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Platelet-derived growth factor receptor (PDGFR) expression in primary spinal cord gliomas

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Abstract

Abnormal signaling through the platelet-derived growth factor receptor (PDGFR) has been proposed as a possible mechanism of spinal cord glioma initiation and progression. However, the extent of PDGFR expression in human spinal cord gliomas remains unknown. In this study we perform immunohistochemical analysis of PDGFRa expression in a series of 33 primary

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intramedullary spinal cord gliomas of different types and grades. PDGFRa was seen to be expressed in a significant subset of these tumors across all major glioma types including ependymoma, oligodendroglioma, pilocytic astrocytoma, astrocytoma, and glioblastoma. These results support the hypothesis that growth factor signaling through the PDGFR may be important for the development of at least a subset of human spinal cord gliomas. Further studies investigating the prognostic significance of PDGFR expression as well as the role of PDGF signaling on the development of intramedullary spinal cord gliomas are warranted.

Keywords

Astrocytoma; Ependymoma; Glioblastoma multiforme; Glioma; Intramedullary; Oligodendroglioma; Pilocytic; Spinal cord; Tumor

Introduction

Primary intramedullary spinal cord gliomas are rare but potentially devastating lesions. The mechanisms of tumorigenesis and the molecular-genetic profile of these tumors remain poorly defined. However, platelet-derived growth factor (PDGF) signaling has been proposed to play an important role in their development. In animal models, induction of PDGF-B expression in the spinal cord has been shown to initiate the formation of intramedullary gliomas [1–3]. These experimental spinal cord gliomas also express the cognate receptor PDGFRa suggesting that signaling downstream of this receptor tyrosine kinase may drive tumor formation. Additionally, evidence of PDGF receptor expression in human spinal cord ependymoma as well as responsiveness to receptor inhibition further implicates a mechanistic role for PDGF signaling in the pathogenesis of intramedullary glioma [4, 5].

Although, a number of reports have demonstrated expression of PDGF receptors in cerebral gliomas [6–10], the extent of PDGF receptor expression in spinal cord gliomas remains unknown. Recognition that PDGF and its receptors are often expressed in cerebral gliomas has provided the framework for the initiation of several clinical trials testing inhibition of this pathway in brain tumors [11–17]. The development and implementation of therapies targeted at PDGF receptor and/or downstream target inhibition in spinal cord gliomas will depend both on the feasibility of receptor profiling and on the frequency with which positive expression is observed. In this study we assess the expression status of platelet-derived growth factor receptor alpha (PDGFR α) by immunohistochemistry in a series of 33 primary spinal cord gliomas. Using this technique we characterize expression within various spinal cord gliomas of different histological types and grades.

Methods

Case selection and pathological samples

The cases for this study were identified within the neuropathology databases maintained by the Columbia University Medical Center, Department of Pathology and within the Department of Neurological Surgery Bartoli Brain Tumor Bank. Cases of primary intramedullary spinal cord glioma with sufficient paraffinized tissue for analysis obtained between 1995 and 2009 were eligible for inclusion. Patients with a diagnosis of myxopapillary ependymoma were excluded. Original surgery and pathology reports were reviewed for each case. After diagnostic confirmation was obtained by review of the original pathological slides, formalin-fixed paraffin-embedded tumor blocks were obtained. Five micron thick sections were cut and mounted on glass slides in preparation for

Immunohistochemistry and microscopy

Glass-mounted 5 micron thick sections were de-paraffinized in xylene and rehydrated with 100% ethanol and distilled water. Antigen retrieval was performed by boiling the samples in 10 mM citrate buffer at a pH of 6. Samples were then blocked in 0.3% peroxide, washed, and blocked in 20% normal goat serum (Vector Laboratories). After washing, the samples were incubated with rabbit anti- PDGFRa (1:500; Cell Signaling). Vectastain ABC kit (Vector laboratories) and DAB chromogen were used for visualization. Sections were then counterstained with hematoxylin, washed, and dehydrated prior to cover slip mounting. Cerebral glioblastomas with known PDGFRa expression status were immunohistochemically processed in parallel with the study samples to serve as positive and negative staining controls.

accordance with policies of the Institutional Review Board.

All photomicrographs were obtained using a Nikon Labphot-2 microscope. Digital images were processed using NIS-Elements imaging software (Nikon). PDGFR α expression was classified as either positive or negative based on whether immunoperoxidase reactivity (brown staining) was observed during microscopic analysis. Due to the small sample sizes and the presence of focal variations in staining intensity, qualitative rather than quantitative immunohistochemical expression profiling was deemed most appropriate.

