were synthesized according to Scheme 1. Thus, amide coupling between amine **7** and acid **8** using HBTU yielded **9**. Bischler-Napieralski reaction of **9** with phosphorus oxychloride in toluene at 90 °C gave the intermediate dihydroisoquinoline,40 which was readily reduced to tetrahydroisoquinoline **10** using sodium borohydride. N-Alkylation with α-bromo-benzylacetoamide gave the desired tetrahydroisoquinoline derivative **11**. 41, 42 The 7-position could then be alkylated as desired using potassium carbonate as base and the addition of tetrabutylammonium iodide as appropriate. For the trifluoroethyl analog **24**, better results were obtained using cesium carbonate at 50 °C. Sulfonates (**22-23, 25-26**) were prepared using the appropriate sulfonyl chloride in dichloromethane with triethylamine as base.

To prepare compounds varying in the N-acetamide (Scheme 2), the base tetramethoxytetrahydroisoquinoline (**31**) was made according to literature procedures and then Nalkylated by ethyl bromoacetate to give ester **32**. ³⁹ The ester could then be hydrolyzed and the acid converted to the amide using BOP and the corresponding amine. Further elaboration of the 4-nitrophenyl derivative gave a series of acylated anilines **57-59**. The benzyl ester **40** was also made by alkylation on the acid with benzyl bromide.

To prepare the 7-alkoxy derivatives, with greater variety in the acetamide, a series of compounds were synthesized via Scheme 3. Phenol **10** was alkylated with ethyl bromoacetate to give the tetrahydroisoquinoline acetate **65** in good yield. The phenol was then alkylated with the appropriate alkyl halide, with either potassium or cesium carbonate as base. Hydrolysis of the ester **66-68** with aqueous sodium hydroxide in ethanol gave the acid, which could then be coupled with the desired amine using BOP as coupling agent.

Activity of the target compounds at the OX_1 and OX_2 receptors was evaluated in a calcium mobilization based functional assay. The apparent dissociation constant Ke was calculated from compound-mediated inhibition of orexin A activity as previously described.³² Briefly, EC50 curves of the agonist, orexin A, were obtained alone and together with the test compound, respectively, and the right-shift of the agonist curve was measured. Ke values were then calculated using the equation Ke = $[L] / ((EC_{50}^-/EC_{50}^+)$ -1), where [L] is the test compound concentration, EC_{50}^- is the EC_{50} of orexin A alone, and EC_{50}^+ is the EC_{50} of orexin A in the presence of test compound. In these assays, The EC_{50} for orexin A at OX_1 and OX₂ is 0.13 ± 0.02 nM and 4.2 ± 0.2 nM, respectively. All the compounds that had OX₁ Ke values $\lt 1 \mu M$ were also tested alone at 10 μM as agonists at the OX₁ receptor. None of them showed any agonist activity. Due to the lack of commercially available radioligands, binding studies were not performed on these compounds.

Our studies on this series focused on two primary areas of the molecule, the 7-position of the tetrahydroisoquinoline moiety itself and the acetamide side chain. Table 1 shows changes at the 7-position. The unsubstituted 7-hydroxy analog **11** showed micromolar potency at both OX_1 and OX_2 . The OX_1 potency increased as the size of the alkoxy group increased (6, 12, **13**), in agreement with the literature report.39 However, the potency started to drop as the chain length further increased (**14, 15**), and the isopentyl (**16**) was equipotent with hexyl (**15**). Finally, compound **17** with the space demanding cyclohexylmethyl group had no $OX₁$ activity up to 10 μ M, suggesting a limited bulk tolerance at this position. In an attempt to probe possible hydrogen bonding effects, heteroatoms as H-acceptors were introduced on the alkyl chain, and this resulted in significant decrease of potency (**18, 19, 20**). Aromatic substituents were then examined. While the benzyl (**21**) was similar in potency to the hexyl (**15**), the two pyridylmethyl analogs (**22, 23**) showed a modest increase in potency over the

benzyl **21**. Interestingly, when a phenoxybutyl group was introduced, **24** displayed equipotency to the butyl analog. This may suggest additional aromatic stacking interaction in this region, probably further away from the tetrahydroisoquinoline core.

Electronic properties at the 7-position were also investigated. The butyryl analog **25** showed similar OX_1 activity as the butyl compound **14**, suggesting electron withdrawing groups are tolerated. Interestingly, two highly electron withdrawing sulfonate analogs (**26, 27**) showed excellent potency, further confirming the preference of electronegativity at this position. Trifluoroethyl (**28**), a group that can be considered a bioisostere for acetyl or mesylate because of its electron deficient character and similar size, gave low nanomolar potency and over 1000-fold selectivity over the OX_2 receptor. This also suggests that the observed high potency is a result of the electronic properties, other than potential hydrogen bonding interactions with the presence of the additional oxygen atoms. Finally, the phenyl (**29**) and tolyl (**30**) sulfonates both had higher potency than the benzyl analog.

At the acetamide position (Table 2), extending the benzyl to a phenethyl (**33**) or phenylpropyl (**34**) lowered potency at OX1. Conformational restriction of the aromatic group led to analogs such as the tetrahydroisoquinoline **35**, the piperidines **36** and **37,** and the piperazine 38 , but none of these compounds showed any appreciable OX_1 activity. It was posited that the NH group might be required for potency, and, indeed, its importance was highlighted by the N-methyl **39** and the benzyl ester **40**, which both had low potency at OX_1 . None of these compounds showed any activity at OX_2 .

Several fused aromatic or biphenyl systems were examined where the second aromatic ring occupies a similar position as that in the benzyl group but in a planar confirmation. The naphthalene **41** had a Ke of 732 nM, around 3.6-fold less potent than the benzyl compound. Heteroatom containing aromatic systems such as the quinolines **42** and **43** gave a further reduction in potency, suggesting a preference for a non-planar conformation for the aromatic ring. We next examined a series of biphenyl analogs **44-46**, with each acting as a phenyl spacer to the second aromatic ring. However, only 2-biphenyl **44** showed sub-micromolar OX_1 potency, with some activity at OX_2 . These results suggest that a non-aromatic spacer is preferred for OX_1 potency. Indeed, the tetrahydronaphthalene (47) gave a Ke of 41 nM, significantly more potent than the naphthalene **41**. Interestingly, the tetrahydroquinoline **48** was inactive, which shows that hydrophobic interactions might be preferred in this region of the molecule.

With the chiral center at the 1-position of the tetrahydroisoquinoline, substitution with a methyl at the α-position of the benzyl group (S-α–methylbenzyl) gave two diastereomers, which were separated by HPLC into compounds 49 and 50 . While 50 showed good $OX₁$ potency and selectivity, **49** was inactive at 10 μM at both receptors. This is consistent with previous findings that the S confirmation at the 1-position of the tetrahydroisoquinoline is required in 2 and its analogs.^{28, 43}

Substituents on the phenyl ring were generally well tolerated, with chloride **51** and fluoride **52** about equipotent with the benzyl (**6**), suggesting that electron withdrawing groups are tolerated. Among the pyridylmethyl analogs, the 3-pyridyl (**54**) showed similar potency as the 2-analog (**53**), both of which were more potent than the 4-analog (**55**). Interestingly, the electron rich dimethylamino (**56**) showed limited OX_1 potency in the μ M range. To determine whether this activity loss is due to electronic or steric reasons, substituents including amides and ureas were introduced onto the nitrogen. Surprisingly, the acetyl group (**57**) resulted in a total loss of OX1 activity. Large amide (**58**) and urea (**59**) showed no or little potency, suggesting limited or no tolerance for size at the 4-position of the benzyl group.

Finally, the requirement for aromaticity was investigated at the acetamide position. The straight chain heptyl analog **60** showed modest potency at 500 nM though was poorly selective over OX2. Attempts to introduce heteroatoms or polar groups (**61-64**) all resulted in dramatic loss of OX_1 potency. Taken together, these findings suggest an aromatic group is generally needed for $OX₁$ potency and selectivity.

Although none of the acetamide substituents gave a significant improvement in potency from the parent benzyl compound **6**, the pyridylmethyl analogs **54** and **55** showed most promise given their improved physicochemical properties due to the possibility of salt formation. Thus, several pyridylmethyl analogs were studied in conjunction with the more potent 7-position substituents (Table 3). Such analogs include 2- and 3-pyridines combined with the 7-ethoxy, propoxy and trifluoroethoxy derivatives (**69-74**). All compounds showed good to excellent OX_1 potency (8-87 nM) and high selectivity over OX_2 , with none of the compounds showing any activity at 10 μM.

Computational Analysis

A comprehensive 3D-pharmacophore for the orexin antagonists evaluated in this study was developed. The pharmacophore was initiated by generating conformational libraries for each analog in this class. This included computation of the 3D-distance metrics between 4 candidate pharmacophore points (variable substituents off the 7-position, the two centroids for each of the aromatic ring substituents and the nitrogen on the tetrahydroisoquinoline core), and determination of the conformations of each ligand giving rise to a common 3D display (Figure 2A and B). Superposition of all ligands in the training set with these four pharmacophore points allowed us to investigate a series of substituent properties that proved important for OX_1 potency. Twenty-one properties, including stereochemical (Verloop parameters,44, 45 volume, area, globularity), electrostatic (positions of electrostatic minima and maxima), polar (polar surface area), electronic (E-HOMO, E-LUMO, polarizability), thermodynamic (vibrational and rotational heat capacity and entropies at 300K) and atom based hydrophobicity (ClogP), were evaluated for ligand conformations that complied with a 3D-pharmacophore overlap rule.⁴⁶ In addition, the same subset of properties was computed for substituents (fragments) alone without the tetrahydroisoquinoline core compliant pharmacophoric region. This dual approach was chosen to allow the addition of substituent independent properties to the QSAR analysis.

Three properties were found to correlate structural modification at the 7-position with alterations in OX1 potency using a three variable QSAR analysis. The Sterimol parameters including spatial extent of the substituent at position 7 and particularly the width (Verloop B1) relative to the long-axis, logP of the substituent, and energy of the lowest unoccupied molecular orbital (E-LUMO) demonstrated statistically significant correlations with changes in OX₁ potency ($r^2 = 0.69$) (Figure 2C). It is apparent from the 3-D QSAR model (Figure 2D) and the functional activity data that size of the substituent plays a considerable role in determining potency at the 7-position. $OX₁$ potency increases monotonically from 1530 nM to 17 nM for compounds **11** (−OH), **6** (−OCH₃), **12** (−OCH₂CH₃), and **13** (−OCH₂CH₂CH₃) respectively at which point the trend begins to reverse. This indicates that substituents of a particular size are required for activity.

Although steric properties play a main role in determining potency at the 7-position, the QSAR analysis also highlights the importance of other non-observable factors such as LUMO energy on antagonist potency. Figure 2E depicts the spatial distribution of the LUMO density on compound **6**. Position 7 is on the aromatic ring most associated with LUMO electron density. Naturally, substitutions at that position would modulate the energy of the LUMO orbital and charge density transfer, a facet plausibly having a role in stabilization of the receptor bound ligand configuration.

Figure 3 depicts a similar 3D-QSAR and COMFA analysis for substituent variation corresponding to the N-alkylation variations summarized in Table 2. Figure 3B illustrates results from a COMFA analysis of the training set superposition shown in Figure 3A. Both steric and electrostatic variations in the ligand substituents contributed relatively equally to the underlying rationale for the $Ke(OX_1)$ variations. This was independently verified in the 2D-QSAR analysis shown in Figure 3D, where coefficients of the B1-Verloop parameter and the polar surface area made significant contributions, along with the energy of the HOMO, and led to a model predictive of $ln Ke(OX_1)$. Analogous to our identification of the E(LUMO) term for position 7 variations, the atomic centers containing the highest occupied molecular orbital were found on the ring system directly attached to the position 2 substitutions. The results from these studies highlight the manner in which substitutions can be rationalized in terms of underlying physical properties keyed to the molecular interactions and binding free energies and provide a basis for a comprehensive 3Dpharmacophore.

Through the establishment of this 3D-pharmacophore model a basic understanding of the impact specific substitutions have on receptor potency and selectivity has been achieved. This information will be important to guide the selection of scaffold modifications to favorably alter receptor potency, subtype selectivity, or ADM. Alternatively, establishing quantitative pharmacophore models allows us to transfer information of the 3D-display of physiochemical properties to new scaffolds with an understanding of those that are critical for activity.

Conditioned Place Preference

Given the well-established activity of OX_1 antagonists in attenuating the rewarding and reinforcing effects of drugs, compound **72** was studied in conditioned place preference (CPP). As shown in Figure 4, vehicle did not produce significant CPP. Cocaine at 10 mg/kg induced robust and significant CPP. Compound **72** at 20 mg/kg did not produce CPP or place aversion but significantly attenuated the development of place preference induced by cocaine.

Conclusions

In an attempt to develop OX_1 selective antagonists, we synthesized and evaluated a series of tetrahydroisoquinolines, a core structure that is present in both the dual $OX₁/OX₂$ antagonist **2** and the OX₂ selective antagonist **5**. SAR studies suggested a preference for steric bulk at the 7-position; however, this tolerance is limited, as the potency decreases with the further size increase of the substituents. Electron deficient groups are also well tolerated at this position. At the acetamide position, an aromatic system was generally preferred, and the pyridylmethyl groups gave the best potency. The pharmacophore model obtained through computational analysis confirmed the steric and electronic requirement at the 7-position. This model also suggests that steric and electrostatic variations in the ligand substituents contributed equally to the underlying rationale for the Ke $(OX₁)$ variations at the acetamide position. In the CPP paradigm, compound 72 , which had excellent OX_1 potency and selectivity over OX_2 in the calcium assay, significantly attenuated cocaine CPP. In summary, several compounds with excellent potency at and selectivity for the $OX₁$ receptor have been identified, and they will serve as probes to further investigate the pharmacology and function of the OX_1 receptor and the orexin system. Modifications at other positions are ongoing and will be reported in due course.

Experimental

Chemistry

All solvents and chemicals were reagent grade. Unless otherwise mentioned, all were purchased from commercial vendors and used as received. Flash column chromatography was done on a Teledyne ISCO CombiFlash Rf system using prepacked columns. Solvents used were hexane, ethyl acetate (EtOAc), dichloromethane, methanol and chloroform:methanol:ammonium hydroxide (80:18:2) (CMA-80). Purity and characterization of compounds was established by a combination of high pressure liquid chromatography (HPLC), thin layer chromatography (TLC), mass spectrometry (MS) and nuclear magnetic resonance (NMR) analysis. 1 H and 13 C NMR spectra were recorded on a Bruker Avance DPX-300 (300 MHz) spectrometer and were determined in chloroform-d or methanol-d4 with tetramethylsilane (TMS) (0.00 ppm) or solvent peaks as the internal reference. Chemical shifts are reported in ppm relative to the reference signal, and coupling constant (J) values are reported in Hz. TLC was done on EMD precoated silica gel 60 F254 plates, and spots were visualized with UV light or iodine staining. Low resolution mass spectra were obtained using a Waters Alliance HT/Micromass ZQ system (ESI). All test compounds were greater than 95% pure (except where noted) as determined by HPLC on an Agilent 1100 system using an Agilent Zorbax SB-Phenyl, 2.1 mm x 150 mm, 5 μm column with gradient elution using the mobile phases (A) H_2O containing 0.1% CF₃COOH and (B) MeCN, with a flow rate of 1.0 mL/min.

