



Published in final edited form as:

*Breast Cancer Res Treat.* 2013 October ; 141(3): 447–459. doi:10.1007/s10549-013-2698-4.

## Genetic variants in microRNAs and breast cancer risk in African American and European American women

Song Yao<sup>1,†,\*</sup>, Kelly Graham<sup>1,†</sup>, Jie Shen<sup>2</sup>, Lara E. Sucheston Campbell<sup>1</sup>, Prashant Singh<sup>3</sup>, Gary Zirpoli<sup>1</sup>, Michelle Roberts<sup>1</sup>, Gregory Ciupak<sup>1</sup>, Warren Davis<sup>1</sup>, Helena Hwang<sup>4</sup>, Thae Khoury<sup>4</sup>, Dana H. Bovbjerg<sup>5</sup>, Lina Jandorf<sup>6</sup>, Karen S. Pawlish<sup>8</sup>, Elisa V. Bandera<sup>9</sup>, Song Liu<sup>10</sup>, Christine B. Ambrosone<sup>1</sup>, and Hua Zhao<sup>2,\*</sup>

<sup>1</sup>Department of Cancer Prevention & Control, Roswell Park Cancer Institute, Buffalo, NY

<sup>2</sup>Department of Epidemiology, The University of Texas, M.D. Anderson Cancer Center, Houston, TX

<sup>3</sup>Department of Pharmacology & Therapeutics, Roswell Park Cancer Institute, Buffalo, NY

<sup>4</sup>Department of Pathology, Roswell Park Cancer Institute, Buffalo, NY

<sup>5</sup>University of Pittsburgh Cancer Institute, University of Pittsburgh, Pittsburgh, PA

<sup>6</sup>Department of Oncological Sciences, Mount Sinai School of Medicine, New York, NY

<sup>7</sup>Department of Medicine, Roswell Park Cancer Institute, Buffalo, NY

<sup>8</sup>New Jersey State Cancer Registry, New Jersey Department of Health, Trenton, NJ

<sup>9</sup>Rutgers Cancer Institute of New Jersey, Robert Wood Johnson Medical School, New Brunswick, NJ

<sup>10</sup>Department of Biostatistics, Roswell Park Cancer Institute, Buffalo, NY

### Abstract

MicroRNAs (miRNAs) are an integral part of the post-transcriptional machinery of gene expression and have been implicated in the carcinogenic cascade. Single nucleotide polymorphisms (SNPs) in miRNAs and risk of breast cancer have been evaluated in populations of European or Asian ancestry, but not among women of African ancestry. Here we examined 145 SNPs in 6 miRNA processing genes and in 78 miRNAs which target genes known to be important in breast cancer among 906 African American (AA) and 653 European American (EA) cases and controls enrolled in the Women's Circle of Health Study (WCHS). Allele frequencies of most SNPs (87%) differed significantly by race. We found a number of SNPs in miRNAs and processing genes in association with breast cancer overall or stratified by estrogen receptor (ER) status. Several associations were significantly different by race, with none of the associations being significant in both races. Using a polygenic risk score to combine the effects of multiple SNPs, we found significant associations with the score in each subgroup analysis. For ER-positive

\*To whom correspondence should be addressed: Tel: 716-845-4968; Fax: 716-845-8487; song.yao@roswellpark.org. Tel: 713-745-7597; hzhao2@mdanderson.org.

†The authors contributed equally to the work.

**Conflict of Interest Statement:** Dr. Kelly Graham is an employee of Susan G. Komen Foundation for the Cure.

cancer, each unit increment of the risk score was associated with a 51% increased risk in AAs (OR=1.51, 95% CI=1.30–1.74,  $p=3.3 \times 10^{-8}$ ) and a 73% increased risk in EAs (OR=1.73, 95% CI=1.45–2.06,  $p=1.4 \times 10^{-9}$ ). These data show, for the first time, that miRNA-related genetic variations may underlie the etiology of breast cancer in both populations of African and European ancestries. Future studies are needed to validate our findings and to explore the underlying mechanisms.

## Keywords

microRNA; SNP; breast cancer; epidemiology; estrogen receptor; polygenic risk score

---

## Introduction

In women, breast cancer is the most common non-skin cancer diagnosed in the United States. In 2011, an estimated 230,480 new cases of breast cancer were diagnosed in women, and an estimated 39,520 women died from the disease [1]. The incidence rates of breast cancer were higher in European American (EA) women while the death rates were higher in African American (AA) women [2]. Studies have also shown that AA women tend to develop estrogen receptor (ER) negative tumors at an earlier age, and EA women are more likely to develop ER positive tumors at later ages [3–8]. The disparities in breast cancer incidence and survival between AA and EA populations have been attributed to several factors, including disease management, access to proper care, and biological influences. A recent study of the disparity in breast cancer mortality between AA and EA women concluded that differences in mortality are driven by higher hazard rates of breast cancer death in AA women, irrespective of ER expression [9]. In that work, the authors suggest that other biological factors may play a role in breast cancer disparities [9].

MicroRNAs (miRNAs) are small, noncoding RNAs that bind to the 3' UTR of target mRNAs, and silence gene expression by inducing degradation of target mRNAs or inhibition of protein translation [10]. Because miRNAs may regulate approximately 60% of human genes [11], the relationship between miRNAs and human diseases has been extensively explored in the last decade. Many studies have demonstrated differential gene expression of miRNAs between normal and diseased tissue in cancer, and specific miRNAs have been linked to carcinogenic properties, including resistance to apoptosis, unchecked proliferation, angiogenesis, limited growth inhibition, and the propensity to invade and metastasize (reviewed in [12]). Aberrant miRNA expression patterns have been identified in breast cancer [13–15]. Of note, racial differences in miRNA expression have been observed in several studies. Son et al found that miRNA expression profiles in non-small cell lung cancer were different between Korean and Western populations [16]. In another study, a number of miRNAs were significantly differentially expressed in uterine leiomyoma between AA and EA women [17]. However, whether miRNA expression profiles are different in breast cancer tissues between AA and EA women is still unknown.

Functional genetic variations can affect gene expression or activity and thereby modify cancer risk. This might be particularly important for genetic variations in miRNA genes,

considering the fact that at least 60% of human protein-encoding genes are regulated by miRNAs. miRNAs can serve as either tumor suppressor genes or oncogenes. Genetic variations in miRNA genes can affect the levels of mature miRNAs, and consequently alter the mRNA expression levels of the target genes. Furthermore, SNPs in miRNA processing genes likely alter the production of miRNAs and SNPs in miRNA binding sites in target genes may change the interaction between miRNAs and target genes and subsequently their expression. SNPs in miRNA genes, miRNA processing genes, and binding sites in target genes have been tested in relation to the risk of several types of cancer, including breast cancer [18–21]. However, to the best of our knowledge, no previous studies have examined these relationships in women of African ancestry.

In this study, we employed a candidate gene approach for identifying SNPs in miRNA processing genes and in miRNA precursors (pre-miRNAs) that target genes known to play a role in breast cancer. We analyzed a total of 145 SNPs that present in 78 pre-miRNAs and 6 miRNA processing genes for associations with breast cancer risk in a large case-control study of AA and EA women.

## Study Population and Methods

### Study population

The Women's Circle of Health Study (WCHS) was designed specifically to study the role of genetic and non-genetic factors in relation to aggressive breast cancer risk in AA and EA women. Study design, enrollment, and collection of data and biospecimens have been described in detail previously [22]. Briefly, women diagnosed with incident breast cancer were identified through both hospital-based case ascertainment in targeted hospitals that had large referral patterns of AAs in four boroughs of the metropolitan New York City area, and using population-based case ascertainment in seven counties in New Jersey (NJ) through the NJ State Cancer Registry. The eligibility criteria for cases were: self-identified AA and EA women, 20–75 years of age at diagnosis, no previous history of cancer other than non-melanoma skin cancer, recently diagnosed with primary, histologically confirmed breast cancer, and English speaking. Controls without a history of any cancer diagnosis other than non-melanoma skin cancer living in the same area as cases were identified through random digit dialing and were matched to cases by self-reported race and 5-year age categories. To increase enrollment of AA women, particularly those with lower socioeconomic status, cases and controls were also invited to participate through community recruitment efforts as described in details previously [23]. Following agreement to participate, in-person interviews were conducted to complete informed consent and to query participants on a number of potential risk factors, including medical history, family history of cancer, diet, physical activity, and other lifestyle factors. Anthropometric measures were taken, and biospecimens were collected. Blood and/or saliva samples were collected for later extraction of DNA. Permission to obtain pathology data, including ER status, as well as tumor tissue blocks was included in the informed consent form. This study was approved by the Institutional Review Boards at Roswell Park Cancer Institute (RPCI), the Cancer Institute of New Jersey (CINJ) – Robert Wood Johnson Medical School (RWJMS), Mount Sinai School of Medicine (MSSM), and the participating hospitals in NYC. At the time of genotyping

(April 2010), DNA and data were available from 491 AA cases and 415 AA controls. We selected 336 EA cases and 317 EA controls from the WCHS by frequency matching them to AA cases and controls by 5-year age group.

