

TABLE 1
Synergistic effects between fractions from
Escherichia coli

Fraction	Total Quantity of Gas (μ L) Evolved or Utilized								
	Formic hy- drogenylase*			Formic dehy- drogenase†			Hydro- genase‡		
	5 min	10 min	15 min	5 min	10 min	15 min	4 min	8 min	13 min
<i>Ps</i>	0	0	0	10	24	41	7	19	36
<i>Pm</i>	0	0	0	6	15	30	13	51	110
<i>SSN</i>	11	30	59	10	18	29	3	9	16
<i>Ps</i> + <i>SSN</i>	74	181	279	40	80	120	36	79	128
<i>Pm</i> + <i>SSN</i>	59	150	240	40	81	121	115	206	—

All Warburg vessels contained 75 μ M sodium-potassium phosphate buffer, pH 6.1, in final volume of 1.2 ml. Temperature, 30 C.

* CO₂ + H₂ production in the presence of 15 μ M sodium formate. Gas phase, helium. Where indicated, 1.86 mg (dry wt) *Ps*, 0.66 mg *Pm*, and 0.1 ml *SSN* were used.

† CO₂ production under the conditions noted for formic hydrogenylase, but with 12 μ M methylene blue added.

‡ Utilization of gas under an atmosphere of H₂ in the presence of 12 μ M of methylene blue. Where indicated, 0.13 mg *Ps*, 0.66 mg *Pm*, and 0.1 ml of a 1:6 dilution of *SSN* were used.

Pm do not decay at the same rates during storage at 4 C. These observations suggest that a partial separation of insoluble cellular fractions with different metabolic activities was achieved.

Aerobic oxidation of citrate, α -ketoglutarate, and several other members of the tricarboxylic

acid cycle was also studied with similar preparations from cells grown as described by Swim and Krampitz (J. Bacteriol., 67, 419, 1954). The particulate fractions alone showed negligible oxidative activity with these substrates. Upon combining particulate and soluble components, disproportionate acceleration of the rates of O₂ utilization was observed. These stimulations are undoubtedly the result of coupling between soluble dehydrogenases and insoluble electron transport enzymes such as the cytochromes. The latter are invariably present in the insoluble fraction of *E. coli* (Gale, 1939; Gest, 1952). Moyed and O'Kane (Arch. Biochem. and Biophys., 39, 457, 1952) have described a very similar cooperative effect between soluble and insoluble components of *Proteus vulgaris* with regard to pyruvate oxidase activity. Considering the results noted in table 1, it seems likely that electron transport enzymes concerned with certain anaerobic processes are also present in the insoluble portion of the cell.

The particulate preparations described are much more stable than animal mitochondria with regard to changes in tonicity of the medium and show excellent retention of certain enzymatic activities upon storage at 4 C. It has been shown in one instance at least (Gest, 1952) that variation in enzymatic composition of intact cells with growth conditions is reflected by the particulate fractions. In view of these properties such preparations should be of considerable value for metabolic and related cytochemical studies.

A CULTURE OF ESCHERICHIA FREUNDII SEROLOGICALLY RELATED TO THE GENUS SALMONELLA

P. R. EDWARDS, ALMA C. McWHORTER, JUSTINE McCURDY, AND R. DAVIS

Communicable Disease Center, Public Health Service, U. S. Department of Health, Education, and Welfare
Atlanta, Georgia, and Bureau of Laboratories, Florida State Board of Health,
Jacksonville, Florida

Received for publication July 6, 1954

Numerous relationships between the genera *Escherichia* and *Salmonella* involving O, K (sheath or envelop), and H antigens have been described. These relationships have been summarized by Kauffmann (*Enterobacteriaceae*, E. Munksgaard, Copenhagen, 1951). Perhaps the best known of these is the occurrence of the Vi antigen of *Salmonella typhi* in cultures of the

Ballerup type of Kauffmann and Møller (J. Hyg., 40, 246, 1940) and in other cultures of *Escherichia freundii* (West and Edwards, Public Health Service Monograph, in press, 1954). The occurrence of *Salmonella* H antigens in *E. freundii* cultures first was noted by Gard (Z. Hyg. Infektionskrankh., 120, 59, 1937) and Gard and Eriksson (Z. Hyg. Infektionskrankh., 122, 54,

1939). Relationships between the O antigens of a variety of *Escherichia* cultures and those of *Salmonella* cultures have been reported by a number of workers. Most striking of these was the occurrence of the antigenic complex of *Salmonella* group B in coliform cultures observed first by Schütze (quoted by Kauffmann, 1951).