2-(3,4-dimethoxyphenyl)-N-[2-(4-hydroxy-3-methoxyphenyl)ethyl]acetamide (9) —3,4-Dimethoxyphenylacetic acid (0.45 g, 2.29 mmol), 4-hydroxy-3-

methoxyphenethylamine hydrochloride (0.47 g, 2.29 mmol) and HBTU (0.96 g, 2.52 mmol) were combined in dry dimethylformamide (20 mL) at RT under N₂. Diisopropylethylamine (1.19 g, 1.6 mL, 9.17 mmol) was added and the reaction stirred at RT overnight. The reaction was diluted with ethyl acetate, washed with 2N hydrochloric acid, sodium bicarbonate solution and brine, dried over MgSO₄ and the solvent removed under reduced pressure to give the desired amide as a yellow oil which solidified upon standing (0.72 g, 91%). 1H NMR (300 MHz, CHLOROFORM-*d*) δ ppm 2.67 (t, *J*=6.88 Hz, 2 H) 3.36 - 3.43 (m, 2 H) 3.45 (s, 2 H) 3.82 (s, 3 H) 3.83 (s, 3 H) 3.88 (s, 3 H) 6.04 (br. s., 1 H) 6.44 - 6.53 (m, 1 H) 6.61 (d, *J*=1.70 Hz, 1 H) 6.66 - 6.77 (m, 3 H) 6.78 - 6.86 (m, 1 H) 7.97 (s, 1 H).

General procedure for Bischler-Napieralski reaction/sodium borohydride reduction 1-[(3,4-dimethoxyphenyl)methyl]-6-methoxy-1,2,3,4-

tetrahydroisoquinolin-7-ol (10)—Amide **9** (0.39 g, 0.91 mmol) was suspended in anhydrous toluene (5 mL) and phosphorous oxychloride (0.70 g, 0.4 mL, 4.56 mmol) added slowly. The reaction was heated to 90 °C for 2 hours. The reaction was cooled, then quenched by slow addition of the reaction mixture to water and stirred vigorously at room temperature for 15 min. Sodium hydroxide solution (2N) was added until pH was 8-9, then the solution was extracted 3 times with dichloromethane. The combined extracts were dried over MgSO4 and the solvent removed under reduced pressure.

The crude dihydroisoquinoline was dissolved in methanol (5 mL) and cooled in an ice bath under N_2 . Sodium borohydride (0.17 g, 4.52 mmol) was added portionwise and the reaction allowed to warm slowly to RT overnight. The reaction was quenched with water then the methanol removed under reduced pressure. The aqueous solution was extracted 3 times with dichloromethane and the combined extracts were dried over $MgSO₄$ and the solvent removed under reduced pressure to give the desired tetrahydroisoquinoline **10** as a frothy solid (0.26 g, 67%). 1H NMR (300 MHz, CHLOROFORM-*d*) δ ppm 2.59 - 2.93 (m, 5 H)

3.11 - 3.24 (m, 1 H) 3.34 - 3.58 (m, 2 H) 3.79 - 3.92 (m, 9 H) 4.08 (dd, *J*=9.42, 3.20 Hz, 1 H) 6.57 (s, 1 H) 6.60 - 6.71 (m, 1 H) 6.72 - 6.88 (m, 3 H).

N-benzyl-2-{1-[(3,4-dimethoxyphenyl)methyl]-7-hydroxy-6-methoxy-1,2,3,4 tetrahydroisoquinolin-2-yl}acetamide (11)—Amine **10** (3.65 g, 11.08 mmol), Nbenzyl-2-bromoacetamide (3.77 g, 16.62 mmol) and tetrabutylammounium iodide (0.82 g, 2.21 mmol) were combined in dry DMF (110 mL) and diisopropylethylamine (3.58 g, 4.8 mL, 27.70 mmol) was added. The reaction was stirred at RT overnight under N_2 . The reaction was diluted with EtOAc, washed with NaHCO₃ solution, water and brine $(x2)$, then dried over MgSO4 and the solvent removed under reduced pressure. The crude was purified by chromatography on silica (0-100% EtOAc in hexane) to obtain the desired product as a frothy white solid (2.98 g, 56%). ¹H NMR (300 MHz, CHLOROFORM-d) δ 7.15 - 7.36 (m, 3H), 7.07 (d, *J* = 7.06 Hz, 2H), 6.89 (br. s., 1H), 6.65 - 6.77 (m, 4H), 6.52 - 6.64 (m, 3H), 5.51 (s, 1H), 4.48 (dd, *J* = 8.24, 15.02 Hz, 1H), 3.88 (s, 3H), 3.80 (s, 3H), 3.73 (s, 3H), 3.33 - 3.68 (m, 3H), 3.06 - 3.32 (m, 2H), 2.78 - 2.99 (m, 4H), 2.46 (d, *J* = 15.82 Hz, 1H). *m/z* 477 (M+H).

General procedure for O-alkylation: N-benzyl-2-{1-[(3,4 dimethoxyphenyl)methyl]-7-ethoxy-6-methoxy-1,2,3,4-tetrahydroisoquinolin-2 yl}acetamide (12)—Phenol **11** (25 mg, 0.025 mmol) and potassium carbonate (22 mg, 0.157 mmol) were combined in dry dimethylformamide, then 1-bromoethane (12 mg, 6 μL, 0.079 mmol) was added and the reaction stirred at RT under N_2 overnight. It was diluted with ethyl acetate, washed with sodium bicarbonate solution and brine, dried over $MgSO₄$ and the solvent was removed under reduced pressure. The compound was purified by chromatography on silica (0-75% EtOAc in hexane) to obtain the desired product as a pale yellow solid (19 mg, 73%). ¹H NMR (300 MHz, CHLOROFORM-d) δ 7.19 - 7.34 (m, 3H), 7.07 - 7.14 (m, 1H), 6.94 - 7.01 (m, 1H), 6.56 - 6.74 (m, 5H), 6.48 (s, 1H), 4.49 (dd, *J* = 8.05, 14.93 Hz, 1H), 4.02 (q, *J* = 6.97 Hz, 2H), 3.85 (s, 3H), 3.81 (s, 3H), 3.74 (s, 3H), 3.57 - 3.70 (m, 2H), 3.34 - 3.48 (m, 1H), 3.12 - 3.34 (m, 2H), 2.79 - 2.99 (m, 4H), 2.41 - 2.54 (m, 1H), 1.45 (t, *J* = 6.97 Hz, 3H). *m/z* 505 (M+H).

N-benzyl-2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}acetamide (6)—Prepared in 51% yield. ¹H NMR (300) MHz, CHLOROFORM-d) δ 7.18 - 7.35 (m, 2H), 7.07 - 7.14 (m, 2H), 6.98 (dd, *J* = 4.94, 7.58 Hz, 1H), 6.62 - 6.74 (m, 3H), 6.59 (s, 1H), 6.45 (s, 1H), 4.50 (dd, *J* = 8.05, 14.93 Hz, 1H), 3.84 - 3.89 (m, 3H), 3.81 (d, *J* = 1.88 Hz, 6H), 3.78 - 3.83 (m, 6H), 3.75 (s, 3H), 3.59 - 3.71 (m, 2H), 3.35 - 3.48 (m, 1H), 3.11 - 3.35 (m, 2H), 2.80 - 3.00 (m, 4H), 2.41 - 2.55 (m, 1H). *m/z* 491 (M+H).

N-benzyl-2-{1-[(3,4-dimethoxyphenyl)methyl]-6-methoxy-7-propoxy-1,2,3,4 tetrahydroisoquinolin-2-.yl}acetamide (13)—Prepared in 48% yield. ¹H NMR (300) MHz, CHLOROFORM-d) δ 7.20 - 7.34 (m, 3H), 7.07 - 7.14 (m, 2H), 6.98 (dd, *J* = 4.95, 7.77 Hz, 1H), 6.61 - 6.75 (m, 3H), 6.59 (s, 1H), 6.47 (s, 1H), 4.50 (dd, *J* = 8.10, 14.98 Hz, 1H), 3.89 (t, *J* = 6.88 Hz, 2H), 3.85 (s, 3H), 3.81 (s, 3H), 3.75 (s, 3H), 3.58 - 3.70 (m, 2H), 3.34 - 3.48 (m, 1H), 3.11 - 3.34 (m, 2H), 2.79 - 2.99 (m, 4H), 2.49 (d, *J* = 15.92 Hz, 1H), 1.77 - 1.92 (m, 2H), 1.00 - 1.09 (m, 3H). *m/z* 519 (M+H).

N-benzyl-2-{7-butoxy-1-[(3,4-dimethoxyphenyl)methyl]-6-methoxy-1,2,3,4 tetrahydroisoquinolin-2-yl}acetamide (14)—Prepared in 75% yield. ¹H NMR (300) MHz, CHLOROFORM-d) δ 7.19 - 7.35 (m, 3H), 7.10 (d, *J* = 6.78 Hz, 2H), 6.93 - 7.02 (m, 1H), 6.61 - 6.75 (m, 3H), 6.58 (s, 1H), 6.47 (s, 1H), 4.50 (dd, *J* = 8.10, 14.98 Hz, 1H), 3.89 - 3.97 (m, 2H), 3.83 - 3.87 (m, 3H), 3.81 (s, 3H), 3.75 (s, 3H), 3.58 - 3.71 (m, 2H), 3.34 - 3.48

(m, 1H), 3.11 - 3.34 (m, 2H), 2.80 - 2.98 (m, 4H), 2.42 - 2.54 (m, 1H), 1.75 - 1.88 (m, 2H), 1.42 - 1.55 (m, 2H), 0.98 (t, *J* = 7.30 Hz, 3H). *m/z* 533 (M+H).

N-benzyl-2-{1-[(3,4-dimethoxyphenyl)methyl]-7-(hexyloxy)-6-methoxy-1,2,3,4 tetrahydroisoquinolin-2-yl}acetamide (15)—Prepared in 64% yield. ¹H NMR (300) MHz, CHLOROFORM-d) δ 7.18 - 7.35 (m, 3H), 7.10 (d, *J* = 6.69 Hz, 2H), 6.97 (dd, *J* = 4.99, 7.44 Hz, 1H), 6.61 - 6.76 (m, 3H), 6.58 (s, 1H), 6.48 (s, 1H), 4.50 (dd, *J* = 8.10, 14.98 Hz, 1H), 3.92 (t, *J* = 6.83 Hz, 2H), 3.82 - 3.87 (m, 3H), 3.81 (s, 3H), 3.74 (s, 3H), 3.58 - 3.69 (m, 2H), 3.34 - 3.49 (m, 1H), 3.11 - 3.34 (m, 2H), 2.80 - 2.99 (m, 4H), 2.41 - 2.54 (m, 1H), 1.75 - 1.89 (m, 2H), 1.30 - 1.53 (m, 6H), 0.91 (t, *J* = 6.90 Hz, 3H). *m/z* 561 (M+H).

N-benzyl-2-{1-[(3,4-dimethoxyphenyl)methyl]-6-methoxy-7-(3 methylbutoxy)-1,2,3,4-tetrahydroisoquinolin-2-yl}acetamide (16)—Prepared in 88% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 7.20 - 7.36 (m, 3H), 7.11 (d, *J* = 6.97 Hz, 2H), 6.95 - 7.04 (m, 1H), 6.62 - 6.76 (m, 3H), 6.59 (s, 1H), 6.48 (s, 1H), 4.51 (dd, *J* = 8.01, 14.98 Hz, 1H), 3.95 (t, *J* = 6.88 Hz, 2H), 3.85 (s, 3H), 3.81 (s, 3H), 3.75 (s, 3H), 3.60 - 3.71 (m, 2H), 3.35 - 3.49 (m, 1H), 3.13 - 3.34 (m, 2H), 2.78 - 2.99 (m, 4H), 2.49 (d, *J* = 15.73 Hz, 1H), 1.84 (td, *J* = 6.62, 13.33 Hz, 1H), 1.71 - 1.77 (m, 2H), 0.98 (d, *J* = 6.50 Hz, 6H). *m/z* 547 (M+H).

N-benzyl-2-[7-(cyclohexylmethoxy)-1-[(3,4-dimethoxyphenyl)methyl]-6 methoxy-1,2,3,4-tetrahydroisoquinolin-2-yl]acetamide (17)—Prepared in 33% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 7.19 - 7.34 (m, 3H), 7.10 (d, *J* = 6.78 Hz, 2H), 7.01 (d, *J* = 6.97 Hz, 1H), 6.61 - 6.76 (m, 3H), 6.58 (s, 1H), 6.45 (s, 1H), 4.49 (dd, *J* = 8.01, 14.98 Hz, 1H), 3.82 (s, 3H), 3.81 (s, 3H), 3.75 (s, 3H), 3.59 - 3.72 (m, 4H), 3.34 - 3.47 (m, 1H), 3.12 - 3.34 (m, 2H), 2.79 - 2.95 (m, 4H), 2.48 (d, *J* = 16.01 Hz, 1H), 1.61 - 1.97 (m, 6H), 0.92 - 1.40 (m, 5H). *m/z* 573 (M+H).

N-Benzyl-2-{1-[(3,4-dimethoxyphenyl)methyl]-7-[2-(dimethylamino)ethoxy]-6 methoxy-1,2,3,4-tetrahydroisoquinolin-2-yl}acetamide (18)—Prepared in 46% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 7.19 - 7.34 (m, 3H), 7.07 - 7.14 (m, 2H), 6.95 (dd, *J* = 4.76, 7.77 Hz, 1H), 6.68 - 6.75 (m, 2H), 6.61 - 6.67 (m, 1H), 6.59 (s, 1H), 6.56 (s, 1H), 4.50 (dd, *J* = 8.19, 14.98 Hz, 1H), 4.01 - 4.12 (m, 2H), 3.84 (s, 3H), 3.82 (s, 3H), 3.75 (s, 3H), 3.57 - 3.68 (m, 2H), 3.36 - 3.50 (m, 1H), 3.12 - 3.34 (m, 2H), 2.83 - 2.99 (m, 4H), 2.75 - 2.82 (m, 2H), 2.44 - 2.54 (m, 1H), 2.38 (s, 6H). *m/z* 548 (M+H).