### Identification of SNPs

For this study, we focused on miRNA genes that are predicted to regulate key breast cancer genes (*BRCA1/2*, *p53*, *PTEN*, *CHEK2*, *ATM*, *NBS1*, *RAD50*, *BRIP1*, *PALB2*, *ER*, *PR* and *ERBB2*). A total of 146 miRNAs fit the criteria. We searched Entrez SNP (<http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?CMD=search&DB=snp>) for SNPs in premiRNA regions of these selected miRNAs and in binding sites of target genes and identified 99 SNPs with minor allele frequencies (MAFs) greater than 0.05 in either EAs or AAs. For miRNA processing genes, including *AGO1*, *AGO4*, *DGCR8*, *XPO5*, *PACT*, and *TARBP2*, we searched an extended genomic region 15kb from both 3' and 5' ends of each gene. Genotype data were downloaded from HapMap (23) and other resequencing projects through the Genome Variation Service at Seattle SNP (<http://gvs.gs.washington.edu/GVS/>), and multi-population tagSNPs to capture variations in both populations of European and African ancestry were selected using the TAGster program [24]. In total, 154 SNPs were selected for genotyping.

### SNP genotyping

Genomic DNA extracted from blood or saliva samples was evaluated and quantified by Nanodrop UV-spectrometer (Thermo Fisher Scientific Inc., Wilmington, DE) and PicoGreen-based fluorometric assay (Molecular Probes, Invitrogen Inc., Carlsbad, CA), and stored at  $-80^{\circ}\text{C}$  until analysis. To control for potential bias due to population admixture, a panel of 108 ancestry informative markers (AIMs) that have been shown to be effective in correcting this bias in case-control studies were chosen [25]. Selected SNPs and AIMs were genotyped by Illumina GoldenGate genotyping assay (Illumina Inc., San Diego, CA) at the Genomics Facility at RPCI. Five percent duplicates and two sets of in-house trio samples were included for genotyping quality control purposes. No SNP violated Mendelian heritability. Six SNPs failed genotyping due to poor clustering or abnormal heterozygosity and were excluded. The average successful genotyping rate for each sample and each SNP was 99%. Three additional SNPs failed Hardy-Weinberg equilibrium and were excluded. As a result, a total of 145 SNPs were analyzed (Supplementary Table S1).

### Statistical analysis

STRUCTURE program was used to estimate the proportion of European ancestry for each woman based on the genotype data of AIMs. Descriptive characteristics were analyzed by student t-test or chi-square test using SAS 9.3 (SAS Institute, Cary, NC). All genotype analyses were performed for AA and EA populations separately, using PLINK program if not otherwise specified. Genotypic (co-dominant) models were assumed for SNP effects. When genotype frequency of the rare homozygote was  $\leq 5\%$  in both populations, it was collapsed with the heterozygote (dominant model) for power considerations. In addition, recessive models were also explored. To test if there was a linear dose-effect of the variant alleles, SNPs were coded as 0, 1 and 2 according to the copy number of the variant allele

and tested using log-additive genetic models. A best model was selected for each SNP by considering both sample size and direction of associations across genotypes. Univariate single SNP analysis was first performed. Covariates, including age, body mass index (BMI), proportion of European ancestry, family history of breast cancer, and education, were then adjusted in multivariate logistic regression models to derive odds ratios (ORs) and 95% confidence intervals (CIs). Multiple comparison error was controlled by 10,000 permutations for SNP analyses. To test whether the associations of SNPs with breast cancer differed between AA and EA women, modification effect by race was examined by including an interactive term between race and each SNP in the model based on all women and tested using likelihood ratio tests. In addition to overall breast cancer risk, the analyses were also stratified by ER status within each race following the same analytical approaches.

### **Polygenic risk score**

In addition to single SNP analysis, we also performed multi-marker analyses by using a modified method of the weighted polygenic risk score as described previously [26]. In brief, this multi-marker risk score was calculated as a sum of the number of risk genotypes (dominant and recessive models) and risk alleles (additive model), depending on the final model chosen for each SNP, weighted by the regression coefficients from logistic regression. For SNPs associated with a decreased risk, the reference and comparison groups were flipped so that the genotypes or alleles counted were associated with an increased risk. For a pair of SNPs located within 500kb on the same chromosome and in high linkage disequilibrium ( $r^2 > 0.8$ ), only one SNP with stronger association from the pair was selected to be included in the polygenic score. For easier interpretation, the final score was standardized by dividing the sum of regression coefficients and then multiplying the expected maximum of number of risk genotypes and alleles, therefore, each unit of the polygenic score equals to one risk genotype or allele. The score was analyzed as a continuous variable in the logistic regression model with adjustment for the same set of covariates as described above.

## **Results**

### **Description of study population**

Table 1 outlines the characteristics of the study population. AA women tended to have a higher BMI than EA women (31.4 vs 27.3 kg/m<sup>2</sup>), were less likely to use HRT (86% non-users in AAs vs. 75% non-users in EAs), and tended to have lower frequencies of family history of breast cancer (13.4% vs 21.0%), when compared to EA cases. The majority of women had college and graduate school education, but rates of women who pursued higher education were lower in AAs (57.6%) vs. EAs (82.2%). As expected, family history of breast cancer was higher in the cases than controls for both AAs and EAs, and controls were more highly educated. Among EAs, HRT use was slightly higher among cases, although the association was not statistically significant.

### **Differences in allele frequencies of SNPs between AA and EA women**

Chromosomal location and minor allele frequency (MAF) of the 145 SNPs included in the final analysis are shown in Supplementary Table S1. MAFs of 126 of the 145 SNPs (87%)

in the controls were significantly different between AA and EA women ( $p < 0.05$ ), 31 of which have the minor allele flipped between AAs and EAs.

### Associations of SNPs in miRNA genes and processing genes with breast cancer risk

Top-ranked significant associations between SNPs in miRNA genes and overall breast cancer risk are shown in Table 2 for AAs and EAs separately. Among AA women, three SNPs, including rs7354931 in *AGO4*, rs12586258 in *hsa-miR-758*, and rs2018562 in *hsa-miR-513a-2* were associated with risk of breast cancer. The most significant SNP rs12586258 was associated with an almost 40% decreased risk in a dominant genetic model (OR=0.61, 95% CI=0.42–0.89,  $p=0.01$ ). Among EA women, we identified seven SNPs, including rs2059691 in *PACT*, rs1527423 in *hsa-miR-106b*, rs1834306 in *hsa-miR-100*, rs11107973 in *hsa-miR-331*, rs10144193 in *hsa-miR-544*, rs1951032 in *hsa-miR-487*, rs5750504 in *hsa-miR-659*, that were significantly associated with breast cancer risk (Table 2). The most significant SNP rs1951032 was associated with an 81% increased risk in a dominant genetic model (OR=1.81, 95% CI=1.26–2.61,  $p=0.001$ ). However, none of the above associations remained significant after correction for multiple comparisons. Interestingly, none of the SNPs showed a significant association with breast cancer in both AA and EA populations. In fact, five out of the above eight SNPs showed differential associations between AA and EA populations with a  $p$  for interaction by race  $<0.05$  (Table 2).

