Recognition and Control of the Progression of Age-Related Hearing Loss

Hong Miao Ren, Jihao Ren, and Wei Liu

Abstract

Recent breakthroughs have provided notable insights into both the pathogenesis and therapeutic strategies for age-related hearing loss (ARHL). Simultaneously, these breakthroughs enhance our knowledge about this neurodegenerative disease and raise the question of whether the disorder is preventable or even treatable. Discoveries relating to ARHL have revealed a unique link between ARHL and the underlying pathologies. Therefore, we need to better understand the pathogenesis or the mechanism of ARHL and learn how to take full advantage of various therapeutic strategies to prevent the progression of ARHL.

Introduction

GE-RELATED HEARING LOSS (ARHL), also known as Apresbycusis, which is a progressive, bilateral, symmetrical hearing decline associated with difficulty in speech discrimination, sound detection, and localization, influences an individual's well being and productivity and causes communication problems, isolation, and social withdrawal.¹ ARHL is generally considered to be a multi-factorial progressive disease caused by numerous factors, such as noise, ototoxic drugs, otologic disease history, trauma, vascular insults, metabolic changes, hormones, diet, and immune system that are superimposed upon an intrinsic, genetically controlled, aging process.^{2,3} The prevalence of ARHL increases proportionally with age. Hearing impairment is one of the most prevalent neurological disorders in humans. According to the World Health Organization (WHO), by 2025 there will be approximately 1.2 billion people in the world over the age of 60; this growth marks a shift in world population to a greater proportion of elderly people. The Medline Plus Medical Encyclopedia estimated that ARHL is projected to affect 25% of people aged 65-75 years and 70%–80% of people over the age of 75.

Although the potential mechanisms underlying ARHL have not been established, the age-related decline in acuity is generally attributed to at least two factors: (1) Progressive peripheral degeneration involving the degeneration of the stria vascularis,⁵ hair cells, spiral ganglion neurons (SGNs),⁶ and fibrocytes,⁷ leading to the disability of the sound afferent, and (2) a central degeneration that involves neuron degeneration in the primary auditory cortex (AI),^{8,9} thalamus,¹⁰

synapses in the hippocampal CA3 region¹¹ and the dorsal cochlear nucleus (DCN)¹² that results in various degrees of the central nervous system (CNS) executive dysfunction to various degrees in many patients with this disorder. The symptoms and signs emerge when these two factors are present. Mouse models are used to mimic different forms of human ARHL. Although the cochlear regions of a given type of pathology in animals may be different from humans, they exhibit similar degeneration of the organ of corti and SGNs, except for the stria vascularis.¹³ Schuknecht¹⁴ reported that approximately 30% of the patients with ARHL exhibit degeneration of the stria vascularis. Thus, animal models are appropriate for studying human ARHL.

The age of clinical onset is variable, but ARHL usually begins after 60 years. Neither the pathogenesis nor the symptoms of the disease have a remarkable onset; however, the disease progresses relentlessly. Recent discoveries related to ARHL have revealed a unique link between ARHL and the underlying pathologies. Therefore, we need to better understand the pathogenesis or the mechanism of ARHL and learn how to take full advantage of various therapeutic strategies to prevent the progression of ARHL. Accumulating evidence15,16 from genetic and electrophysiological studies postulates that reactive oxygen species (ROS)-related mitochondrial (mtDNA mutations and other gene mutations contribute to ARHL; a significant load of acquired mtDNA mutations have been shown in ARHL auditory tissue. Recently, several lines of evidence indicate that oxidative phosphorylation dysfunction is the common pathway of ARHL.¹⁷ Amounts of correlative data from morphological, bioenergetic, biochemical, and genetic studies that support the mitochondrial theory of aging¹⁸

Department of Otolaryngology Head and Neck Surgery, the Second Xiangya Hospital, Central South University, Changsha, Hunan, P.R. China.

Mutant molecules	Chromosome	Type (deafness)	Genus	References
Enzyme				
SŎD1	21	ARHL, NIHL	Mice	39, 43, 44
mtDNA				
mtDNA 4977 bp	(1)	ARHL	Human, Rats	51, 52
mtDNA 4834 bp	2	ARHL	Rats	53
mt-DNA A3243G	3	ARHL	Rats	54, 55
mt-Tr	4	ARHL	Mice	17
Gene				
Cdh23	10	ARHL	Mice	57, 58, 59
mdfw	10	ARHL	Mice	60, 61
waltzer (v)	10	ARHL	Mice	60
Ahl	10	ARHL, NIHL	Mice	62
Ahl2	5	ARHL	Mice	63
Ahl3	10	ARHL, NIHL	Mice	64, 65
Ahl4	10	ARHL	Mice	68
Ahl5	10	ARHL	Mice	69
Ahl6	18	ARHL	Mice	69
Ahl8	11	ARHL	Mice	70
Phl1	17	ARHL	Mice	71
Phl2	10	ARHL	Mice	71

 TABLE 1. GENETIC CAUSES OF AGE-RELATED HEARING LOSS

SOD1, superoxide dismutase 1; mtDNA, mitochondrial DNA; ARHL, age-related hearing loss; NIHL, noise-induced hearing loss; mt-Tr, a single nucleotide insertion in the *tRNA-Arg* gene; *mdfw*, *modifier of deaf waddler* ① nucleotide positions 13,447–13,459 and 8470–28,482 ② deletion junction (bp 8103, bp 12,937–12,952); ③ tRNA^{Leu(UUR)} ④, an adenine at position 9826 in mt-Tr.

are as follows. Normal metabolism causes ROS production by the electron transport chain; subsequently, ROS, which are generated inside the mitochondria, damage lipids, proteins,¹⁹ and the nucleic acids in mitochondria.²⁰ As a result, the ROSinduced mtDNA mutations lead to the synthesis of functionally impaired respiratory chain subunits, which cause respiratory chain dysfunction and augmented ROS production.²¹ Finally, the accumulation of this damage ultimately leads to permanent age-related mitochondrial dysfunction, which in turn leads to apoptosis and contributes to ARHL.²² Thus, we may speculate that inherited gene mutations or the ROS-predisposed mtDNA variants play a major role in the progression of ARHL.

Previous investigations have reported that caloric restriction (CR), which retards several aspects of the aging process in multiple species and slows the progression of ARHL in C57BL/6J mice, reduces the levels of apoptosis in the cochlea.²³ Apoptosis may play a key role in the age-related decline of physiological function in the aging cochlea through suppressing the age-related increases in capase-3, capase-9,²⁴ and Bcl-2 family members, such as Bak.

In the present review, we begin by summarizing the initiating events by which ROS induce apoptosis. Then we outline several types of mtDNA mutations and other gene mutations (Table 1) and review recent insights into their mechanism (Fig. 1), and emerging themes in their pathogenesis followed by discussions of the current preventative strategies. Finally, we discuss the challenges and opportunities that lie ahead for ARHL.

ARHL: Oxidative Phosphorylation Dysfunction-Related Apoptosis

Since mitochondrial disease was first described by Rolf Luft and his colleagues in $1959_{,}^{25}$ more than 150 distinct

genetic mitochondrial syndromes have been defined, such as lactic acidosis, skeletal myopathy, blindness, sub-acute neurodegeneration, intestinal dysmotility, and peripheral neuropathy, especially deafness.²⁶ Mitochondria are a major source of ROS-induced oxidative damage, which is consistent with the mitochondrial free radical theory of aging in which aging results from the accumulated oxidative damage caused by ROS, originating from the mitochondrial respiratory chain.²⁷ Various of studies have shown that mitochondrial ROS play a critical role in cochlear aging, even in ARHL.²²

As our ability to define the genetic and environmental bases of mitochondrial disorders has accelerated, new questions have arisen. How can ROS result in mitochondrial dysfunction? The main pathway relevant to the two events awaits further study. Apoptosis may make a great contribution to the age-related decline of physiological function in the cochlea.²⁸ Moreover, the progression of ARHL in C57BL/6J mice increases the levels of apoptosis in the cochlea and reduces the levels of the mitochondrial proapoptotic Bcl-2 family member Bak.²⁹ It has also became clear that either Bcl-2 and other members of the family are essential for apoptosis.^{28,30,31} Bak and Bax, either directly or indirectly, induce the release of cytochrome c and mitochondrial fragmentation.³² Subsequently, cytochrome c diffuses from the inter-membrane space into the cytosol³³ and leads to caspase activation^{34,35} and apoptosis. All of the factors mentioned above that are sufficient to induce or inhibit the apoptosis pathway are similar to what has been described for Alzheimer disease, amyotrophic lateral scle-rosis (ALS), and so on.^{36,37} Aggregates of mtDNA mutations that promote apoptosis may be a central mechanism that drives ARHL during mammalian aging. By interfering with various cellular functions, such as intracellular transport³⁸ and mitochondrial metabolism, cellular stress ultimately gives rise to mtDNA mutations.

