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***Eya1* is required for the morphogenesis of mammalian thymus, parathyroid and thyroid**

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SUMMARY

Eyes absent (Eya) genes regulate organogenesis in both vertebrates and invertebrates. Mutations in human *EYA1* cause congenital Branchio-Oto-Renal (BOR) syndrome, while targeted inactivation of murine *Eya1* impairs early developmental processes in multiple organs, including ear, kidney and skeletal system. We have now examined the role of *Eya1* during the morphogenesis of organs derived from the pharyngeal region, including thymus, parathyroid and thyroid. The thymus and parathyroid are derived from 3rd pharyngeal pouches and their development is initiated via inductive interactions between neural crest-derived arch mesenchyme, pouch endoderm, and possibly the surface ectoderm of 3rd pharyngeal clefts. *Eya1* is expressed in all three cell types during thymus and parathyroid development from E9.5 and the organ primordia for both of these structures failed to form in *Eya1*^{-/-} embryos. These results indicate that *Eya1* is required for the initiation of thymus and parathyroid gland formation. *Eya1* is also expressed in the 4th pharyngeal region and ultimobranchial bodies. *Eya1*^{-/-} mice show thyroid hypoplasia, with severe reduction in the number of parafollicular cells and the size of the thyroid lobes and lack of fusion between the ultimobranchial bodies and the thyroid lobe. These data indicate that *Eya1* also regulates mature thyroid gland formation. Furthermore, we show that *Six1* expression is markedly reduced in the arch mesenchyme, pouch endoderm and surface ectoderm in the pharyngeal region of *Eya1*^{-/-} embryos, indicating that *Six1* expression in those structures is *Eya1* dependent. In addition, we show that in *Eya1*^{-/-} embryos, the expression of *Gcm2* in the 3rd pouch endoderm is undetectable at E10.5, however, the expression of *Hox* and *Pax* genes in the pouch endoderm is preserved at E9.5–10.5. Finally, we found that the surface ectoderm of the 3rd and 4th pharyngeal region show increased cell death at E10.5 in *Eya1*^{-/-} embryos. Our results indicate that *Eya1* controls critical early inductive events involved in the morphogenesis of thymus, parathyroid and thyroid.

Keywords

Eya1; Thymus; Parathyroid; Thyroid; Morphogenesis; *Hox*; *Pax*; *Six1*; *Gcm2*; Neural crest; Endoderm; Ectoderm; Apoptosis; Mouse

INTRODUCTION

The morphogenesis of mammalian organs derived from the pharyngeal region is complex (Hilfer and Brown, 1984). The thymus and parathyroids form as lateral evaginations of the 3rd pharyngeal pouches, which are derived from the foregut endoderm. The thyroid is formed by the fusion of two structures of separate embryonic origin: the thyroid diverticulum, derived from the endoepithelium in the floor of the pharynx; and the ultimobranchial bodies, formed as evaginations of the 4th pouches (Hilfer, 1968; Rogers, 1927; Rogers, 1971). For each of these endodermal evaginations, there is a corresponding ectodermal invagination or pharyngeal cleft. The ectodermal and endodermal epithelia, placed back to back, form the pharyngeal membrane (Cordier and Haumont, 1980). During their descent in the neck, these pharyngeal organs are surrounded by mesenchyme derived from the cranial neural crest (LeLievre and LeDouarin, 1975). The organ primordia for thymus, parathyroid and ultimobranchial body are formed through epithelial-mesenchymal interactions at approximately E10.5 (Smith, 1965; Moore and Owen, 1967; Owen and Ritter, 1969; LeDouarin and Joterean, 1975; Cordier and Haumont, 1980; LeDouarin et al., 1984; Manley, 2000; Owen et al., 2000). Chick-quail chimera experiments suggested that the endoderm provides the initiating signals for induction of the thymus, and is sufficient for the development of the major epithelial subtypes (LeDouarin and Joterean, 1975). All of these organs, the thyroid, thymus, parathyroid, and ultimobranchial body, develop concurrently and migrate from their sites of origin in the pharynx and pharyngeal pouches to their final positions along the ventral midline of the pharyngeal and upper thoracic region. During their migration, the organ primordia interact with one another to ensure their normal development (Manley and Capecchi, 1998). At present, the molecular basis for control of these events is largely unknown.

The cranial neural crest has been shown to play a critical role during the development of thymus, parathyroid and thyroid, initially by providing the mesenchymal cells that populate the pharyngeal region. Ablation of neural crest cells in chick results in aplasia or dysplasia of the thymus and abnormal development of other pharyngeal pouch derivatives, facial features, and the heart (Bockman and Kirby, 1984). It has been demonstrated that as development proceeds, the neural crest mesenchyme contributes connective tissue elements to organs developing in the pharyngeal region, including thymus and parathyroid, and forms the calcitonin-producing cells of thyroid gland (LeLievre and LeDouarin, 1975; LeDouarin, 1977; Noden, 1983; Noden, 1984). However, the precise role of neural crest cells in the development of organs derived from the pharyngeal region remains unclear.

The ectoderm has also been shown to play an important role in the development of thymus. In the chick embryos, ablation of the ectoderm of the 3rd and 4th clefts before it fuses with the endoderm of the 3rd and 4th pouches results in a rudimentary thymus, while the parathyroid and the ultimobranchial body develop normally (Hammond, 1954). It has been demonstrated that similarly to man, the entire thymus in the mouse is of a mixed ecto-endodermal origin and that the arrested development of the 3rd ectodermal cleft at E11.5 in nude mice results in thymic dysgenesis (Cordier and Haumont, 1980). Recently, it has been described that the *Foxn1* gene, which encodes a transcription factor of the winged helix/forkhead class, is mutated in the nude mice (Nehls et al., 1994). However, the precise role of the ectoderm in thymus development is also not clear.

Several genes have been implicated in the development of the pharyngeal organs. *Hoxa3*, a member of the Hox gene family, which specifies positional identity in the developing embryo (Krumlauf, 1994), is expressed in both 3rd and 4th pharyngeal pouch endoderm and neural crest mesenchyme. Inactivation of the *Hoxa3* gene in mice results in parathyroid and athymia and persistent ultimobranchial bodies (Manley and Capecchi, 1995). *Pax1* and *Pax9*

are closely related members of the paired-box gene family, which play critical roles in the development of multiple organs (Strachan and Read, 1994; Stuart et al., 1994; Neubuser et al., 1995; Dahl et al., 1997). Inactivation of *Pax9* results in early failure of thymus, parathyroid and ultimobranchial body formation (Peters et al., 1998), while *Pax1* mutants have hypoplastic parathyroid and thymus and disturbed thymocyte maturation (Wallin et al., 1996; Su et al., 2001). Recently, *Gcm2*, homologous to the *Drosophila Glial cells missing* gene encoding a transcription factor with a novel DNA binding domain, has been shown to play a key role specifically for the organogenesis of parathyroid glands (Akiyama et al., 1996; Kim et al., 1998; Gordon et al., 2001). In addition to these transcription factors, retinoid signaling has been shown to be essential for the formation of the 3rd and 4th pharyngeal arches (Dupe et al., 1999; Wendling et al., 2000). Although these studies have started to define specific genes controlling early pharyngeal organ development, the identity of the regulatory pathways has not been established.

Four mammalian *Eya* genes, homologues of the *Drosophila eyes absent (eya)* gene (Bonini et al., 1993), have recently been isolated (Xu et al., 1997a; Abdelhak et al., 1997; Duncan et al., 1997; Zimmerman et al., 1997; Borsani et al., 1999). The *eya* gene products contain a divergent N-terminal transactivation domain (Xu et al., 1997b) and a highly conserved 271 amino acid C-terminal Eya domain that participates in protein-protein interactions with So and Dac, the gene products encoded by the *Drosophila sine oculis (so)* and *dachshund (dac)* genes, respectively (Chen et al., 1997; Pignoni et al., 1997). In mice, *Eya1* and *Eya2* are widely expressed in the cranial sensory placodes and at the sites of inductive tissue interactions during organogenesis, while *Eya3* and *Eya4* are expressed in peri-placodal mesenchyme and dermamyotome, respectively (Xu et al., 1997a; Borsani et al., 1999). These features suggest major roles for *Eya* genes in the development of vertebrate organs and sensory systems, potentially by mediating the expression of inductive signals that act between tissue layers. In addition in many developing tissues, *Eya* genes are widely and strikingly co-expressed with *Pax* and *Six* genes, the latter representing the mammalian homologs of *so* (Oliver et al., 1995a; Oliver et al., 1995b; Xu et al., 1997a), suggesting possible interactions between their gene products. Besides the cranial placodes and sensory systems, *Eya* genes are also strongly expressed in the pharyngeal region and its derivatives from E9.5 during mouse development (Xu et al., 1997a). However, the function of *Eya* genes in pharyngeal development has not been previously described.