### Stratified analysis by ER status

When stratified by ER status, eight SNPs were associated with ER-positive cancer risk in AAs, with only one SNP, rs2018562 in *hsa-miR-513a-2*, being previously associated with overall breast cancer risk in AAs (OR=1.51, 95% CI=1.03–2.22,  $p=0.04$ ) (Table 3). This same SNP was also associated with ER-negative cancer risk in AAs (OR=1.74, 95% CI=1.06–2.86,  $p=0.03$ ), indicating that the association was not restricted to either subtype by ER status. Among EAs, nine SNPs were associated with ER-positive cancer risk (Table 3). These include the five SNPs previously associated with overall cancer risk in EAs (rs1834306 in *hsa-miR-100*, rs11107973 in *hsa-miR-331*, rs10144193 in *hsa-miR-544*, rs1951032 in *hsa-miR-487*, and rs5750504 in *hsa-miR-659*). The risk allele of the most significant SNP, rs5750504 in *hsa-miR-659*, was associated with an increased risk (per copy of the A allele: OR=1.45, 95% CI=1.12–1.87,  $p=0.005$  in an additive genetic model). When tested for interaction by race, 3 of the 17 SNPs associated with ER-positive cancer risk in either AAs or EAs showed differential associations by race ( $p$  for interaction by race  $<0.05$ ; Table 3).

For ER-negative cancer, four SNPs, which were not found in association with overall cancer risk, were associated specifically with ER-negative cancer risk in AAs, including the most significant SNP rs107822 in *hsa-miR-219* (OR=1.99, 95% CI=1.24–3.19,  $p=0.004$ ) (Table 4). Three other SNPs were found in significant association with ER-negative breast cancer in EAs, including the most significant SNP rs2281611 in *hsa-miR-495* (OR=2.29, 95% CI=1.19–4.39,  $p=0.01$ ) (Table 4). Only 1 of the 7 SNPs, rs2281611 in *hsa-miR-495*, associated with ER-negative cancer risk in either AAs or EAs showed differential associations by race ( $p$  for interaction by race  $<0.05$ ; Table 4).



### Associations with polygenic risk score

A polygenic risk score was derived to examine the combined effects of significant single SNPs in relation to overall breast cancer risk in AA and in EA women, and stratified by ER status within each race category. The SNPs, designated risk allele or genotype, expected range of the polygenic score, mean and standard deviation of the score in cases and controls, and risk estimates per unit of the score are shown in Table 5. In each subgroup, breast cancer patients had higher polygenic risk score than controls, and per unit increment of the score was associated with significantly increased risk. The most significant results were found for ER+ cancer risk. Among AAs, per unit of the polygenic score was associated with a more than 50% increased risk of ER+ cancer risk (OR=1.51, 95% CI=1.30–1.74,  $p=3.3 \times 10^{-8}$ ). Among EAs, per unit of the polygenic score was associated with a more than 70% increased risk of ER+ cancer risk (OR=1.73, 95% CI=1.45–2.06,  $p=1.4 \times 10^{-9}$ ).

### Discussion

In this study, we aimed to examine potential relationships between miRNA genetic variants in AA and EA women in relation to breast cancer risk. To achieve this goal, we analyzed 145 SNPs in miRNAs and miRNA processing genes associated with breast cancer from 906 AA women and 653 EA women enrolled in the WCHS study. There were marked differences in allele frequencies of SNPs in miRNAs examined in this study between populations of African and European ancestry. We found a number of SNPs in miRNAs and processing genes in association with overall breast cancer risk and stratified by ER status in either EA or AA women. Given the sample size, none of the associations remained significant after controlling for multiple comparisons. Nevertheless, using a polygenic risk score to combine the number of risk alleles or genotypes weighted by their effect sizes, we found highly significant associations between the risk score and breast cancer overall and by ER status. To our knowledge, this is the first study of its kind to investigate miRNA gene variants in breast cancer in both AA and EA women.

The associations between SNPs in miRNA genes and breast cancer risk have been previously studied, with rs11614913 in *hsa-miR-196a2* and rs2910164 in *hsa-miR-146a* found to be significant [27,28,18,19,29], although the associations are not consistent across the studies [30–32]. We included both of these SNPs in our study but, unfortunately, rs2910164 in *hsa-miR-146a* was dropped out due to low genotyping quality. For rs11614913 in *hsa-miR-196a2*, no association was observed in either the EA or AA population or by ER status. Our results are consistent with the report from Catucci *et al.* In their analysis of 1,894 German and Italian breast cancer patients and 2,760 healthy women, they failed to observe a significant association between rs11614913 in *hsa-miR-196a2* and breast cancer [32]. Lack of association was also observed in a recent study in an EA case-control study in Australia [30].

Overall, we observed 10 SNPs that were significantly associated with breast cancer in either EA or AA women. None of these SNPs have been previously investigated in relation to breast cancer. Interestingly, three of these significant SNPs, rs12586258 in *hsa-miR-758*, rs10144193 in *hsa-miR-544* and rs1951032 in *hsa-miR-487*, are located in a shared miRNA cluster on chromosome 14, which may potentially be the largest tumor suppressor miRNA

cluster. In addition, a fourth SNP in the chromosome 14 miRNA cluster, rs2281611 in *hsa-miR-495*, was associated with ER negative breast cancer in EA women in our study. This cluster has been shown to be down-regulated through epigenetic alteration in ovarian tumors, hepatocellular carcinomas, and gliomas [33–36]. Further studies should be conducted to identify the potential role of this miRNA cluster in breast cancer etiology. Although all three SNPs are located in the primary miRNA regions, not the precursor or mature miRNA regions, studies have shown that genetic variations in primary miRNA regions might affect the secondary structures of the primary transcripts of miRNAs and thereby alter the production of mature miRNAs. More interestingly, the effect of these three SNPs might not be limited to the miRNAs which they are close to. Since miRNAs tend to be transcribed together (e.g. miRNAs in the chromosome 14 cluster) and then spliced into individual miRNAs, genetic variations in primary miRNA regions have the potential to modify the expression of multiple mature miRNAs originating from the same primary transcripts.

The biogenesis of miRNAs is a complex process involving multiple proteins and RNAs (10). The major players include DROSHA, DGCR8, RAN, XPO5, DICER, and AGO family members. In this study, we evaluated tagSNPs in *AGO1*, *AGO4*, *DGCR8*, *XPO5*, *PACT*, and *TARBP2* genes. We found that *AGO4* gene variants were consistently associated with breast cancer among AA women (overall and ER positive), which have not been previously reported. The main function of the AGO protein family is to act cooperatively to silence both perfectly and partially complementary target RNAs bearing multiple small RNA-binding sites. Because of the potential overlap in function among the family members (e.g. AGO1, AGO3 and AGO4), the exact role of AGO4 in miRNA function is still unclear. Notably, *AGO4* was found to be overexpressed in colon cancers with distant metastases [37]. This finding may provide some insight into the aggressive behavior of breast tumors in AA women.

There were significant interactions between SNPs and race on breast cancer risk in our analyses. It is notable that no associations were observed in both AA and EA populations. Although the significant racial difference could be due to chance because of our moderate sample size, the findings may reflect the fact that the genomic structures are quite different between EAs and AAs. In fact, 126 of 145 genotyped SNPs showed significant MAF differences between EA and AA women in the control group. These differences in associations by race are also commonly reported in the literature. For example, among 11 EA GWAS SNPs, Zheng et al found that only two SNPs (2q35 and FGFR2 loci) were associated with breast cancer in AA women [38]. Thus, our results provide an impetus to pursue more genetic susceptibility studies in the AA populations.

One limitation of our study is the lack of a validation population. Our sample size became relatively limited after stratification by race and ER status, and none of the associations remained significant after correcting for multiple comparisons. Although the associations of the polygenic risk score and breast cancer risk were highly significant in our analyses, the SNPs chosen to be included in the scores have not been validated in an independent population, which may include false positive SNPs, and the effect sizes of the SNPs may have been inflated given our sample size. Therefore, we cannot exclude the possibility of



false positive findings in our study. However, the highly significant associations with the polygenic risk scores may provide support to our hypothesis of a role of miRNA genetic variations in breast cancer etiology. AA women are more likely to be diagnosed at a younger age than EAs. We enrolled all eligible AA women, but randomly frequency matched eligible EAs by 5 year age categories. We also initially limited eligibility to women 65 years or younger because of low participation of older women without breast cancer to case-control studies. Thus, the overall study population is relatively younger than many other studies. The high proportion of premenopausal women in this study needs to be considered in relation to generalizability of our findings to populations of older age.