N-benzyl-2-{1-[(3,4-dimethoxyphenyl)methyl]-7-[4-(dimethylamino)butoxy]-6 methoxy-1,2,3,4-tetrahydroisoquinolin-2-yl}acetamide (19)—Phenol **11** (0.30 g, 0.63 mmol) and potassium carbonate (0.35 g, 1.26 mmol) were combined in dimethylformamide (6 mL) and 1-bromo-4-chlorobutane (0.22 g, 0.15 mL, 1.26 mmol) was added. The reaction was heated to 50 °C overnight. The reaction was cooled, diluted with water and extracted twice with EtOAc. The combined extracts were washed with brine, dried over $MgSO₄$ and the solvent removed under reduced pressure. It was then purified by chromatography on silica (0-80% EtOAc in hexane) to obtain the desired chloride (0.22 g, 61%) for use in the following reaction. 1H NMR (300 MHz, CHLOROFORM-*d*) δ ppm 7.20 - 7.35 (m, 3 H) 7.11 (d, *J*=7.16 Hz, 2 H) 6.93 - 7.01 (m, 1 H) 6.62 - 6.75 (m, 3 H) 6.60 (s, 1 H) 6.48 (s, 1 H) 4.50 (dd, *J*=14.93, 8.05 Hz, 1 H) 3.97 (t, *J*=5.51 Hz, 2 H) 3.85 (s, 3 H) 3.80 (s, 3 H) 3.75 (s, 3 H) 3.58 - 3.70 (m, 4 H) 3.36 - 3.55 (m, 1 H) 3.12 - 3.34 (m, 2 H) 2.79 - 2.98 (m, 4 H) 2.42 - 2.56 (m, 1 H) 1.94 - 2.03 (m, 4 H).

The chloride (30 mg, 0.053 mmol), dimethylamine hydrochloride (7 mg, 0.079 mmol), potassium carbonate (18 mg, 0.132 mmol) and tetrabutylammonium iodide (4 mg, 0.011

mmol) were combined in dimethylformamide (0.5 mL) and heated at 50 °C overnight. The reaction was cooled, diluted with water and extracted 3 times with EtOAc. The combined extracts were washed with brine, dried over $MgSO₄$ and the solvent removed under reduced pressure. The compound was purified by chromatography on silica (0-50% CMA-80 in EtOAc) to give the desired product (6 mg, 19%). ¹H NMR (300 MHz, CHLOROFORM-d) δ 7.19 - 7.35 (m, 3H), 7.07 - 7.15 (m, 2H), 6.91 - 7.00 (m, 1H), 6.62 - 6.76 (m, 3H), 6.59 (s, 1H), 6.51 (s, 1H), 4.50 (dd, *J* = 8.15, 15.02 Hz, 1H), 3.98 (t, *J* = 6.50 Hz, 2H), 3.84 (s, 3H), 3.82 (s, 3H), 3.75 (s, 3H), 3.57 - 3.68 (m, 2H), 3.36 - 3.49 (m, 1H), 3.11 - 3.34 (m, 2H), 2.81 - 2.99 (m, 4H), 2.43 - 2.54 (m, 1H), 2.33 - 2.42 (m, 2H), 2.27 (s, 6H), 1.79 - 1.92 (m, 2H), 1.62 - 1.76 (m, 2H). *m/z* 576 (M+H).

N-benzyl-2-{1-[(3,4-dimethoxyphenyl)methyl]-6-methoxy-7-[2-(piperidin-1 yl)ethoxy]-1,2,3,4-tetrahydroisoquinolin-2-yl}acetamide (20)—Prepared in 24% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 7.20 - 7.35 (m, 3H), 7.10 (d, *J* = 7.06 Hz, 2H), 6.93 (dd, *J* = 4.71, 7.82 Hz, 1H), 6.68 - 6.76 (m, 2H), 6.61 - 6.66 (m, 1H), 6.59 - 6.61 (m, 1H), 6.58 (s, 1H), 4.49 (dd, *J* = 8.24, 15.02 Hz, 1H), 4.11 (t, *J* = 6.45 Hz, 2H), 3.85 (s, 3H), 3.81 - 3.84 (m, 3H), 3.74 (s, 3H), 3.55 - 3.65 (m, 2H), 3.36 - 3.51 (m, 1H), 3.11 - 3.33 (m, 2H), 2.75 - 2.98 (m, 6H), 2.41 - 2.61 (m, 5H), 1.56 - 1.71 (m, 4H), 1.48 (d, *J* = 4.99 Hz, 2H). *m/z* 588 (M+H).

N-benzyl-2-[7-(benzyloxy)-1-[(3,4-dimethoxyphenyl)methyl]-6-methoxy-1,2,3,4 tetrahydroisoquinolin-2-yl]acetamide (21)—Prepared in 78% yield. ¹H NMR (300) MHz, CHLOROFORM-d) δ 7.39 (td, *J* = 7.11, 15.16 Hz, 3H), 7.19 - 7.33 (m, 4H), 7.10 (d, *J* = 6.69 Hz, 2H), 6.91 - 7.01 (m, 1H), 6.57 - 6.70 (m, 5H), 6.49 (s, 1H), 5.06 (s, 2H), 4.48 (dd, *J* = 8.05, 14.93 Hz, 1H), 3.87 (s, 3H), 3.79 (s, 3H), 3.74 (s, 3H), 3.52 - 3.70 (m, 2H), 3.32 - 3.45 (m, 1H), 3.10 - 3.32 (m, 2H), 2.79 - 2.98 (m, 3H), 2.67 - 2.78 (m, 1H), 2.41 - 2.53 (m, 1H). *m/z* 567 (M+H).

N-benzyl-2-{1-[(3,4-dimethoxyphenyl)methyl]-6-methoxy-7-(pyridin-2 ylmethoxy)-1,2,3,4-tetrahydroisoquinolin-2-yl}acetamide (22)—Prepared in 61% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 8.60 (d, *J* = 4.71 Hz, 1H), 7.68 - 7.77 (m, 1H), 7.58 (d, *J* = 7.82 Hz, 1H), 7.17 - 7.35 (m, 5H), 7.08 (d, *J* = 7.06 Hz, 2H), 6.89 (br. s., 1H), 6.56 - 6.75 (m, 4H), 5.27 (s, 2H), 4.48 (dd, *J* = 8.19, 14.98 Hz, 1H), 3.90 (s, 3H), 3.82 (s, 3H), 3.74 (s, 3H), 3.50 - 3.62 (m, 2H), 3.33 - 3.48 (m, 1H), 3.08 - 3.31 (m, 2H), 2.68 - 3.00 (m, 4H), 2.48 (d, *J* = 15.64 Hz, 1H). *m/z* 568 (M+H).

N-benzyl-2-{1-[(3,4-dimethoxyphenyl)methyl]-6-methoxy-7-(pyridin-3 ylmethoxy)-1,2,3,4-tetrahydroisoquinolin-2-yl}acetamide (23)—Prepared in 47% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 8.68 (s, 1H), 8.58 (d, *J* = 4.71 Hz, 1H), 7.79 (d, *J* = 7.82 Hz, 1H), 7.18 - 7.37 (m, 4H), 7.11 (d, *J* = 7.06 Hz, 2H), 6.98 (br. s., 1H), 6.57 - 6.73 (m, 4H), 6.50 (s, 1H), 5.05 (s, 2H), 4.49 (dd, *J* = 8.01, 14.98 Hz, 1H), 3.87 (s, 3H), 3.81 (s, 3H), 3.75 (s, 3H), 3.54 - 3.71 (m, 2H), 3.34 - 3.48 (m, 1H), 3.10 - 3.33 (m, 2H), 2.72 - 3.01 (m, 4H), 2.43 - 2.57 (m, 1H). *m/z* 568 (M+H).

N-benzyl-2-{1-[(3,4-dimethoxyphenyl)methyl]-6-methoxy-7-(4 phenoxybutoxy)-1,2,3,4-tetrahydroisoquinolin-2-yl}acetamide (24)—Prepared in 82% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 7.20 - 7.36 (m, 5H), 7.11 (d, *J* = 6.78 Hz, 2H), 6.87 - 7.01 (m, 4H), 6.62 - 6.76 (m, 3H), 6.60 (s, 1H), 6.51 (s, 1H), 4.51 (dd, *J* = 8.05, 14.93 Hz, 1H), 3.98 - 4.11 (m, 4H), 3.84 (s, 3H), 3.81 (s, 3H), 3.75 (s, 3H), 3.58 - 3.70 (m, 2H), 3.35 - 3.51 (m, 1H), 3.11 - 3.34 (m, 2H), 2.81 - 3.02 (m, 4H), 2.42 - 2.56 (m, 1H), 1.94 - 2.11 (m, 4H). *m/z* 624 (M+H).

2-[(benzylcarbamoyl)methyl]-1-[(3,4-dimethoxyphenyl)methyl]-6 methoxy-1,2,3,4-tetrahydroisoquinolin-7-yl butanoate (25)—Phenol 11 (50 mg, 0.105 mmol), butyric acid (9 mg, 0.105 mmol) and BOP (46 mg, 0.105 mmol) were combined in dichloromethane (1 mL). Diisopropylethylamine (34 mg, $46 \mu L$, 0.262 mmol) was added and the reaction stirred at RT under N_2 overnight. The reaction was diluted with EtOAc, washed with 2M HCl, NaHCO₃ solution and brine, dried over $MgSO₄$ and the solvent removed under reduced pressure. The crude was purified by chromatography on silica (0-75% EtOAc in hexane) to give the desired product as a pale yellow solid (54 mg, 95%). 1H NMR (300 MHz, CHLOROFORM-d) δ 7.18 - 7.34 (m, 4H), 7.10 (d, *J* = 6.69 Hz, 2H), 6.92 (d, *J* = 6.97 Hz, 1H), 6.74 (s, 1H), 6.56 - 6.71 (m, 4H), 4.47 (dd, *J* = 8.01, 15.07 Hz, 1H), 3.80 (s, 3H), 3.79 (s, 3H), 3.74 (s, 3H), 3.59 - 3.70 (m, 2H), 3.11 - 3.44 (m, 3H), 2.77 - 3.01 (m, 4H), 2.55 (t, *J* = 7.00 Hz, 2H), 2.49 (d, *J* = 2.64 Hz, 1H), 1.72 - 1.87 (m, 2H), 1.05 (t, *J* = 7.39 Hz, 3H). *m/z* 547 (M+H).

General procedure for sulfonate formation: 2-[(benzylcarbamoyl)methyl]-1- [(3,4-dimethoxyphenyl)methyl]-6-methoxy-1,2,3,4-tetrahydroisoquinolin-7-yl methanesulfonate (26)—To a solution of phenol **11** (30 mg, 0.063 mmol) in

dichloromethane (0.5 mL) cooled in ice under N₂ was added triethylamine (16 mg, 22 μ L, 0.157 mmol), and methanesulfonyl chloride (14 mg, $10 \mu L$, 0.126 mmol). The reaction was allowed to warm slowly to RT overnight. Solvent was removed under reduced pressure and the crude purified by chromatography on silica (0-100% EtOAc in hexane) to give the desired product (35 mg, 54%). ¹H NMR (300 MHz, CHLOROFORM-d) δ 7.20 - 7.36 (m, 3H), 7.03 - 7.12 (m, 3H), 6.86 (br. s., 1H), 6.67 - 6.76 (m, 3H), 6.60 - 6.65 (m, 1H), 4.48 (dd, *J* = 8.10, 14.98 Hz, 1H), 3.89 (s, 3H), 3.82 (s, 3H), 3.74 (s, 3H), 3.54 - 3.72 (m, 2H), 3.24 - 3.48 (m, 2H), 3.19 (s, 3H), 3.10 - 3.17 (m, 1H), 2.82 - 3.05 (m, 4H), 2.48 - 2.61 (m, 1H). *m/z* 555 (M+H).

2-[(benzylcarbamoyl)methyl]-1-[(3,4-dimethoxyphenyl)methyl]-6 methoxy-1,2,3,4-tetrahydroisoquinolin-7-yl trifluoromethanesulfonate (27)— Prepared in 18% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 7.21 - 7.36 (m, 4H), 7.10 (d, *J* = 8.01 Hz, 2H), 6.90 (s, 1H), 6.87 (br. s., 1H), 6.75 (s, 1H), 6.59 - 6.72 (m, 2H), 4.49 (dd, *J* = 8.05, 15.02 Hz, 1H), 3.90 (s, 3H), 3.80 (s, 3H), 3.74 (s, 3H), 3.60 - 3.73 (m, 2H), 3.33 - 3.45 (m, 1H), 3.11 - 3.33 (m, 2H), 2.77 - 3.04 (m, 4H), 2.56 (d, *J* = 18.46 Hz, 1H). 19F NMR (282 MHz, CHLOROFORM-d) δ -73.7. *m/z* 609 (M+H).

N-benzyl-2-{1-[(3,4-dimethoxyphenyl)methyl]-6-methoxy-7-(2,2,2 trifluoroethoxy)-1,2,3,4-tetrahydroisoquinolin-2-yl}acetamide (28)—Phenol **11** (100 mg, 0.24 mmol) and cesium carbonate (235 mg, 0.72 mmol) were combined in dry dimethylformamide (1.5 mL) and 2,2,2-trifluoroiodoethane (101 mg, 47 μ L, 0.48 mmol) was added. The reaction was heated to 50 °C overnight until TLC (4:1 EtOAc/hexane) showed all starting material gone. The reaction was cooled, diluted with ethyl acetate and then washed with sodium bicarbonate solution and brine. The solution was then dried over MgSO4 and the solvent removed under reduced pressure. The compound was purified by chromatography on silica (0-50% EtOAc in hexane) to obtain the desired product as a pale yellow solid (86 mg, 72%). 1H NMR (300 MHz, CHLOROFORM-d) δ 7.19 - 7.34 (m, 3H), 7.09 (d, *J* = 6.69 Hz, 2H), 6.94 (d, *J* = 5.09 Hz, 1H), 6.59 - 6.75 (m, 5H), 4.48 (dd, *J* = 8.15, 15.02 Hz, 1H), 4.23 - 4.41 (m, 2H), 3.87 (s, 3H), 3.81 (s, 3H), 3.74 (s, 3H), 3.57 - 3.68 (m, 2H), 3.34 - 3.50 (m, 1H), 3.09 - 3.34 (m, 2H), 2.76 - 3.00 (m, 4H), 2.44 - 2.57 (m, 1H). *m/z* 559 (M+H).