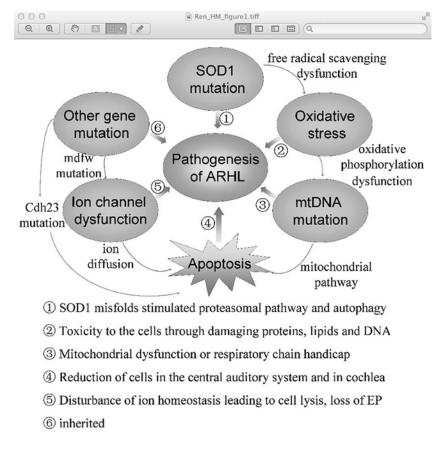


FIG. 1. The mechanism of age-related hearing loss (ARHL). The *SOD1* mutation, as an initial event accelerating ARHL through stimulating the proteasomal pathway and autophagy, directly leads to free radical scavenging dysfunction, which in turn stimulates mitochondrial (mt) DNA mutation. mtDNA mutations are not only toxic to the auditory cells by damaging proteins, lipids, and DNA in cells but also results in cell apoptosis through the mitochondrial pathway, which untimately leads to ARHL. Other gene mutations such as the Cdh23 mutation may be harmful to the stereociliary tip link and cause hair cell apoptosis from the base to apex, leading to early onset of ARHL. Meanwhile, the *mdfw* mutation leads to the plasma membrane Ca^{2+} ATPase2 dysfunction, finally resulting in cell apoptosis and even the appearance of ARHL.

SOD1 mutation : Initial event accelerating mtDNA mutation

Superoxide dismutase 1 (SOD1), composed of 153 amino acids, is a cytosolic enzyme that converts the superoxide radical to hydrogen peroxide and O₂. SOD is the only eukaryotic enzyme that is capable of detoxifying ROS and is involved in free radical scavenging.³⁹ A study showed that *sod-12345 Caenorhabditis elegans* worms (a *sod* quintuple mutant that lacks all five *sod* genes) exhibit a specific sensitivity to ROS and use multiple compensatory mechanisms to maintain low levels of oxidative damage.⁴⁰ Previously, the mechanism of SOD1 in the degenerative diseases such as ALS has been illustrated. Mutant SOD1 misfolds and is targeted for degradation through ubiquitylation. Additionally, mutant SOD1 seems to have adverse effects against the cell's degradation machinery by impairing the proteasomal pathway and autophagy.^{41,42}

Various types of genetic study have shown that deletion of SOD1 leads to accelerated ARHL with underlying pathologies of severe SGN loss, reduced size of the stria vascularis, and accelerated out hair cell (OHC) loss.⁴³ Meanwhile, another investigation demonstrated that SOD1 appeared to be important for the survival of cochlear neurons and the stria

vascularis; however, even half the amount is sufficient and an over-abundance does not provide much protection from ARHL.³⁹ Interestingly, genetic over-expression of SOD1 has not been shown to provide a protective benefit against ARHL.^{39,44} Further interventions may focus on increasing the activity of anti-oxidant enzymes or decreasing the production of free radicals. This endeavor could be an effective method for reducing age-related degeneration of sensory cells in the cochlea. Collectively, all of these studies may lead to a conclusion that it is critical to maintain the balance of anti-oxidant/oxidative stress for ARHL.

mtDNA mutations

mtDNA mutations are hypothesized to cause aging and have been found to increase in a variety of tissues in aging individuals.⁴⁵ The "common" deletion (CD) was initially identified in patients with Kearns–Sayre syndrome⁴⁶ and was detected in heart muscle, brain, liver, and skeletal muscle of elderly individuals.^{47–49} The results from animal experiments and human autopsy studies have suggested that somatic mtDNA mutations, including CD, are considered to be the significant underlying factor in the progression of ARHL.^{47,50,51} Previous studies have revealed that the mtDNA 4977-bp mutation, known as the CD, plays an important role in ARHL.⁵¹ Numerous investigations have been conducted to demonstrate the association between elevated mtDNA mutation levels and ARHL. It is reported that 14 out of 17 patients with presbycusis had detectable levels of the mtDNA 4977-bp deletion in archival temporal bone, whereas only eight of the 17 human specimens presented the mtDNA 4977-bp deletion with normal audiograms.⁵² However, the mtDNA 4977-bp deletion also occurs in young people and in children with hearing loss. All of those reports indicated that the CD occurred not only in elderly individuals with normal hearing but also in young people. So, we surmise that the effect of the CD on ARHL is still poorly understand. Meanwhile, a similar mtDNA 4834-bp deletion that resulted in the down-regulation of the activity of anti-oxidant enzymes and the up-regulation of lipid peroxidation was also detected to occur in naturally aging rats.53 However, several studies have indicated that the mtDNA 4834-bp and mtDNA A3243G deletions⁵⁴ increase in aged rats and have a great relevance with ARHL,55 whereas Kong reported that the mtDNA 4834-bp deletion cannot directly induce the hearing loss, but can act as a predisposing factor that can greatly enhance the sensitivity of the inner ear to the aminoglycoside antibiotic.56 Subsequently, the association between the mtDNA A3243G deletion, diabetes mellitus, and hearing loss was confirmed in population studies of patients with diabetes. On the basis of these controversial results, we proposed that the mtDNA mutation might also occur in people without ARHL and the CD might not directly induce ARHL. Therefore, the exact mechanism of mtDNA deletion on ARHL needs further basic and clinical studies.

Gene mutations

Cadherin 23 (CDH23), which is encoded by CdH23, is an important constituent of the hair cell tip link in the organ of Corti. Thus, mutations in CdH23 lead to mechano-electrical transduction dysfunction and are associated with ARHL in humans and C57BL/6J mice.57-59 A series of loci on chromosome 10, such as *mdfw*, *Ahl*, and *waltzer* (v), are also associated with ARHL. The mdfw locus on chromosome 10 modifies the hearing loss phenotype in mice in heterozygous for Atp2b2 (plasma membrane Ca²⁺ATPase2 gene).⁶⁰ The genetic complementation tests indicate that *mdfw* may be allelic with Ahl.61 The Cdh23 gene is localized in the mdfw and ahl interstitial genomic region and responsible for ARHL.⁵⁷ The predisposition to early-onset ARHL that is conferred by Cdh23 depends on the effects of mitochondrial mutation mt-Tr in A/J mice.¹⁷ Combination of the "accelerating alleles" with *Cdh23* is sufficient to induce ARHL onset. A study of Cdh23^{nmf308/nmf308} in mice reported that the CdH23 mutation may be harmful to the stereociliary tip link and causes hair cell apoptosis from the base to apex, which leads to early onset of ARHL.59 All of those reported data indicated that homozygosity with respect to CdH23 significantly increases the susceptility to ARHL, but it is not the only cause for its pathogenesis and symptoms in phenotypic manifestation. We may conclude that the occurrence of ARHL not only attributed to the Cdh23 mutation but also to the additive or stochastic interactions of secondary factors with Cdh23. The genetic architecture of ARHL may provide a paradigm for predisposition to ARHL in humans and defines a presbycusis model to explore therapeutic avenues, such as stem cell therapy.

In addition to the discovery of the mtDNA mutation, predisposing genetic factors are also responsible for most forms of ARHL. Ahl is the first and major gene (locus on chromosome 10) that affects late-onset of ARHL and has been mapped in mice.⁶² The Ahl gene has an epistatic interaction with A/J-derived mtDNA, wherein homozygosity for the predisposing Ahl allele is a prerequisite for the manifestation of the mtDNA effect on hearing loss.¹⁷ Furthermore, it may increase the susceptibility of mice to noiseinduced hearing loss (NIHL) so that preventative measures can be taken, such as minimizing exposure to loud sounds. Therefore, Ahl not only directly affects the onset of ARHL but it also indirectly plays a role in NIHL by interacting with mtDNA and up-regulating the susceptibility to loud sounds. The same author later reported the genetic mapping of *Ahl*2 (a locus on chromosome 5) that contributes to the difference in ARHL onset times between NOD/LtJ (NOD) and C57BL/ 6J (B6) mice. Moreover, the influence of Ahl2 on ARHL seems to be more strain specific than that of Ahl, and may be restricted to NOD-derived strains.⁶³

The molecular identification of ARHL-contributing genes, such as Ahl and Ahl2, in experimentally amenable inbred mouse strains will help to identify similar genes in human populations. In 2007, a study demonstrated that Ahl3 (a locus on chromosome 17) exists within the region between D17Mit274 and D17Mit183 and affects not only ARHL but also renders in B6 mice more susceptible to NIHL, which is highly similar to Ahl. The interaction of Ahl1 or Ahl3 on ARHL is attributed to the resistant alleles in either locus, such as *persephin* (Pspn), neurturin (Nrtn), and solute carrier family members (Slc25a23 and Slc5a7).64,65 Studies of knockout mice for anti-oxidant enzyme copper/zinc superoxide dismutase (Cu/Zn SOD) and cellular glutathione peroxidase (GPX) genes and ion regulators showed they were more vulnerable to ARHL. Of note, the candidates of Ahl3 factors described above seem to be functionally related to Cu/Zn SOD and GPX and ion regulator genes.66,67 Meanwhile, Ahl4, a locus on chromosome 10 contributes to the early-onset, rapidly progressing hearing loss of A/J mice because of its earlier and more severe hair cell loss in A/J mice than in B6 mice during the first 5 months of age.⁶⁸ The authors also reported the effects of Ahl5, and Ahl6,⁶⁹ Ahl8,⁷⁰ Phl1, and Phl2⁷¹ on ARHL and indicated that they can be tested as candidate genes for ARHL susceptibility. Gene identification could provide entry to the molecular pathways of pathogenesis and pathophysiology of ARHL and may contribute to the development of diagnostics, which suggests a possible means of preventative interventions and therapies for human presbycusis.