To the best of our knowledge, this is the first study to investigate the role of SNPs in miRNA genes and miRNA processing genes in the etiology of breast cancer in a large population with both AA and EA women. We found prevalent differences in allele frequencies in SNPs in miRNAs, and significant associations between risk of breast cancer risk and a number of SNPs in miRNA genes and in miRNA processing genes, as well as a polygenic risk score, in either AA or EA women. Some of the SNPs showed significant interactions with ancestry. Additional studies are needed to confirm the associations and explore the genetic basis and underlying molecular mechanisms of the associations.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

## Acknowledgement

This work was supported by grants from the U.S. Army Medical Research and Materiel Command (USARMMC) (DAMD-17-01-1-0334 and W81XWH-08-1-0379), the National Cancer Institute (R01 CA100598, R01 CA136483 and R25 CA114101), the Breast Cancer Research Foundation and a gift from Philip L. Hubbell family. Samples were stored and managed by the RPCI DataBank and BioRepository (DBBR) and genotyping was performed in the RPCI Genomics Core Facility, both CCSG shared resources, supported by P30 CA016056-32. The New Jersey State Cancer Registry (NJSCR) is a participant in the Centers for Disease Control and Prevention's National Program of Cancer Registries (NPCR) and is a National Cancer Institute SEER Expansion Registry. The NJSCR is supported by the Centers for Disease Control and Prevention under cooperative agreement 1U58DP00039311-01 awarded to the New Jersey Department of Health. The collection of State of New Jersey cancer incidence data is also supported by the National Cancer Institute's Surveillance, Epidemiology, and End Results (SEER) Program under contract N01-PC-2010-00027 and the State of New Jersey. The funding agents played no role in design, in the collection, analysis, and interpretation of data, in the writing of the manuscript, or in the decision to submit the manuscript for publication. We are grateful to the women who participated in this study, and to colleagues, physicians and clinical staff at participating hospitals in New York who facilitated identification and enrollment of cases into the study: Drs. Kandace Amend (i3 Drug Safety), Helena Furberg (Memorial Sloan-Kettering Cancer Center), Thomas Rohan and Joseph Sparano (Albert Einstein College of Medicine), Kitwaw Demissie (University of Medicine and Dentistry of New Jersey), Paul Tartter and Alison Estabrook (St. Luke's Roosevelt Hospital), James Reilly (Kings County Hospital Center), Benjamin Pace, George Raptis and Christina Wetz (Mount Sinai School of Medicine), Maria Castaldi (Jacob Medical Center), Sheldon Feldman (New York-Presbyterian), and Margaret Kemeny (Queens Hospital Center).

## References

1. Siegel R, Ward E, Brawley O, Jemal A. Cancer statistics, the impact of eliminating socioeconomic and racial disparities on premature cancer deaths. *CA Cancer J Clin.* 2011; 61(4):212–236. [PubMed: 21685461]
2. Gerend MA, Pai M. Social determinants of Black-White disparities in breast cancer mortality: a review. *Cancer Epidemiol Biomarkers Prev.* 2008; 17(11):2913–2923. [PubMed: 18990731]

3. Ademuyiwa FO, Edge SB, Erwin DO, Orom H, Ambrosone CB, Underwood W 3rd. Breast cancer racial disparities: unanswered questions. *Cancer Res.* 71(3):640–644. [PubMed: 21135114]
4. Carey LA, Perou CM, Livasy CA, Dressler LG, Cowan D, Conway K, Karaca G, Troester MA, Tse CK, Edmiston S, Deming SL, Geradts J, Cheang MC, Nielsen TO, Moorman PG, Earp HS, Millikan RC. Race, breast cancer subtypes, and survival in the Carolina Breast Cancer Study. *JAMA.* 2006; 295(21):2492–2502. [PubMed: 16757721]
5. Cunningham JE, Butler WM. Racial disparities in female breast cancer in South Carolina: clinical evidence for a biological basis. *Breast Cancer Res Treat.* 2004; 88(2):161–176. [PubMed: 15564799]
6. Jemal A, Siegel R, Xu J, Ward E. Cancer statistics 2010. *CA Cancer J Clin.* 60(5):277–300. [PubMed: 20610543]
7. Lund MJ, Trivers KF, Porter PL, Coates RJ, Leyland-Jones B, Brawley OW, Flagg EW, O'Regan RM, Gabram SG, Eley JW. Race and triple negative threats to breast cancer survival: a population-based study in Atlanta, GA. *Breast Cancer Res Treat.* 2009; 113(2):357–370. [PubMed: 18324472]
8. Stead LA, Lash TL, Sobieraj JE, Chi DD, Westrup JL, Charlot M, Blanchard RA, Lee JC, King TC, Rosenberg CL. Triple-negative breast cancers are increased in black women regardless of age or body mass index. *Breast Cancer Res.* 2009; 11(2):R18. [PubMed: 19320967]
9. Menashe I, Anderson WF, Jatoi I, Rosenberg PS. Underlying causes of the black-white racial disparity in breast cancer mortality: a population-based analysis. *J Natl Cancer Inst.* 2009; 101(14):993–1000. [PubMed: 19584327]
10. Cheng AM, Byrom MW, Shelton J, Ford LP. Antisense inhibition of human miRNAs and indications for an involvement of miRNA in cell growth and apoptosis. *Nucleic Acids Res.* 2005; 33(4):1290–1297. [PubMed: 15741182]
11. Berezikov E. Evolution of microRNA diversity and regulation in animals. *Nature reviews Genetics.* 2011; 12(12):846–860.
12. Negrini M, Nicoloso MS, Calin GA. MicroRNAs and cancer--new paradigms in molecular oncology. *Curr Opin Cell Biol.* 2009; 21(3):470–479. [PubMed: 19411711]
13. Blenkiron C, Goldstein LD, Thorne NP, Spiteri I, Chin SF, Dunning MJ, Barbosa-Morais NL, Teschendorff AE, Green AR, Ellis IO, Tavaré S, Caldas C, Miska EA. MicroRNA expression profiling of human breast cancer identifies new markers of tumor subtype. *Genome Biol.* 2007; 8(10):R214. [PubMed: 17922911]
14. Iorio MV, Ferracin M, Liu CG, Veronese A, Spizzo R, Sabbioni S, Magri E, Pedriali M, Fabbri M, Campiglio M, Menard S, Palazzo JP, Rosenberg A, Musiani P, Volinia S, Nenci I, Calin GA, Querzoli P, Negrini M, Croce CM. MicroRNA gene expression deregulation in human breast cancer. *Cancer Res.* 2005; 65(16):7065–7070. [PubMed: 16103053]
15. Sempere LF, Christensen M, Silahatoglu A, Bak M, Heath CV, Schwartz G, Wells W, Kauppinen S, Cole CN. Altered MicroRNA expression confined to specific epithelial cell subpopulations in breast cancer. *Cancer Res.* 2007; 67(24):11612–11620. [PubMed: 18089790]
16. Son JW, Kim YJ, Cho HM, Lee SY, Jang JS, Choi JE, Lee JU, Kang MG, Lee YM, Kwon SJ, Choi E, Na MJ, Park JY. MicroRNA Expression Profiles in Korean Non-Small Cell Lung Cancer. *Tuberc Respir Dis.* 2009; 67(5):413.
17. Wang T, Zhang X, Obijuru L, Laser J, Aris V, Lee P, Mittal K, Soteropoulos P, Wei JJ. A micro-RNA signature associated with race, tumor size, and target gene activity in human uterine leiomyomas. *Genes Chromosomes Cancer.* 2007; 46(4):336–347. [PubMed: 17243163]
18. Hu Z, Liang J, Wang Z, Tian T, Zhou X, Chen J, Miao R, Wang Y, Wang X, Shen H. Common genetic variants in pre-microRNAs were associated with increased risk of breast cancer in Chinese women. *Hum Mutat.* 2009; 30(1):79–84. [PubMed: 18634034]
19. Shen J, Ambrosone CB, DiCioccio RA, Odunsi K, Lele SB, Zhao H. A functional polymorphism in the miR-146a gene and age of familial breast/ovarian cancer diagnosis. *Carcinogenesis.* 2008; 29(10):1963–1966. [PubMed: 18660546]
20. Shen J, Ambrosone CB, Zhao H. Novel genetic variants in microRNA genes and familial breast cancer. *Int J Cancer.* 2009; 124(5):1178–1182. [PubMed: 19048628]