2-[(benzylcarbamoyl)methyl]-1-[(3,4-dimethoxyphenyl)methyl]-6 methoxy-1,2,3,4-tetrahydroisoquinolin-7-yl benzenesulfonate (29)—Prepared in

64% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 7.87 - 7.94 (m, 2H), 7.62 - 7.71 (m, 1H), 7.48 - 7.58 (m, 2H), 7.19 - 7.35 (m, 3H), 7.09 (d, *J* = 6.88 Hz, 2H), 6.96 (s, 1H), 6.83 (d, *J* = 2.83 Hz, 1H), 6.58 - 6.73 (m, 3H), 6.55 (s, 1H), 4.48 (dd, *J* = 8.24, 15.02 Hz, 1H), 3.82 (s, 3H), 3.74 (s, 3H), 3.54 - 3.67 (m, 2H), 3.51 (s, 3H), 3.32 - 3.47 (m, 1H), 3.07 - 3.31 (m, 2H), 2.73 - 2.99 (m, 4H), 2.41 - 2.56 (m, 1H). *m/z* 617 (M+H).

2-[(benzylcarbamoyl)methyl]-1-[(3,4-dimethoxyphenyl)methyl]-6-

methoxy-1,2,3,4-tetrahydroisoquinolin-7-yl 4-methylbenzene-1-sulfonate (30) —Prepared in 63% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 7.78 (d, *J* = 8.19 Hz, 2H), 7.19 - 7.37 (m, 5H), 7.08 (d, *J* = 6.97 Hz, 2H), 6.94 (s, 1H), 6.79 - 6.86 (m, 1H), 6.59 - 6.72 (m, 3H), 6.56 (s, 1H), 4.49 (dd, *J* = 8.19, 14.98 Hz, 1H), 3.82 (s, 3H), 3.74 (s, 3H), 3.58 - 3.67 (m, 2H), 3.54 (s, 3H), 3.32 - 3.45 (m, 1H), 3.07 - 3.31 (m, 2H), 2.73 - 2.98 (m, 4H), 2.47 -2.54 (m, 1H), 2.45 (s, 3H). *m/z* 631 (M+H).

General procedure for N-alkylation with acetate: 2-{1-[(3,4 dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-tetrahydroisoquinolin-2-

yl}acetate (32)—Tetrahydroisoquinoline **31** (0.26 g, 0.63 mmol), ethyl bromoacetate (0.16 g, 0.11 mL, 0.95 mmol) and tetrabutylammonium iodide (47 mg, 0.13 mmol) were combined in anhydrous dimethylformamide (6 mL) and diisopropylethylamine (0.20 g, 0.28 mL, 1.58 mmol) was added. The reaction was stirred at RT under $N₂$ overnight. The reaction was diluted with ethyl acetate, washed with sodium bicarbonate solution and brine, dried over $MgSO₄$ and solvent removed under reduced pressure. The compound was purified by chromatography on silica (0-50% EtOAc/hexane) to give the desired product as a frothy solid (0.17 g, 55%). 1H NMR (300 MHz, CHLOROFORM-*d*) δ ppm 6.73 - 6.79 (m, 1 H) 6.60 - 6.68 (m, 2 H) 6.55 (s, 1 H) 6.01 (s, 1 H) 4.11 - 4.22 (m, 2 H) 3.93 (dd, *J*=7.39, 5.23 Hz, 1 H) 3.85 (s, 3 H) 3.83 (s, 3 H) 3.80 (s, 3 H) 3.57 (s, 3 H) 3.39 - 3.55 (m, 2 H) 3.28 (ddd, *J*=12.88, 9.68, 5.04 Hz, 1 H) 3.14 (dd, *J*=13.37, 4.99 Hz, 1 H) 2.98 (dt, *J*=12.60, 4.77 Hz, 1 H) 2.73 - 2.87 (m, 2 H) 2.45 - 2.59 (m, 1 H) 1.26 (t, *J*=7.11 Hz, 3 H).

General procedure for ester hydrolysis: 2-{1-[(3,4 dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-tetrahydroisoquinolin-2-

yl}acetic acid—Ester (80 mg, 0.16 mmol) was dissolved in ethanol (2 mL) and aqueous sodium hydroxide solution (2N, 0.3 mL, 0.64 mmol) was added. The reaction was stirred at RT for 2 hours. The pH of the solution was adjusted to between 8 and 9 with 2N HCl, then all solvents were removed under reduced pressure. The crude solid was redissolved as far as possible in methanol, the remaining solids removed by filtration and the filtrate concentrated. The desired acid was obtained as a white solid $(75 \text{ mg}, 100\%)$. ¹H NMR (300) MHz, METHANOL-*d*4) δ ppm 6.92 (d, *J*=8.10 Hz, 1 H) 6.82 (s, 1 H) 6.68 - 6.78 (m, 2 H) 5.96 (s, 1 H) 4.67 (dd, *J*=9.04, 5.09 Hz, 1 H) 3.81 - 3.82 (m, 3 H) 3.76 (s, 3 H) 3.71 - 3.85 (m, 6 H) 3.46 (s, 3 H) 3.38 - 3.66 (m, 2 H) 2.95 - 3.18 (m, 3 H).

General procedure for final coupling: 2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7 dimethoxy-1,2,3,4-tetrahydroisoquinolin-2-yl}-N-(2-phenylethyl)acetamide (33)

—Acid (30 mg, 0.075 mmol), phenethylamine (9 mg, 9 μL, 0.075 mmol) and BOP (33 mg, 0.075 mmol) were combined in anhydrous dimethylformamide (1 mL) and diisopropylethylamine (24 mg, 33 μL, 0.187 mmol) was added. The reaction was stirred at RT under N_2 overnight. The reaction was diluted with ethyl acetate, washed with sodium bicarbonate solution and brine, dried over $MgSO₄$ and the solvent removed under reduced pressure. The compound was purified by chromatography on silica (0-75% EtOAc/hexane) to give the desired product as a pale yellow solid $(10 \text{ mg}, 26\%)$. ¹H NMR $(300 \text{ MHz},$ CHLOROFORM-d) δ 7.18 - 7.33 (m, 3H), 7.11 - 7.18 (m, 2H), 6.66 - 6.83 (m, 4H), 6.58 (s, 1H), 6.41 (s, 1H), 3.87 (s, 6H), 3.85 (s, 3H), 3.80 (s, 3H), 3.59 (dd, *J* = 5.56, 9.23 Hz, 1H),

3.28 - 3.47 (m, 2H), 3.03 - 3.28 (m, 2H), 2.73 - 2.99 (m, 5H), 2.39 - 2.67 (m, 3H). *m/z* 505 $(M+H)$.

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-N-(3-phenylpropyl)acetamide (34)—Prepared in 64% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 7.23 - 7.33 (m, 2H), 7.11 - 7.22 (m, 3H), 6.69 - 6.85 (m, 3H), 6.61 - 6.68 (m, 1H), 6.59 (s, 1H), 6.42 (s, 1H), 3.86 (s, 3H), 3.85 (s, 3H), 3.81 (s, 3H), 3.80 (s, 3H), 3.60 (dd, *J* = 5.37, 9.23 Hz, 1H), 3.01 - 3.50 (m, 4H), 2.78 - 3.00 (m, 4H), 2.60 - 2.78 (m, 1H), 2.42 - 2.57 (m, 3H), 1.45 - 1.69 (m, 2H). *m/z* 519 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4 tetrahydroisoquinolin-2-yl}-1-(1,2,3,4-tetrahydroisoquinolin-2-yl)ethan-1-one (35)—Prepared in 32% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 7.01 - 7.24 (m, 3H), 6.53 - 6.90 (m, 5H), 6.27 (d, J = 11.87 Hz, 1H), 4.24 - 4.89 (m, 2H), 3.65 - 4.01 (m, 12H), 3.22 - 3.64 (m, 5H), 2.73 - 3.10 (m, 6H), 2.35 - 2.72 (m, 2H). *m/z* 517.4 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-1-(3-phenylpiperidin-1-yl)ethan-1-one (36)— Prepared in 42% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 7.13 - 7.41 (m, 6H), 6.88 - 7.10 (m, 1H), 6.52 - 6.83 (m, 3H), 4.64 (d, *J* = 7.72 Hz, 1H), 3.76 - 3.95 (m, 12H), 3.53 - 3.75 (m, 4H), 3.20 - 3.52 (m, 4H), 2.71 - 3.15 (m, 5H), 2.30 - 2.66 (m, 4H). *m/z* 545 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-1-(4-phenylpiperidin-1-yl)ethan-1-one (37)— Prepared in 52% yield. ¹H NMR (300 MHz, CHLOROFORM-d) δ 7.02 - 7.40 (m, 5H), 6.63 - 6.83 (m, 3H), 6.59 (s, 1H), 6.31 (s, 1H), 4.53 - 4.85 (m, 1H), 3.77 - 3.91 (m, 9H), 3.72 (s, 3H), 3.62 (s, 1H), 3.21 - 3.55 (m, 4H), 2.75 - 3.18 (m, 5H), 2.36 - 2.72 (m, 4H), 1.86 (br. s., 1H), 1.33 - 1.59 (m, 2H). *m/z* 545 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-1-(4-phenylpiperazin-1-yl)ethan-1-one (38)— Prepared in 42% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 7.18 - 7.39 (m, 3H), 6.89 (s, 2H), 6.71 (s, 3H), 6.59 (s, 1H), 6.33 (s, 1H), 3.77 - 3.94 (m, 9H), 3.73 (s, 3H), 3.47 (d, *J* = 12.81 Hz, 3H), 3.34 (d, *J* = 12.81 Hz, 5H), 2.70 - 3.07 (m, 8H), 2.47 (dd, *J* = 3.86, 16.48 Hz, 1H). *m/z* 546 (M+H).

N-benzyl-2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4 tetrahydroisoquinolin-2-yl}-N-methylacetamide (39)—Prepared in 47% yield. ¹H NMR (300 MHz, CHLOROFORM-d) δ 7.17 - 7.37 (m, 4H), 7.01 (d, *J* = 7.16 Hz, 1H), 6.64 - 6.80 (m, 3H), 6.58 (d, *J* = 4.90 Hz, 1H), 6.18 - 6.34 (m, 1H), 4.40 - 4.74 (m, 1H), 3.89 - 4.33 (m, 2H), 3.78 - 3.88 (m, 9H), 3.65 - 3.76 (m, 3H), 3.28 - 3.58 (m, 3H), 2.83 - 3.15 (m, 4H), 2.70 - 2.82 (m, 3H), 2.39 - 2.60 (m, 1H). *m/z* 505 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}acetate (40)—Acid (30 mg, 0.075 mmol), potassium carbonate (26 mg, 0.187 mmol) and tetrabutylammonium iodide (6 mg, 0.015 mmol) were combined in dimethylformamide (1 mL) and benzyl bromide (19 mg, 13 μ L, 0.112 mmol) was added. The reaction was heated to 50 $^{\circ}$ C under N₂ overnight. The reaction was cooled and diluted with EtOAc. It was washed with NaHCO₃ solution and brine, dried over $MgSO₄$ and the solvent was removed under reduced pressure. The material was purified by chromatography on silica (0-50% EtOAc in hexane) to give the desired ester (9 mg, 24%). 1H NMR (300 MHz, CHLOROFORM-d) δ 7.35 (s, 5H), 6.73 (d, *J* = 8.01 Hz, 1H),

6.57 - 6.66 (m, 2H), 6.55 (s, 1H), 6.00 (s, 1H), 5.16 (s, 2H), 3.90 - 4.03 (m, 1H), 3.84 (s, 6H), 3.78 (s, 3H), 3.45 - 3.65 (m, 5H), 3.22 - 3.36 (m, 1H), 3.09 - 3.20 (m, 1H), 2.94 - 3.08 (m, 1H), 2.71 - 2.90 (m, 2H), 2.53 (d, *J* = 16.39 Hz, 1H). *m/z* 492 (M+H). HPLC purity 91.0%.

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-N-(naphthalen-1-yl)acetamide (41)—Prepared in 64% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 9.25 (s, 1H), 7.84 (d, *J* = 7.63 Hz, 2H), 7.66 (d, *J* = 8.10 Hz, 1H), 7.33 - 7.54 (m, 4H), 6.63 - 6.72 (m, 2H), 6.59 (s, 1H), 6.46 (s, 1H), 6.30 (d, *J* = 8.10 Hz, 1H), 3.87 - 3.92 (m, 3H), 3.90 (s, 1H), 3.80 (s, 3H), 3.65 (s, 3H), 3.36 - 3.61 (m, 3H), 3.27 (s, 3H), 2.92 - 3.15 (m, 4H), 2.64 (d, *J* = 15.82 Hz, 1H). *m/z* 527 $(M+H)$.

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-N-(8-fluoroquinolin-4-yl)acetamide (42)—Prepared in 27% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 8.88 (d, *J* = 4.99 Hz, 1H), 8.31 (d, *J* = 4.99 Hz, 1H), 7.54 (dd, *J* = 1.60, 8.10 Hz, 1H), 7.28 - 7.46 (m, 2H), 6.87 (d, *J* = 8.38 Hz, 1H), 6.61 - 6.71 (m, 2H), 6.53 (d, *J* = 1.79 Hz, 1H), 6.43 (s, 1H), 6.37 (d, *J* = 8.10 Hz, 1H), 4.01 (t, *J* = 6.73 Hz, 1H), 3.90 (s, 3H), 3.75 - 3.81 (m, 3H), 3.68 (s, 3H), 3.39 - 3.55 (m, 6H), 2.89 - 3.20 (m, 4H), 2.62 - 2.76 (m, 1H). *m/z* 546 (M+H). HPLC purity 93.0%.

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-N-(2-methylquinolin-4-yl)acetamide (43)—Prepared in 28% yield. ¹H NMR (300 MHz, CHLOROFORM-d) δ 9.89 (s, 1H), 8.13 (s, 1H), 8.00 (d, *J* = 8.48 Hz, 1H), 7.66 (t, *J* = 7.30 Hz, 1H), 7.33 (t, *J* = 7.35 Hz, 1H), 7.11 (d, *J* = 8.38 Hz, 1H), 6.61 - 6.69 (m, 2H), 6.47 - 6.54 (m, 1H), 6.40 (s, 1H), 6.36 (d, *J* = 8.19 Hz, 1H), 3.99 (t, *J* = 6.64 Hz, 1H), 3.90 (s, 3H), 3.79 - 3.87 (m, 2H), 3.76 (s, 3H), 3.62 - 3.69 (m, 3H), 3.43 - 3.54 (m, 2H), 3.39 (s, 3H), 2.87 - 3.21 (m, 4H), 2.73 (s, 3H). *m/z* 542 (M+H). HPLC purity 93.0%.