We have recently reported that *Eya1*-null animals lack ears and kidneys due to defective inductive tissue interactions and apoptotic regression of organ primordia (Xu et al., 1999a). Interestingly, numerous defects in a continuum of cranial structures that are dependent on neural-crest, beginning in the orbital region and extending through the face and neck into the heart, were observed in *Eya1* mutants (Xu et al., 1999a) (P.-X. Xu, unpublished). We have now examined the development of organs derived from the pharyngeal region, including thymus, parathyroid and thyroid in *Eya1*^{-/-} animals. In *Eya1*^{-/-} embryos, the organ primordia for both thymus and parathyroids fail to form. A thyroid defect is also present, which results from a severe reduction in the number of parafollicular and follicular cells; these cells do not interact with each other owing to a lack of fusion between the ultimobranchial bodies and the thyroid lobe. Furthermore, we show that in *Eya1*^{-/-} embryos, *Six1* expression in pharyngeal arch mesenchyme, pouch endoderm and surface ectoderm was markedly reduced and that *Gcm2* expression in the 3rd pouch endoderm was undetectable. Finally, we have shown that increased cell death was observed in the surface ectoderm of the 3rd and 4th pharyngeal region in *Eya1*^{-/-} embryos of E10.5, indicating that *Eya1* is required for the ectodermal cell survival. Our data indicate that *Eya1* plays an essential role in the neural crest, endodermal and ectodermal cells that are required for the morphogenesis of mammalian thymus, parathyroid and thyroid.

MATERIALS AND METHODS

Animals and genotyping

Mice heterozygous for a targeted disruption of the *Eya1* gene were intercrossed to produce embryos of all three possible genotypes. The mutant allele contained a *neo* cassette replacing most of the conserved Eya domain region (Xu et al., 1999a). This mutant allele is maintained in three different genetic backgrounds (129, BALB/c, C57BL/6J). Genotyping of mice and embryos was performed as described (Xu et al., 1999a).

The *Six1* mutant allele was created by replacement of the endogenous start codon as well as the exon 1 with a promoterless *E. coli* ATG-*lacZ*-poly(A) cassette and the *PGK-neo* gene, and will be presented elsewhere (C. L. and P. M., unpublished). Mutant mice carrying *Six1* mutant allele, *Six1^{lacZ}*, were obtained using gene targeting technology. *Eya1/Six1* double heterozygous mutant mice were generated by crossing mice carrying mutant alleles of *Eya1* and *Six1* (*Six1^{lacZ}*). A set of *lacZ* primers are used to detect the mutant allele and a set of primers within the exon 1 of *Six1* gene are used to detect *Six1* wild-type allele.

Phenotype analyses and in situ hybridization

Embryos for histology and in situ hybridization were dissected out in PBS and fixed with 4% PFA at 4°C overnight. Embryonic membranes were saved in DNA isolation buffer for genotyping. Histology was performed as described previously (Xu et al., 1999a). To visualize *Six1^{lacZ}* expression, mutant embryos were stained with X-gal and sectioned as described previously (Xu et al., 1999b).

For in situ hybridization, we used 3 wild-type or mutant embryos at each stage for each probe as described previously (Xu et al., 1997a).

Immunohistochemistry

Whole-mount immunostaining with Hoxb1-specific antibody (BabCO) and section staining with calcitonin-specific antibody (ICN) were performed as described previously (Manley and Capecchi, 1995). Both antibodies were detected using HRP-coupled secondary antisera (Vector Laboratories) and staining with diaminobenzidine (DAB). Immunostained sections were counterstained with diluted Ehrlich's Hematoxylin, mounted in Permount and viewed using DIC.

TUNEL analysis

We performed TUNEL assay for detecting apoptotic cell death using the ApopTag detection kit (Intergen). We used 3 wild-type or mutant embryos for this assay.

RESULTS

Eya1 is required for normal development of the thymus and parathyroid

Eya1 is strongly expressed in the pharyngeal arches (a1–a4), the pouch endoderm (p2–p4) and the surface ectoderm including clefts (2nd, 3rd and 4th) in the pharyngeal region at E9.5–10.5 (Fig. 1A,B and data not shown) (Kalatzis et al., 1998). Subsequently, *Eya1* expression was observed in the structures that are derived from the pharyngeal region, including thymus and parathyroid (Fig. 1C,D). To study whether *Eya1* plays any role during the formation of thymus and parathyroid, serial histological sections of the neck and upper trunk region of *Eya1^{-/-}* embryos from E14.5 to P0 were analyzed. The thymus and parathyroid, both derivatives of the 3rd pharyngeal pouches, are absent in *Eya1^{-/-}* animals on both BALB/c and C57BL/6J backgrounds (Fig. 1E–H). These results suggest that *Eya1*

expression in the developing thymus and parathyroid glands is required for their normal development.

To determine the onset of phenotypic abnormalities during pharyngeal pouch development, we analyzed *Eya1*^{-/-} embryos at E11.5–12.5. In wild-type embryos, the 3rd pouches evaginate and then separate as epithelial buds to form the primordia of thymus/parathyroid at approximately E12.0 (Fig. 2A). In *Eya1*^{-/-} embryos, however the organ primordia of thymus/parathyroid were never seen (Fig. 2B; other data not shown). In contrast, the primordia of the ultimobranchial bodies derived from 4th pouches formed with slight indentations in both sides of the head in four out of six *Eya1*^{-/-} BALB/c embryos (arrow, Fig. 2B).

Pax1 and *Pax9* genes are expressed in pharyngeal pouch endoderm from E9.5 and in their derivatives (Fig. 2C,E) (Neubuser et al., 1995; Wallin et al., 1996). In *Eya1*^{-/-} embryos, both *Pax1* and *Pax9* expression in the pouch endoderm is not detectably altered at E9.5–10.5, before the budding and outgrowth of thymus/parathyroid rudiments (Fig. 6C–F). However, at E11.5–12.5, *Pax1* and *Pax9* expression was only detected in the rudiments of the ultimobranchial bodies in *Eya1*^{-/-} embryos (Fig. 2C–F). The absence of *Pax1* and *Pax9* expression in the prospective region of the thymus/parathyroid rudiments further confirms the absence of these structures in *Eya1*^{-/-} embryos (Fig. 2C–F).

To determine whether there are immunological deficits in *Eya1* heterozygous mice, we have examined peripheral blood lymphocytes from *Eya1*^{+/-} mice on 129, BALB/c and C57BL/6J inbred strains using flow immunocytometry. Monoclonal antibodies were used to assess the proportions of cells expressing antigens B220 (B cells), CD3 and CD4 (T cells), CD11 (macrophages), and NK1 (natural killer cells). Responsiveness to T cell and B cell mitogens in culture (4 days stimulation with conA and overnight stimulation with LPS) was evaluated by examining expression of the early activation antigen CD69 on Thy 1-positive T cells and B220-positive B cells. None of the three strains differed from wild-type mice (data not shown), suggesting that *Eya1*^{+/-} mice are immunocompetent. In addition, the thymus in the three strains of *Eya1*^{+/-} mice had normal morphology (data not shown).

***Eya1* regulates mature thyroid formation**

The thyroid gland is formed by the fusion of the thyroid diverticulum from the floor of the pharynx and the ultimobranchial body from 4th pharyngeal pouch (Hilfer, 1968; Pearse and Polak, 1971; Rogers, 1927; Rogers, 1971; LeLievre and LeDouarin, 1975; Fontaine, 1979). *Eya1* is expressed in the pouch endoderm, the ectodermal clefts and the arch mesenchyme in the 4th pharyngeal region (Fig. 1A and data not shown). Although the 4th pouches are slightly indented in some *Eya1*^{-/-} animals (Fig. 2B), they do separate as buds to form the rudiments of the ultimobranchial bodies and these rudiments express both *Pax1* and *Pax9* (Fig. 2B,D,F).