21. Wu M, Jolicoeur N, Li Z, Zhang L, Fortin Y, L'Abbe D, Yu Z, Shen SH. Genetic variations of microRNAs in human cancer and their effects on the expression of miRNAs. *Carcinogenesis*. 2008; 29(9):1710–1716. [PubMed: 18356149]
22. Ambrosone CB, Ciupak GL, Bandera EV, Jandorf L, Bovbjerg DH, Zirpoli G, Pawlish K, Godbold J, Furberg H, Fatone A, Valdimarsdottir H, Yao S, Li Y, Hwang H, Davis W, Roberts M, Sucheston L, Demissie K, Amend KL, Tartter P, Reilly J, Pace BW, Rohan T, Sparano J, Raptis G, Castaldi M, Estabrook A, Feldman S, Weltz C, Kemeny M. Conducting Molecular Epidemiological Research in the Age of HIPAA: A Multi-Institutional Case-Control Study of Breast Cancer in African-American and European-American Women. *J Oncol*. 2009; 2009:871250. [PubMed: 19865486]
23. Bandera EV, Chandran U, Zirpoli G, McCann SE, Ciupak G, Ambrosone CB. Rethinking sources of representative controls for the conduct of case-control studies in minority populations. *BMC medical research methodology*. 2013; 13:71
24. Xu Z, Kaplan NL, Taylor JA. TAGster: Efficient Selection of LD tag SNPs in Single or Multiple Populations. *Bioinformatics*. 2007
25. Tsai HJ, Choudhry S, Naqvi M, Rodriguez-Cintron W, Burchard EG, Ziv E. Comparison of three methods to estimate genetic ancestry and control for stratification in genetic association studies among admixed populations. *Hum Genet*. 2005; 118(3–4):424–433. [PubMed: 16208514]
26. Speliotes EK, Willer CJ, Berndt SI, Monda KL, Thorleifsson G, Jackson AU, Lango Allen H, Lindgren CM, Luan J, Magi R, Randall JC, Vedantam S, Winkler TW, Qi L, Workalemahu T, Heid IM, Steinthorsdottir V, Stringham HM, Weedon MN, Wheeler E, Wood AR, Ferreira T, Weyant RJ, Segre AV, Estrada K, Liang L, Nemesh J, Park JH, Gustafsson S, Kilpelainen TO, Yang J, Bouatia-Naji N, Esko T, Feitosa MF, Kutalik Z, Mangino M, Raychaudhuri S, Scherag A, Smith AV, Welch R, Zhao JH, Aben KK, Absher DM, Amin N, Dixon AL, Fisher E, Glazer NL, Goddard ME, Heard-Costa NL, Hoesel V, Hottenga JJ, Johansson A, Johnson T, Ketkar S, Lamina C, Li S, Moffatt MF, Myers RH, Narisu N, Perry JR, Peters MJ, Preuss M, Ripatti S, Rivadeneira F, Sandholt C, Scott LJ, Timpson NJ, Tyrer JP, van Wingerden S, Watanabe RM, White CC, Wiklund F, Barlassina C, Chasman DI, Cooper MN, Jansson JO, Lawrence RW, Pelliikka N, Prokopenko I, Shi J, Thiering E, Alavere H, Alibrandi MT, Almgren P, Arnold AM, Aspelund T, Atwood LD, Balkau B, Balmforth AJ, Bennett AJ, Ben-Shlomo Y, Bergman RN, Bergmann S, Biebermann H, Blakemore AI, Boes T, Bonnycastle LL, Bornstein SR, Brown MJ, Buchanan TA, Busonero F, Campbell H, Cappuccino FP, Cavalcanti-Proenca C, Chen YD, Chen CM, Chines PS, Clarke R, Coin L, Connell J, Day IN, den Heijer M, Duan J, Ebrahim S, Elliott P, Elosua R, Eiriksdottir G, Erdos MR, Eriksson JG, Facheris MF, Felix SB, Fischer-Posovszky P, Folsom AR, Friedrich N, Freimer NB, Fu M, Gagel S, Gejman PV, Geus EJ, Gieger C, Gjesing AP, Goel A, Goyette P, Grallert H, Grassler J, Greenawalt DM, Groves CJ, Gudnason V, Guiducci C, Hartikainen AL, Hassanali N, Hall AS, Havulinna AS, Hayward C, Heath AC, Hengstenberg C, Hicks AA, Hinney A, Hofman A, Homuth G, Hui J, Igl W, Iribarren C, Isomaa B, Jacobs KB, Jarick I, Jewell E, John U, Jorgensen T, Jousilahti P, Jula A, Kaakinen M, Kajantie E, Kaplan LM, Kathiresan S, Kettunen J, Kinnunen L, Knowles JW, Kolcic I, Konig IR, Koskinen S, Kovacs P, Kuusisto J, Kraft P, Kvaloy K, Laitinen J, Lantieri O, Lanzani C, Launer LJ, Lecoeur C, Lehtimaki T, Lettre G, Liu J, Lokki ML, Lorentzon M, Luben RN, Ludwig B, Manunta P, Marek D, Marre M, Martin NG, McArdle WL, McCarthy A, McKnight B, Meitinger T, Melander O, Meyre D, Midtjell K, Montgomery GW, Morken MA, Morris AP, Mulic R, Ngwa JS, Nelis M, Neville MJ, Nyholt DR, O'Donnell CJ, O'Rahilly S, Ong KK, Oostra B, Pare G, Parker AN, Perola M, Pichler I, Pietilainen KH, Platou CG, Polasek O, Pouta A, Rafelt S, Raitakari O, Rayner NW, Ridderstrale M, Rief W, Ruukonen A, Robertson NR, Rzehak P, Salomaa V, Sanders AR, Sandhu MS, Sanna S, Saramies J, Savolainen MJ, Scherag S, Schipf S, Schreiber S, Schunkert H, Silander K, Sinisalo J, Siscovick DS, Smit JH, Soranzo N, Sovio U, Stephens J, Surakka I, Swift AJ, Tammesoo ML, Tardif JC, Teder-Laving M, Teslovich TM, Thompson JR, Thomson B, Tonjes A, Tuomi T, van Meurs JB, van Ommen GJ, Vatin V, Viikari J, Visvikis-Siest S, Vitart V, Vogel CI, Voight BF, Waite LL, Wallaschofski H, Walters GB, Widen E, Wiegand S, Wild SH, Willemssen G, Witte DR, Witteman JC, Xu J, Zhang Q, Zgaga L, Ziegler A, Zitting P, Beilby JP, Farooqi IS, Hebebrand J, Huikuri HV, James AL, Kahonen M, Levinson DF, Macciardi F, Nieminen MS, Ohlsson C, Palmer LJ, Ridker PM, Stumvoll M, Beckmann JS, Boeing H, Boerwinkle E, Boomsma DI, Caulfield MJ, Chanock SJ, Collins FS, Cupples LA, Smith GD, Erdmann J, Froguel