Other factors inducing ARHL

 Ca^{2+} . An *in vitro* study detected that overstimulation of isolated outer hair cells (OHCs) will up-regulate the level of cytoplasmic [Ca²⁺].⁷² Phosphatidylinositol phosphate kinase type 1_{γ} (PIPKI_{γ}) is a key enzyme in the generation of phosphatidylinositol 4,5-bisphosphate (PI[4,5]P₂) and is primarily responsible for the synthesis of the receptor-regulated phospholipase C (PLC)-sensitive PI(4,5)P₂ pool in the cell syncytia that supports auditory hair cells. Homozygous

knockout mice lacking this enzyme die post-natally within 24 hr. PIPKI_v catalyzes the phosphorylation of phosphatidylinositol 4-phosphate (PI[4]P) at the D-5 position of the inositol ring, ultimately resulting in $PI(4,5)P_2$ formation, a minor glycerophospholipid of the inner leaflet of the plasma membrane involved in cell signaling cascades. Then, $PI(4,5)P_2$, either combined with the $PI(4,5)P_2$ -binding proteins or acting as the substrate of $PI(3,4,5)P_3$ kinase,⁷³ is converted into diacylglycerol and inositol 1,4,5-triphosphate (IP₃) generated by PLC-dependent hydrolysis of PI(4,5)P₂. IP3 activates Ca²⁺ efflux from the endoplasmic reticulum, raising the cytosolic free Ca^{2+} concentration ($[Ca^{2+}]_i$) by binding to its receptors (IP_3R)⁷⁴ or through gap junction (GJ) channels such as P2x7 receptors (P2x7Rs) and pannexin-1 channels,^{75,76} which have been implicated in adenosine triphosphate (ATP) secretion from glial cells during Ca²⁺ wave propagation⁷⁷ and amplification of astrocytic ICS.⁷⁸ The coexpression of Px1 and P2x7R produces a signaling complex that, when activated by ATP, in turn opens a cation channel, which creates a trans-membrane pathway that is sufficiently large to allow the passage of large-molecular-weight fluorescent dyes (up to 900 Da), and cell lysis.^{79,80}

The formation of vast functional syncytia coupling transfer of ions, signaling molecules, and nutrients through GJ channels contributes to the functional maturation of the cochlea.⁸¹ Therefore, alterations of IP3R Ca²⁺ signaling in non-sensory cells of the developing cochlea are linked to impaired hearing acquisition in mice. The PIPKI γ deficiency in mice results in a sizeable and spatially graded down-regulation of IP₃R Ca²⁺ signaling along the sensory epithelium of the cochlea. On the basis of the primary role played by PIPKI γ in IP₃ signaling, we surmised that a reduction of PI(4,5)P₂ levels may affect the development of hearing capability. Accordingly, transgenic mice with PIPKI γ deficiency study showed that PIPKI γ plays a key role in the inner ear and that it has an essential function for the acquisition of hearing.⁸²

The disturbance of neuronal calcium homeostasis has long been suggested to be an important factor underlying agerelated impairment of neuronal function.⁸³ Numerous investigations have focused on the effect of T-type calcium channels in ARHL of SGNs. B6 mice homozygous or heterozygous for a null allele of the α 1H calcium channel subunit can significantly slow age-related auditory brainstem response (ABR) threshold shifts, after a dramatic decrease in age-related SGNs loss.³⁸ Thus, anti-epileptic drugs that block T-type calcium channel may lead to better hearing thresholds in females during aging,⁸⁴ which suggests that calcium dysregulation contributes to ARHL.

The large conductance voltage- and Ca²⁺-activated potassium (BK) channel, which is composed of the pore-forming α -subunit co-expressed with the auxiliary β 1-subunit, has been suggested to play an important role in the signal transduction process of cochlear IHC. Consistent with the phenotype elicited by the pharmacologic blockade of KCNQ4 channels, BK channel α -subunit (BK $\alpha^{-/-}$) and β 1-subunit (BK β 1^{-/-}) knockout mice were accompanied by OHC dysfunction. The human BK-coding *slo1* gene mutation as a susceptibility factor for progressive deafness is similar to KCNQ4 potassium channel mutations.^{85,86}

 K^+ . Several K^+ channels and transporters occur in various compartments of the stria vasularis with a K^+ -rich

extracellular fluid known as endolymph, which is essential for preserving the sensory structures and supporting transduction and has been shown to be involved in EP formation. Maintaining the endolymph homeostasis is critical to sustain auditory functions. Na⁺, K⁺-ATPase, and Na⁺, K⁺, 2Cl⁻ co-transporter (NKCC) are localized on the basolateral membranes of marginal cells^{87–90} and are linked to the maintenance of the inner ear osmotic balance⁹¹ and ARHL.⁹² The loss of EPs, which results from a decrease of Na, K-ATPase activity, is thought to be the causative factor of ARHL in aged Mongolian gerbils. Subsequently, the loss of ATP in the strial cells may lead to permanent cell damage and/or dysfunction of the Na⁺-K⁺-ATPase pump, with the resulting in a long-term reduction of the EP.

Vibration of the basilar membrane opens the mechanosensitive channels in hair-cell stereocilia; the ensuing K⁺ influx⁹³ and the permeation of Ca²⁺ amplify the motility of hair bundles.^{94,95} After K⁺ exits hair cells on their basolateral surfaces, it must cycle back to the endolymph through a pathway comprising perilymph, supporting cells, the spiral ligament, and the stria vascularis, including MCs.^{90,96,97} One study⁹⁸ reported that intrastrial potential (ISP) represents the K⁺ diffusion potential, which also contributes to the evoked potential (EP) across the apical membranes of intermediate cells through Ba²⁺-sensitive K⁺ channels. EP originates from the ISP, which indicates that the potential difference across the membranes of intercellular space (IC) can be explained largely as K⁺ diffusion potential and underlies the ISP^{99,100} interference with any of these elements to interrupt hearing.

ATP α is the sole Na pump α -subunit in auditory organs. ATP α preferentially localized to the scolopale cell ablumenal membrane and consistent with a role in pumping K⁺ ions into the scolopale cell en route to the receptor lymph, which resembles its contribution to generating vertebrate inner ear endolymph. Furthermore, the nrv2 and nrv3 of ATP α subunits show cell-type-specific expression and functions to scolopale cells and neurons, respectively. ATP α is required for hearing.¹⁰¹

Fibrocytes. Spiral ligament (SL) fibrocytes in the lateral wall of the cochlea supply K^+ to the strial pump vial gap junctions by recycling back into the stria vascularis. Na-K-ATPase–rich strial marginal cells could pump the K^+ ions into the cells, with K^+ subsequently flowing down its concentration gradient into the endolymph. The fibrocytes will take K^+ from the perilymph to the endolymph.¹⁰² A recent study in 129S6 and BALB/c mice found that dysfunction of stellate fibrocytes could disrupt a medial K^+ recycling pathway that serves to remove potassium from around the inner hair cells along this pathway. Perhaps a widespread degeneration of cells in the cochlea can critically impair ion and fluid homeostasis.¹³

Inflammation. Once the inner ear is exposed to bacteria, endotoxin and an abundance of monocytes and macrophages are transported into the perilymph.¹⁰² Additionally, the increase in the generation of ROS combined with the imbalance of anti-oxidant enzymes such as manganese superoxide dismutase (MnSOD) and GPX may cause a persistent inflammation.^{7,103} The release of ROS will activate the cell signaling pathways including mitogen-activated protein kinase/extracellular signal-regulated kinase (MAPK/ERK)

kinase (MEK), and ERK phosphorylation and activation of specific transcription factors (*e.g.*, nuclear factor- κ B [NF κ B], AP-1) are also increased along with the release of ROS. In response to ROS, mitochondrial dysfunction, increased gene expression of death receptors, and/or their ligands (tumor necrosis factor- α [TNF α], FasL) appeared. All of the factors activated by the ROS in the perilymph might act directly on the vascular endothelial cells and the SL fibrocytes, which will produce various inflammatory mediators. Finally, those pro-inflammatory cytokines could cause the SL fibrocyte damage that is seen with cochlear inflammation. It is reasonable to think that chronic inflammation is one of the causal factors that might trigger premature ARHL.

Therapeutic Strategies for ARHL Derived from the Biological Pathogenesis

The development of effective therapeutic strategies to prevent or treat ARHL has proven to be a relatively difficult progress, which is demonstrated by the lack of the restorative medicines and technologies. To the best of our knowledge, ARHL is primarily attributed to the inner ear HC, subsequently leading to SGN degeneration. The ensuing HC loss is irreversible because of its limited capacity to regenerate in mammals. Therefore, we would like to explore the advantages and limitations associated with the currently available strategies for inner ear restoration, *i.e.*, stem cellbased therapy, gene therapy, anti-oxidant defense, and operant training.

