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Complete Genome Sequence of *Exiguobacterium* sp. Strain MH3, Isolated from Rhizosphere of *Lemna minor*

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We report the complete genome sequence of *Exiguobacterium* sp. strain MH3, isolated from the rhizosphere of duckweed. The genome assembly is 3.16 Mb, with a G+C content of 47.24%, and it may provide useful information about plant-microbe interactions and the genetic basis for the tolerance of the strain to various environmental stresses.

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Members of the genus *Exiguobacterium* are widely distributed in diverse habitats, including plant rhizospheres (1), biofilms (2), freshwater (3), marine waters (4), hot/cold environments (5, 6), and extreme conditions (7, 8). To date, 14 species of the genus have been isolated and characterized. They show great potential in industrial applications, including biotechnology, agriculture, food, and pharmaceutical industries, among others (6, 8, 9). Seven strains have had their genomes sequenced recently (6, 7), but none were isolated from a rhizosphere.

Duckweed, the smallest flowering plant, has been used widely for environmental monitoring and bioremediation and, lately, for bioenergy (10). Employing an ultrasonic method, *Exiguobacterium* sp. strain MH3, a psychrophilic facultative anaerobic bacterium, was isolated from the rhizosphere of a duckweed strain, *Lemna minor*, collected from Shenzhen, China (22°35′5.47″N, 113°57′36.37″E). On the basis of its 16S rRNA sequence, we determined that it is a member of the *Exiguobacterium* genus. MH3 was characterized as Gram-positive, non-spore-forming, having good mobility with flagella, and being able to grow between 4 and 40°C, at pH levels from 5 to 9.5, and with up to 10% NaCl (wt/vol).

Whole-genome shotgun sequencing of *Exiguobacterium* sp. MH3 was performed at the Beijing Genomics Institute (BGI) (Shenzhen, China) using Illumina HiSeq 2000 on a combination of 0.5-kb and 6.5-kb DNA libraries. This generated a total of 5,566,668 filtered paired-end reads, providing 140-fold coverage of the genome. The SOAP*denovo* alignment tool (11) assembled the reads into 1 scaffold, comprising 3 contigs. The gaps within the scaffold were confirmed and closed using long-distance PCR amplification by Sanger sequencing. Finally, a single replicon (total, 3,164,195 bp), with a G+C content of 47.24%, was obtained for the genome of *Exiguobacterium* sp. MH3.

Gene prediction and genome annotation were performed on the RAST server and the NCBI PAPPC (12, 13). tRNA and rRNA sequences were identified using tRNAscan-SE and RNAmmer, respectively (14, 15). In total, 3,203 coding sequences (CDSs) representing 398 predicted SEED subsystem features were predicted, including 2,479 proteins with identified functions. Nine rRNA operons and 60 tRNAs were also annotated. As expected, many stress tolerance-related genes were identified, including putative genes encoding a Pho regulon for high-affinity phosphate uptake (6), carbon starvation, oxidative stress, detoxification, and cold and heat shock proteins. Interestingly, genes related to auxin biosynthesis, siderophores, and iron acquisition and metabolism are also found in the MH3 genome, which may contribute to the promotion of plant growth, a definite benefit to its plant host.

Further analysis of the genome, including functional, molecular, and biochemical studies and plant-microbe relationship analyses, will be used to understand the mechanisms of MH3 for environmental stress tolerance, as well as its possible functions in promoting plant growth and stress resistance. This is the eighth draft genome for the genus *Exiguobacterium* but the only one isolated from a rhizosphere. It will provide a reference for many further phylogenetic, comparative genomic, metagenomic, and functional studies of this widely distributed genus.

Nucleotide sequence accession number. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession no. CP006866.