Next, we used *Hoxb1* expression as an early marker of the 4th pouch endoderm (Frohman et al., 1990; Manley and Capecchi, 1995). Immunostaining revealed that at E10.5–11.5, the expression level of *Hoxb1* in the 4th pouches (p4) was indistinguishable from that of wild type (brown stain in Fig. 3A,B, and data not shown). Therefore, our results indicate that the 4th pouches form the primordia of the ultimobranchial body in the absence of *Eya1*.

To confirm our observation, we used an antibody against calcitonin to identify parafollicular or C cells that are believed to be neural crest-derived in the ultimobranchial bodies on transverse sections of *Eya1*^{-/-} and wild-type newborn animals. In wild-type mice, the thyroid is a bilobed structure, with two large lateral lobes joined across the ventral midline at their posterior aspects by the isthmus. Fig. 3C shows a transverse section through one lateral

lobe of a control animal. Calcitonin-producing C cells (brown stain) were numerous and could be found throughout the lobe. In contrast, *Eya1*^{-/-} mice had hypoplastic lobes and fewer calcitonin-producing cells, with 4 BALB/c animals showing a reduction of approximately 60% in the number of calcitonin-positive cells and all the calcitonin-positive cells are localized in an anterodorsally placed vesicular structure (Fig. 3D). These vesicles varied in size and were associated with, but had no visible connection to, the thyroid, which lacked calcitonin-positive cells. In addition, we have observed follicular structures in the calcitonin-positive vesicles (Fig. 3D). H and E stained sections of *Eya1*^{-/-} mutants showed that these follicular structures did contain colloid, indicating that thyroglobulin is produced and secreted to the lumen of the follicles (data not shown). These calcitonin-positive vesicles probably present persistent ultimobranchial bodies that failed to fuse with the thyroid diverticulum, similar to that seen in *Hoxa3* mutant mice (Manley and Capecchi, 1995) and in *Echidna*, in which ultimobranchial bodies never fuse with thyroid gland and also have colloid-containing follicles (Maurer, 1899).

We next examined the expression of the thyroid-specific genes *Pax8* and *Tf1* in *Eya1*^{-/-} thyroid morphogenesis. *Pax8* is expressed in the developing thyroid gland from E9.0 and inactivation of *Pax8* results in a total absence of the thyroid follicular cells (Lazzaro et al., 1991; Mansouri et al., 1998). The homeobox gene *Tf1* is expressed in both the ultimobranchial body and the developing thyroid from E9.0 and mice lacking *Tf1* lack both C cells and follicular cells (Lazzaro et al., 1991; Kimura et al., 1996; Mansouri et al., 1998). Both gene products have been shown to cooperate in transcriptional activation of the thyroglobulin gene promoter in follicular cells (Pasca di Magliano et al., 2000). In E15.5 wild-type embryos, although the ultimobranchial bodies are already embedded in the thyroid lobes, they are still visible as discrete cell populations, which show weak *Pax8* expression within the thyroid (arrow, Fig. 3E). Parathyroid glands were visible on the sections in which the ultimobranchial body cell populations were evident (Fig. 3E). However, in *Eya1*^{-/-} embryos, no ultimobranchial body cell population was observed within the thyroid lobes and *Pax8* was strongly expressed throughout the lobe (Fig. 3F). The ultimobranchial bodies showing weak *Pax8* expression were observed at the anterodorsal end of *Eya1*^{-/-} thyroid lobes (Fig. 3H). The *Tf1* probe showed similar results with strong expression in both the ultimobranchial bodies and the thyroid lobes of *Eya1*^{-/-} embryos (Fig. 3I-L) (Mansouri et al., 1998). The ultimobranchial bodies in *Eya1*^{-/-} animals were detected as separate structures located at the anterodorsal end of the thyroid lobes (Fig. 3D,H,L). We examined 4 BALB/c animals at E15.5 with a *Pax8* marker and 4 BALB/c newborn animals with calcitonin staining; 4 showed persistent ultimobranchial bodies on both sides and 4 showed persistent ultimobranchial body on one side only (Table 1).

The structure of the thyroid lobe itself was also affected in *Eya1*^{-/-} newborn mice. In all cases examined the thyroid lobe was reduced by approximately 40–60% (Fig. 3D and data not shown) and three animals lacked an isthmus (Table 1). Follicles were formed, but the follicular cells were reduced in number. To determine whether *Eya1* plays a direct role in the morphogenesis of thyroid gland, we analyzed the thyroid development in *Eya1*^{-/-} embryos and *Eya1* expression in the thyroid from earlier stages. The morphogenesis of thyroid glands begins at around E10.5 when a small region of the floor of the pharyngeal endoderm invaginates to form the thyroid bud (Fig. 4A). The thyroid bud migrates dorsocaudally to the ventral midline (Fig. 4C). In *Eya1*^{-/-} embryos, the thyroid primordia form and express both *Pax8* and *Tf1* (Fig. 4B,D,F,H). We next performed in situ hybridization on both transverse and sagittal sections of 6 wild-type embryos at each stage to determine whether *Eya1* is expressed in the thyroid. We failed to detect convincing *Eya1* expression in the thyroid at E9.5–14.5 (Fig. 4I,J and Fig. 1D and data not shown), indicating that *Eya1* is not expressed in the thyroid gland at these stages. Therefore, it is possible that the defect detected in the thyroid lobe of *Eya1*^{-/-} animals is due to the lack of fusion with

the ultimobranchial body. Nonetheless, our results show that *Eya1* is required for the formation of a mature thyroid gland.

***Eya1* is required for the expression of *Six1* in pharyngeal arch mesenchyme**

The formation of the thymus, parathyroid and ultimobranchial body involves interactions between several embryonic cell types (Auerbach, 1960; Rogers, 1971; Bockman and Kirby, 1984). These morphogenetic events require the properly timed interactions of neural crest-derived arch mesenchyme, pouch endoderm, and possibly surface ectoderm from pharyngeal clefts (Auerbach, 1960; Cordier and Haumont, 1980; Bockman and Kirby, 1984; Kuratani and Bockman, 1990). *Eya1* is expressed in all three cell types from early embryonic stages (Fig. 1A,B). To determine the earliest stage at which a defect in thymus, parathyroid and thyroid development could be detected, we examined the expression of genes that are known to be expressed in the neural crest, endodermal or ectodermal cells as molecular markers. Embryos were initially analyzed at E9.5 and 10.5, after neural crest migration is complete (E10.5), but before overt thymus and parathyroid development is detectable.

We first examined the expression of *Hoxa3*, *Msx1* and *Six1* in the neural crest cells in *Eya1*^{-/-} embryos. *Hoxa3* is expressed in premigratory neural crest cells in hindbrain rhombomeres 5, 6 and migratory neural crest cells that populate the 3rd and 4th pharyngeal arches (Manley and Capecchi, 1995). No significant difference of *Hoxa3* expression in the neural tube, migratory neural crest cells or 3rd and 4th pharyngeal arches was observed in *Eya1*^{-/-} embryos at E9.5–10.5 (Fig. 5A,B and data not shown). *Msx1*, another homeobox gene, is normally expressed in the distal pharyngeal arch and below the epithelial layer at E10.5 (Brown et al., 1993), which overlaps with the expression domain of *Eya1*. No significant difference of *Msx1* expression in the pharyngeal arches was observed between wild-type and *Eya1*^{-/-} embryos at E9.5 (Fig. 5C,D). At E10.5, after completion of neural crest migration, the *Msx1* expression domain became smaller in *Eya1*^{-/-} embryos (data not shown). Since *Eya1*^{-/-} embryos show hypoplasia of the pharyngeal arches, determined by H and E staining of 12 E10.5 *Eya1*^{-/-} embryos on both transverse and sagittal sections (data not shown), this indicates that the reduction of the *Msx1* expression domain could be a result of the size reduction of *Eya1*^{-/-} pharyngeal arches. Nonetheless, our results indicate that *Eya1* is not required for the expression of *Msx1* in the pharyngeal arch mesenchyme.