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-N-(2-phenylphenyl)acetamide (44)—Prepared in 40% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 9.41 (s, 1H), 8.35 (d, *J* = 8.29 Hz, 1H), 7.35 - 7.45 (m, 1H), 7.15 - 7.32 (m, 8H), 6.63 (d, *J* = 8.29 Hz, 1H), 6.51 (s, 1H), 6.36 - 6.45 (m, 2H), 5.85 (s, 1H), 3.87 (s, 3H), 3.73 (d, *J* = 11.68 Hz, 6H), 3.57 - 3.66 (m, 1H), 3.54 (s, 3H), 3.30 - 3.45 (m, 1H), 3.17 - 3.29 (m, 1H), 2.93 - 3.08 (m, 1H), 2.60 (d, *J* = 8.29 Hz, 4H), 2.27 - 2.39 (m, 1H). *m/z* 553 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-N-(3-phenylphenyl)acetamide (45)—Prepared in 27% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 8.68 (s, 1H), 7.57 (d, *J* = 7.91 Hz, 2H), 7.39 - 7.53 (m, 3H), 7.29 - 7.37 (m, 1H), 7.23 (d, *J* = 8.29 Hz, 3H), 6.71 (d, *J* = 1.51 Hz, 3H), 6.63 (s, 1H), 6.48 (s, 1H), 3.89 (s, 3H), 3.81 (d, *J* = 6.40 Hz, 6H), 3.76 (s, 3H), 3.45 - 3.57 (m, 1H), 3.24 - 3.45 (m, 2H), 2.89 - 3.09 (m, 4H), 2.52 - 2.64 (m, 1H). *m/z* 553 (M+H)

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-N-(4-phenylphenyl)acetamide (46)—Prepared in 16% yield. ¹H NMR (300 MHz, CHLOROFORM-d) δ 8.74 (br. s., 1H), 7.59 (d, $J = 6.78$ Hz, 3H), 7.28 - 7.51 (m, 4H), 7.06 (br. s., 1H), 6.58 - 6.92 (m, 4H), 6.45 (s, 1H), 4.12 (q, *J* = 7.16 Hz, 1H), 3.77 - 3.97 (m, 6H), 3.56 - 3.76 (m, 6H), 3.23 - 3.54 (m, 4H), 2.85 - 3.13 (m, 3H), 2.58 (d, *J* = 15.82 Hz, 1H). *m/z* 553 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-N-(1,2,3,4-tetrahydronaphthalen-1-yl)acetamide (47)—Prepared in 63% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 7.06 - 7.22 (m, 4H), 6.53 - 6.63 (m, 2H), 6.18 - 6.49 (m, 3H), 5.00 - 5.21 (m, 1H), 3.79 - 3.88 (m, 3H), 3.74 (d, *J* = 6.03 Hz, 6H), 3.67 (s, 3H), 3.11 - 3.49 (m, 3H), 2.62 - 3.05 (m, 7H), 2.42 - 2.56 (m, 1H), 1.66 - 2.02 (m, 4H). *m/z* 531 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4 tetrahydroisoquinolin-2-yl}-N-(2-methyl-1,2,3,4-tetrahydroquinolin-4-

yl)acetamide (48)—Quinoline **43** (20 mg, 0.037 mmol) and 10% palladium on carbon (20 mg) were combined in ethanol (5 mL) and chloroform (1 mL) and hydrogenated at 50 psi of H_2 in a Parr shaker for 3 days. The reaction was filtered through Celite and the solvent was removed under reduced pressure. The crude was purified by chromatography on silica $(0-20\% \text{ MMA-80 in EtOAc})$ to give the tetrahydroisoquinoline derivative $(3 \text{ mg}, 15\%)$. ¹H NMR (300 MHz, CHLOROFORM-d) δ 9.09 (s, 1H), 7.78 (s, 1H), 7.26 (s, 3H), 6.57 - 6.67 (m, 3H), 6.51 (s, 1H), 6.28 (s, 1H), 3.82 - 3.99 (m, 5H), 3.59 - 3.81 (m, 10H), 3.27 - 3.54 (m, 4H), 3.04 - 3.19 (m, 1H), 2.79 - 3.03 (m, 5H), 2.58 - 2.70 (m, 1H), 2.48 (s, 3H). *m/z* 546 (M $+H$).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4 tetrahydroisoquinolin-2-yl}-N-[(1S)-1-phenylethyl]acetamide (49 and 50)—

Prepared from tetrahydroisoquinoline 11 and 2-bromo-N-[(1S)-1-phenylethyl]acetamide as above as a mixture of diastereomers in 54% yield. Diastereomers were separated by reverse phase HPLC; the first to elute was 49 and the second was 50, 49: 1 H NMR (300 MHz, CHLOROFORM-d) δ ppm 8.24 (br. s., 1 H) 7.20 - 7.37 (m, 5 H) 6.74 (d, *J*=8.10 Hz, 1 H) 6.62 (s, 1 H) 6.58 (s, 1 H) 6.49 (d, *J*=8.19 Hz, 1 H) 5.86 (br. s., 1 H) 4.97 - 5.09 (m, 1 H) 4.60 - 4.70 (m, 1 H) 3.88 - 4.03 (m, 2 H) 3.86 (s, 3 H) 3.83 - 3.85 (m, 3 H) 3.75 (s, 3 H) 3.61 (dd, *J*=11.73, 6.64 Hz, 2 H) 3.53 (s, 3 H) 3.45 - 3.50 (m, 1 H) 3.19 (d, *J*=10.83 Hz, 1 H) 2.83 - 3.04 (m, 2 H) 1.48 (d, *J*=6.97 Hz, 3 H). *m/z* 505 (M+H). 50: 1H NMR (300 MHz, CHLOROFORM-d) δ ppm 8.49 (d, *J*=7.63 Hz, 1 H) 7.21 - 7.39 (m, 5 H) 6.73 (d, *J*=8.19 Hz, 1 H) 6.62 (s, 1 H) 6.54 (d, *J*=1.70 Hz, 1 H) 6.43 (d, *J*=8.10 Hz, 1 H) 5.78 (s, 1 H) 5.05 (s, 1 H) 4.52 (d, *J*=6.31 Hz, 1 H) 3.86 (s, 3 H) 3.84 (s, 3 H) 3.78 - 3.95 (m, 2 H) 3.75 (s, 3 H) 3.56 - 3.69 (m, 2 H) 3.51 - 3.54 (m, 1 H) 3.49 (s, 3 H) 3.10 - 3.33 (m, 1 H) 2.83 - 3.04 (m, 2 H) 1.50 (d, *J*=6.97 Hz, 3 H). *m/z* 505 (M+H).

N-[(4-chlorophenyl)methyl]-2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7 dimethoxy-1,2,3,4-tetrahydroisoquinolin-2-yl}acetamide (51)—Prepared in 60% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 7.21 - 7.29 (m, 2H), 7.03 (d, *J* = 8.38 Hz, 2H), 6.91 (t, *J* = 6.31 Hz, 1H), 6.63 - 6.78 (m, 3H), 6.59 (s, 1H), 6.48 (s, 1H), 4.41 (dd, *J* = 8.05, 15.02 Hz, 1H), 3.87 (s, 3H), 3.84 (s, 3H), 3.82 (s, 3H), 3.77 (s, 3H), 3.52 - 3.65 (m, 2H), 3.36 - 3.51 (m, 1H), 3.09 - 3.34 (m, 2H), 2.80 - 3.01 (m, 4H), 2.42 - 2.55 (m, 1H). *m/z* 527, 525 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4 tetrahydroisoquinolin-2-yl}-N-[(4-fluorophenyl)methyl]acetamide (52)—Prepared in 64% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 7.02 - 7.11 (m, 2H), 6.87 - 7.02 (m, 3H), 6.64 - 6.78 (m, 3H), 6.59 (s, 1H), 6.47 (s, 1H), 4.42 (dd, *J* = 8.10, 14.79 Hz, 1H), 3.87 (s, 3H), 3.83 (s, 3H), 3.82 (s, 3H), 3.78 (s, 3H), 3.52 - 3.66 (m, 2H), 3.36 - 3.51 (m, 1H), 3.09 - 3.34 (m, 2H), 2.80 - 3.00 (m, 4H), 2.42 - 2.56 (m, 1H). *m/z* 509 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4 tetrahydroisoquinolin-2-yl}-N-(pyridin-2-ylmethyl)acetamide (53)—Prepared in 57

% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 8.52 (d, *J* = 4.52 Hz, 1H), 7.53 - 7.69 (m, 2H), 7.16 (dd, *J* = 5.18, 7.16 Hz, 1H), 7.09 (d, *J* = 7.82 Hz, 1H), 6.62 - 6.72 (m, 3H), 6.60 (s, 1H), 6.29 (s, 1H), 4.53 (dd, *J* = 6.55, 16.25 Hz, 1H), 4.17 (dd, *J* = 5.32, 16.25 Hz, 1H), 3.86 (s, 3H), 3.77 (s, 3H), 3.67 - 3.75 (m, 7H), 3.17 - 3.51 (m, 3H), 2.82 - 3.11 (m, 4H), 2.45 - 2.60 (m, 1H). *m/z* 492 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4 tetrahydroisoquinolin-2-yl}-N-(pyridin-3-ylmethyl)acetamide (54)—Prepared in 49% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 8.50 (dd, *J* = 1.37, 4.76 Hz, 1H), 8.33 (d, *J* = 1.70 Hz, 1H), 7.43 (d, *J* = 7.82 Hz, 1H), 7.22 (dd, *J* = 4.80, 7.72 Hz, 1H), 6.89 - 6.98 (m, 1H), 6.64 - 6.79 (m, 3H), 6.60 (s, 1H), 6.51 (s, 1H), 4.41 (dd, *J* = 8.01, 15.26 Hz, 1H), 3.87 (s, 3H), 3.85 (s, 3H), 3.84 (s, 3H), 3.77 (s, 3H), 3.56 - 3.70 (m, 2H), 3.38 - 3.54 (m, 1H), 3.09 - 3.34 (m, 2H), 2.82 - 3.02 (m, 4H), 2.43 - 2.57 (m, 1H). *m/z* 492 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-N-(pyridin-4-ylmethyl)acetamide (55)—Prepared in 89% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 8.48 - 8.56 (m, 2H), 6.96 - 7.02 (m, 2H), 6.92 (dd, *J* = 5.13, 7.68 Hz, 1H), 6.71 - 6.80 (m, 2H), 6.58 - 6.68 (m, 2H), 6.53 (s, 1H), 4.40 (dd, *J* = 8.10, 15.92 Hz, 1H), 3.88 (s, 3H), 3.86 (s, 3H), 3.83 (s, 3H), 3.69 (s, 3H), 3.55 - 3.66 (m, 2H), 3.41 - 3.54 (m, 1H), 3.10 - 3.37 (m, 2H), 2.82 - 3.04 (m, 4H), 2.53 (d, *J* = 16.29 Hz, 1H). *m/z* 492 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4 tetrahydroisoquinolin-2-yl}-N-{[4-(dimethylamino)phenyl]methyl}acetamide (56)—Prepared in 7% yield. ¹H NMR (300 MHz, CHLOROFORM-d) δ 7.03 (d, $J = 8.57$ Hz, 2H), 6.92 - 6.99 (m, 1H), 6.62 - 6.73 (m, 5H), 6.58 (s, 1H), 6.38 (s, 1H), 4.38 - 4.49 (m, 1H), 3.86 (s, 3H), 3.83 (s, 3H), 3.82 (s, 3H), 3.78 (s, 3H), 3.52 - 3.69 (m, 2H), 3.31 - 3.45 (m, 1H), 3.21 (q, *J* = 16.80 Hz, 2H), 2.90 - 2.96 (m, 6H), 2.77 - 2.90 (m, 4H), 2.41 - 2.53 (m, 1H). *m/z* 534 (M+H).

2-bromo-N-[(4-nitrophenyl)methyl]acetamide—To a solution of 4-nitrobenzylamine hydrochloride (1.89 g, 10 mmol) and triethylamine (1.01 g, 1.39 mL, 10 mmol) in dichloromethane (10 mL) cooled in an ice bath was added dropwise bromoacetyl bromide (1.01 g, 0.44 mL, 5 mmol). A precipitate formed immediately. The ice bath was removed and the reaction stirred at RT for 2 hr. The precipitate was removed by filtration and the solution washed twice with 2N HCl, dried over MgSO₄ and the solvent removed under reduced pressure to give the bromide as a white solid $(1.4 \text{ g}, 100\%)$. ¹H NMR (300 MHz, CHLOROFORM-d) δ 8.22 (d, *J* = 8.67 Hz, 2H), 7.47 (d, *J* = 8.48 Hz, 2H), 4.57 - 4.64 (m, 2H), 4.15 (s, 2H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-N-[(4-nitrophenyl)methyl]acetamide—Prepared as per general alkylation procedure using 2-bromo-N-[(4-nitrophenyl)methyl]acetamide in 63% yield. ¹H NMR (300 MHz, CHLOROFORM-d) δ 8.15 (d, $J = 8.76$ Hz, 2H), 7.23 (d, $J =$ 8.67 Hz, 2H), 6.90 - 7.00 (m, 1H), 6.65 - 6.81 (m, 3H), 6.61 (s, 1H), 6.53 (s, 1H), 4.47 (dd, *J* $= 8.01, 15.54$ Hz, 1H), 3.87 (s, 3H), 3.87 (s, 3H), 3.83 (s, 3H), 3.70 - 3.75 (m, 3H), 3.42 -3.71 (m, 3H), 3.11 - 3.35 (m, 2H), 2.83 - 3.03 (m, 4H), 2.45 - 2.57 (m, 1H).

N-[(4-aminophenyl)methyl]-2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-

dimethoxy-1,2,3,4-tetrahydroisoquinolin-2-yl}acetamide—To a solution of nitrophenyl (197 mg, 0.37 mmol) and hydrazine monohydrate (0.15 mL) in ethanol (5 mL) warmed to 50 °C was added Raney nickel (2800, as a slurry in water, 25 mg) and the

reaction heated at 50 °C for 1 hr. It was cooled and filtered through Celite and the solvent removed under reduced pressure to give the aniline (155 mg, 83%). ¹H NMR (300 MHz, CHLOROFORM-d) δ 6.92 (d, *J* = 8.29 Hz, 2H), 6.70 (s, 2H), 6.56 - 6.68 (m, 4H), 6.40 (s, 1H), 4.39 (dd, *J* = 7.91, 14.51 Hz, 1H), 3.86 (s, 3H), 3.82 (s, 6H), 3.79 (s, 3H), 3.51 - 3.67 (m, 3H), 3.32 - 3.44 (m, 1H), 3.10 - 3.31 (m, 2H), 2.78 - 2.98 (m, 4H), 2.42 - 2.53 (m, 1H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-N-[(4-acetamidophenyl)methyl]acetamide (57)—To the aniline (30 mg, 0.059 mmol) in dichloromethane (1 mL) was added acetic anhydride (12 mg, $11 \mu L$, 0.119 mmol) and diisopropylethylamine (19 mg, $26 \mu L$, 0.148 mmol) and the reaction stirred at RT overnight. The reaction was diluted with EtOAc, washed with $NaHCO₃$ solution. The aqueous was extracted once with EtOAc and the combined organics were washed with brine, dried over $MgSO₄$ and the solvent removed under reduced pressure. Purified by chromatography on silica (0-100% EtOAc in hexane) to give the amide (34 mg, 100%). 1H NMR (300 MHz, CHLOROFORM-d) δ 7.41 (d, *J* = 8.38 Hz, 2H), 7.23 (s, 1H), 7.05 (d, *J* = 8.38 Hz, 2H), 6.89 - 6.99 (m, 1H), 6.64 - 6.79 (m, 3H), 6.59 (s, 1H), 6.46 (s, 1H), 4.41 (dd, *J* = 7.91, 15.07 Hz, 1H), 3.87 (s, 3H), 3.81 - 3.84 (m, 6H), 3.78 (s, 3H), 3.54 - 3.67 (m, 2H), 3.35 - 3.50 (m, 1H), 3.09 - 3.33 (m, 2H), 2.78 - 2.99 (m, 4H), 2.42 - 2.56 (m, 1H), 2.17 (s, 3H). *m/z* 548 (M+H).