Stem cell-based therapy

Stem cell-based regenerative therapy is a potential cellular therapeutic strategy for patients with incurable brain diseases. The stem cell transplantation strategy has received significant attention as an alternative therapy for ARHL in pre-clinical studies. The cellular and biological mechanisms underlying the observed effects of a stem cell-based treatment in an animal model need to be determined.¹⁰⁴ First, it might be possible to replace lost neurons or cochlear cells by transplantation of stem cell-derived cells that have been predifferentiated in vitro to various stages of maturation. Second, cell replacement might also be achieved by inducing endogenous stem cells in the adult CNS or cochlear to form new neurons, HCs, and SGNs. Finally, the grafted stem cells and their derivatives could induce optimal recovery that could also be of clinical value by releasing therapeutic molecules for immunomodulation, trophic actions, neuroprotection, and stimulation of angiogenesis.¹⁰⁵

In a study of senescence-accelerated mice (SAMP1),¹⁰⁶ allogeneic bone marrow cells derived from BALB/c(H-2^K) mice were transplanted in the SAMP1 mice with ARHL. Allogeneic bone marrow transplantation (BMT) was found to prevent the development of immunological dysfunction, hearing loss, and apoptosis of the spinal ganglion cells by replacing abnormal hematopoietic stem cells (HSC_S) with normal HSC_S. Finally, the author demonstrated that the accelerated presbycusis resulted not from the defects in the cochlea, but from the defects in the HSCs, which are differentiated into immunocompetent cells. Although several studies have revealed that autoimmune mechanisms participate in the occurrence of ARHL, this factor may be one of the mechanisms that resulted in ARHL; however, it is not the

only factor. Just as we mentioned before, ARHL is attributed to the pathology of the CNS and peripheral system including cochlea.

The conclusions in the SAMP1 mice study and the other studies seem to be contradictory.¹⁰⁷ We have accessed the capability of intra-cerebral embryonic neural stem cell (NSC) transplantation for C57BL/6J mice with ARHL, showing that auditory function of C57BL/6J mice can be partially restored by intra-cerebral embryonic NSCs transplantation. Just as we surmised, despite the achievements in the treatment of the degenerative diseases described above, ARHL is more suitable for stem cell therapy because of its relatively better environment for graft survival without inflammation compared to stoke and rare complications concerning of Parkinson disease after regeneration therapy. In addition to the transplantation of stem cells into the central nervous system, several studies focused on the regeneration of the cochlear cell by stem cell transplantation. It has been observed that transplantation of various types of stem cells derived from embryonic, neonatal, and adult animals have been observed to differentiate into HC or SGNs containing similar markers and proteins with functional properties. Four weeks after transplantation of induced pluripotent stem cells (iPSCs) into the cochlea of C57BL/6J mice, most parts of the cochlea exhibited the settlement of iPSCs.¹⁰⁸ Previously, embryonic stem cells (ESCs) have also been used to generate neurons that make synaptic contacts with HC.¹⁰⁹ Recently, a novel protocol demonstrated that two types of otic progenitors that differentiated from human (h) ESCs can engraft, differentiate, and significantly improve auditory-evoked response thresholds after being transplanted into an auditory neuropathy model.¹¹⁰

Despite the effectiveness we currently observed, future studies must face the challenge of finding the mechanisms underlying the observed improvements: (1) How can we make best use of stem cells that may be better compliant mechanically and structurally with native tissues and integrate the transplanted tissue with the native tissue without immunological rejection?¹¹¹ (2) How can we make them grow where and how we want? (3) What is the correlation between the grafted cells and the native cells and how do the responses to downstream networks affect the computations of behavioral functions?¹¹² More solid pre-clinical studies have to be conducted prior to further patient applications.

Gene therapy

Gene therapeutic strategies have long been considered to be a promising and novel approach for delivering therapeutic genes or RNA interference (RNAi) to treat ARHL; gene strategies have been applied in the inner ear pathologies both *in vivo* and *in vitro*. On the basis of the reported findings, the *Atoh1* gene is most commonly applied to stimulate HC production and to provide modest improvements in hearing function.¹¹³ A number of genes driving the morphogenesis of inner HC differentiation, especially reduction and delayed deletion of *Atoh1*, result in the progressive degeneration of the majority of the HC within 3 weeks after birth.^{114,115} Interestingly, RNAi-mediated gene suppression is typically superior to the methods of gene knockdown or direct gene therapy for its exquisite sensitivity to the target sites.¹¹⁶ The transfer results of the *Akv-1* and

apoA-II genes were previously reported. Both of these genes are potential genetic markers, indicating that are potentially useful for ARHL diagnosis.^{117,118} However, the therapeutic genes can also be mediated by stem cells and then delivered to animal models. Compared with direct gene transfer, an advantage of cell-based gene delivery is that production of the trophic factor continues even if the disease process destroys the endogenous cells. In an in vitro study, an adenovectormediated Atoh1 gene induced HC-like cells after being delivered into human umbilical cord mesenchymal stromal cells (hUCMSCs).¹¹⁹ Nevertheless, vectors used to deliver the gene in animal models have been the subject of intense speculation and controversy for several years, as they played an important role in the efficiency of gene therapy. Further investigations need to take into account their characteristics of stability and target selection, off-target effects, immune activation, and toxicity to inner ear.¹²⁰

Anti-oxidant defense

Caloric restriction. We are all aware that CR, which entails reducing food consumption by 25%–60% without malnutrition,¹²¹ consistently extends the life span and health span of a variety of species and ARHL in mammals.^{23,122} As we have discussed in this review, ARHL is linked to accumulated ROS-induced oxidative damage after mitochondrial function decline.¹²³ Simultaneously, the accumulation of mtDNA mutations may also initiate apoptosis in the development of ARHL.¹²⁴

Numerous studies have suggested that CR can reduce the oxidative damage, enhance anti-oxidant defense,¹²⁵ upregulation of the sirtuin pathway, and stress responseinduced up-regulation of heat shock proteins and so on. Recent studies have suggested that the loss of critical cells through apoptosis is an important mechanism of ARHL in mammals and that CR can retard apoptotic cell death in the mouse cochlea and prevents the late onset of presbycusis.^{23,126}

The SIRT family of genes helps regulate gene silencing, DNA repair, rDNA recombination, apoptosis, 29,127 and mitochondrial energetics.¹²⁸ Shinichi Someya reported that CR may directly stimulate Sirt3, a member of the sirtuin family, to deacetylate; this, in turn, activates mitochondrial isocitrate dehydrogenase 2 (Idh2). Idh2 is an enzyme that converts 2-oxoaldehyde dehydrogenase (NADP⁺) to nicotinamide adenine dinucleotide phosphate (NADPH) in the mitochondria and leads to increased NADPH levels and an increased ratio of reduced oxidized/glutathione, a major redox couple in the cell. CR fails to modify cell phenotypes in mice in cultured cells lacking Sirt3, which is recognized to be an essential player in enhancing the mitochondrial glutathione anti-oxidant defense system during CR. Therefore, Someya et al. proposed that Sirt3-dependent mitochondrial adaptations may be a central mechanism for aging retardation in mammals.¹²⁴ In addition to using CR to prevent ARHL, another intriguing possible approach is to introduce pharmaceutical interventions mimicking CR. Those interventions would induce the sirtuin pathway in multiple tissues by increasing the oxidative stress resistance and preventing the mitochondrial decay associated with aging. Resveratrol (trans-3,5,4'-trihydroxystilbene), a naturally occurring phytoalexin produced by a wide variety of plants, represents an attractive medicinal source in the treatment of ARHL.129

Resveratrol has been reported to produce an anti-aging effect,¹³⁰ expansion of life span,¹³¹ and up-regulation of mitochondrial biogenesis.¹³² Resveratrol has recently been found to delay hearing loss in a F344/NHsd rat model.¹³³ Resveratrol therapy is a promising avenue to prevent the progression of ARHL.

Anti-oxidant enhancement

Vitamin C, vitamin E, and melatonin have been thought to be protective factors for ARHL.^{126,134} Animal models that administered with anti-oxidants vitamin C and vitamin E exhibited efficiency in ARHL.135 Melatonin, an indole hormone secreted primarily by the pineal gland, functions as a regulator of circadian rhythms and has been postulated to enhance the defense capacity of many anti-oxidative enzymes and thus to protect DNA, membrane lipids, and possibly cytosolic proteins against oxidative damage, thus functioning as an important neural anti-oxidant and free radical scavenger.134,136,137 A cross-sectional study was carried out to determine the correlation between the hearing threshold and the plasma melatonin and ascorbic acid (vitamin C) levels. Low plasma levels of melatonin are significantly associated with high-frequency hearing loss (HL) among the elderly with ARHL.¹³⁷ Melatonin also plays a role in the appearance of autoimmune deafness by acting upon lymphocytes.¹³⁸

Lecithin, as a polyunsaturated phosphatidylcholine (PPC), is an important component of biological membranes. PPC played a rate-limiting role in the activation of SOD and glutathione, which, in turn, protect cell membranes from damage by ROS. Increasing evidence has shown that lecithin may preserve cochlear mitochondrial function and protect against hearing loss associated with aging.¹³⁹ Moreover, it has been found that aged gerbils that were chronically pretreated with the spin-trapping compound N-tert-butyl- α phenylnitrone (PBN) had a significantly decreased level of oxidized protein and increased glutamine synthetase (GS) and neutral protease activities as compared to young adult gerbils. If PBN administration was stopped after 2 weeks, the significantly decreased level of oxidized protein and increased GS and neutral protease activities in old gerbils will return to its primary levels.140 Compared with the aforementioned compounds, the findings regarding SOD1 may provide an even more powerful therapy for ARHL. Mice with a complete loss of SOD1 exhibit an acceleration of ARHL, whereas there is no significant effect on ARHL from either the reduction or the over-expression of SOD1.39,135 Thus, in a study in which there was chronic administration of SOD1 mimetics, the Eukarion experimental compounds EUK-189 and EUK-207 almost completely reversed the cognitive deficits and increase in oxidative stress. Particularly, protein oxidation was completely improved.140 The demonstration of these methods signified the potential application of one or more medicine compounds to treat ARHL.