Six1, a member of the *Six* gene family, is normally co-expressed with *Eya1* in the pharyngeal arch mesenchyme (Fig. 5E), and its expression depends on *Eya1* function during otic development (Xu et al., 1999a). Similar to otic development, *Six1* expression was markedly reduced in *Eya1*^{-/-} pharyngeal arches at E9.5 and 10.5 (Fig. 5F and data not shown). Therefore, our results show that *Eya1* is required for the normal expression of *Six1* in the neural crest of pharyngeal arches.

***Eya1* is required for the expression of *Six1* in the pouch endoderm and the surface ectoderm**

We next examined the expression of *Hoxa3*, *Pax1*, *Pax9*, *Fgf3* and *Six1* in the pharyngeal pouch endoderm at E9.5–10.5. *Hoxa3* is also expressed in 3rd and 4th pouch endoderm and neural crest mesenchyme, and absence of *Hoxa3* results in aparathyroid, athymia and thyroid hypoplasia (Manley and Capecchi, 1995). Similar to that seen in neural crest cells, *Hoxa3* expression is not affected in the pouch endoderm in *Eya1*^{-/-} embryos (Fig. 6A,B). *Pax1* and *Pax9* are normally expressed in pouch endoderm and deletion of *Pax1* results in parathyroid and thymus hypoplasia, while deletion of *Pax9* results in absence of all derivatives from 3rd and 4th pharyngeal pouches (Dietrich and Gruss, 1995; Neubuser et al., 1995; Wallin et al., 1996; Peters et al., 1998). At E9.5–10.5, both *Pax1* and *Pax9* expression in pouch endoderm is not detectably altered in *Eya1*^{-/-} embryos (Fig. 6C–F and data not

shown). *Fgf3* is expressed in the posterior half of the pouch endoderm (Fig. 6G) (Wilkinson et al., 1989), and its expression depends on *Eya1* function during otic development (Xu et al., 1999a). However, no significant difference in *Fgf3* expression was detected in the pharyngeal pouches between control and mutant embryos (Fig. 6G,H). Taken together, these results indicate that *Hoxa3*, *Pax1*, *Pax9* and *Fgf3* function either upstream or independently of *Eya1*.

In contrast, the expression of *Six1*, also co-expressed with *Eya1* in pharyngeal pouches at E9.5–10.5, was significantly reduced in the pouch endoderm of *Eya1*^{-/-} embryos (arrows in Fig. 6I,J and data not shown). In addition, the expression of *Six1* in the ectodermal clefts was also markedly reduced (arrowheads, Fig. 6I,J). To further confirm our observation, we then crossed the *Six1*^{lacZ} allele into *Eya1*^{+/-} mice and generated *Eya1*^{-/-}*Six1*^{lacZ/+} embryos to determine the expression of *Six1* in *Eya1*^{-/-} pouch endoderm and surface ectoderm using X-gal staining for *Six1*^{lacZ}. As shown in Fig. 6K, *Six1*^{lacZ} was strongly expressed in the 2nd, 3rd and 4th pouch endoderm and the surface ectoderm including 2nd, 3rd and 4th pharyngeal clefts. However, its expression in *Eya1*^{-/-} embryos was markedly reduced in both the pouch endoderm and the surface ectoderm (arrows and arrowheads, Fig. 6L). The expression of *Six1*^{lacZ} surrounding the arteries of pharyngeal arches remained unaffected in *Eya1*^{-/-} embryos (open arrowheads, Fig. 6K,L). These results indicate that the expression of *Six1* also requires *Eya1* function in both the pouch endoderm and the surface ectoderm.

Gcm2 expression in the 3rd pharyngeal pouch endoderm is also Eya1-dependent

To further investigate the molecular defects in the formation of thymus and parathyroid primordia in *Eya1*^{-/-} embryos, we examined the expression of the parathyroid-specific *Gcm2* and thymus-specific *Foxn1* genes. Mice lacking *Gcm2* have no identifiable parathyroid glands, but have normal thymus (Gunther et al., 2000). *Gcm2* is expressed in the 3rd pouch endoderm from E9.5 and its expression becomes restricted to a small domain within the 3rd pouch endoderm by E10.5 (Fig. 7A and data not shown) (Gordon et al., 2001). However, *Gcm2* expression was not detectable in the 3rd pouches of *Eya1*^{-/-} embryos at E9.5–11.5 (Fig. 7B and data not shown), indicating that *Gcm2* expression in the 3rd pouch endoderm is also *Eya1* dependent.

Foxn1 is required cell-autonomously for thymic epithelial cell differentiation (Nehls et al., 1994; Blackburn et al., 1996), but is not required for the initiation of thymus organogenesis (Nehls et al., 1996). *Foxn1* is first expressed in the common primordia of thymus/parathyroid after it forms at around E11.5 and its expression marks the presumptive thymus-specific region in a complementary pattern to the parathyroid marker *Gcm2* (Fig. 7C) (Gordon et al., 2001). In *Eya1*^{-/-} embryos, the 3rd pouches do not separate from the pharyngeal endoderm as epithelial buds and no *Foxn1* expression was detected in the 3rd pharyngeal region (Fig. 7D and data not shown). The absence of *Foxn1* expression in the prospective region of the thymus/parathyroid primordia in *Eya1*^{-/-} embryos further suggests that the formation of the primordia is not initiated in *Eya1*^{-/-} embryos.

Abnormal induction of apoptosis of the pharyngeal surface ectoderm

Since we have previously observed that *Eya1* is required for cell survival in the otic epithelium and metanephric mesenchyme, we analyzed whether the hypoplasia of the pharyngeal arches and the defective formation of pharyngeal organs in *Eya1*^{-/-} embryos could partly result from programmed cell death (apoptosis). Apoptotic cells were detected in the pouch endoderm and arch mesenchyme in wild-type embryos at E10.5–11.5 (Fig. 8A–C and data not shown). In *Eya1*^{-/-} embryos, the number of apoptotic cells detected by TUNEL assay was slightly increased in the proximal region of the 1st and 2nd pharyngeal arches (arrows, Fig. 8D,E). However, no significant changes of apoptotic cells were found in the

region where *Eya1* is strongly expressed in the 1st and 2nd pharyngeal arches of *Eya1*^{-/-} embryos (Fig. 8A,B,D,E). We have also not detected significant changes of apoptotic cells in the 3rd and 4th pharyngeal arch mesenchyme and pouch endoderm (Fig. 8B,C,E,F and data not shown), indicating that apoptosis was not altered in the 3rd and 4th pharyngeal arches and pouches in the absence of *Eya1*.

In contrast, abnormal apoptosis was observed in the surface ectoderm including 3rd and 4th clefts in *Eya1*^{-/-} embryos (reflexed arrows, Fig. 8E,F and data not shown). This indicates that *Eya1* is required for the pharyngeal ectodermal cell survival.

DISCUSSION

Development of the thymus, parathyroid and thyroid normally occurs between E10.5 and E15.5 of mouse embryogenesis. In the absence of *Eya1*, the failure of thymus and parathyroid development occurs coincidentally with the initial induction of organogenesis. Our data indicate that *Eya1* is expressed and plays a role in the neural crest, endodermal and ectodermal cells that are required for the induction of thymus and parathyroid organogenesis.