N-{4-[(2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}acetamido)methyl]phenyl}hexanamide (58)—Aniline (23 mg, 0.046 mmol), hexanoic acid (5 mg, 0.046 mmol) and BOP (20 mg, 0.046 mmol) were combined in dichloromethane (1 mL) and diisopropylethylamine (12 mg, 16 μL, 0.091 mmol) was added. The reaction was stirred at RT overnight and diluted with EtOAc. It was washed with NaHCO₃ solution and brine, dried over $MgSO₄$ and the solvent was removed under reduced pressure. It was purified by chromatography on silica (0-100% EtOAc in hexane) to give the amide (30 mg, 100%). ¹H NMR (300 MHz, CHLOROFORM-d) δ 7.43 (d, *J* = 8.29 Hz, 2H), 7.21 - 7.28 (m, 1H), 7.04 (d, *J* = 8.38 Hz, 2H), 6.94 (dd, *J* = 5.09, 7.72 Hz, 1H), 6.65 - 6.75 (m, 3H), 6.59 (s, 1H), 6.46 (s, 1H), 4.41 (dd, *J* = 8.05, 14.83 Hz, 1H), 3.86 (s, 3H), 3.81 - 3.84 (m, *J* = 0.80 Hz, 6H), 3.78 (s, 3H), 3.54 - 3.66 (m, 2H), 3.35 - 3.53 (m, 1H), 3.09 - 3.33 (m, 2H), 2.79 - 2.99 (m, 4H), 2.43 - 2.56 (m, 1H), 2.34 (t, *J* = 7.54 Hz, 2H), 1.64 - 1.79 (m, 2H), 1.31 - 1.39 (m, 4H), 0.87 - 0.95 (m, 3H). *m/z* 604 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4 tetrahydroisoquinolin-2-yl}-N-({4-

[(hexylcarbamoyl)amino]phenyl}methyl)acetamide (59)—To the aniline (40 mg, 0.079 mmol) in toluene (1 mL) was added n-hexyl isocyanate (11 mg, $13 \mu L$, 0.087 mmol) and the mixture heated to 75 °C for 2 hr. The reaction was cooled, the solvent was removed under reduced pressure and the crude was purified by chromatography on silica (0-100% EtOAc in hexane) to give the urea (38 mg, 76%). ¹H NMR (300 MHz, CHLOROFORM-d) δ 7.12 (d, *J* = 8.48 Hz, 2H), 7.00 (dd, *J* = 5.27, 7.72 Hz, 1H), 6.93 (d, *J* = 8.48 Hz, 2H), 6.87 (s, 1H), 6.66 - 6.77 (m, 3H), 6.60 (s, 1H), 6.49 (s, 1H), 5.29 (t, *J* = 5.56 Hz, 1H), 4.33 (dd, *J* $= 7.82, 14.98$ Hz, 1H), 3.87 (s, 3H), 3.84 (s, 3H), 3.82 (s, 3H), 3.75 (s, 3H), 3.54 - 3.66 (m, 2H), 3.36 - 3.51 (m, 1H), 3.07 - 3.32 (m, 4H), 2.79 - 3.01 (m, 4H), 2.43 - 2.56 (m, 1H), 1.39 - 1.54 (m, 2H), 1.21 - 1.36 (m, 6H), 0.82 - 0.90 (m, 3H). *m/z* 633 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-N-heptylacetamide (60)—Prepared in 63% yield. ¹H NMR (300 MHz, CHLOROFORM-d) δ 6.67 - 6.91 (m, 3H), 6.59 (m, 2H), 6.42 (m, 1H), 3.74 - 3.96 (m, 12H), 3.62 (m, 1H), 3.33 - 3.49 (m, 1H), 3.03 - 3.30 (m, 3H), 2.78 - 3.02 (m,

4H), 2.62 (m, 1H), 2.51 (d, *J* = 16.29 Hz, 1H), 1.08 - 1.38 (m, 10H), 0.89 (m, 3H). *m/z* 499 $(M+H)$.

2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4 tetrahydroisoquinolin-2-yl}-N-[3-(dimethylamino)propyl]acetamide (61)— Prepared in 57% yield. ¹H NMR (300 MHz, CHLOROFORM-d) δ 6.86 (d, $J = 8.10$ Hz,

2H), 6.72 - 6.79 (m, 1H), 6.70 (d, *J* = 1.79 Hz, 1H), 6.59 (s, 1H), 6.35 (s, 1H), 3.89 (s, 3H), 3.87 (s, 3H), 3.86 (s, 3H), 3.77 (s, 3H), 3.58 - 3.65 (m, 1H), 3.32 - 3.46 (m, 1H), 3.05 - 3.31 (m, 3H), 2.78 - 3.03 (m, 4H), 2.59 - 2.74 (m, 1H), 2.44 - 2.58 (m, 1H), 2.12 - 2.22 (m, 6H), 2.10 - 2.24 (m, 2H), 1.35 - 1.53 (m, 2H). *m/z* 486 (M+H).

2-(2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}acetamido)acetate (62)—Prepared in 55% yield. ¹H NMR (300 MHz, CHLOROFORM-d) δ 7.27 (d, *J* = 2.35 Hz, 1H), 6.94 (br. s., 1H), 6.73 - 6.86 (m, 2H), 6.59 (d, *J* = 2.17 Hz, 1H), 6.41 (d, *J* = 2.07 Hz, 1H), 4.06 (s, 1H), 3.84 - 3.91 (m, 8H), 3.75 - 3.83 (m, 5H), 3.68 - 3.75 (m, 3H), 3.38 - 3.57 (m, 2H), 3.11 - 3.33 (m, 2H), 2.82 - 3.00 (m, 4H), 2.46 - 2.59 (m, 1H). *m/z* 473 (M+H).

2-(2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}acetamido)acetic acid—Ester (60 mg, 0.127 mmol) was dissolved in methanol (5 mL) and 2N sodium hydroxide solution (0.25 mL, 0.508 mmol) was added. The reaction was stirred at RT overnight. The pH was adjusted to 7-8 with 2N HCl, then all solvent was removed under reduced pressure. The solids were redissolved as far as possible in methanol, the solids were removed by decantation, then the solvents were removed under reduced pressure to give the acid as a white solid $(93 \text{ mg}, 100\%)$. ¹H NMR (300 MHz, METHANOL-d4) δ 6.78 - 6.96 (m, 3H), 6.69 (s, 1H), 6.51 (s, 1H), 3.77 - 3.86 (m, 9H), 3.63 - 3.77 (m, 6H), 3.46 (d, *J* = 5.18 Hz, 1H), 3.17 - 3.28 (m, 2H), 2.83 - 3.04 (m, 4H), 2.47 - 2.61 (m, 1H).

2-(2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}acetamido)-N-methylacetamide (63)—The acid (30 mg, 0.065 mmol), methylamine hydrochloride (9 mg, 0.131 mmol) and BOP (38 mg, 0.085 mmol) were combined in dry DMF (1 mL) and diisopropylethylamine (42 mg, 57 μL, 0.327 mmol) was added. The reaction was stirred at RT overnight and then diluted with EtOAc. It was washed with NaHCO₃ solution and brine, dried over $MgSO₄$ and the solvents were removed under reduced pressure. The crude was purified by chromatography on silica $(0-50\% \text{ CMA-80 in EtOAc})$ to give the desired amide $(13 \text{ mg}, 42 \text{ %})$. ¹H NMR $(300 \text{ MHz},$ METHANOL-d4) δ 6.80 - 6.96 (m, 3H), 6.69 (s, 1H), 6.62 (s, 1H), 3.78 - 3.85 (m, 10H), 3.75 (s, 3H), 3.60 - 3.73 (m, 3H), 3.42 - 3.59 (m, 1H), 3.01 - 3.09 (m, 1H), 2.84 - 2.99 (m, 6H), 2.72 (s, 3H), 2.53 (dd, *J* = 5.09, 15.73 Hz, 1H). *m/z* 472 (M+H).

N-(carbamoylmethyl)-2-{1-[(3,4-dimethoxyphenyl)methyl]-6,7-

dimethoxy-1,2,3,4-tetrahydroisoquinolin-2-yl}acetamide (64)—Prepared from acid using ammonium chloride in 23% yield. ¹H NMR (300 MHz, METHANOL-d₄) δ 6.79 -6.95 (m, 3H), 6.69 (s, 1H), 6.57 - 6.61 (m, 1H), 3.83 (s, 3H), 3.82 (s, 3H), 3.80 (s, 3H), 3.73 (s, 3H), 3.65 - 3.76 (m, 2H), 3.43 - 3.57 (m, 1H), 3.26 - 3.37 (m, 2H), 3.01 - 3.11 (m, 1H), 2.84 - 3.00 (m, 4H), 2.53 (dd, *J* = 5.04, 15.49 Hz, 1H). *m/z* 458 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6-methoxy-7-propoxy-1,2,3,4 tetrahydroisoquinolin-2-yl}-N-(pyridin-2-ylmethyl)acetamide (69)—Prepared in 52% yield. ¹H NMR (300 MHz, CHLOROFORM-d) δ 8.52 (d, J = 4.80 Hz, 1H), 7.52 - 7.69 (m, 2H), 7.13 - 7.21 (m, 1H), 7.08 (d, J = 7.82 Hz, 1H), 6.62 - 6.74 (m, 3H), 6.59 (s, 1H),

6.33 (s, 1H), 4.53 (dd, J = 6.69, 16.20 Hz, 1H), 4.16 (dd, J = 5.27, 16.20 Hz, 1H), 3.85 (s, 3H), 3.77 (s, 3H), 3.72 (s, 3H), 3.66 - 3.83 (m, 3H), 3.17 - 3.51 (m, 3H), 2.81 - 3.09 (m, 4H), 2.46 - 2.59 (m, 1H), 1.73 - 1.88 (m, 2H), 1.01 (t, J = 7.39 Hz, 3H). m/z 520 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6-methoxy-7-propoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-N-(pyridin-3-ylmethyl)acetamide (70)—Prepared in 43% yield. 1H NMR (300 MHz, CHLOROFORM-d) δ 8.45 - 8.52 (m, 1H), 8.33 (s, 1H), 7.43 (d, *J* = 7.82 Hz, 1H), 7.22 (dd, *J* = 4.85, 7.77 Hz, 1H), 6.89 - 6.99 (m, 1H), 6.63 - 6.78 (m, 3H), 6.59 (s, 1H), 6.53 (s, 1H), 4.40 (dd, *J* = 8.01, 15.26 Hz, 1H), 3.93 (t, *J* = 6.88 Hz, 2H), 3.85 (s, 3H), 3.83 (s, 3H), 3.76 (s, 3H), 3.54 - 3.70 (m, 2H), 3.37 - 3.54 (m, 1H), 3.09 - 3.33 (m, 2H), 2.80 - 2.99 (m, 4H), 2.42 - 2.55 (m, 1H), 1.79 - 1.93 (m, 2H), 1.05 (t, *J* = 7.44 Hz, 3H). *m/z* 520 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6-methoxy-7-(2,2,2 trifluoroethoxy)-1,2,3,4-tetrahydroisoquinolin-2-yl}-N-(pyridin-2 ylmethyl)acetamide (71)—Prepared in 31% yield. ¹H NMR (300 MHz,

CHLOROFORM-d) δ 8.52 (d, *J* = 3.96 Hz, 1H), 7.63 (t, *J* = 7.63 Hz, 1H), 7.51 (br. s., 1H), 7.13 - 7.22 (m, 1H), 7.07 (d, *J* = 7.63 Hz, 1H), 6.55 - 6.73 (m, 4H), 6.50 (s, 1H), 4.50 (dd, *J* $= 6.55, 15.97$ Hz, 1H), $4.08 - 4.35$ (m, 3H), 3.86 (s, 3H), 3.77 (s, 3H), 3.72 (s, 3H), 3.66 -3.75 (m, 1H), 3.14 - 3.49 (m, 3H), 2.78 - 3.09 (m, 4H), 2.47 - 2.62 (m, 1H). *m/z* 560 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-6-methoxy-7-(2,2,2 trifluoroethoxy)-1,2,3,4-tetrahydroisoquinolin-2-yl}-N-(pyridin-3 ylmethyl)acetamide (72)—Prepared in 29% yield. ¹H NMR (300 MHz, CHLOROFORM-d) δ 8.49 (d, *J* = 4.71 Hz, 1H), 8.32 (s, 1H), 7.42 (d, *J* = 7.82 Hz, 1H), 7.22 (dd, *J* = 4.85, 7.68 Hz, 1H), 6.81 - 6.92 (m, 1H), 6.62 - 6.77 (m, 5H), 4.24 - 4.49 (m,

3H), 3.86 (s, 3H), 3.84 (s, 3H), 3.76 (s, 3H), 3.54 - 3.66 (m, 2H), 3.38 - 3.53 (m, 1H), 3.07 - 3.34 (m, 2H), 2.77 - 3.02 (m, 4H), 2.45 - 2.61 (m, 1H). *m/z* 560 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-7-ethoxy-6-methoxy-1,2,3,4-

tetrahydroisoquinolin-2-yl}-N-(pyridin-2-ylmethyl)acetamide (73)—Prepared in 36% yield. ¹H NMR (300 MHz, CHLOROFORM-d) δ 8.49 - 8.55 (m, 1H), 7.63 (dt, J = 1.84, 7.70 Hz, 1H), 7.55 (t, J = 5.79 Hz, 1H), 7.16 (dd, J = 5.32, 7.02 Hz, 1H), 7.08 (d, J = 7.82 Hz, 1H), 6.61 - 6.72 (m, 3H), 6.59 (s, 1H), 6.34 (s, 1H), 4.53 (dd, J = 6.69, 16.20 Hz, 1H), 4.14 (dd, J = 5.27, 16.20 Hz, 1H), 3.88 - 4.00 (m, 2H), 3.85 (s, 3H), 3.77 (s, 3H), 3.72 (s, 3H), 3.65 - 3.70 (m, 1H), 3.17 - 3.48 (m, 3H), 2.81 - 3.08 (m, 4H), 2.46 - 2.57 (m, 1H), 1.41 (t, $J = 6.97$ Hz, 2H). m/z 506 (M+H).