Operant training

ARHL is currently described as a degenerative disease with characteristics of age-related hearing decline associated with difficulty in speech discrimination, recognition, sound detection, and localization. Fortunately, both the animal and human studies have shown that extended training mostly improved discrimination thresholds¹⁴¹ through the change in auditory

pathways and the amygdala.^{142,143} The improvement is specific to the trained stimuli, which suggests that plasticity occurs in areas of the brain that have relevant information about lowlevel (for example, physical) properties of the stimuli.¹⁴⁴ It has been shown that attention and reward can modulate perceptual learning and influence its specificity.¹⁴⁵

The perceptual and cognitive declines resulting from ARHL cannot be explained solely by a dysfunction of peripheral sensory organs, and they frequently translate to slowed perceptual processing and difficulty in accurately identifying stimuli.146 A recent study on age-related cortical processing deficits in the primary auditory cortex of aging and young rats showed a nearly complete reversal of the majority of previously observed functional and structural cortical impairments. A significant positive impact on the frequency resolution and signal-in-noise processing of a series of cortical processing abnormalities appeared in both young and aged rats after a variety of operant training, for example, oddball training, whereas the response modulation was clearer and could also be the major contributor to the restoration of temporal selectivity in the aged auditory cortex. The authors concluded that age-related cognitive decline is a tightly regulated plastic process and demonstrated that most of these age-related changes are, by their fundamental nature, reversible.147 The underlying mechanisms with operant training may be specific refinement of cortical representations,¹⁴⁸ changes in protein expression,¹⁴⁹ and alterations in the cortical inhibitory circuitry¹⁵⁰ and supporting glia.¹⁵¹

Uncertainty surrounding other therapeutics

Various investigations have applied salicylate to treat noise-induced hearing loss. Although one author has mentioned that the possibility of actually improving hearing in the inner ear with ARHL is an exciting idea with significant clinical implications, but there is insufficient evidence to support the hypothesis to date.

Challenges and Opportunities

With the rapid development of technology, ARHL turns out to be a sophisticated disorder and this may explain why there are insufficient therapeutic strategies for ARHL until now. Moreover, the mutations in *SOD1* have paved the way for mtDNA mutations, which lead to the apoptosis in the auditory system. The discovery of *Cdh23*, *Ahl*, and other gene mutations may also give rise to progressive hearing loss in several patients as well as the entire families. Additionally, not only the gene mutations but also the diversity of ion diffusion through ion channels all contribute to the pathogenesis of ARHL. Therefore, the exact mechanisms underlying the peripheral degeneration and central degeneration are multi-factorial, thus posing a great challenge for both preclinical and clinical research on ARHL. Nevertheless, ARHL also offers many opportunities for therapeutic intervention.

Over the past few years, there has been continuous progress in our understanding the mechanisms underlying ARHL and developing approaches to ameliorate the auditory function so that the quality of life can be improved among elderly people suffering from this type of hearing loss. However, many essential questions await answers. How do we take full advantage of the cell- and gene-based therapies, drug delivery, operant training, and electrophysiologic strategies to induce the hearing curve back to its normal levels and make complete behavioral recovery in animal models? How do we evaluate the safety and efficacy of the strategies discussed above? The clinical application of these therapeutic interventions and acceptance of patients with pre-clinical evidence of efficacy and safety, as required for those strategies, is unacceptable and will delay the development of clinically useful therapies to some extent. Some patients with ARHL have few or no therapeutic options and are prepared to test any new approaches, whereas a large proportion of patients with ARHL have abandoned the chance of treatment. Thus, it is hoped that scientists and clinicians can act together responsibly to translate knowledge regarding the mechanisms of ARHL and the basic research into disease-modifying treatments that will make a substantial difference for patients.

Acknowledgments

This research was supported by Hunan Provincial Innovation Foundation for Postgraduate (CX2013B114).

Author Disclosure Statement

All of the authors declare that no competing financial interests exist during the process of writing and publication. The subjects' written was approved by the Second Xiangya Hospital, Central South University.

References

- Fetoni AR, Picciotti PM, Paludetti G, Troiani D. Pathogenesis of presbycusis in animal models: A review. Exp Gerontol 2011;46:413–425.
- Bao J, Ohlemiller KK. Age-related loss of spiral ganglion neurons. Hear Res 2010;264:93–97.
- 3. Huang Q, Tang J. Age-related hearing loss or presbycusis. Eur Arch Otorhinolaryngol 2010; **267:** 1179–91.
- Sprinzl G, Riechelmann H. Current trends in treating hearing loss in elderly people: A review of the technology and treatment options—a mini-review. Gerontology 2010; 56:351–358.
- Lang H, Jyothi V, Smythe NM, Dubno JR, Schulte BA, Schmiedt RA. Chronic reduction of endocochlear potential reduces auditory nerve activity: Further confirmation of an animal model of metabolic presbyacusis. J Assoc Res Otolaryngol 2010;11:419–434.
- Engle JR, Tinling S, Recanzone GH. Age-related hearing loss in rhesus monkeys is correlated with cochlear histopathologies. PloS One 2013;8:e55092.
- Menardo J, Tang Y, Ladrech S, Lenoir M, Casas F, Michel C, Bourien J, Ruel J, Rebillard G, Maurice T. Oxidative stress, inflammation, and autophagic stress as the key mechanisms of premature age-related hearing loss in SAMP8 mouse cochlea. Antioxid Redox Signal 2012;16:263–274.
- Hughes LF, Turner JG, Parrish JL, Caspary DM. Processing of broadband stimuli across A1 layers in young and aged rats. Hear Res 2010;264:79–85.
- Rauschecker JP, Tian B. Mechanisms and streams for processing of "what" and "where" in auditory cortex. Proc Natl Acad Sci USA 2000;97:11800–11806.
- Barkat TR, Polley DB, Hensch TK. A critical period for auditory thalamocortical connectivity. Nat Neurosci 2011; 14:1189–1194.

- 11. Yu Y-F, Zhai F, Dai C-F, Hu J-J. The relationship between age-related hearing loss and synaptic changes in the hip-pocampus of C57BL/6J mice. Exp Gerontol 2011;46: 716–722.
- Pilati N, Ison MJ, Barker M, Mulheran M, Large CH, Forsythe ID, Matthias J, Hamann M. Mechanisms contributing to central excitability changes during hearing loss. Proc Natl Acad Sci USA 2012;109:8292–8297.
- Ohlemiller KK, Gagnon PM. Cellular correlates of progressive hearing loss in 129S6/SvEv mice. J Comp Neurol 2004;469:377–390.
- 14. Schuknecht HF, Gacek MR. Cochlear pathology in presbycusis. Ann Otol Rhinol Laryngol 1993;102:1–16.
- 15. Loeb LA, Wallace DC, Martin GM. The mitochondrial theory of aging and its relationship to reactive oxygen species damage and somatic mtDNA mutations. Proc Natl Acad Sci USA 2005;102:18769–18770.
- Crawley BK, Keithley EM. Effects of mitochondrial mutations on hearing and cochlear pathology with age. Hear Res 2011;280:201–208.
- Johnson KR, Zheng QY, Bykhovskaya Y, Spirina O, Fischel-Ghodsian N. A nuclear-mitochondrial DNA interaction affecting hearing impairment in mice. Nat Genet 2001;27:191–194.
- Trifunovic A, Hansson A, Wredenberg A, Rovio AT, Dufour E, Khvorostov I, Spelbrink JN, Wibom R, Jacobs HT, Larsson N-G. Somatic mtDNA mutations cause aging phenotypes without affecting reactive oxygen species production. Proc Natl Acad Sci USA 2005;102:17993–17998.
- Oliver CN, Ahn B-w, Moerman EJ, Goldstein S, Stadtman ER. Age-related changes in oxidized proteins. J Biol Chem 1987;262:5488–5491.
- Mecocci P, MacGarvey U, Kaufman AE, Koontz D, Shoffner JM, Wallace DC, Beal MF. Oxidative damage to mitochondrial DNA shows marked age-dependent increases in human brain. Ann Neurol 1993;34:609–616.
- 21. Harman D. The biologic clock: the mitochondria? J Am Geriatr Soc 1972;20:145–147.
- Wallace DC. A mitochondrial paradigm of metabolic and degenerative diseases, aging, and cancer: A dawn for evolutionary medicine. Annu Rev Genet 2005;39:359–407.
- Someya S, Yamasoba T, Weindruch R, Prolla TA, Tanokura M. Caloric restriction suppresses apoptotic cell death in the mammalian cochlea and leads to prevention of presbycusis. Neurobiol Aging 2007;28:1613–1622.
- 24. Shelke RR, Leeuwenburgh C. Lifelong caloric restriction increases expression of apoptosis repressor with a caspase recruitment domain (ARC) in the brain. FASEB J 2003; 17:494–496.
- 25. Ernster L, Ikkos D, Luft R. Enzymic activities of human skeletal muscle mitochondria: A tool in clinical metabolic research. Nature 1959;184:1851.
- Vafai SB, Mootha VK. Mitochondrial disorders as windows into an ancient organelle. Nature 2012;491:374–383.
- Balaban RS, Nemoto S, Finkel T. Mitochondria, oxidants, and aging. Cell 2005;120:483–495.
- Youle RJ, Strasser A. The BCL-2 protein family: Opposing activities that mediate cell death. Nat Rev Mol Cell Biol 2008;9:47–59.
- Someya S, Xu J, Kondo K, Ding D, Salvi RJ, Yamasoba T, Rabinovitch PS, Weindruch R, Leeuwenburgh C, Tanokura M. Age-related hearing loss in C57BL/6J mice is mediated by Bak-dependent mitochondrial apoptosis. Proceed Natl Acad Sci USA 2009;106:19432–19437.