The role of *Eya1* in the development of neural crest-derived pharyngeal arch mesenchyme

Since the neural crest mesenchymal structures derived from the 3rd and 4th pharyngeal arches and pouches are affected similarly by neural crest ablation in chick, and by loss of *Eya1*, we conclude that *Eya1* is required for normal neural crest development in pharyngeal arches (Bockman and Kirby, 1984; Bockman et al., 1989; Kuratani and Bockman, 1990). *Eya1* is not expressed in premigratory neural crest cells and our marker analysis of *Hoxa3* shows that there is no significant change of *Hoxa3* expression in premigratory, migratory neural crest cells or in the 3rd and 4th pharyngeal arches in *Eya1*^{-/-} embryos. Thus, the neural crest defects seen in *Eya1* mutants must occur at the postmigratory stage. Interestingly, our analysis has shown that *Six1*, which is normally co-expressed with *Eya1* in pharyngeal arch mesenchyme, was markedly reduced in *Eya1*^{-/-} embryos. Besides the defects detected in neural crest mesenchyme of pharyngeal arches, *Eya1*^{-/-} mice show multiple defects in cranial structures which are dependent on neural crest, including lack of multiple cranial ganglia and nerves, malformation of the skull, abnormal development of eyelids, middle ear ossicles, multiple skeleton components and other derivatives from the pharyngeal arches and pouches (including the parafollicular cells), cleft secondary palate formation, malformation of the tongue and heart (P-X. Xu, unpublished). Thus, *Eya1* probably plays a critical role in neural crest development. The neural crest defects seen in *Eya1*^{-/-} embryos may be caused by inappropriate cell proliferation. Consistent with this view, no abnormal cell death in both 3rd and 4th pharyngeal arches was observed. Nonetheless, the neural crest defect detected in the pharyngeal arches of *Eya1*^{-/-} embryos could contribute to the defective formation of thymus and parathyroid.

A *Pax-Eya-Six* regulatory hierarchy in pharyngeal pouch endoderm

Since *Eya1* is expressed in the 3rd pouch endoderm before the initiation of thymus and parathyroid formation and our marker gene analyses show that the expression of *Six1* was undetectable in the *Eya1*^{-/-} pouch endoderm, indicating that *Eya1* also plays a role in the pouch endoderm. Our data also suggest that similar to the otic and kidney induction (Xu et al., 1999a), *Eya1* functions in an evolutionarily conserved cassette downstream of *Pax* genes and upstream of *Six* genes during the development of pharyngeal pouch endoderm. It has been shown that deletion of *Pax9* results in lack of 3rd and 4th pouches, while deletion of *Pax1* results in parathyroid and thymus abnormalities at later stages (Dietrich and Gruss, 1995; Wallin et al., 1996; Peters et al., 1998; Su et al., 2001). In *Eya1*^{-/-} pouch endoderm,

Pax1 and *Pax9* expression was not affected. We have also assayed *Eya1* expression in *Pax9* mutant embryos and found no significant changes of *Eya1* expression in the *Pax9*-deficient pharyngeal pouch endoderm at E9.5 (data not shown). Although *Pax1* apparently does not compensate for *Pax9*-deficiency during embryonic parathyroid and thymus formation, it is possible that *Pax1* and *Pax9* function redundantly in the pharyngeal pouches and that a critical threshold of Pax1/Pax9 protein expression in pharyngeal endoderm regulates *Eya1* expression. Such compensatory up-regulation has been noted for *Pax1* and *Pax9* in the formation of other organs (Peters et al., 1999). Consistent with this genetic relationship between *Pax1*, *Pax9* and *Eya1*, our data suggest that *Eya1* also functions downstream of *Hoxa3* in the 3rd and 4th pouch endoderm. *Hoxa3* expression is unaffected in the 3rd and 4th pouch endoderm of *Eya1* mutant embryos, and *Pax1* and *Pax9* expression is down-regulated in *Hoxa3* mutants, which also lack parathyroid and thymus development (Chisaka and Capecchi, 1991; Manley and Capecchi, 1995). During parathyroid organogenesis, recent studies have shown that *Hoxa3* is required for the initiation of *Gcm2* expression in the 3rd pouch endoderm and both *Hoxa3* and *Pax1* are required for the maintenance of *Gcm2* expression (Su et al., 2001) (S. Ellis, J. Koushik, J. Gordon and N. R. Manley, unpublished). Our data indicate that *Gcm2* requires *Eya1* function for its expression in the 3rd pouch endoderm before the formation of thymus/parathyroid primordia. Therefore, it is possible that *Gcm2* expression is regulated by the *Hoxa3*-*Pax1/9*-*Eya1* pathway during parathyroid organogenesis. *Gcm2* expression is initiated normally at E9.5–10.5 in *Pax1* single mutants (Su et al., 2001). This could be the result of *Eya1* expression being correctly initiated in the 3rd pouch endoderm in *Pax1* single mutants, similar to that seen in *Pax9* single mutants (data not shown). The genetic relationship between *Eya* and *Pax* genes may be pertinent in other developmental contexts, since both *Eya1* and *Pax9* mutants exhibit related palate defects (Peters et al., 1998), and *Pax9* expression was unaffected in *Eya1* mutant palatal shelves (P.-X. Xu, unpublished). We are in the process of further clarifying the genetic relationship between *Pax* and *Eya1* genes by examining the *Eya1* expression in pharyngeal pouches and palatal shelves of *Pax1/Pax9* double mutants, and in the otic epithelium and kidney of *Pax2/Pax8* double mutants.

The role of *Eya1* in the development of pharyngeal surface ectoderm

Since *Eya1* is also expressed in the surface ectoderm, including that of the 2nd, 3rd and 4th clefts, and in the absence of *Eya1* the surface ectoderm undergoes abnormal apoptosis, it would appear that *Eya1* also plays a role in the development of pharyngeal surface ectoderm.

Previous studies have demonstrated the importance of the ectoderm in the thymus development. It has been suggested that most of the ectoderm is formed by the proliferation of the 3rd cleft, which covers the endodermal part at around E11.0 (Cordier and Haumont, 1980). In addition, the cervical vesicle, formed by an invagination of the dorsal segment of the ectoderm between the 3rd and 4th pharyngeal clefts, also contributes to the formation of the thymus (Cordier and Haumont, 1980). In the chick embryo, after removal of the ectoderm of the 3rd and 4th clefts before it fuses with the pouch endoderm, the endoderm produces only a rudimentary thymus, while the parathyroid and the ultimobranchial body develop normally (Hammond, 1954). In *Foxn1* mutant nude mice, the arrested development of the 3rd ectodermal cleft resulted in thymic dysgenesis, while the parathyroid develops normally (Cordier and Haumont, 1980). It has been proposed that the thymic dysgenesis in the nude mice is caused by the fact that the endoderm is no longer covered by the ectoderm and is deprived of its normal inducing agent (Cordier and Haumont, 1980). In *Eya1*^{-/-} embryos, we found that *Six1* expression in the surface ectoderm, including the 3rd cleft, was significantly reduced compared to wild-type embryos. This indicates that *Eya1* may function along with *Six1* in a molecular pathway within the ectoderm, which may provide inducing

signals for the development of pouch endoderm. In addition, our data indicate that *Eya1* is required for ectodermal cell survival. Thus, the thymic agenesis detected in *Eya1*^{-/-} animals may partly result from the defects in the ectoderm.

Thyroid phenotype

The thyroid phenotype observed in *Eya1* mutants does not occur as early as the defects in the thymus and parathyroid development. Although *Eya1* mutant mice show reduction in the number of C cells in the thyroid, *Eya1* is not required for the differentiation of either C or follicular cells. Interestingly, C cells are only observed in the persistent ultimobranchial bodies, while follicular cells are found in both the persistent ultimobranchial bodies and the thyroid lobes, similar to the phenotype observed in *Hoxa3* mutant mice (Manley and Capecchi, 1995; Manley and Capecchi, 1998).

Previous studies have shown that *Pax8* is required for the differentiation of thyroid follicular cells (Plachov et al., 1990; Mansouri et al., 1998). In *Pax8*^{-/-} mice, the ultimobranchial bodies form and calcitonin-positive C cells are identified, however, both thyroglobulin and thyroperoxidase expression was not detected in *Pax8*^{-/-} ultimobranchial bodies (Mansouri et al., 1998), indicating that there are no follicular cells. Previous studies have also shown that *Pax8* and *Ttf1* gene products cooperate to activate the expression of thyroglobulin gene in the follicular cells (Pasca di Magoliano et al., 2000). Interestingly, we have found that *Pax8* expression is ectopically turned on in *Eya1*^{-/-} ultimobranchial bodies at E14.5–15.5, when the thyroglobulin gene begins to be expressed (Lazzaro et al., 1991). This suggests that the ectopic expression of *Pax8* may be required for the expression of thyroglobulin in the follicular cells of *Eya1*^{-/-} ultimobranchial bodies. Consistent with this, H&E stained sections of *Eya1*^{-/-} persistent ultimobranchial bodies showed the presence of colloid-containing follicles, indicating that thyroglobulin is produced and secreted into the lumen of the follicles (data not shown). Since *Eya1* is not expressed in the thyroid primordia at E10.5–14.5, it is possible that the defect detected in the thyroid lobe is due to the lack of fusion with the ultimobranchial body.