- P, Gronberg H, Gyllensten U, Hall P, Hansen T, Harris TB, Hattersley AT, Hayes RB, Heinrich J, Hu FB, Hveem K, Illig T, Jarvelin MR, Kaprio J, Karpe F, Khaw KT, Kiemeny LA, Krude H, Laakso M, Lawlor DA, Metspalu A, Munroe PB, Ouwehand WH, Pedersen O, Penninx BW, Peters A, Pramstaller PP, Quertermous T, Reinehr T, Rissanen A, Rudan I, Samani NJ, Schwarz PE, Shuldiner AR, Spector TD, Tuomilehto J, Uda M, Uitterlinden A, Valle TT, Wabitsch M, Waeber G, Wareham NJ, Watkins H, Wilson JF, Wright AF, Zillikens MC, Chatterjee N, McCarroll SA, Purcell S, Schadt EE, Visscher PM, Assimes TL, Borecki IB, Deloukas P, Fox CS, Groop LC, Haritunians T, Hunter DJ, Kaplan RC, Mohlke KL, O'Connell JR, Peltonen L, Schlessinger D, Strachan DP, van Duijn CM, Wichmann HE, Frayling TM, Thorsteinsdottir U, Abecasis GR, Barroso I, Boehnke M, Stefansson K, North KE, McCarthy MI, Hirschhorn JN, Ingelsson E, Loos RJ. Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. *Nat Genet.* 2010; 42(11):937–948. [PubMed: 20935630]
27. Gao LB, Bai P, Pan XM, Jia J, Li LJ, Liang WB, Tang M, Zhang LS, Wei YG, Zhang L. The association between two polymorphisms in pre-miRNAs and breast cancer risk: a meta-analysis. *Breast Cancer Res Treat.* 2011; 125(2):571–574. [PubMed: 20640596]
28. Hoffman AE, Zheng T, Yi C, Leaderer D, Weidhaas J, Slack F, Zhang Y, Paranjape T, Zhu Y. microRNA miR-196a-2 and breast cancer: a genetic and epigenetic association study and functional analysis. *Cancer Res.* 2009; 69(14):5970–5977. [PubMed: 19567675]
29. Xu W, Xu J, Liu S, Chen B, Wang X, Li Y, Qian Y, Zhao W, Wu J. Effects of common polymorphisms rs11614913 in miR-196a2 and rs2910164 in miR-146a on cancer susceptibility: a meta-analysis. *PLoS one.* 2011; 6(5):e20471. [PubMed: 21637771]
30. Jedlinski DJ, Gabrovska PN, Weinstein SR, Smith RA, Griffiths LR. Single nucleotide polymorphism in hsa-mir-196a-2 and breast cancer risk: a case control study. *Twin research and human genetics : the official journal of the International Society for Twin Studies.* 2011; 14(5): 417–421. [PubMed: 21962133]
31. Garcia AI, Cox DG, Barjhoux L, Verny-Pierre C, Barnes D, Antoniou AC, Stoppa-Lyonnet D, Sinilnikova OM, Mazoyer S. The rs2910164:G>C SNP in the MIR146A gene is not associated with breast cancer risk in BRCA1 and BRCA2 mutation carriers. *Hum Mutat.* 2011
32. Catucci I, Yang R, Verderio P, Pizzamiglio S, Heesen L, Hemminki K, Sutter C, Wappenschmidt B, Dick M, Arnold N, Bugert P, Niederacher D, Meindl A, Schmutzler RK, Bartram CC, Ficarazzi F, Tizzoni L, Zaffaroni D, Manoukian S, Barile M, Pierotti MA, Radice P, Burwinkel B, Peterlongo P. Evaluation of SNPs in miR-146a, miR196a2 and miR-499 as low-penetrance alleles in German and Italian familial breast cancer cases. *Hum Mutat.* 2010; 31(1):E1052–E1057. [PubMed: 19847796]
33. Zhang L, Volinia S, Bonome T, Calin GA, Greshock J, Yang N, Liu CG, Giannakakis A, Alexiou P, Hasegawa K, Johnstone CN, Megraw MS, Adams S, Lassus H, Huang J, Kaur S, Liang S, Sethupathy P, Leminen A, Simossis VA, Sandaltzopoulos R, Naomoto Y, Katsaros D, Gimotty PA, DeMichele A, Huang Q, Butzow R, Rustgi AK, Weber BL, Birrer MJ, Hatzigeorgiou AG, Croce CM, Coukos G. Genomic and epigenetic alterations deregulate microRNA expression in human epithelial ovarian cancer. *Proceedings of the National Academy of Sciences of the United States of America.* 2008; 105(19):7004–7009. [PubMed: 18458333]
34. Lavon I, Zrihan D, Granit A, Einstein O, Fainstein N, Cohen MA, Zelikovitch B, Shoshan Y, Spektor S, Reubinoff BE, Felig Y, Gerlitz O, Ben-Hur T, Smith Y, Siegal T. Gliomas display a microRNA expression profile reminiscent of neural precursor cells. *Neuro-oncology.* 2010; 12(5): 422–433. [PubMed: 20406893]
35. Toffanin S, Hoshida Y, Lachenmayer A, Villanueva A, Cabellos L, Minguez B, Savic R, Ward SC, Thung S, Chiang DY, Alsinet C, Tovar V, Roayaie S, Schwartz M, Bruix J, Waxman S, Friedman SL, Golub T, Mazzaferro V, Llovet JM. MicroRNA-based classification of hepatocellular carcinoma and oncogenic role of miR-517a. *Gastroenterology.* 2011; 140(5):1618–1628. e1616. [PubMed: 21324318]
36. Shen J, Wang D, Gregory SR, Medico L, Hu Q, Yan L, Odunsi K, Lele SB, Ambrosone CB, Liu S, Zhao H. Evaluation of microRNA expression profiles and their associations with risk alleles in lymphoblastoid cell lines of familial ovarian cancer. *Carcinogenesis.* 2012
37. Ganz PA, Land SR, Geyer CE Jr, Cecchini RS, Costantino JP, Pajon ER, Fehrenbacher L, Atkins JN, Polikoff JA, Vogel VG, Erban JK, Livingston RB, Perez EA, Mamounas EP, Wolmark N,

- Swain SM. Menstrual history and quality-of-life outcomes in women with node-positive breast cancer treated with adjuvant therapy on the NSABP B-30 trial. *J Clin Oncol.* 2011; 29(9):1110–1116. [PubMed: 21300930]
38. Zheng W, Cai Q, Signorello LB, Long J, Hargreaves MK, Deming SL, Li G, Li C, Cui Y, Blot WJ. Evaluation of 11 breast cancer susceptibility loci in African-American women. *Cancer Epidemiol Biomarkers Prev.* 2009; 18(10):2761–2764. [PubMed: 19789366]

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

**Table 1**  
Descriptive characteristics of African American and European American breast cancer cases and controls

| Characteristics             | African American |                 |      | European American |                 |       |
|-----------------------------|------------------|-----------------|------|-------------------|-----------------|-------|
|                             | Case (n=491)     | Control (n=415) | P    | Case (n=336)      | Control (n=317) | P     |
|                             | Mean (SD)        | Mean (SD)       |      | Mean (SD)         | Mean (SD)       |       |
| Age                         | 50.8 (10.0)      | 50.2 (8.9)      | 0.40 | 50.5 (8.4)        | 50.4 (8.3)      | 0.93  |
| Body mass index             | 31.2 (6.8)       | 31.6 (7.7)      | 0.47 | 26.9 (6.0)        | 27.7 (7.4)      | 0.15  |
| % European ancestry         | 0.09 (0.15)      | 0.10 (0.16)     | 0.13 | 0.98 (0.07)       | 0.99 (0.03)     | 0.09  |
|                             | Count (%)        | Count (%)       |      | Count (%)         | Count (%)       |       |
| Menopausal status           |                  |                 | 0.07 |                   |                 | 0.44  |
| Premenopausal               | 300 (61.1)       | 229 (55.2)      |      | 203 (60.6)        | 182 (57.6)      |       |
| Postmenopausal              | 191 (38.9)       | 186 (44.8)      |      | 132 (39.4)        | 134 (42.4)      |       |
| Family history              |                  |                 | 0.29 |                   |                 | 0.003 |
| Yes                         | 71 (14.5)        | 50 (12.0)       |      | 86 (25.6)         | 51 (16.1)       |       |
| No                          | 420 (85.5)       | 365 (88.0)      |      | 250 (74.4)        | 266 (83.9)      |       |
| Education                   |                  |                 | 0.07 |                   |                 | 0.002 |
| Less than high school       | 71 (14.5)        | 49 (11.8)       |      | 7 (2.1)           | 4 (1.3)         |       |
| High school                 | 154 (31.4)       | 110 (26.5)      |      | 73 (21.7)         | 32 (10.1)       |       |
| College and graduate school | 266 (54.2)       | 256 (61.7)      |      | 256 (76.2)        | 281 (88.6)      |       |
| Hormone replacement therapy |                  |                 | 0.94 |                   |                 | 0.35  |
| Yes                         | 66 (13.6)        | 57 (13.7)       |      | 88 (26.2)         | 73 (23.0)       |       |
| No                          | 421 (86.4)       | 358 (86.3)      |      | 248 (73.8)        | 244 (77.0)      |       |

Footnote:

\* P derived from student t-test for continuous variables and chi-square test or Fisher's exact test for categorical variables.

Abbreviation: SD: standard deviation.