- Lindsten T, Ross AJ, King A, Zong W-X, Rathmell JC, Shiels HA, Ulrich E, Waymire KG, Mahar P, Frauwirth K. The combined functions of proapoptotic Bcl-2 family members bak and bax are essential for normal development of multiple tissues. Mol Cell 2000;6:1389–1399.
- Degenhardt K, Sundararajan R, Lindsten T, Thompson C, White E. Bax and Bak independently promote cytochrome *c* release from mitochondria. J Biol Chem 2002;277:14127– 14134.
- 32. Martinou J, Youle R. Which came first, the cytochrome *c* release or the mitochondrial fission? Cell Death Differ 2006;13:1291–1295.
- Marsden VS, O'Connor L, O'Reilly LA, Silke J, Metcalf D, Ekert PG, Huang DC, Cecconi F, Kuida K, Tomaselli KJ. Apoptosis initiated by Bcl-2-regulated caspase activation independently of the cytochrome c/Apaf-1/caspase-9 apoptosome. Nature 2002;419:634–637.
- 34. Wang X. The expanding role of mitochondria in apoptosis. Genes Dev 2001;15:2922–2933.
- 35. Shi Y. Mechanical aspects of apoptosome assembly. Curr Opin Cell Biol 2006;18:677–684.
- 36. Zhang Y, Cook A, Kim J, Baranov SV, Jiang J, Smith K, Cormier K, Bennett E, Browser RP, Day AL. Melatonin inhibits the caspase-1/cytochrome c/caspase-3 cell death pathway, inhibits MT1 receptor loss and delays disease progression in a mouse model of amyotrophic lateral sclerosis. Neurobiol Dis 2013;55:26–35.
- Gaggelli E, Kozlowski H, Valensin D, Valensin G. Copper homeostasis and neurodegenerative disorders (Alzheimer's, prion, and Parkinson's diseases and amyotrophic lateral sclerosis). Chem Rev 2006;6:1995–2044.
- Lei D, Gao X, Perez P, Ohlemiller KK, Chen C-C, Campbell KP, Hood AY, Bao J. Anti-epileptic drugs delay age-related loss of spiral ganglion neurons via T-type calcium channel. Hear Res 2011;278:106–112.
- Keithley EM, Canto C, Zheng QY, Wang X, Fischel-Ghodsian N, Johnson KR. Cu/Zn superoxide dismutase and age-related hearing loss. Hear Res 2005;209:76–85.
- Van Raamsdonk JM, Hekimi S. Superoxide dismutase is dispensable for normal animal lifespan. Proc Natl Acad Sci USA 2012;109:5785–5790.
- Chen S, Zhang X, Song L, Le W. Autophagy dysregulation in amyotrophic lateral sclerosis. Brain Pathol 2012;22: 110–116.
- 42. Bendotti C, Marino M, Cheroni C, Fontana E, Crippa V, Poletti A, De Biasi S. Dysfunction of constitutive and inducible ubiquitin-proteasome system in amyotrophic lateral sclerosis: Implication for protein aggregation and immune response. Progr Neurobiol 2012;97:101–126.
- 43. McFadden SL, Ding D, Reaume AG, Flood DG, Salvi RJ. Age-related cochlear hair cell loss is enhanced in mice lacking copper/zinc superoxide dismutase. Neurobiol Aging 1999;20:1–8.
- Coling DE, Yu KC, Somand D, Satar B, Bai U, Huang T-T, Seidman MD, Epstein CJ, Mhatre AN, Lalwani AK. Effect of SOD1 overexpression on age-and noise-related hearing loss. Free Radic Biol Med 2003;34:873–880.
- 45. Linnane A, Ozawa T, Marzuki S, Tanaka M. Mitochondrial DNA mutations as an important contributor to ageing and degenerative diseases. Lancet 1989;333:642–645.
- 46. Schon EA, Rizzuto R, Moraes CT, Nakase H, Zeviani M, DiMauro S. A direct repeat is a hotspot for large-scale deletion of human mitochondrial DNA. Science 1989;244: 346–349.

- 47. Cortopassi G, Shibata D, Soong N, Arnheim N. A pattern of accumulation of a somatic deletion of mitochondrial DNA in aging human tissues. Proc Natl Acad Sci USA 1992;89:7370–7374.
- Picard M, Turnbull DM. Linking the metabolic state and mitochondrial DNA in chronic disease, health, and aging. Diabetes 2013;62:672–678.
- 49. Signer RA, Morrison SJ. Mechanisms that regulate stem cell aging and life span. Cell Stem Cell 2013;12:152–165.
- Corral-Debrinski M, Horton T, Lott MT, Shoffner JM, Beal MF, Wallace DC. Mitochondrial DNA deletions in human brain: Regional variability and increase with advanced age. Nat Genet 1992;2:324–329.
- Markaryan A, Nelson EG, Hinojosa R. Detection of mitochondrial DNA deletions in the cochlea and its structural elements from archival human temporal bone tissue. Mutat Res 2008;640:38–45.
- 52. Edris W, Burgett B, Colin Stine O, Filburn CR. Detection and quantitation by competitive PCR of an age-associated increase in a 4.8-kb deletion in rat mitochondrial DNA. Mutat Res 1994;316:69–78.
- Bai U, Seidman MD, Hinojosa R, Quirk WS. Mitochondrial DNA deletions associated with aging and possibly presbycusis: A human archival temporal bone study. Otol Neurotol 1997;18:449–453.
- Deschauer M, Muller T, Wieser T, Schulte-Mattler W, Kornhuber M, Zierz S. Hearing impairment is common in various phenotypes of the mitochondrial DNA A3243G mutation. Arch Neurol 2001;58:1885.
- Gadaleta M, Rainaldi G, Lezza A, Milella F, Fracasso F, Cantatore P. Mitochondrial DNA copy number and mitochondrial DNA deletion in adult and senescent rats. Mutat Res 1992;275:181–193.
- Kong W-J, Hu Y-J, Wang Q, Wang Y, Han Y-C, Cheng H-M, Kong W, Guan M-X. The effect of the mtDNA4834 deletion on hearing. Biochem Biophys Res Commun 2006;344:425–430.
- Noben-Trauth K, Zheng QY, Johnson KR. Association of cadherin 23 with polygenic inheritance and genetic modification of sensorineural hearing loss. Nat Genet 2003;35:21–23.
- Kane KL, Longo-Guess CM, Gagnon LH, Ding D, Salvi RJ, Johnson KR. Genetic background effects on age-related hearing loss associated with Cdh23 variants in mice. Hear Res 2012;283:80–88.
- 59. Liu S, Li S, Zhu H, Cheng S, Zheng QY. A mutation in the cdh23 gene causes age-related hearing loss in Cdh23 (nmf308/nmf308) mice. Gene 2012;499:309–317.
- 60. Erway LC, Willott JF, Archer JR, Harrison DE. Genetics of age-related hearing loss in mice: I. Inbred and F1 hybrid strains. Hear Res 1993;65:125–132.
- 61. Noben-Trauth K, Zheng QY, Johnson KR, Nishina PM. mdfw: A deafness susceptibility locus that interacts with deaf waddler (dfw). Genomics 1997;44:266–272.
- Johnson KR, Zheng QY, Erway LC. A major gene affecting age-related hearing loss is common to at least ten inbred strains of mice. Genomics 2000;70:171–180.
- 63. Johnson KR, Zheng QY. Ahl2, a second locus affecting agerelated hearing loss in mice. Genomics 2002;80:461–464.
- 64. Morita Y, Hirokawa S, Kikkawa Y, Nomura T, Yonekawa H, Shiroishi T, Takahashi S, Kominami R. Fine mapping of Ahl3 affecting both age-related and noise-induced hearing loss. Biochem Biophys Res Commun 2007;355:117–121.
- 65. Nemoto M, Morita Y, Mishima Y, Takahashi S, Nomura T, Ushiki T, Shiroishi T, Kikkawa Y, Yonekawa H, Kominami

R. Ahl3, a third locus on mouse chromosome 17 affecting age-related hearing loss. Biochem Biophys Res Commun 2004;324:1283–1288.