In summary, our results suggest that the *Eya1* protein level or activity critically affects the development of the thymus, parathyroid and thyroid and that *Eya1* controls critical early inductive events that are required for the morphogenesis of mammalian thymus, parathyroid and thyroid.

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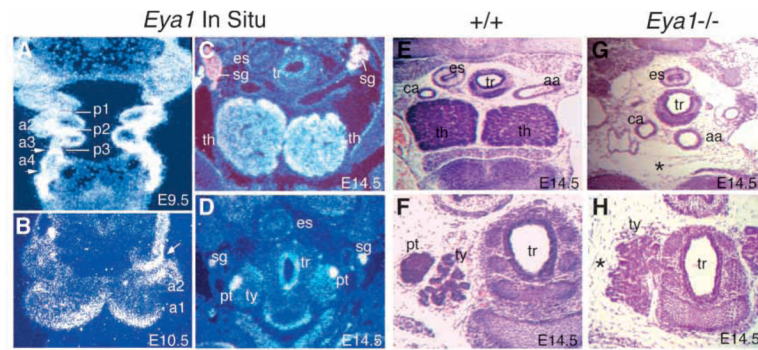


Fig. 1. *Eya1*^{-/-} embryos lack thymus and parathyroid glands. (A) A frontal section showing *Eya1* expression in pharyngeal arch (a1–4) mesenchyme, pouch endoderm (p1–3) and pharyngeal clefts (arrows) at E9.5. Cranial is up. (B) A transverse section showing strong *Eya1* expression in the distal part of the pharyngeal arches and the surface ectoderm of 2nd cleft (arrow) at E10.5. (C,D) Transverse sections showing *Eya1* expression in the thymic lobes (th) and parathyroid glands (pt) at E14.5. In addition, *Eya1* is also expressed in sympathetic ganglia (sg) and trachea (tr). *Eya1* is not expressed in the thyroid lobe (ty). (E–H) H and E-stained transverse sections of the neck region at E14.5. (E,F) In wild-type embryos, two thymic lobes are present and the parathyroid glands are associated with the thyroid gland (ty). (G,H) In *Eya1*^{-/-} embryos, no thymus and parathyroid formation (*) was found at the same level or in other regions of the neck and upper trunk. For B–H, dorsal is up. aa, aortic arches; ca, carotid artery; es, esophagus.

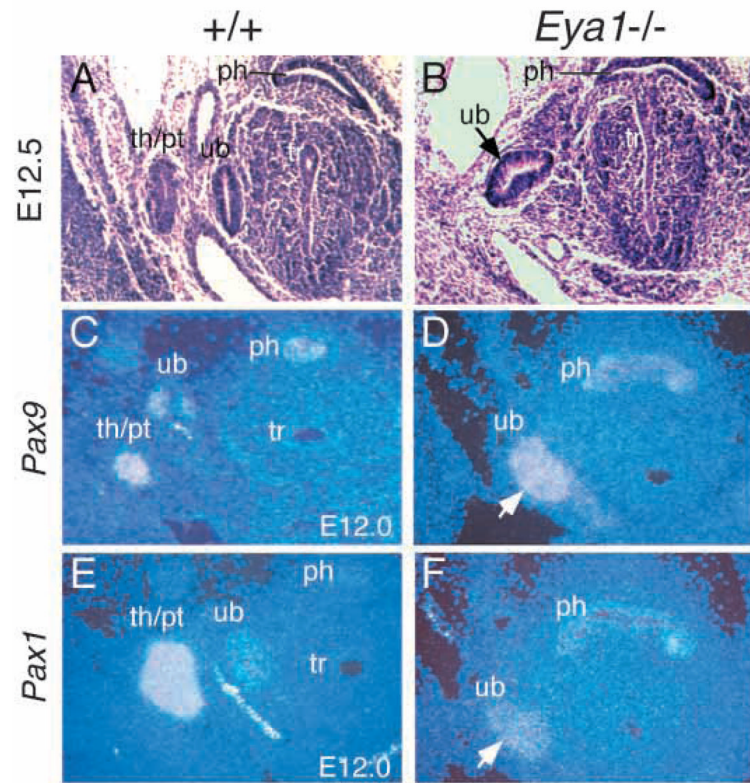


Fig. 2.

The 3rd pharyngeal pouches do not separate as buds to form the primordia of thymus/parathyroid. (A,B) Transverse sections showing that in wild-type embryos (A), the 3rd pouches evaginate and then separate as buds to form the primordia of thymus/parathyroid (th/pt) at around E12.5, while the 4th pouches separate to form the rudiments of the ultimobranchial bodies (ub) at the same time; however in *Eya1*^{-/-} embryos (B), the primordia of thymus/parathyroid failed to form, while the rudiments of ultimobranchial bodies (ub) were formed but with a slight indentation (arrow in B). (C–F) Transverse sections showing *Pax1* and *Pax9* expression in the rudiments of the thymus/parathyroid and ultimobranchial bodies at E12.0 of wild-type embryos (C,E). In *Eya1*^{-/-} embryos, the expression of *Pax1* and *Pax9* was only detected in the rudiments of ultimobranchial bodies (arrows). The absence of *Pax1* and *Pax9* expression in the prospective region of the thymus/parathyroid rudiments confirms the absence of these structures in *Eya1*^{-/-} embryos (D,F). Dorsal is up. ph, pharynx; tr, trachea.

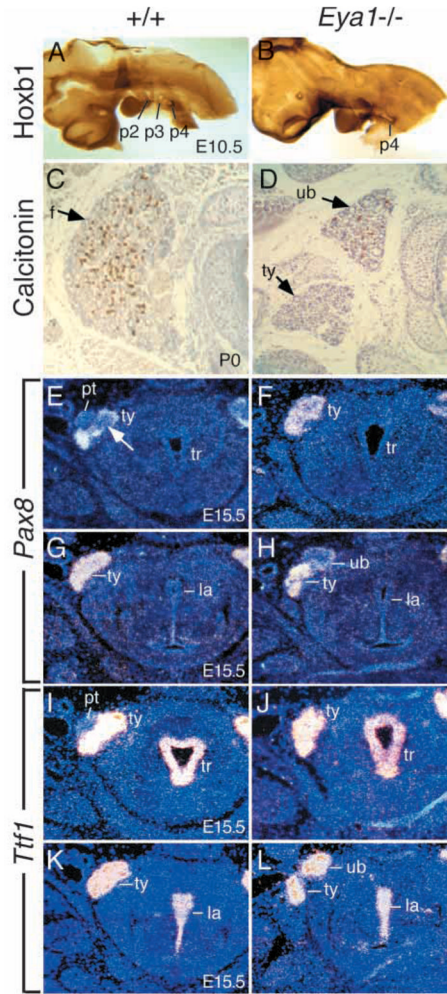


Fig. 3. Ultimobranchial body and thyroid lobe defects in *Eya1*^{-/-} mice. (A,B) The embryos of E10.5, after completion of neural crest migration, were stained with Hoxb1 antibody to label the 4th pharyngeal pouches (p4). The 4th pharyngeal pouches express Hoxb1 (brown stain) in both wild-type (A) and *Eya1*^{-/-} embryos (B). No significant difference of Hoxb1 expression in the 4th pouch endoderm was observed in *Eya1* mutants. (C,D) Transverse sections through newborns, stained with anti-calcitonin antibody (brown staining). (C) A wild-type thyroid with numerous calcitonin-positive cells and follicular cells (f, arrow) throughout the lobe. (D) An *Eya1*^{-/-} animal with bilateral persistent ultimobranchial bodies and malformed thyroid lobes. Only a few follicles are formed in the main body of the thyroid (ty, arrow). Note that the dorsally placed vesicle which is strongly positive for calcitonin represents a persistent ultimobranchial body (ub) in *Eya1*-mutants, not fusing with the thyroid lobe. This ultimobranchial body also contains follicle-like structures (arrow). No isthmus was present in this animal. (E–H) Transverse sections at E15.5 showing *Pax8* expression in the thyroid lobes in wild-type and *Eya1*^{-/-} embryos. (E) The ultimobranchial body cell populations showing weak *Pax8* expression (arrow) were visible within the thyroid lobes at this stage. Parathyroid (pt) was also visible. (F) No ultimobranchial body cell population was observed within the *Eya1*^{-/-} thyroid lobes on the same level and parathyroid was also absent. However, the ultimobranchial bodies showing weak *Pax8* expression were observed at the anterior end dorsal to the thyroid lobe in *Eya1*^{-/-} embryos

(H). No ultimobranchial bodies were found in wild-type embryos on the same level (G). (I–L) Transverse sections showing *Ttf1* expression in the thyroid lobes in wild-type embryos at E15.5 (I,K) and in both the persistent ultimobranchial bodies and the thyroid lobes of *Eya1^{-/-}* embryos (J,L). Similarly, the persistent ultimobranchial bodies were observed as separate structures located anterodorsally to the thyroid lobes in *Eya1^{-/-}* animals (L). For C–L, dorsal is up. tr, trachea; la, larynx.