2-{1-[(3,4-dimethoxyphenyl)methyl]-7-ethoxy-6-methoxy-1,2,3,4 tetrahydroisoquinolin-2-yl}-N-(pyridin-3-ylmethyl)acetamide (74)—Prepared in

41% yield. ¹H NMR (300 MHz, CHLOROFORM-d) δ 8.49 (dd, J = 1.55, 4.76 Hz, 1H), 8.33 $(d, J = 1.70 \text{ Hz}, 1H), 7.43 \text{ (td, } J = 1.93, 7.82 \text{ Hz}, 1H), 7.18 - 7.25 \text{ (m, 1H)}, 6.93 \text{ (dd, } J = 5.23,$ 7.58 Hz, 1H), 6.71 - 6.78 (m, 2H), 6.64 - 6.70 (m, 1H), 6.60 (s, 1H), 6.54 (s, 1H), 4.40 (dd, J $= 8.01, 15.26$ Hz, 1H), 4.06 (q, J = 7.00 Hz, 2H), 3.86 (s, 3H), 3.83 (s, 3H), 3.76 (s, 3H), 3.55 - 3.68 (m, 2H), 3.38 - 3.54 (m, 1H), 3.10 - 3.33 (m, 2H), 2.77 - 3.01 (m, 4H), 2.43 - 2.56 (m, 1H), 1.47 (t, J = 6.97 Hz, 3H). m/z 506 (M+H).

Functional determinations

Activity of the target compounds at the OX_1 and OX_2 receptors was assayed using RD-HGA16 cells (Molecular Devices), a CHO cell line stably over-expressing the promiscuous Gq-protein Ga16. Two individual cell lines were used that stably express either OX_1 or OX_2 receptors. Cells were cultured overnight in black, clear bottom, 96-well tissue culture plates

at 25,000 cells per well in 100 μL/well Ham's F12 medium supplemented with 10% FBS, 1x pen/strep, 0.1 μg/mL Normocin, 400 μg/mL Geneticin, and 200 μg/mL Hygromycin. Medium was then removed and cells were loaded with $200 \mu L/well$ of a calcium sensitive fluorescent dye (calcium 5, Molecular Devices) in assay buffer (1x HBSS, 20 μM HEPES plus 2.7 mM probenecid) as per manufacturer's instructions for 45 min. at 37 °C. Cells were then pretreated by addition of $25 \mu L/well 10\%$ DMSO (for intrinsic activity measurements) or 25 μL 10% DMSO including 10x final concentration of test compound (for antagonist assays) for 15 min. at 37 °C. Activity was then assayed on a FlexStation II at 37 °C. Baseline fluorescence was measured every 1.52 seconds for 19 seconds at which time 25 μl of 100μ M test compound (for intrinsic activity) or an 8-point half-log concentration curve of Orexin A (for antagonist activity) was added by the FlexStation II. Fluorescence was measured every 1.52 seconds for another 41 seconds. Activity was measured as change in fluorescence intensity by setting the average baseline value of each well to zero and measuring peak fluorescence intensity following compound addition. EC_{50} values were calculated for orexin A (EC_{50}^-) and orexin A + test compound (EC_{50}^+), and these were used to calculate the test compound Ke. The concentration/response data were fit to a threeparameter logistic equation using GraphPad Prism (v5 for Windows; GraphPad Software, San Diego, CA) to calculate the EC_{50} values. For OX_1 , at least two different concentrations of test compound were used for these experiments, and these were chosen such that they caused a 4-fold or greater rightward shift in the orexin A EC_{50} . For OX_{2} , test compounds were run at 10 μ M due to the low potency of these compounds at OX₂. The Ke was calculated from the formula $Ke = [L]/((EC_{50}/EC_{50}^+)$ -1), where [L] equals the test compound concentration, EC_{50} ⁻ equals the EC_{50} of orexin A alone, and EC_{50} ⁺ equals the $EC₅₀$ of orexin A in the presence of test compound. The data represent the mean from at least three independent experiments.

Conformational Libraries and 3D-Pharmacophores

Initial 3D geometries of the ligands were generated from SMILES strings using CORINA (Molecular Networks, Erlangen, Germany). Conformational libraries were generated using 12 Genetic Algorithm (GA) and Energy Minimization runs, each based on a distinct random number seed, employing BALLOON as the GA-engine.⁴⁷ An initial population size of 200 was employed for every GA run, each consisting of 200 generations. The energy (fitness) function was evaluated using the full MMFF94 forcefield and a continuum dielectric of 80 for dielectric response screening electrostatics. The GA runs for each ligand were then pooled and unique conformers retained. For the purposes of this study, a unique conformer was defined as any structure differing by less than 30 degrees for any torsion (excluding those terminating in hydrogens).

3D-pharmacophores were developed using MOLMOD.^{48, 49} This code takes as input 1) pharmacophore hypotheses, 2) conformational libraries for each of the ligands, 3) distance and tolerance criteria for each set of pharmacophore points defining pharmacophore metric compliance and 4) an energy window for consideration of conformations from each library. Given the relative homogeneity of the present study chemotype, the objective of pharmacophore analysis was primarily the development of an overlap rule for those structural commonalities in the scaffold common to all ligands, thus enabling property evaluation and multivariate statistical analysis (2D and 3D QSAR) of the pharmacophore compliant conformations. Should one or more pharmacophores be found, MOLMOD outputs both distance metrics for each and a quaternion least squares superposition of conformations of each of the ligands complying with the pharmacophore.

Property Evaluation: Properties of both whole ligand and fragments (substituents) were evaluated for each of the conformations complying with the pharmacophore(s) using semiempirical QM (MOPAC) models. Post-computation analysis of results was done with

GRAPHA, a utility which extracts QM-energetic, electrostatic, hydrophobic, frontier-orbital energetic, Sterimol, shape, polar-non-polar surface area, volume and thermodynamic descriptors.

Fragment/2D/3D QSAR: Properties compiled were analyzed to develop quantitative structure relationships in an effort to ascertain the key physiochemical properties modulating orexin Ke values as a function of substitution. Multivariate least squares approaches embodied in the R PROJECT software for Statistical Computing ([http://www.r-project.org/\)](http://www.r-project.org/) and Tripos Sybyl were employed.

Conditioned Place Preference

Subjects

Adult Sprague-Dawley rats (Harlan, Indianapolis, IN) were housed individually on a 12/12 h light/dark cycle with free access to water and food except during experimental sessions. Testing occurred during the light period. Animal maintenance and experiments were done in accordance with the Institutional Animal Care and Use Committee, University at Buffalo, and the 2010 Guide for the Care and Use of Laboratory Animals (Institute of Laboratory Animal Resources on Life Sciences, National Research Council, National Academy of Sciences, Washington DC).

Apparatus

The standard experimental chamber for automated assessment of CPP (LE890, Harvard Apparatus, Holliston, MA) consisted of two Perspex compartments of the same size (30 cm \times 30 cm \times 34 cm) interconnected by a central corridor (8 cm \times 10 cm \times 34 cm). Both side compartments were equipped with a grey Perspex guillotine door that separated the end compartment from the central grey corridor. Each compartment had distinct, yet neutral, visual and tactile cues: One had black walls and a white rough floor, the other had white walls and a black smooth floor. The corridor had grey walls and grey floor. Time spent in each compartment was monitored by a video camera located on the ceiling and controlled by video-tracking software (Smart Junior, Harvard Apparatus, Holliston, MA).

Procedure

Details were described previously (Thorn et al., 2012). Briefly, the experiment started with a 30-min pretest session during which rats had free access to both compartments to verify the absence of preference for either. Only the first 15 min (900 sec) of the pretest session was recorded and analyzed. Rats spending more than 75 % (> 675 sec) or less than 25 % (< 225 sec) of the total time in a compartment were eliminated from further testing (only 1 rat was excluded from the studies). While this was an unbiased setup, individual rats somewhat preferred one side over the other. On average individual rats spent 58% of total pretest time in one chamber and 38% in the other chamber with the remaining 4% being spent in the central corridor. Following the pre-test phase, rats underwent place conditioning across 8 days, with alternating treatment-compartment pairings. Rats received vehicle-compartment pairing on odd days and drug (cocaine, compound **72** or cocaine + compound **72**) compartment pairing on even days. For a conditioning session, vehicle or drug was administered, and the rat was placed immediately in the paired compartment with no access to the other compartment for 30 min. This conditioning time was chosen because it was used in other studies and was established in our laboratory.50 Drug and compartment pairing were counterbalanced. On the test day (24 hours after last conditioning session), rats were randomly placed into one compartment without treatment and had access to both compartments during the 15-min observation period, and time spent in each compartment was recorded. For the combination group, compound **72** was administered 10 min before

cocaine. This pretreatment time was chosen because initial studies found that 10 min was adequate for compound **72** to be behaviorally active in a drug discrimination assay (data not shown).

Data Analyses

The magnitude of place preference was presented as the preference score, which was defined as time spent in the drug-associated compartment on the test day minus time spent in the drug-associated compartment on pretest day. Data were submitted to one-way analysis of variance with treatment (four groups) as the factor, followed by Bonferroni's post hoc test. An effect was considered significant if $p < .05$.

Drugs

Cocaine hydrochloride (Research Technology Branch, National Institute on Drug Abuse, Rockville, MD) and compound **72** were dissolved in 0.9 % physiological saline and administered i.p. Dose was expressed as milligrams of the form indicated above per kilogram of body weight (mg/kg). Injection volume was 1 mL/kg.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

We are grateful to the National Institute of Health/National Institute on Drug Abuse (grants DA032837 and DA026582 to YZ, DA034806 to JXL) for the financial support of this research. We thank Ms. Tiffany Langston, Mr. Keith Warner, Ms. Allyson Smith, Dr. Angela Giddings and Dr. Ann Decker for their valuable technical assistance. We also thank Dr. Scott Runyon and Dr. Gerald Pollard for their critical reading and editing of the manuscript.

ABBREVIATIONS

References

- 1. de Lecea L, Kilduff TS, Peyron C, Gao X, Foye PE, Danielson PE, Fukuhara C, Battenberg EL, Gautvik VT, Bartlett FS 2nd, Frankel WN, van den Pol AN, Bloom FE, Gautvik KM, Sutcliffe JG. The hypocretins: hypothalamus-specific peptides with neuroexcitatory activity. Proc Natl Acad Sci U S A. 1998; 95:322–327. [PubMed: 9419374]
- 2. Sakurai T, Amemiya A, Ishii M, Matsuzaki I, Chemelli RM, Tanaka H, Williams SC, Richardson JA, Kozlowski GP, Wilson S, Arch JR, Buckingham RE, Haynes AC, Carr SA, Annan RS, McNulty DE, Liu WS, Terrett JA, Elshourbagy NA, Bergsma DJ, Yanagisawa M. Orexins and orexin receptors: a family of hypothalamic neuropeptides and G protein-coupled receptors that regulate feeding behavior. Cell. 1998; 92:573–585. [PubMed: 9491897]
- 3. Gotter AL, Webber AL, Coleman PJ, Renger JJ, Winrow CJ. International Union of Basic and Clinical Pharmacology. LXXXVI. Orexin receptor function, nomenclature and pharmacology. Pharmacol Rev. 2012; 64:389–420. [PubMed: 22759794]
- 4. Kukkonen JP. Physiology of the orexinergic/hypocretinergic system: a revisit in 2012. Am J Physiol Cell Physiol. 2013; 304:C2–32. [PubMed: 23034387]
- 5. Date Y, Ueta Y, Yamashita H, Yamaguchi H, Matsukura S, Kangawa K, Sakurai T, Yanagisawa M, Nakazato M. Orexins, orexigenic hypothalamic peptides, interact with autonomic, neuroendocrine and neuroregulatory systems. Proc Natl Acad Sci U S A. 1999; 96:748–753. [PubMed: 9892705]
- 6. Nambu T, Sakurai T, Mizukami K, Hosoya Y, Yanagisawa M, Goto K. Distribution of orexin neurons in the adult rat brain. Brain Res. 1999; 827:243–260. [PubMed: 10320718]
- 7. Peyron C, Tighe DK, van den Pol AN, de Lecea L, Heller HC, Sutcliffe JG, Kilduff TS. Neurons containing hypocretin (orexin) project to multiple neuronal systems. J Neurosci. 1998; 18:9996– 10015. [PubMed: 9822755]
- 8. Sakurai T. Roles of orexin/hypocretin in regulation of sleep/wakefulness and energy homeostasis. Sleep Med Rev. 2005; 9:231–241. [PubMed: 15961331]
- 9. Sutcliffe JG, de Lecea L. The hypocretins: setting the arousal threshold. Nat Rev Neurosci. 2002; 3:339–349. [PubMed: 11988773]
- 10. Marcus JN, Aschkenasi CJ, Lee CE, Chemelli RM, Saper CB, Yanagisawa M, Elmquist JK. Differential expression of orexin receptors 1 and 2 in the rat brain. J Comp Neurol. 2001; 435:6– 25. [PubMed: 11370008]
- 11. Trivedi P, Yu H, MacNeil DJ, Van der Ploeg LH, Guan XM. Distribution of orexin receptor mRNA in the rat brain. FEBS Lett. 1998; 438:71–75. [PubMed: 9821961]
- 12. Scammell TE, Winrow CJ. Orexin receptors: pharmacology and therapeutic opportunities. Annu Rev Pharmacol Toxicol. 2011; 51:243–266. [PubMed: 21034217]
- 13. Kilduff TS, Peyron C. The hypocretin/orexin ligand-receptor system: implications for sleep and sleep disorders. Trends Neurosci. 2000; 23:359–365. [PubMed: 10906799]
- 14. Ohno K, Sakurai T. Orexin neuronal circuitry: role in the regulation of sleep and wakefulness. Front Neuroendocrinol. 2008; 29:70–87. [PubMed: 17910982]
- 15. Boutrel B, de Lecea L. Addiction and arousal: the hypocretin connection. Physiol Behav. 2008; 93:947–951. [PubMed: 18262574]
- 16. Harris GC, Wimmer M, Aston-Jones G. A role for lateral hypothalamic orexin neurons in reward seeking. Nature. 2005; 437:556–559. [PubMed: 16100511]
- 17. Tsujino N, Sakurai T. Orexin/hypocretin: a neuropeptide at the interface of sleep, energy homeostasis, and reward system. Pharmacol Rev. 2009; 61:162–176. [PubMed: 19549926]
- 18. Harris GC, Aston-Jones G. Arousal and reward: a dichotomy in orexin function. Trends Neurosci. 2006; 29:571–577. [PubMed: 16904760]
- 19. Boutrel B, Kenny PJ, Specio SE, Martin-Fardon R, Markou A, Koob GF, de Lecea L. Role for hypocretin in mediating stress-induced reinstatement of cocaine-seeking behavior. Proc Natl Acad Sci U S A. 2005; 102:19168–19173. [PubMed: 16357203]
- 20. Narita M, Nagumo Y, Hashimoto S, Narita M, Khotib J, Miyatake M, Sakurai T, Yanagisawa M, Nakamachi T, Shioda S, Suzuki T. Direct involvement of orexinergic systems in the activation of the mesolimbic dopamine pathway and related behaviors induced by morphine. J Neurosci. 2006; 26:398–405. [PubMed: 16407535]