- 66. McFadden SL, Ding D, Burkard RF, Jiang H, Reaume AG, Flood DG, Salvi RJ. Cu/Zn SOD deficiency potentiates hearing loss and cochlear pathology in aged 129, CD-1 mice. J Comp Neurol 1999;413:101–112.
- 67. Ohlemiller KK, McFadden SL, Ding D-L, Lear PM, Ho Y-S. Targeted mutation of the gene for cellular glutathione peroxidase (Gpx1) increases noise-induced hearing loss in mice. J Assoc Res Otolaryngol 2000;1:243–254.
- Zheng QY, Ding D, Yu H, Salvi RJ, Johnson KR. A locus on distal chromosome 10 (ahl4) affecting age-related hearing loss in A/J mice. Neurobiol Aging 2009;30:1693–1705.
- 69. Drayton M, Noben-Trauth K. Mapping quantitative trait loci for hearing loss in Black Swiss mice. Hear Res 2006;212:128–139.
- Johnson KR, Longo-Guess C, Gagnon LH, Yu H, Zheng QY. A locus on distal chromosome 11 (ahl8) and its interaction with Cdh23, ahl underlie the early onset, agerelated hearing loss of DBA/2J mice. Genomics 2008;92: 219–225.
- Mashimo T, Erven AE, Spiden SL, Guénet J-L, Steel KP. Two quantitative trait loci affecting progressive hearing loss in 101/H mice. Mamm Genome 2006;17:841–850.
- Fridberger A, Ulfendahl M. Acute mechanical overstimulation of isolated outer hair cells causes changes in intracellular calcium levels without shape changes. Acta Otolaryngol (Stockh) 1996;116:17–24.
- 73. Di Paolo G, De Camilli P. Phosphoinositides in cell regulation and membrane dynamics. Nature 2006;443:651–657.
- Berridge MJ, Bootman MD, Roderick HL. Calcium signalling: Dynamics, homeostasis and remodelling. Nat Rev Mol Cell Biol 2003;4:517–529.
- Zhao H-B, Yu N, Fleming CR. Gap junctional hemichannelmediated ATP release and hearing controls in the inner ear. Proc Natl Acad Sci USA 2005;102:18724–18729.
- 76. Anselmi F, Hernandez VH, Crispino G, Seydel A, Ortolano S, Roper SD, Kessaris N, Richardson W, Rickheit G, Filippov MA. ATP release through connexin hemichannels and gap junction transfer of second messengers propagate Ca²⁺ signals across the inner ear. Proc Natl Acad Sci USA 2008;105:18770–18775.
- Barbe MT, Monyer H, Bruzzone R. Cell-cell communication beyond connexins: The pannexin channels. Physiology 2006;21:103–114.
- Suadicani SO, Brosnan CF, Scemes E. P2X7 receptors mediate ATP release and amplification of astrocytic intercellular Ca²⁺ signaling. J Neurosci 2006;26:1378–1385.
- Pelegrin P, Surprenant A. Pannexin-1 mediates large pore formation and interleukin-1β release by the ATP-gated P2X7 receptor. EMBO J 2006;25:5071–5082.
- Locovei S, Scemes E, Qiu F, Spray DC, Dahl G. Pannexin1 is part of the pore forming unit of the P2X₇ receptor death complex. FEBS Lett 2007;581:483–488.
- Kelly MC, Chen P. Development of form and function in the mammalian cochlea. Curr Opin Neurobiol 2009;19: 395–401.
- 82. Rodriguez L, Simeonato E, Scimemi P, Anselmi F, Calì B, Crispino G, Ciubotaru CD, Bortolozzi M, Ramirez FG, Majumder P. Reduced phosphatidylinositol 4, 5-bisphosphate synthesis impairs inner ear Ca²⁺ signaling and highfrequency hearing acquisition. Proc Natl Acad Sci USA 2012;109:14013–14018.

- Buchholz JN, Behringer EJ, Pottorf WJ, Pearce WJ, Vanterpool CK. Age-dependent changes in Ca²⁺ homeostasis in peripheral neurones: Implications for changes in function. Aging Cell 2007;6:285–296.
- Mills JH, Matthews LJ, LEE FS, Dubno JR, Schulte BA, Weber PC. Gender-specific effects of drugs on hearing levels of older persons. Ann NY Acad Sci 1999;884:381–388.
- 85. Rüttiger L, Sausbier M, Zimmermann U, Winter H, Braig C, Engel J, Knirsch M, Arntz C, Langer P, Hirt B. Deletion of the Ca²⁺⁻activated potassium (BK) α-subunit but not the BKβ1-subunit leads to progressive hearing loss. Proc Natl Acad Sci USA 2004;101:12922–12927.
- 86. Kharkovets T, Hardelin J-P, Safieddine S, Schweizer M, El-Amraoui A, Petit C, Jentsch TJ. KCNQ4, a K⁺ channel mutated in a form of dominant deafness, is expressed in the inner ear and the central auditory pathway. Proc Natl Acad Sci USA 2000;97:4333–4338.
- Bielefeld EC, Coling D, Chen G-D, Li M, Tanaka C, Hu B-H, Henderson D. Age-related hearing loss in the Fischer 344/ NHsd rat substrain. Hear Res 2008;241:26–33.
- Nakazawa K, Spicer SS, Schulte BA. Ultrastructural localization of Na, K-ATPase in the gerbil cochlea. J Histochem Cytochem 1995;43:981–991.
- Crouch JJ, Sakaguchi N, Lytle C, Schulte BA. Immunohistochemical localization of the Na-K-Cl co-transporter (NKCC1) in the gerbil inner ear. J Histochem Cytochem 1997;45:773–778.
- 90. Wangemann P. K⁺ cycling and the endocochlear potential. Hear Res 2002;165:1–9.
- Bartolami S, Gaboyard S, Quentin J, Travo C, Cavalier M, Barhanin J, Chabbert C. Critical roles of transitional cells and Na/K-ATPase in the formation of vestibular endolymph. J Neurosci 2011;31:16541–16549.
- Diaz RC, Vazquez AE, Dou H, Wei D, Cardell EL, Lingrel J, Shull GE, Doyle KJ, Yamoah EN. Conservation of hearing by simultaneous mutation of Na, K-ATPase and NKCC1. J Association Res Otolaryngol 2007; 8: 422–34.
- Hudspeth AJ. How the ear's works work. Nature 1989; 341:397–404.
- Hibino H, Kurachi Y. Molecular and physiological bases of the K⁺ circulation in the mammalian inner ear. Physiology 2006;21:336–345.
- Chan DK, Hudspeth A. Ca²⁺ current–driven nonlinear amplification by the mammalian cochlea in vitro. Nat Neurosci 2005;8:149–155.
- Kikuchi T, Adams JC, Miyabe Y, So E, Kobayashi T. Potassium ion recycling pathway via gap junction systems in the mammalian cochlea and its interruption in hereditary nonsyndromic deafness. Med Electron Microsc 2000; 33: 51–56.
- Spicer SS, Schulte BA. Evidence for a medial K⁺ recycling pathway from inner hair cells. Hear Res 1998;118:1–12.
- Nin F, Hibino H, Doi K, Suzuki T, Hisa Y, Kurachi Y. The endocochlear potential depends on two K⁺ diffusion potentials and an electrical barrier in the stria vascularis of the inner ear. Proc Natl Acad Sci USA 2008;105:1751–1756.
- Salt AN, Mleichar I, Thalmann R. Mechanisms of endocochlear potential generation by stria vascularis. Laryngoscope 1987;97:984–991.
- 100. Takeuchi S, Ando M, Kakigi A. Mechanism generating endocochlear potential: role played by intermediate cells in stria vascularis. Biophys J 2000;79:2572–2782.
- Roy M, Sivan-Loukianova E, Eberl DF. Cell-type–specific roles of Na⁺/K⁺ ATPase subunits in Drosophila auditory

mechanosensation. Proc Natl Acad Sci USA 2013;110: 181–186.