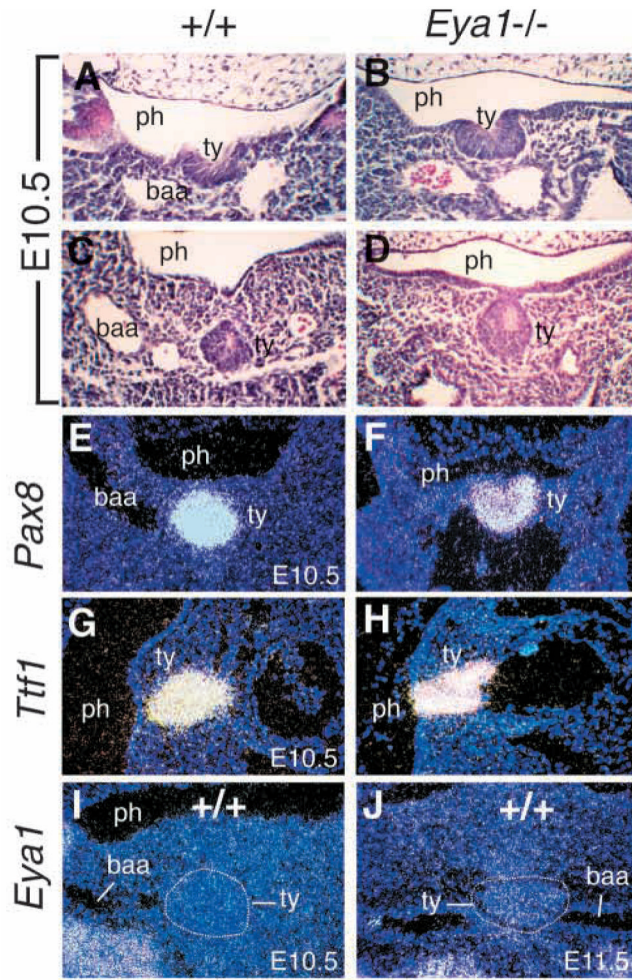


Fig. 4. Normal development of the thyroid primordia in *Eya1*^{-/-} embryos. (A–D) Transverse sections showing the thyroid primordia (ty) in wild-type (A,C) and *Eya1*^{-/-} (B,D) embryos at E10.5. The thyroid primordia were evident in *Eya1*^{-/-} embryos. (E,F) Transverse sections showing *Pax8* expression in the thyroid primordia of wild-type and *Eya1*^{-/-} embryos at E10.5. (G,H) Sagittal sections showing *Ttf1* expression in wild-type and *Eya1*^{-/-} thyroid primordia at E10.5. (I,J) Transverse sections showing that *Eya1* expression was not detectable in the thyroid primordia at E10.5–11.5 in wild-type embryos. ph, pharynx; baa, 2nd branchial arch artery. For transverse sections, dorsal is up; for sagittal sections, cranial is up and dorsal is to the left.

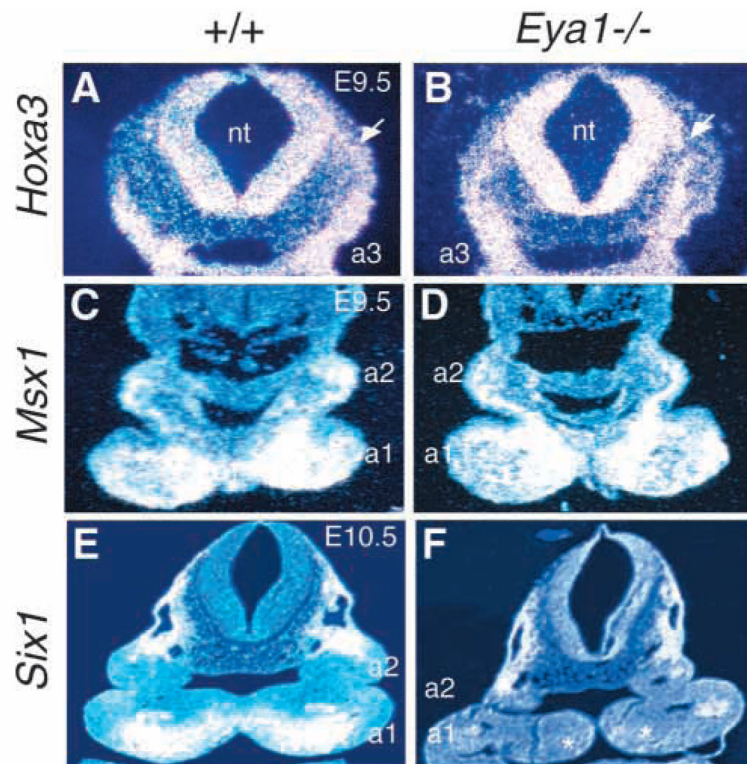


Fig. 5. Neural crest defects in the pharyngeal arches of *Eya1*^{-/-} embryos. (A,B) Transverse sections of wild-type and *Eya1*^{-/-} embryos at E9.5 showing *Hoxa3* expression in the hindbrain neural tube, migrating neural crest cells (arrow) and 3rd pharyngeal arches (a3). No significant change was found in *Eya1* mutants. (C,D) Transverse sections showing *Msx1* expression in the neural crest cells in the pharyngeal arches of wild-type and *Eya1*^{-/-} embryos at E9.5. No significant difference of *Msx1* expression was observed at E9.5 in *Eya1*^{-/-} embryos. (E,F) Transverse sections showing *Six1* expression in the distal edge of arch mesenchyme in wild-type and *Eya1*^{-/-} embryos at E10.5. *Six1* expression was not detectable in *Eya1*^{-/-} embryos (asterisks in F). Dorsal is up.