In spite of the numerous reports in the literature involving antigenic relationships between the *Salmonella* and *Escherichia* groups, no coliform culture yet has been reported which contained both O and H antigens in common with *Salmonella* strains. The present report deals with a culture (4598-53) which was a typical strain of *E. freundii* and which contained both O and H antigens found in *Salmonella* cultures. Culture 4598-53 was isolated from the stool of a "normal" adult in the course of a survey of food handlers.

Culture 4598-53 produced H₂S, acidified Jordan's tartrate agar, and grew in Simmons' citrate medium. It failed to produce indole, to hydrolyze urea, or to liquefy gelatin. The methyl red test was positive, and the Voges-Proskauer test was negative. In tests for growth in KCN medium and in tests for decarboxylase activity, the culture behaved as a typical strain of *E. freundii*.¹ Acid and gas were produced promptly from glucose, arabinose, xylose, maltose, lactose, trehalose, rhamnose, mannitol, and sorbitol. Salicin was fermented after 14 days of incubation. Sucrose, raffinose, dulcitol, inositol, and adonitol were not attacked.

The organism was strongly agglutinated by O serum *Salmonella* group E₂ (3, 15) and to a lesser degree by serums for groups E₁ (3, 10) and E₃ (1, 3, 19). The relationship of the O antigen of 4598-53 to the O antigen of group E₂ is shown in table 1. Except for a very small fraction of antigen 3, which is common to groups E₁ and E₂, 4598-53 possessed the whole antigenic complex of group E₂. Further, as shown by absorption of its serum, 4598-53 possessed no O antigen other than that found in *Salmonella* group E₂.

The H antigens 4598-53 were monophasic and were related to those of the *Salmonella* serotypes known as Weslaco and Macallen, both of which are represented by the symbol z₃₆. The relationships of these antigens are detailed in table 2. None of the three cultures possessed identical H antigens. The two *Salmonella* types shared an

¹ The writers are indebted to Dr. F. Kauffmann and Dr. Vagn Møller for the performance of the KCN and decarboxylase tests.

TABLE 1
O Antigens of *Escherichia freundii*, 4598-53

Serums	Antigens		
	Salmonella E ₁ (3, 10)	Salmonella E ₂ (3, 15)	<i>E. freundii</i> 4598-53
<i>Salmonella</i> E ₂ (3, 15)			
Unabsorbed.....	1,000	4,000	4,000
Absorbed by			
4598-53.....	100	100	<20
4598-53 + <i>Salmonella</i> E ₁	<20	<20	<20
<i>E. freundii</i> , 4598-53			
Unabsorbed.....	1,000	2,000	4,000
Absorbed by <i>Salmonella</i> E ₂ (3, 15).....	<20	<20	<20

TABLE 2
H Antigens of *Escherichia freundii*, 4598-53

Serums	Antigens		
	Weslaco (z ₃₆)	Macallen (z ₃₆)	4598-53
Weslaco (z ₃₆)			
Unabsorbed.....	4,000	2,000	2,000
Absorbed by			
Macallen (z ₃₆).....	1,000	<100	<100
4598-53.....	2,000	1,000	<100
Macallen (z ₃₆)			
Unabsorbed.....	4,000	8,000	8,000
Absorbed by			
Weslaco (z ₃₆).....	<100	4,000	500
4598-53.....	2,000	4,000	<100
4598-53			
Unabsorbed.....	1,000	4,000	16,000
Absorbed by			
Weslaco (z ₃₆).....	<100	500	8,000
Macallen (z ₃₆).....	<100	<100	4,000

antigen not found in 4598-53 and Macallen, and 4598-53 contained an antigen not shared by Weslaco. For practical purposes the antigenic formula of the *E. freundii* culture may be expressed by the *Salmonella* formula 3, 15: z₃₆.

SUMMARY

A typical culture of *Escherichia freundii* isolated from a normal food handler was found to have both O and H antigens in common with the *Salmonella* group. The antigens of the culture could be expressed by the *Salmonella* formula 3, 15: z₃₆.