- Ichimiya I, Yoshida K, Hirano T, Suzuki M, Mogi G. Significance of spiral ligament fibrocytes with cochlear inflammation. Int J Pediatr Otorhinolaryngol 2000;56:45–51.
- 103. Fubini B, Hubbard A. Reactive oxygen species (ROS) and reactive nitrogen species (RNS) generation by silica in inflammation and fibrosis. Free Radic Biol Med 2003;34:1507– 1516.
- 104. Lindvall O, Kokaia Z. Stem cells in human neurodegenerative disorders—time for clinical translation? J Clin Invest 2010;120:29.
- 105. Zhang ZG, Chopp M. Neurorestorative therapies for stroke: Underlying mechanisms and translation to the clinic. Lancet Neurol 2009;8:491.
- 106. Iwai H, Lee S, Inaba M, Sugiura K, Tomoda K, Yamashita T, Ikehara S. Prevention of accelerated presbycusis by bone marrow transplantation in senescence-accelerated mice. Bone Marrow Transplant 2001;28:323–328.
- 107. Ren H, Chen J, Wang Y, Zhang S, Zhang B. Intracerebral neural stem cell transplantation improved the auditory of mice with presbycusis. Int J Clin Exp Pathol 2013;6:230.
- Nishimura K, Nakagawa T, Sakamoto T, Ito J. Fates of murine pluripotent stem cell-derived neural progenitors following transplantation into mouse cochleae. Cell Transplant 2012;21:763–771.
- Matsumoto M, Nakagawa T, Kojima K, Sakamoto T, Fujiyama F, Ito J. Potential of embryonic stem cell-derived neurons for synapse formation with auditory hair cells. J Neurosci Res 2008;86:3075–3085.
- Chen W, Jongkamonwiwat N, Abbas L, Eshtan SJ, Johnson SL, Kuhn S, Milo M, Thurlow JK, Andrews PW, Marcotti W. Restoration of auditory evoked responses by human EScell-derived otic progenitors. Nature 2012;490:278–282.
- 111. Shevde N. Stem cells: Flexible friends. Nature 2012;483: S22–S26.
- 112. Jaramillo S, Zador AM. The auditory cortex mediates the perceptual effects of acoustic temporal expectation. Nat Neurosci 2010;14:246–251.
- 113. Zheng JL, Gao W-Q. Overexpression of Math1 induces robust production of extra hair cells in postnatal rat inner ears. Nat Neurosci 2000;3:580–586.
- 114. Fekete DM. Cell fate specification in the inner ear. Curr Opin Neurobiol 1996;6:533–541.
- 115. Fekete DM. Development of the vertebrate ear: Insights from knockouts and mutants. Trends Neurosci 1999;22: 263–269.
- 116. Behlke MA. Progress towards in vivo use of siRNAs. Mol Ther 2006;13:644–670.
- 117. Kitado H, Higuchi K, Takeda T. Molecular genetic characterization of the senescence-accelerated mouse (SAM) strains. J Gerontol 1994;49:B247–B254.
- Hosono M, Hanada K, Toichi E, Naiki H, Higuchi K, Hosokawa T. Immune abnormality in relation to nonimmune diseases in SAM mice. Exp Gerontol 1997;32:181–195.
- 119. Devarajan K, Forrest ML, Detamore MS, Staecker H. Adenovector-mediated gene delivery to human umbilical cord mesenchymal stromal cells induces inner ear cell phenotype. Cell Reprogram 2013;15:43–54.
- 120. Kesser B, Lalwani A. Gene therapy and stem cell transplantation: Strategies for hearing restoration. Adv Otorhinolaryngol 2009;66:64–86.
- 121. Koubova J, Guarente L. How does calorie restriction work? Genes Dev 2003;17:313–321.

- 122. Colman RJ, Anderson RM, Johnson SC, Kastman EK, Kosmatka KJ, Beasley TM, Allison DB, Cruzen C, Simmons HA, Kemnitz JW. Caloric restriction delays disease onset and mortality in rhesus monkeys. Science 2009;325:201– 204.
- 123. Jiang H, Talaska AE, Schacht J, Sha S-H. Oxidative imbalance in the aging inner ear. Neurobiol Aging 2007;28:1605– 1612.
- 124. Someya S, Yu W, Hallows WC, Xu J, Vann JM, Leeuwenburgh C, Tanokura M, Denu JM, Prolla TA. Sirt3 mediates reduction of oxidative damage and prevention of age-related hearing loss under caloric restriction. Cell 2010;143:802–812.
- 125. Masoro EJ. Caloric restriction and aging: an update. Exp Gerontol 2000;35:299–305.
- 126. Seidman MD. Effects of dietary restriction and antioxidants on presbyacusis. Laryngoscope 2000;110:727–738.
- 127. Pallas M, Verdaguer E, Tajes M, Gutierrez-Cuesta J, Camins A. Modulation of sirtuins: New targets for antiageing. Recent Pat CNS Drug Discov 2008;3:61–69.
- 128. Guarente L. Sirtuins in aging and disease. Cold Spring Harb Symp Quant Biol 2007;72:483–488.
- 129. Burns J, Yokota T, Ashihara H, Lean ME, Crozier A. Plant foods and herbal sources of resveratrol. J Agric Food Chem 2002;50:3337–3340.
- 130. Aggarwal BB, Takada Y, Oommen OV. From chemoprevention to chemotherapy: common targets and common goals. Exp Opin Investig Drugs 2004;13:1327–1338.
- 131. Baur JA, Pearson KJ, Price NL, Jamieson HA, Lerin C, Kalra A, Prabhu VV, Allard JS, Lopez-Lluch G, Lewis K. Resveratrol improves health and survival of mice on a high-calorie diet. Nature 2006;444:337–342.
- 132. Lagouge M, Argmann C, Gerhart-Hines Z, Meziane H, Lerin C, Daussin F, Messadeq N, Milne J, Lambert P, Elliott P. Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1α. Cell 2006;127:1109–1122.
- Seidman M, Babu S, Tang W, Naem E, Quirk WS. Effects of resveratrol on acoustic trauma. Otolaryngol Head Neck Surg 2003;129:463–470.
- 134. Seidman MD, Ahmad N, Joshi D, Seidman J, Thawani S, Quirk WS. Age-related hearing loss and its association with reactive oxygen species and mitochondrial DNA damage. Acta Otolaryngol (Stockh) 2004;124:16–24.
- 135. Le T, Keithley EM. Effects of antioxidants on the aging inner ear. Hear Res 2007;226:194–202.
- 136. Takumida M, Anniko M. Nitric oxide in the inner ear. Curr Opin Neurol 2002;15:11–15.
- 137. Lasisi AO, Fehintola FA. Correlation between plasma levels of radical scavengers and hearing threshold among elderly subjects with age-related hearing loss. Acta Otolaryngol (Stockh) 2011;131:1160–1164.
- 138. Lopez-Gonzalez MA, M Guerrero J, Sanchez B, Delgado F. Melatonin induces hyporeactivity caused by type II collagen in peripheral blood lymphocytes from patients with autoimmune hearing losses. Neurosci Lett 1997;239:1–4.
- 139. Seidman MD, Khan MJ, Tang WX, Quirk WS. Influence of lecithin on mitochondrial DNA and age-related hearing loss. Otolaryngol Head Neck Surg 2002;127:138–144.

- 140. Carney J, Starke-Reed P, Oliver C, Landum R, Cheng M, Wu J, Floyd R. Reversal of age-related increase in brain protein oxidation, decrease in enzyme activity, and loss in temporal and spatial memory by chronic administration of the spin-trapping compound *N*-tert-butyl-alpha-phenylnitrone. Proc Natl Acad Sci USA 1991;88:3633–3636.
- 141. Antunes R, Moita MA. Discriminative auditory fear learning requires both tuned and nontuned auditory pathways to the amygdala. J Neurosci 2010;30:9782–9787.
- 142. Armony J, Servan-Schreiber D, Romanski LM, Cohen JD, LeDoux JE. Stimulus generalization of fear responses: Effects of auditory cortex lesions in a computational model and in rats. Cereb Cortex 1997;7:157–165.
- 143. Chavez CM, McGaugh JL, Weinberger NM. The basolateral amygdala modulates specific sensory memory representations in the cerebral cortex. Neurobiol Learn Mem 2009; 91:382–392.
- 144. Wright BA, Zhang Y. A review of the generalization of auditory learning. Philos Trans R Soc Lond B Biol Sci 2009; 364:301–311.
- 145. Resnik J, Sobel N, Paz R. Auditory aversive learning increases discrimination thresholds. Nat Neurosci 2011;14: 791–796.
- 146. Salthouse TA. The processing-speed theory of adult age differences in cognition. Psychol Rev 1996;103:403–427.
- 147. de Villers-Sidani E, Alzghoul L, Zhou X, Simpson KL, Lin RC, Merzenich MM. Recovery of functional and structural age-related changes in the rat primary auditory cortex with operant training. Proc Natl Acad Sci USA 2010;107:13900– 13905.
- 148. Bao S, Chang EF, Woods J, Merzenich MM. Temporal plasticity in the primary auditory cortex induced by operant perceptual learning. Nat Neurosci 2004;7:974–981.
- 149. Klintsova AY, Dickson E, Yoshida R, Greenough WT. Altered expression of BDNF and its high-affinity receptor TrkB in response to complex motor learning and moderate exercise. Brain Res 2004;1028:92–104.
- 150. Tokarski K, Urban-Ciecko J, Kossut M, Hess G. Sensory learning-induced enhancement of inhibitory synaptic transmission in the barrel cortex of the mouse. Eur J Neurosci 2007;26:134–141.
- 151. Bengtsson SL, Nagy Z, Skare S, Forsman L, Forssberg H, Ullén F. Extensive piano practicing has regionally specific effects on white matter development. Nat Neurosci 2005; 8:1148–1150.

Address correspondence to: Jihao Ren Department of Otolaryngology Head and Neck Surgery The Second Xiangya Hospital Central South University No.139 Middle Renmin Road Changsha, Hunan 410011 P.R. China

E-mail: jihao5114@sina.com

Received: April 17, 2013 Accepted: August 5, 2013