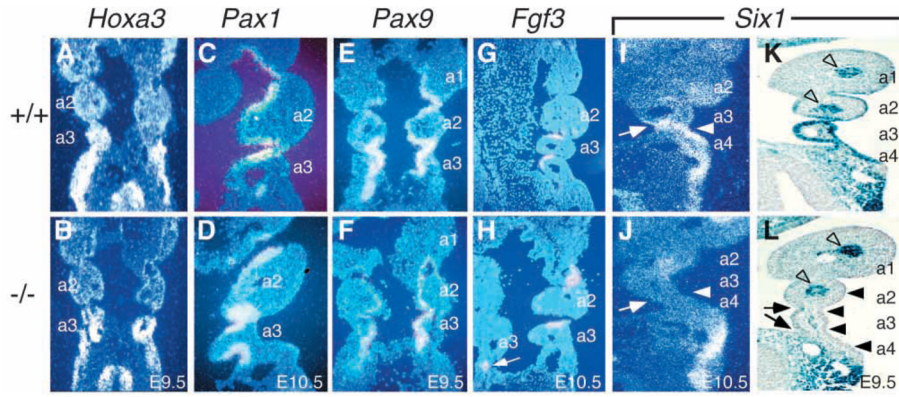


Fig. 6. Expression of *Six1* in the pharyngeal pouch endoderm and the ectoderm of pharyngeal clefts was markedly reduced in *Eya1*^{-/-} embryos. (A,B) Coronal sections showing that *Hoxa3* is normally expressed in the 3rd and 4th pharyngeal pouches and arches (A), and its expression was unaffected in *Eya1*^{-/-} embryos at E9.5 (B). (C,D) Coronal sections showing that *Pax1* is normally expressed in the pouch endoderm at E9.5–10.5 (C), and its expression is preserved in *Eya1*^{-/-} embryos (D). (E,F) Coronal sections showing that *Pax9* is also expressed in the pouch endoderm at E9.5–10.5 (E) and its expression is also preserved in *Eya1*^{-/-} embryos (F). (G,H) Coronal sections showing that *Fgf3* is normally expressed in the posterior half of the pouch endoderm at E10.5 (G) and its expression was unaffected in *Eya1*^{-/-} embryos (H). (I,J) Coronal sections showing that *Six1* is normally co-expressed with *Eya1* in the 3rd pouch endoderm (arrow in I) and the surface ectoderm of 3rd clefts (arrowhead in I), and its expression was markedly reduced in both structures in *Eya1*^{-/-} embryos (arrows and arrowheads in J). (K,L) Coronal sections showing strong *Six1*^{lacZ} expression in wild type (K) in the pouch endoderm and surface ectoderm including 2nd, 3rd and 4th pharyngeal clefts; however, in *Eya1*^{-/-} embryos, *Six1*^{lacZ} expression was significantly reduced in the pouch endoderm (arrows) and surface ectoderm (arrowheads, L) in the 2nd, 3rd and 4th pharyngeal regions (a2–a4). In contrast, its expression surrounding the arteries of pharyngeal arches remains unaffected (open arrowheads).

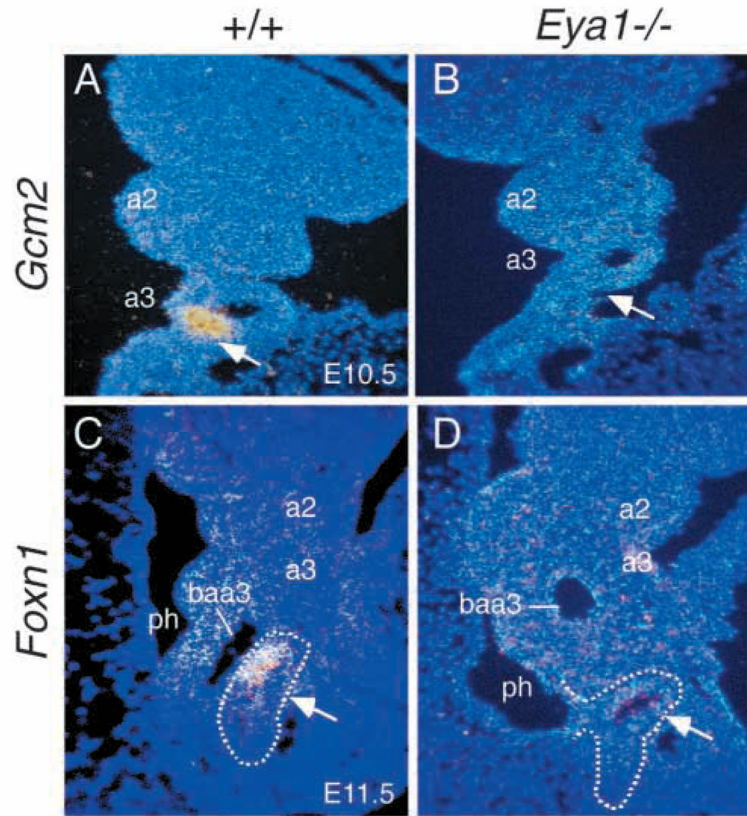


Fig. 7. *Gcm2* and *Foxn1* expression was not detectable in *Eya1*^{-/-} embryos. (A,B) Coronal sections through the 3rd pharyngeal pouch of E10.5 wild-type and *Eya1*^{-/-} embryos showing that *Gcm2* is strongly expressed in the 3rd pouch endoderm in wild-type embryos (arrow, A) and its expression was not detectable in *Eya1*^{-/-} embryos (arrow, B). (C,D) Sagittal sections through the 3rd pharyngeal region showing that *Foxn1* is expressed in the ventral region of the thymus/parathyroid primordia at E11.5 of wild-type embryos (arrow, C), but its expression was not detectable in *Eya1*^{-/-} embryos (arrow, D). Note that the 3rd pouch is attached to the pharyngeal endoderm and does not separate as buds to form the primordia of thymus/parathyroid in *Eya1*^{-/-} embryos. Cranial is up (A–D) and dorsal is to the left (C,D). a, pharyngeal arches; baa3, 3rd branchial arch arteries; ph, pharynx.

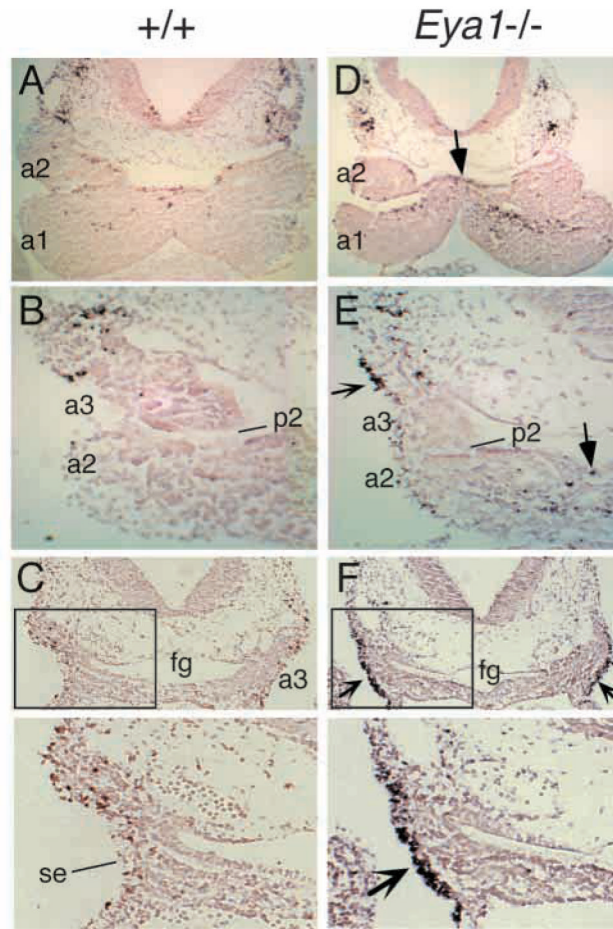


Fig. 8. TUNEL analysis of transverse sections through the pharyngeal region in wild-type and *Eya1*^{-/-} embryos at E10.5. (A–C) Apoptotic cells were observed in the pharyngeal arches (a) and pouches (p) in wild-type embryos. (D–F) In *Eya1*^{-/-} embryos, there were more apoptotic cells in the proximal region of the 1st and 2nd arches (arrows in D,E, other data not shown). No significant changes of apoptotic cells were observed either in the neural crest mesenchyme or the pouch endoderm in the 3rd and 4th pharyngeal regions (E,F and other data not shown). Note that numerous apoptotic cells were detected in the surface ectoderm (se) of the 3rd (reflexed arrows in E,F) and 4th pharyngeal regions (data not shown). The panels below C and F are higher magnification of the boxed areas. Dorsal or cranial is up. fg, pharyngeal region of foregut.

Table 1Summary of thyroid and ultimobranchial body defects in *Eya1*^{-/-} animals

No. of animals analyzed	No. of ub defects	Types of ub defects	Thyroid isthmus defects
4 at E15.5*	4/4	2 persistent on both sides 2 persistent on one side and absent on the other side	
4 newborns†	4/4	2 persistent on both sides 2 persistent on one side and absent on the other side	3/4 absent

* Animals at E15.5 were analyzed by *Pax8* expression. Ultimobranchial bodies (ub) showing weak *Pax8* expression and located as separate structures anterodorsal to the thyroid lobes were scored as persistent. Absent ubs were scored as absence of both a persistent ub and weak *Pax8* expression within the thyroid lobe on a single side.

† Animals at newborn stage were analyzed by calcitonin staining. ubs were scored as absent if there was no persistent ub and no calcitonin-positive cells within the thyroid lobe on a single side. Three animals lacked an isthmus.