**Table 2**  
Top ranked SNPs associated with breast cancer risk in African American and European American women

| Gene/miRNA     | SNP        | Genotype          | AA             |                  |      | EA             |                  |       | P for interaction with race |
|----------------|------------|-------------------|----------------|------------------|------|----------------|------------------|-------|-----------------------------|
|                |            |                   | # Case/Control | OR (95% CI)      | P    | # Case/Control | OR (95% CI)      | P     |                             |
| AGO4           | rs7354931  | CC                | 429/356        | 1.00             | 0.03 | 326/306        | 1.00             | 0.99  | 0.47                        |
|                |            | CA/AA             | 42/55          | 0.63 (0.41–0.96) |      |                |                  |       |                             |
| PACT           | rs2059691  | GG/GA             | 445/389        | 1.00             | 0.92 | 283/283        | 1.00             | 0.04  | 0.22                        |
|                |            | AA                | 24/21          | 1.03 (0.56–1.89) |      | 46/27          | 1.72 (1.03–2.87) |       |                             |
| hsa-miR-106b   | rs1527423  | AA                | 37/37          | 1.00             | 0.64 | 90/110         | 1.00             | 0.02  | 0.37                        |
|                |            | AG/GG             | 436/375        | 1.12 (0.69–1.82) |      | 238/200        | 1.50 (1.07–2.11) |       |                             |
| hsa-miR-100    | rs1834306  | GG/GA             | 248/221        | 1.00             | 0.77 | 267/230        | 1.00             | 0.03  | 0.05                        |
|                |            | AA                | 225/191        | 1.04 (0.8–1.36)  |      | 62/79          | 0.64 (0.44–0.95) |       |                             |
| hsa-miR-331    | rs11107973 | AA/AG             | 305/285        | 1.00             | 0.18 | 256/267        | 1.00             | 0.02  | 0.18                        |
|                |            | GG                | 169/127        | 1.22 (0.92–1.62) |      | 72/42          | 1.65 (1.08–2.52) |       |                             |
| hsa-miR-758    | rs12586258 | GG                | 417/336        | 1.00             | 0.01 | 169/170        | 1.00             | 0.33  | 0.01                        |
|                |            | GA/AA             | 56/76          | 0.61 (0.42–0.89) |      | 159/140        | 1.17 (0.85–1.6)  |       |                             |
| hsa-miR-544    | rs10144193 | AA                | 187/167        | 1.00             | 0.81 | 198/219        | 1.00             | 0.004 | 0.03                        |
|                |            | AT/TT             | 287/244        | 1.03 (0.79–1.36) |      | 130/91         | 1.65 (1.18–2.31) |       |                             |
| has-mir-487    | rs1951032  | GG                | 430/364        | 1.00             | 0.23 | 176/198        | 1.00             | 0.001 | 0.005                       |
|                |            | GA/AA             | 38/44          | 0.75 (0.47–1.19) |      | 110/73         | 1.81 (1.26–2.61) |       |                             |
| hsa-miR-659    | rs750504   | TT                | 145/110        | 1.00             | 0.19 | 98/118         | 1.00             | 0.03  | 0.01                        |
|                |            | TA/AA             | 328/302        | 0.82 (0.61–1.10) |      | 231/192        | 1.45 (1.04–2.03) |       |                             |
| hsa-miR-513a-2 | rs2018562  | AA                | 133/143        | 1.00             | 0.04 | 190/178        | 1.00             | 0.97  | 0.36                        |
|                |            | AG                | 225/193        | 1.25 (0.92–1.70) |      | 117/113        | 0.97 (0.70–1.36) |       |                             |
|                |            | GG                | 115/75         | 1.64 (1.13–2.39) |      | 22/19          | 1.04 (0.54–2.01) |       |                             |
|                |            | Per copy G allele |                | 1.28 (1.06–1.54) | 0.01 |                | 1.00 (0.77–1.29) | 0.98  | 0.12                        |

\* Footnote: Odds ratios and p-values adjusted for age, body mass index (BMI), proportion of European ancestry, family history of breast cancer, and education.

Abbreviations: OR: odds ratio; CI: confidence interval.

**Table 3** Top ranked SNPs associated with estrogen receptor (ER) positive breast cancer risk in African American and European American women

| Gene/miRNA     | SNP        | Genotype | AA             |                  |      | EA             |                  |      | P for interaction with race |
|----------------|------------|----------|----------------|------------------|------|----------------|------------------|------|-----------------------------|
|                |            |          | # Case/Control | OR (95% CI)      | P    | # Case/Control | OR (95% CI)      | P    |                             |
| AGO4           | rs16822342 | AA       | 155/229        | 1.00             | 0.04 | 180/277        | 1.00             | 0.84 | 0.42                        |
|                |            | AG/GG    | 87/183         | 0.71 (0.51–0.99) |      | 20/33          | 0.94 (0.52–1.69) |      |                             |
| AGO4           | rs3820276  | GG       | 111/232        | 1.00             | 0.01 | 188/287        | 1.00             | 0.52 | 0.11                        |
| XPO5           | rs11077    | GC/CC    | 131/179        | 1.51 (1.1–2.08)  |      | 12/23          | 0.79 (0.38–1.63) |      | 0.06                        |
|                |            | AA       | 39/45          | 1.00             | 0.05 | 76/127         | 1.00             | 0.68 |                             |
|                |            | AC/CC    | 203/366        | 0.63 (0.4–1)     |      | 124/183        | 1.08 (0.75–1.56) |      | 0.04                        |
| hsa-miR-548a-2 | rs878175   | AA       | 59/110         | 1.00             | 0.58 | 154/210        | 1.00             | 0.02 | 0.04                        |
|                |            | AG/GG    | 183/302        | 1.11 (0.77–1.61) |      | 46/100         | 0.6 (0.4–0.91)   |      |                             |
| hsa-miR-106b   | rs1527423  | AA       | 23/37          | 1.00             | 0.88 | 55/110         | 1.00             | 0.05 | 0.19                        |
|                |            | AG/GG    | 219/375        | 0.96 (0.55–1.66) |      | 144/200        | 1.49 (1.01–2.21) |      |                             |
| hsa-miR-455    | rs2060133  | CC       | 49/65          | 1.00             | 0.12 | 149/209        | 1.00             | 0.04 | 0.9                         |
|                |            | CG/GG    | 193/345        | 0.72 (0.47–1.09) |      | 50/101         | 0.66 (0.44–0.99) |      |                             |
| hsa-miR-606    | rs12266981 | GG       | 186/344        | 1.00             | 0.02 | 199/309        | 1.00             | 0.99 | 0.98                        |
|                |            | GA/AA    | 55/66          | 1.6 (1.07–2.4)   |      | 1/0            |                  |      |                             |
| hsa-miR-100    | rs1834306  | GG/GA    | 125/221        | 1.00             | 0.67 | 167/230        | 1.00             | 0.01 | 0.02                        |
|                |            | AA       | 116/191        | 1.07 (0.78–1.49) |      | 33/79          | 0.56 (0.35–0.88) |      |                             |
| hsa-miR-331    | rs11107973 | AA/AG    | 161/285        | 1.00             | 0.58 | 155/267        | 1.00             | 0.03 | 0.13                        |
|                |            | GG       | 81/127         | 1.1 (0.78–1.55)  |      | 44/42          | 1.7 (1.06–2.73)  |      |                             |
| hsa-miR-544    | rs10144193 | AA       | 86/167         | 1.00             | 0.23 | 125/219        | 1.00             | 0.04 | 0.44                        |
|                |            | AT/TT    | 156/244        | 1.22 (0.88–1.7)  |      | 74/91          | 1.49 (1.01–2.19) |      |                             |
| has-mir-487    | rs1951032  | GG       | 216/364        | 1.00             | 0.88 | 118/198        | 1.00             | 0.05 | 0.2                         |
|                |            | GA/AA    | 24/44          | 0.96 (0.56–1.64) |      | 62/73          | 1.53 (1.01–2.31) |      |                             |
| hsa-miR-628    | rs8041885  | AA       | 92/124         | 1.00             | 0.03 | 160/245        | 1.00             | 0.75 | 0.29                        |
|                |            | AG/GG    | 150/288        | 0.69 (0.49–0.97) |      | 39/64          | 0.93 (0.59–1.46) |      |                             |
| hsa-miR-628    | rs8041044  | CC       | 95/123         | 1.00             | 0.01 | 161/245        | 1.00             | 0.66 | 0.23                        |
|                |            | CA/AA    | 147/288        | 0.65 (0.46–0.91) |      | 39/65          | 0.9 (0.58–1.42)  |      |                             |