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REFERENCES

- Rodrigues DF, Tiedje JM. 2007. Multi-locus real-time PCR for quantitation of bacteria in the environment reveals *Exiguobacterium* to be prevalent in permafrost. FEMS Microbiol. Ecol. 59:489–499.
- Carneiro AR, Ramos RT, Dall'Agnol H, Pinto AC, de Castro Soares S, Santos AR, Guimarães LC, Almeida SS, Baraúna RA, das Graças DA, Franco LC, Ali A, Hassan SS, Nunes CI, Barbosa MS, Fiaux KK, Aburjaile FF, Barbosa EG, Bakhtiar SM, Vilela D, Nóbrega F, dos Santos AL, Carepo MS, Azevedo V, Schneider MP, Pellizari VH, Silva A. 2012. Genome sequence of *Exiguobacterium antarcticum* B7, isolated from a biofilm in Ginger Lake, King George Island, Antarctica. J. Bacteriol. 194:6689–6690.
- 3. Raichand R, Pareek S, Singh NK, Mayilraj S. 2012. Exiguobacterium aquaticum sp. nov., a member of the genus Exiguobacterium. Int. J. Syst. Evol. Microbiol. 62:2150–2155.

- 4. Kim IG, Lee MH, Jung SY, Song JJ, Oh TK, Yoon JH. 2005. *Exiguobacterium aestuarii* sp. nov. and *E. marinum* sp. nov., isolated from tidal flat of the yellow sea in Korea. Int. J. Syst. Evol. Microbiol. 55:885–889.
- Vishnivetskaya TA, Lucas S, Copeland A, Lapidus A, Glavina del Rio T, Dalin E, Tice H, Bruce DC, Goodwin LA, Pitluck S, Saunders E, Brettin T, Detter C, Han C, Larimer F, Land ML, Hauser LJ, Kyrpides NC, Ovchinnikova G, Kathariou S, Ramaley RF, Rodrigues DF, Hendrix C, Richardson P, Tiedje JM. 2011. Complete genome sequence of the thermophilic bacterium *Exiguobacterium* sp. AT1b. J. Bacteriol. 193: 2880–2881.
- White RA III, Grassa CJ, Suttle CA. 2013. Draft genome sequence of Exiguobacterium pavilionensis strain RW-2, with wide thermal, salinity, and pH tolerance, isolated from modern freshwater microbialites. Genome Announc. 1(4):e00597-13. doi:10.1128/genomeA.00597-13.
- Ordoñez OF, Lanzarotti E, Kurth D, Gorriti MF, Revale S, Cortez N, Vazquez MP, Farías ME, Turjanski AG. 2013. Draft genome sequence of the polyextremophilic *Exiguobacterium* sp. strain S17, isolated from hyperarsenic lakes in the Argentinian Puna. Genome Announc. 1(4): e00480-13. doi:10.1128/genomeA.00480-13.
- Jiang X, Xue Y, Wang L, Yu B, Ma Y. 2013. Genome sequence of a novel polymer-grade L-lactate-producing alkaliphile, *Exiguobacterium* sp. strain 8-11-1. Genome Announc. 1(4):e00616-13. doi:10.1128/genomeA.00616 -13.
- Margesin R, Feller G. 2010. Biotechnological applications of psychrophiles. Environ. Technol. 31:835–844.

- 10. Xu J, Zhao H, Stomp AM, Cheng JJ. 2012. The production of duckweed as a source of biofuels. Biofuels 3:589–601.
- 11. Luo R, Liu B, Xie Y, Li Z, Huang W, Yuan J, He G, Chen Y, Pan Q, Liu Y, Tang J, Wu G, Zhang H, Shi Y, Liu Y, Yu C, Wang B, Lu Y, Han C, Cheung DW, Yiu SM, Peng S, Zhu X, Liu G, Liao X, Li Y, Yang H, Wang J, Lam TW, Wang J. 2012. SOAPdenovo2: an empirically improved memory-efficient short-read de novo assembler. GigaScience 1:18. doi:10.1186/2047-217X-1-18.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. doi:10.1186/1471-2164-9-75.
- Pruitt KD, Tatusova T, Klimke W, Maglott DR. 2009. NCBI reference sequences: current status, policy and new initiatives. Nucleic Acids Res. 37:D32–D36. doi:10.1093/nar/gkn721.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res. 25: 955–964.
- Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res. 35:3100–3108.