- 21. Hara J, Yanagisawa M, Sakurai T. Difference in obesity phenotype between orexin-knockout mice and orexin neuron-deficient mice with same genetic background and environmental conditions. Neurosci Lett. 2005; 380:239–242. [PubMed: 15862893]
- 22. Lawrence AJ, Cowen MS, Yang HJ, Chen F, Oldfield B. The orexin system regulates alcoholseeking in rats. Br J Pharmacol. 2006; 148:752–759. [PubMed: 16751790]
- 23. Aston-Jones G, Smith RJ, Moorman DE, Richardson KA. Role of lateral hypothalamic orexin neurons in reward processing and addiction. Neuropharmacology. 2009; 56(Suppl 1):112–121. [PubMed: 18655797]
- 24. Richards JK, Simms JA, Steensland P, Taha SA, Borgland SL, Bonci A, Bartlett SE. Inhibition of orexin-1/hypocretin-1 receptors inhibits yohimbine-induced reinstatement of ethanol and sucrose seeking in Long-Evans rats. Psychopharmacology (Berl). 2008; 199:109–117. [PubMed: 18470506]
- 25. Hollander JA, Lu Q, Cameron MD, Kamenecka TM, Kenny PJ. Insular hypocretin transmission regulates nicotine reward. Proc Natl Acad Sci U S A. 2008; 105:19480–19485. [PubMed: 19033203]
- 26. Boss C, Brisbare-Roch C, Jenck F. Biomedical application of orexin/hypocretin receptor ligands in neuroscience. J Med Chem. 2009; 52:891–903. [PubMed: 19199652]
- 27. Roecker AJ, Coleman PJ. Orexin receptor antagonists: medicinal chemistry and therapeutic potential. Curr Top Med Chem. 2008; 8:977–987. [PubMed: 18673167]
- 28. Gatfield J, Brisbare-Roch C, Jenck F, Boss C. Orexin receptor antagonists: a new concept in CNS disorders? ChemMedChem. 2010; 5:1197–1214. [PubMed: 20544785]
- 29. Kodadek T, Cai D. Chemistry and biology of orexin signaling. Mol Biosyst. 2010; 6:1366–1375. [PubMed: 20532299]
- 30. Smart D, Sabido-David C, Brough SJ, Jewitt F, Johns A, Porter RA, Jerman JC. SB-334867-A: the first selective orexin-1 receptor antagonist. Br J Pharmacol. 2001; 132:1179–1182. [PubMed: 11250867]
- 31. Porter RA, Chan WN, Coulton S, Johns A, Hadley MS, Widdowson K, Jerman JC, Brough SJ, Coldwell M, Smart D, Jewitt F, Jeffrey P, Austin N. 1,3-Biarylureas as selective non-peptide antagonists of the orexin-1 receptor. Bioorg Med Chem Lett. 2001; 11:1907–1910. [PubMed: 11459658]
- 32. Perrey DA, Gilmour BP, Runyon SP, Thomas BF, Zhang Y. Diaryl urea analogues of SB-334867 as orexin-1 receptor antagonists. Bioorg Med Chem Lett. 2011; 21:2980–2985. [PubMed: 21478014]
- 33. McElhinny CJ Jr. Lewin AH, Mascarella SW, Runyon S, Brieaddy L, Carroll FI. Hydrolytic instability of the important orexin 1 receptor antagonist SB-334867: possible confounding effects on in vivo and in vitro studies. Bioorg Med Chem Lett. 2012; 22:6661–6664. [PubMed: 23031594]
- 34. Langmead CJ, Jerman JC, Brough SJ, Scott C, Porter RA, Herdon HJ. Characterisation of the binding of [3H]-SB-674042, a novel nonpeptide antagonist, to the human orexin-1 receptor. Br J Pharmacol. 2004; 141:340–346. [PubMed: 14691055]
- 35. Jiang R, Song X, Bali P, Smith A, Bayona CR, Lin L, Cameron MD, McDonald PH, Kenny PJ, Kamenecka TM. Disubstituted piperidines as potent orexin (hypocretin) receptor antagonists. Bioorg Med Chem Lett. 2012; 22:3890–3894. [PubMed: 22617492]
- 36. Stasi, PL.; Rovati, L. Spiro amino compounds suitable for the treatment of inter alia sleep disorders and drug addiction. 2011. WO2011006960
- 37. Steiner MA, Gatfield J, Brisbare-Roch C, Dietrich H, Treiber A, Jenck F, Boss C. Discovery and Characterization of ACT-335827, an Orally Available, Brain Penetrant Orexin Receptor Type 1 Selective Antagonist. ChemMedChem. 2013; 8:898–903. [PubMed: 23589487]
- 38. Hirose M, Egashira S, Goto Y, Hashihayata T, Ohtake N, Iwaasa H, Hata M, Fukami T, Kanatani A, Yamada K. N-acyl 6,7-dimethoxy-1,2,3,4-tetrahydroisoquinoline: the first orexin-2 receptor selective non-peptidic antagonist. Bioorg Med Chem Lett. 2003; 13:4497–4499. [PubMed: 14643355]

- 39. Koberstein R, Aissaoui H, Bur D, Clozel M, Fischli W, Jenck F, Mueller C, Nayler O, Sifferlen T, Treiber A, Weller T. Tetrahydroisoquinolines as orexin receptor antagonists: Strategies for lead optimization by solution-phase chemistry. Chimia. 2003; 57:270–275.
- 40. Rice KC, Brossi A. Expedient Synthesis of Racemic and Optically-Active NNorreticuline and N-Substituted and 6'-Bromo-N-Norreticulines. Journal of Organic Chemistry. 1980; 45:592–601.
- 41. Nilsson BL, Kiessling LL, Raines RT. Staudinger ligation: A peptide from a thioester and azide. Organic Letters. 2000; 2:1939–1941. [PubMed: 10891196]
- 42. Caddick S, Afonso CAM, Candeias SX, Hitchcock PB, Jenkins K, Murtagh L, Pardoe D, Santos AG, Treweeke NR, Weaving R. Synthesis of alpha-amino esters by dynamic kinetic resolution of alpha-haloacyl imidazolidinones. Tetrahedron. 2001; 57:6589–6605.
- 43. Neubauer DN. Almorexant, a dual orexin receptor antagonist for the treatment of insomnia. Curr Opin Investig Drugs. 2010; 11:101–110.
- 44. Verloop A, Tipker J. QSAR in Drug Design and Toxicology. Elsevier. 97:1987.
- 45. Verloop, A. The STERIMOL Approach to Drug Design. Marcel Dekker; New York: 1987.
- 46. Kantola A, Villar HO, Loew GH. Atom Based Parametrization for a Conformationally Dependent Hydrophobic Index. Journal of Computational Chemistry. 1991; 12:681–689.
- 47. Vainio MJ, Johnson MS. Generating conformer ensembles using a multiobjective genetic algorithm. J Chem Inf Model. 2007; 47:2462–2474. [PubMed: 17892278]
- 48. Harris D, Clayton T, Cook J, Sahbaie P, Halliwell RF, Furtmüller R, Huck S, Sieghart W, DeLorey TM. Selective influence on contextual memory: Physiochemical properties associated with selectivity of benzodiazepine ligands at GABAA receptors containing the a5 subunit. J. Med. Chem. 2008; 51:3788–3803. [PubMed: 18537233]
- 49. Harris DL, DeLorey TM, He X, Cook JM, Loew GH. Determinants of recognition of ligands binding to benzodiazepine receptor/GABA(A) receptors initiating sedation. Eur J Pharmacol. 2000; 401:271–287. [PubMed: 10936485]
- 50. Thorn DA, An XF, Zhang Y, Pigini M, Li JX. Characterization of the hypothermic effects of imidazoline I(2) receptor agonists in rats. Br J Pharmacol. 2012; 166:1936–1945. [PubMed: 22324428]

Perrey et al. Page 26

Figure 1. Orexin antagonists.

Perrey et al. Page 27

Figure 2.

A) Ligands superimposed at 3D-pharmacophore points defined by B) the abstract pharmacophore representation computed from 21 ligands in Table 1. C) a plot of the predicted $ln Ke(OX_1)$ vs. experimental for a fragment (2D) QSAR illustrating that contributions from the width (Verloop Sterimol "B1"), polar surface area, ClogP and lowest unoccupied (LUMO) energy of the substituent give a robust analysis (r2=0.7/F=8.2 (n1=4,n2=18), p=0.001/ 2.8B1+0.7*E(LUMO)-0.13logP-5.5)). D) 3D-COMFA/QSAR illustration of the increased steric bulk contributions region of the substituent variation highlighted by green correlate with low Ke values with minor modulation due to

electrostatic contributions. E) a depiction of the location of the LUMO density on compound **6**.

Perrey et al. Page 29

Figure 3.

Depiction of COMFA steric (left) and electrostatic fields (right) about compound **6** derived from the analysis of **6, 20, 21, 23, 30, 41, 44, 47, 50, 51, 52, 55** and **60**. The analysis $(R2=0.761/F[n1=1, n2=11)]$ 35.108, SE=0.61) for lnKe(OX₁) using the 3D-pharmacophore quaternion least squares superposition of pharmacophore points illustrated in Figure 2.

Figure 4.

Effects of compound **72** on cocaine-induced conditioned place preference in rats (n=8-9). Data presented as mean \pm S.E.M. * P < 0.05 as compared to vehicle treated rats. P < 0.05 as compared to 10 mg/kg cocaine treated rats.

^aReagents and Conditions: (a) HBTU, iPr₂EtN, DMF; (b) (i) POCl₃, toluene, 90 °C; (ii) NaBH₄, MeOH; (c) BrCH₂CONHBn, iPr₂EtN, Bu₄NI, DMF; (d) R₁-Br, K₂CO₃, Bu₄NI, DMF; (e) R₁-I, Cs₂CO₃, DMF, 50 °C; (f) R-SO₂Cl, Et₃N, CH₂Cl₂ (R₁=R-SO₂).

Scheme 1.

Synthesis of 6-Methoxy-7-Alkoxy-N-benzylacetamidotetrahydroisoquinolines^a

^aReagents and Conditions: (a) BrCH₂-CO₂Et, iPr₂EtN, Bu₄NI, DMF; (b) (i) 2N NaOH, EtOH; (ii) R_2 -NH₂ (or R_2 -NHMe), BOP, iPr₂EtN, DMF or BnBr, K₂CO₃, Bu₄NI, DMF; (c) (i) Raney nickel, $NH_2NH_2\bullet H_2O$, EtOH; (ii) R_3 -CO₂H and BOP, iPr₂EtN, CH₂Cl₂ or R_3CO_2O , iPr₂EtN, CH₂Cl₂ or R₃NCO, toluene.

Scheme 2.

Synthesis of N-acetamidotetrahydroisoquinolines^a

^aReagents and Conditions: (a) BrCH₂CO₂Et, iPr₂EtN, Bu₄NI, DMF; (b) R₁-X, K₂CO₃ or Cs_2CO_3 , Bu₄NI (for X=Br), DMF; (c) (i) 2N NaOH, EtOH; (ii) R₂-NH₂, BOP, iPr₂EtN, DMF.

Scheme 3.

Synthesis of tetrahydroisoquinoline derivatives varied at both acetamide and 7 positions^a

Table 1

Effect of the 7-position substitution on OX antagonism

 a $<$ 30% inhibition at 10 μ M

b Values are the mean of at least three independent experiments in duplicate

 c Values are the mean of at least two independent experiments in duplicate; for compounds with Ke < 100 nM at OX1 at least three independent experiments in duplicate were performed.

Table 2

Effect of N-alkylation on OX antagonism

	MeO ∣ N、 $x^{\mathcal{R}}$ MeO MeO MeO						
No.	$X-R$	Ke $\left(\mathbf{OX}_{1},\mathbf{n}\mathbf{M}\right)^{\pmb{b}}$	$\mathrm{Ke}\left(\mathrm{OX}_2,nM\right)^{\mathcal{C}}$	OX_2/OX_1			
$\bf 6$		199±47	>10000	>50.3			
33	H	$3250{\pm}230$	\boldsymbol{a}				
34		2370±240	\boldsymbol{a}				
35		$3030{\pm}170$	9000 ± 760	3.0			
36		$3530{\pm}650$	9030 ± 3800	$2.6\,$			
$\bf 37$		>10000	\boldsymbol{a}				
38	Ν	>10000	\boldsymbol{a}				
39		2090±330	$>\!\!10000$	>4.8			

J Med Chem. Author manuscript; available in PMC 2013 December 04.

Perrey et al. Page 36

NIH-PA Author Manuscript NIH-PA Author Manuscript

J Med Chem. Author manuscript; available in PMC 2013 December 04.

Perrey et al. Page 37

NIH-PA Author Manuscript NIH-PA Author Manuscript

MeO. ۱. N $x^{\sim R}$ MeO MeO MeO						
No.	$X-R$	Ke $\left(\text{OX}_1, \text{nM} \right)^b$	$\mathrm{Ke}\left(\mathrm{O}\mathrm{X}_2,\mathrm{nM}\right)^{\mathcal{C}}$	$\mathbf{OX}_{2}/\mathbf{OX}_{1}$		
47		41.1 ± 6.0	655±190	15.9		
48	NΗ	$7170{\pm}390$	\boldsymbol{a}			
49	N H	>10000	\boldsymbol{a}			
50	١	$80.0{\pm}12$	>10000	>125		
51	н СI	$162{\pm}16$	$2270{\pm}110$	$14.0\,$		
52	F	279 ± 81	3460±1000	12.4		
53		$261{\pm}120$	$>\!\!10000$	>38.3		
54	Ń	152 ± 41	>10000	>65.8		

J Med Chem. Author manuscript; available in PMC 2013 December 04.

NIH-PA Author Manuscript NIH-PA Author Manuscript

 a $<$ 30% inhibition at 10 μ M

b Values are the mean of at least three independent experiments in duplicate

Table 3

Effect of multiple changes on OX antagonism

*a*Values are the mean of at least three independent experiments in duplicate

b Values are the mean of at least three independent experiments in duplicate.