| Gene/miRNA     | SNP       | Genotype          | AA             |                  |      | EA             |                  |       | P for interaction with race |
|----------------|-----------|-------------------|----------------|------------------|------|----------------|------------------|-------|-----------------------------|
|                |           |                   | # Case/Control | OR (95% CI)      | P    | # Case/Control | OR (95% CI)      | P     |                             |
| hsa-miR-122a   | rs17669   | AA/AG             | 212/338        | 1.00             | 0.05 | 188/291        | 1.00             | 0.90  | 0.24                        |
|                |           | GG                | 29/73          | 0.63 (0.39–1)    |      | 12/18          | 1.05 (0.49–2.28) |       |                             |
| DGCR8          | rs9606241 | AA/AG             | 228/382        | 1.00             | 0.62 | 94/159         | 1.00             | 0.03  | 0.26                        |
|                |           | GG                | 14/28          | 0.85 (0.43–1.64) |      | 26/22          | 1.99 (1.09–3.64) |       |                             |
| hsa-miR-659    | rs750504  | TT                | 76/110         | 1.00             | 0.08 | 57/118         | 1.00             | 0.02  | 0.05                        |
|                |           | TA                | 104/215        | 0.71 (0.49–1.04) |      | 94/144         | 1.33 (0.88–2.02) |       |                             |
| hsa-miR-513a-2 | rs2018562 | AA                | 62/87          | 1.06 (0.68–1.65) |      | 49/48          | 2.13 (1.27–3.56) |       | 0.04                        |
|                |           | Per copy A allele |                | 1.01 (0.81–1.26) | 0.93 |                | 1.45 (1.12–1.87) | 0.005 |                             |
|                |           | AA/AG             | 180/336        | 1.00             | 0.04 | 188/291        | 1.00             | 0.93  |                             |
|                |           | GG                | 61/75          | 1.51 (1.03–2.22) |      | 12/19          | 0.97 (0.45–2.06) |       | 0.28                        |

*Footnote:* Odds ratios and p-values adjusted for age, body mass index (BMI), proportion of European ancestry, family history of breast cancer, and education.

*Abbreviations:* OR: odds ratio; CI: confidence interval; ER: estrogen receptor

**Table 4**

Top ranked SNPs associated with estrogen receptor (ER) negative breast cancer risk in African American and European American women

| Gene           | SNP       | Genotype | AA             |                  |       | EA             |                  |      | P for interaction with race |
|----------------|-----------|----------|----------------|------------------|-------|----------------|------------------|------|-----------------------------|
|                |           |          | # Case/Control | OR (95% CI)      | P     | # Case/Control | OR (95% CI)      | P    |                             |
| hsa-miR-219    | rs107822  | GG       | 29/174         | 1.00             | 0.004 | 24/180         | 1.00             | 0.17 | 0.5                         |
|                |           | GA/AA    | 78/235         | 1.99 (1.24–3.19) |       | 27/130         | 1.52 (0.84–2.77) |      |                             |
| hsa-mir-595    | rs4909238 | AA/AG    | 77/338         | 1.00             | 0.03  | 37/248         | 1.00             | 0.16 | 0.81                        |
|                |           | GG       | 27/70          | 1.76 (1.05–2.95) |       | 14/61          | 1.64 (0.82–3.26) |      |                             |
| hsa-miR-204    | rs7861254 | GG       | 37/106         | 1.00             | 0.04  | 28/166         | 1.00             | 0.77 | 0.3                         |
|                |           | vs. GG   | 70/306         | 0.62 (0.39–0.99) |       | 23/144         | 0.91 (0.5–1.67)  |      |                             |
| hsa-miR-608    | rs4919510 | CC       | 43/153         | 1.00             | 0.42  | 41/204         | 1.00             | 0.03 | 0.18                        |
|                |           | CG/GG    | 64/259         | 0.83 (0.54–1.29) |       | 10/106         | 0.45 (0.21–0.94) |      |                             |
| hsa-miR-758    | rs7141987 | AA       | 2/19           | 1.00             | 0.36  | 8/94           | 1.00             | 0.03 | 0.83                        |
|                |           | AG/GG    | 104/392        | 2.01 (0.45–8.93) |       | 43/210         | 2.41 (1.08–5.37) |      |                             |
| hsa-miR-495    | rs2281611 | CC       | 62/233         | 1.00             | 0.92  | 15/151         | 1.00             | 0.01 | 0.03                        |
|                |           | CA/AA    | 45/179         | 0.98 (0.63–1.51) |       | 36/159         | 2.29 (1.19–4.39) |      |                             |
| hsa-miR-513a-2 | rs2018562 | AA       | 25/143         | 1.00             | 0.03  | 26/178         | 1.00             | 0.33 | 0.46                        |
|                |           | AG/GG    | 82/268         | 1.74 (1.06–2.86) |       | 25/132         | 1.35 (0.74–2.47) |      |                             |

\* *Footnote:* Odds ratios and p-values adjusted for age, body mass index (BMI), proportion of European ancestry, family history of breast cancer, and education.  
*Abbreviations:* OR: odds ratio; CI: confidence interval; ER: estrogen receptor

**Table 5**  
Polygenic risk score and breast cancer risk in African American and European American women

| Subgroup               | SNPs (risk genotype or allele)   | Expected range of polygenic risk score | Mean (SD) of polygenic score in cases | Mean (SD) of polygenic score in controls | OR (95% CI) per unit of composite genetic score | P                    |
|------------------------|--|--|---------------------------------------|--|---|----------------------|
| Overall risk in AAs    | rs7354931(CC), rs12586258(GG), rs2018562(G allele)   | 0–4.0                                  | 3.0 (0.8)                             | 2.8 (0.8)                                | 1.44 (1.21–1.71)                                | 3.3*10 <sup>-5</sup> |
| Overall risk in EAs    | rs2059691(AA), rs1527423(AG/GG), rs1834306(GG/GA), rs11107973(GG), rs1951032(GA/AA), rs5750504(TA/AA)                                | 0–6.0                                  | 3.7 (1.2)                             | 3.3 (1.2)                                | 1.37 (1.18–1.59)                                | 3.2*10 <sup>-5</sup> |
| ER+ in AAs             | rs16822342(AA), rs3820276(GC/CC), rs11077(AA), rs12266981(GA/AA), rs8041044(CC), rs17669(AA/AG), rs2018562(GG)                       | 0–7.0                                  | 3.0 (1.2)                             | 2.5 (1.1)                                | 1.51 (1.30–1.74)                                | 3.3*10 <sup>-8</sup> |
| ER+ in EAs             | rs878175(AA), rs1527423(AG/GG), rs2060133(CC), rs1834306(GG/GA), rs11107973(GG), rs1951032(GA/AA), rs9606241(GG), rs750504(A allele) | 0–9.0                                  | 4.6 (1.3)                             | 3.9 (1.2)                                | 1.73 (1.45–2.06)                                | 1.4*10 <sup>-9</sup> |
| ER– in AAs             | rs107822(GA/AA), rs4909238(GG), rs7861254(GG), rs2018562(AG/GG)  | 0–4.0                                  | 2.1 (0.8)                             | 1.7 (0.9)                                | 1.73 (1.35–2.22)                                | 1.4*10 <sup>-5</sup> |
| ER– in EAs             | rs4919510(CC), rs7141987(AG/GG), rs2281611(CA/AA)  | 0–3.0                                  | 2.4 (0.9)                             | 1.9 (1.0)                                | 1.73 (1.22–2.45)                                | 2.0*10 <sup>-3</sup> |
| Triple negative in AAs | rs107822 (GA/AA), rs4909238(GG), rs7861254(GG), rs2185743(AA), 3008372(GA/AA), rs543412(GG), rs2217653(AG/GG)                        | 0–7.0                                  | 3.5 (1.3)                             | 2.7 (1.4)                                | 1.49 (1.21–1.83)                                | 2.0*10 <sup>-4</sup> |
| Triple negative in EAs | rs11100610(AA), rs5905010(GC/CC)   | 0–2.0                                  | 1.1 (0.6)                             | 0.6 (0.7)                                | 2.86 (1.47–5.55)                                | 2.0*10 <sup>-3</sup> |

*Footnote:* Odds ratios and p-values adjusted for age, body mass index (BMI), proportion of European ancestry, family history of breast cancer, and education. Weighted composite genetic score is calculated as the sum of the number of risk allele or genotype weighted by their effect sizes of top ranked SNPs in each subgroup, which is then standardized to make 1 unit of the score equal to 1 risk allele or genotype, depending on the genetic model used for each SNP.

*Abbreviations:* SD: standard deviation; OR: odds ratio; CI: confidence interval