Results

Study cohort

A total of 33 histologically confirmed cases of primary intramedullary spinal cord glioma were included in this study (Table 1). The patients include 15 women and 18 men with a median age of 40 (range 2–81) years at the time of diagnosis. The pathological diagnoses include 26 WHO grade II ependymomas, 1 WHO grade II oligodendroglioma, 1 pilocytic astrocytoma, 1 anaplastic pilocytic astrocytoma, 3 WHO grade II astrocytomas, and 1 glioblastoma multiforme. The tumors had no segmental level predilection and were seen throughout the cervical, thoracic, and lumbar spinal cord.

Histological and immunohistochemical analysis

PDGFRα localization and expression—A total of 10 out of 33 (30.3%) spinal cord gliomas analyzed were seen to express PDGFRα (Table 1). PDGFRα immunoreactivity localized to both the cytoplasm and cell membranes of tumor cells within positive samples. Staining of variable intensity was often seen in a discrete subset of cells. The density and distribution of PDGFRα-expressing cells varied from tumor to tumor and from region to region within tumors.

Ependymoma—Ependymoma was the most commonly identified intra-medullary spinal cord tumor within the queried tumor banks and 26 were included in this study. On histological examination all 26 tumors were classified World Health Organization (WHO) grade II ependymoma with three being subtyped tanycytic. In addition to histological phenotyping, the diagnosis of ependymoma was supported by the presence of characteristic dot-like epithelial membrane antigen (EMA) immunoreactivity in 8 of 9 tested samples (not shown). PDGFRα immunoreactivity was detected in 19% (5/26) of the spinal cord ependymoma cases studied. Representative examples are shown in Fig. 1.

Oligodendroglioma—WHO grade II intramedullary oligodendroglioma is rarely diagnosed and one case was identified within the cohort. A characteristic "Indian file"

pattern of cellular infiltration was observed in the pathological sample (Fig. 2a). Areas of calcification were also seen. PDGFR α immunoreactivity was seen to highlight the linear bands of tumor cells within an a cellular background (Fig. 2b).

Pilocytic astrocytoma—Both a WHO grade I (Fig. 3a–b) and an anaplastic WHO grade III (Fig. 3c–d) pilocytic astrocytoma were among the cohort of tumors samples in this series. The anaplastic pilocytic astrocytoma exhibited areas with distinct features of malignant progression (Fig. 3c, inset) with other areas showing lower grade histology. The grade I pilocytic astrocytoma was PDGFRa negative (Fig. 3b) while the anaplastic pilocytic astrocytom astrocytoma (Fig. 3d).

Astrocytoma and glioblastoma—PDGFRa was expressed in 2 of 3 WHO grade II spinal cord astrocytomas and in the 1 spinal cord glioblastoma case analyzed (Fig. 4). The low cell density of the astrocytoma shown in (Fig. 4b) illustrates cytoplasmic and cell membrane PDGFRa immunoreactivity that highlights the boundaries between individual cells. Conversely, cellular borders are obscured within the glioblastoma due to the high density of neoplastic cells and dysplastic vasculature (Fig. 4f).

Discussion

Abnormal PDGF signaling is well recognized to be an important mechanism of gliomagenesis in the brain [18, 19]. However, relatively little is known about the role of PDGF and its receptors in spinal gliomagenesis. In this study we present the largest and most comprehensive series of human spinal cord gliomas surveyed for PDGFRa expression to date. Using immnohistochemical analysis we demonstrate that a significant subset of spinal cord gliomas express receptors for the growth factor PDGF and show that these receptors localize to tumor cell membranes and cytoplasm. Approximately 30% of all tumors and 20% of ependymomas-the largest subset of gliomas included in this studywere immunopositive for PDGFRa. This is in contrast to the relatively high rate of PDGFR α immunopositivity (5 of 7 tumors) seen in a small series of spinal ependymomas by Barton et al. [5]. Within each diagnostic category the cohort sizes were too small to make statistical inferences, however, it is important to note that PDGFR α expression was seen in all major categories of glioma found in the spinal cord. These results have implications regarding the identification of the cell types that give rise to spinal cord gliomas, determining the mechanism of spinal cord glioma development, and delineating the potential therapeutic options for patients with these tumors.

A number of animal and human studies suggest that autocrine and paracrine signaling through the PDGF cell surface receptor is responsible for the initiation and progression of brain gliomas [7, 9, 10, 20–25]. Expression profiling of human cerebral gliomas indicates that PDGF and its receptors (PDGFR) are often co-expressed [7, 20]. In vitro studies have shown that PDGF is a powerful mitogen for glioma cells and that small molecule inhibitors of PDGF will block glioma cell proliferation and survival [26, 27]. Animal studies have shown that infecting glial progenitors in the subventricular zone (SVZ), subcortical white matter, or brainstem with PDGF-expressing retroviruses will induce the formation of gliomas [22, 28, 29]. Furthermore, viruses that express higher levels of PDGF drive cerebral gliomas to form faster, more consistently, and with more malignant features, suggesting that PDGF-driven gliomagenesis is a dose dependent phenomena [30].

Thus far, evidence primarily from studies in animal models implicates PDGF receptor signaling in spinal cord gliomas as well [1–3]. This study provides the first evidence from human spinal cord glioma tissue that PDGF receptor expression may play an important role in the development of at least a subset of spinal cord gliomas. Interestingly, the human brain

initiating glioma formation in the presence of PDGF [31–33]. These glial progenitors are thought to possess an inherent capacity to proliferate massively in response to PDGF stimulation enabling even genetically normal progenitors to become tumorigenic if exposed to sufficient levels of PDGF [31]. Confirmation that this is also true in the human spinal cord awaits the results of future studies.

While maximal, safe resection is the initial treatment strategy utilized for most low grade gliomas of the spinal cord, adjuvant therapy may be useful in the management of residual, recurrent, or high-grade tumors. However, there is little evidence that current adjuvant therapies for spinal cord gliomas are effective [34–36]. A better strategy in treating these patients may be the rational use of targeted chemotherapies that block specific signal transduction pathways in tumor cells such as those downstream of the PDGF receptor. Though the promising pre-clinical results from studies evaluating PDGF receptor blockade with agents such as imatinib, sunitinib, and sorafenib in cerebral gliomas have yet to convincingly show clinical benefit, this represents an active area of research [11–17]. This strategy has not been explored in the treatment of spinal cord gliomas save for a single case report of partial remission of a PDGFR-expressing spinal ependymoma with use of imatinib [4]. As nearly a third of spinal cord gliomas in this series were shown to express PDGFR α , initiating therapies that target this receptor may be helpful in a significant subset of patients.

Limitations of this study stem from the small cohort sizes, especially for the non-ependymal tumors, from the qualitative immunohistochemical approach used for expression analysis, and from the inherent heterogeneity within gliomas. As primary spinal cord tumors are rare being 10–15 times less common than intracranial tumors [37] the accumulation of larger cohorts in future studies may require grouping of samples from multiple institutions. Our use of immunohistochemical analysis was low-cost, technically straightforward, and proved very reliable for screening the small pieces of tissue typical of spinal cord tumor biopsies. The primacy of neurological function preservation during spinal cord tumor biopsies means that extensive analysis of intratumoral expression variability may only be possible on autopsy samples.

Conclusion

PDGFR α is expressed in a subset of intramedullary spinal cord gliomas including ependymoma, oligodendroglioma, pilocytic astrocytoma, astrocytoma, and glioblastoma. This finding supports previous studies which suggest that PDGF signaling can play an important role in spinal cord gliomagenesis. Validation of these results in larger cohorts using quantitative expression analysis will be important in future studies. The prognostic significance of PDGFR α expression in spinal cord gliomas as well as its implications for the effectiveness of targeted therapies remains to be determined.

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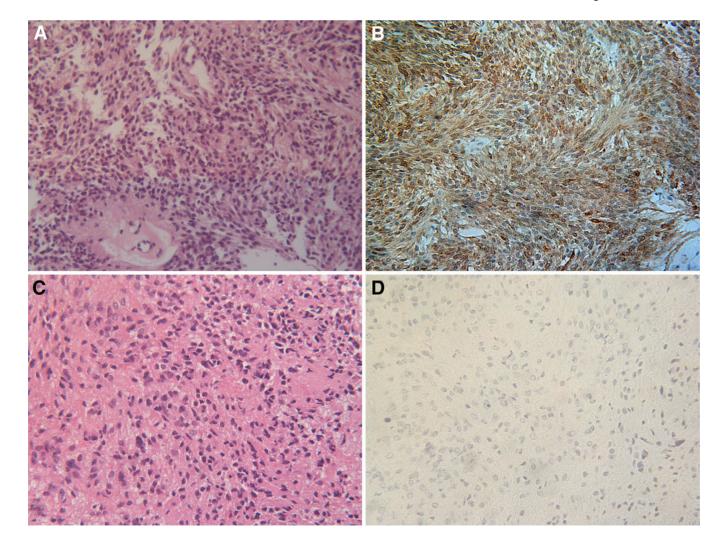


Fig. 1.

Spinal cord ependymomas. Representative H&E (**a**, **c**) and corresponding PDGFR α immunoperoxidase (**b**, **d**) stained sections from cases 1 and 16, respectively are shown. Positive (**b**) and negative (**d**) PDGFR α immunoreactivity (*brown*) is demonstrated. Magnification in all panels 200×

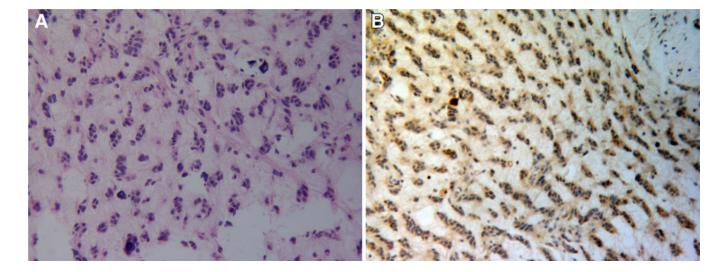


Fig. 2.

Spinal cord oligodendroglioma (case 27). H&E staining shows a moderately cellular neoplasm within a mucinous background. The tumor cells exhibit a palisading growth pattern and are aligned in linear bands (**a**). The majority of tumor cells show PDGFR α immunoreactivity (*brown*) (**b**). Magnification in all panels 200×

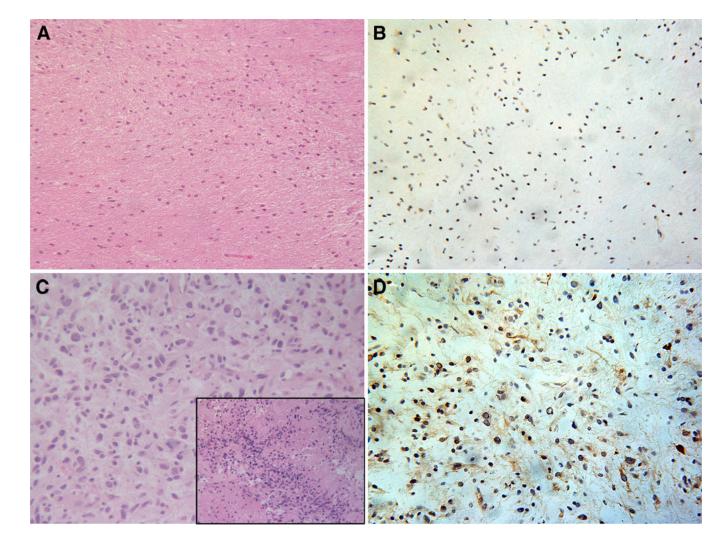


Fig. 3.

WHO grade I pilocytic astrocytoma (case 28) and anaplastic pilocytic astrocytoma of the spinal cord (case 29). H&E stained section from a WHO grade I pilocytic astrocytoma shows a low cellularity glial neoplasm within an eosinophilic background (**a**). No PDGFRα immunoreactivity is demonstrated within this tumor (**b**). In contrast, H&E section from a WHO grade III anaplastic pilocytic astrocytoma demonstrates a more cellular and pleomorphic tumor with areas of incipient necrosis (*inset*) (**c**). Immunoperoxidase staining demonstrates PDGFRα immunoreactivity (*brown*) within this tumor (**d**). Magnification in all panels 200X

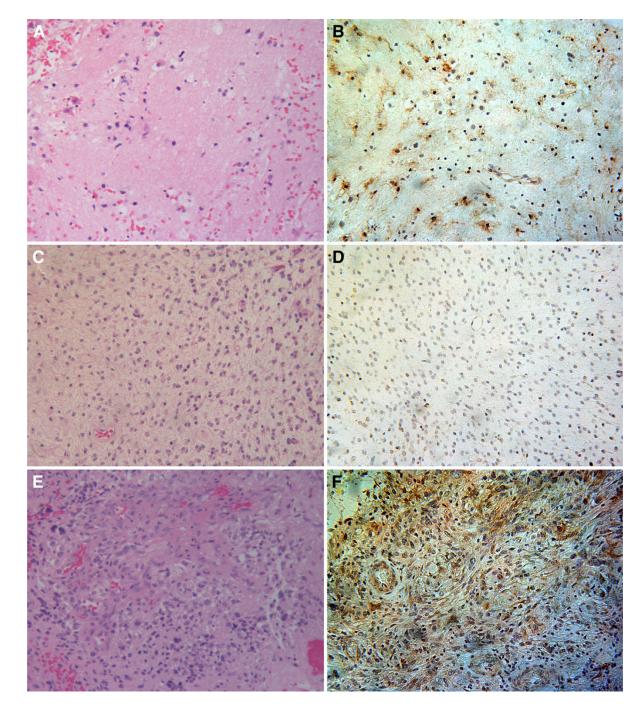


Fig. 4.

WHO grade II astrocytoma and glioblastoma (WHO grade IV) of the spinal cord. H&E section from a WHO grade II astrocytoma (case 30) demonstrates a low density of infiltrative glial cells within the spinal cord white matter (**a**). A subset of tumor cells distributed throughout the specimen show PDGFR α immunoreactivity (*brown*) (**b**). H&E section from another sample (case 32) of WHO grade II astrocytoma (**c**) with negative PDGFR α staining (**d**) is shown for comparison. A high-grade lesion with prominent glomeruloid vascular proliferation, hemorrhage, and necrosis consistent with a diagnosis of glioblastoma is demonstrated on H&E staining from case 33 of this series (**e**). The majority

of cells in this tumor stain strongly positive for PDGFRa (*brown*) (f). Magnification in all panels 200X

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Spinal cord glioma cases

Case	Gender	\mathbf{Age}	Diagnosis	WHO grade	Spinal cord level	PDGFRa status
_	М	68	Ependymoma	Π	Thoracic	Positive
5	Μ	21	Ependymoma	Π	Thoracolumbar	Positive
3	ц	29	Ependymoma	Π	Cervical	Positive
4	Μ	48	Ependymoma	Π	Cervical	Positive
5	Μ	33	Ependymoma	Π	Thoracolumbar	Positive
9	ц	71	Ependymoma	Π	Lumbar	Negative
	Ц	25	Ependymoma	Π	Lumbar	Negative
×	ц	59	Ependymoma	Π	Thoracic	Negative
6	Μ	46	Ependymoma	Π	Thoracic	Negative
10	Ц	44	Ependymoma	Π	Cervicothoracic	Negative
Ξ	Ц	39	Ependymoma	Π	Cervical	Negative
12	Μ	60	Ependymoma	Π	Cervical	Negative
13	Μ	48	Ependymoma	Π	Thoracic	Negative
14	Ц	56	Ependymoma	Π	Cervical	Negative
15	Μ	44	Ependymoma	Π	Thoracic	Negative
16	Μ	51	Ependymoma	Π	Thoracolumbar	Negative
17	Μ	71	Ependymoma	Π	Cervical	Negative
18	Μ	32	Ependymoma	Π	Cervical	Negative
19	Μ	40	Ependymoma	Π	Cervicothoracic	Negative
20	Ц	31	Ependymoma	Π	Lumbar	Negative
21	Μ	19	Ependymoma	Π	Thoracic	Negative
22	ц	19	Ependymoma	Π	Lumbar	Negative
23	Μ	47	Ependymoma	Π	Cervical	Negative
24	Μ	40	Ependymoma (tanycytic)	Π	Thoracic	Negative
25	Μ	39	Ependymoma (tanycytic)	Π	Lumbar	Negative
26	ц	50	Ependymoma (tanycytic)	Π	Cervicothoracic	Negative
27	Μ	46	Oligodendroglioma	Π	Thoracic	Positive
28	ц	8	Pilocytic astrocytoma	Ι	Cervical	Negative
29	Σ	81	Anaplastic pilocytic astrocytoma	Ш	Cervical	Positive

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Case	Gender	Age	Case Gender Age Diagnosis	WHO grade	WHO grade Spinal cord level PDGFRG status	PDGFRa statu
30	Ч	33	Astrocytoma	II	Thoracic	Positive
31	Ц	2	Astrocytoma	II	Thoracic	Positive
32	Ц	23	Astrocytoma	II	Thoracic	Negative
33	ц	27	Glioblastoma multiforme	IV	Thoracolumbar	Positive

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