

Research Article

Clinical Characteristics of Turkish Women with *Candida krusei* Vaginitis and Antifungal Susceptibility of the *C. krusei* Isolates

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Objective. *Candida krusei* causes approximately 1% of vulvovaginal candidiasis (VVC) cases and is naturally resistant to fluconazole. Antifungal testing may be required if *C. krusei* vaginitis fails to respond to non-fluconazole therapy, particularly in patients with recurrent infections. **Design.** We investigated the clinical characteristics and antifungal susceptibility profile of vaginal *C. krusei* isolates. Between 2009 and 2012, we identified 560 unrelated *Candida* spp.-positive vaginal cultures, of which 28 (5.0%) were *C. krusei*. These isolates were analyzed according to host factors and the clinical forms of VVC, and their *in vitro* susceptibility to 10 antifungal agents was tested using a reference microdilution method. **Results.** We observed that perineal laceration and increased age (>50 years) were significant predictors of *C. krusei* in vaginal samples ($P < 0.05$). All isolates were susceptible to amphotericin B, caspofungin, ketoconazole, and miconazole. Additionally, susceptible dose-dependent and resistant rates were found for fluconazole as 42.9% and 57.1%, respectively. Remarkably, only 42.9% and 67.9% of the isolates were susceptible to itraconazole and voriconazole, respectively. **Conclusions.** Understanding local susceptibility patterns, especially those of non-*C. albicans* *Candida* species, can significantly aid in the selection of an effective antifungal agent. The *in vivo* response of *C. krusei* vaginitis to various antifungal therapeutics remains unknown and requires further research.

1. Introduction

Vulvovaginal candidiasis (VVC) is a common illness attributed to an overgrowth of *Candida* species, and it is estimated that 75% of all women will experience an episode of VVC in their lifetimes. *C. albicans* accounts for 80–95% of all episodes of VVC worldwide [1, 2]. The prevalence of VVC due to non-*C. albicans* *Candida* species previously ranged from 5 to 20%; however, the number of reported cases has increased sharply over the last two decades, particularly for cases of *C. glabrata* [3, 4]. Therefore, the possibility of antifungal resistant strains of non-*C. albicans* *Candida* species in *Candida* vaginitis should be considered in clinics. The emergence of resistance may be attributed to the following factors (i) the widespread use of over-the-counter (OTC) medications; (ii) long-term use of suppressive azoles; and (iii) the frequent use

of courses of antifungal medications [1, 3] or (iv) the increase use of vaginal cultures for reliable diagnoses [2, 5]. There is no evidence to suggest the followings: (i) certain women may be more susceptible to infection by particular *Candida* species over other species, or (ii) there are epidemiologic factors that may predispose women to acute VVC (AVVC) versus recurrent VVC (RVVC) [1, 2].

VVC is also, albeit infrequently, caused by *C. parapsilosis*, *C. tropicalis*, and *C. krusei* [1, 6, 7]. The decreased susceptibility of bloodstream *C. krusei* isolates to amphotericin B and 5-flucytosine as determined using the broth microdilution method is well documented [8]. However, *in vitro* susceptibility testing has not been used to evaluate the clinical response of *C. krusei* vaginitis [9]. In addition, little is known about vaginal *C. krusei* infections because they are relatively rare. However, *C. krusei* is known to

be inherently resistant to one of the most commonly used antifungal drugs, fluconazole. The signs and symptoms of *C. krusei* vaginitis appear to be indistinguishable from the signs and symptoms of VVC cases caused by other *Candida* species [6, 10]. Although rare, *C. krusei* is an intractable cause of RVVC. Furthermore, most institutions have had limited experience with *C. krusei* vaginitis [6]. Thus, the present study aims to fill this gap in the literature. Here, we retrospectively analyzed the epidemiological characteristics of 28 vaginal *C. krusei* isolates, including host and risk factors. In addition, we investigated the antifungal susceptibility profiles of these isolates to 10 antifungal drugs to determine the most appropriate therapeutic choice(s) in women with *C. krusei* vaginitis.

2. Materials and Methods

2.1. Vaginal *C. krusei* Isolates. We examined 1,543 vaginal samples from unrelated women, of which 560 (36.3%) were culture-positive and 983 (63.7%) were culture-negative for *Candida* yeasts, and the medical records of these cases were reviewed. Among the 560 vaginal yeast isolates, *C. albicans* was the most common species and identified in 242 (43.2%) isolates, followed by *C. glabrata* in 155 (27.7%), *C. krusei* in 28 (5.0%), *C. kefyr* in 20 (3.6%), and in 115 (20.5%) representing several species of *Candida*. Women who had *C. krusei* in their vagina were included in the study. The definitions of the clinical presentations of VVC for each group were as follows: AVVC (group 1), currently asymptomatic women with initial or sporadic episodes of symptomatic vaginitis, that is, occurring fewer than four times per year ($n = 8$); RVVC (group 2), symptomatic patients with a history of four or more clinical episodes of VVC per year ($n = 13$); and controls (group 3), women who incidentally carried a normal level of *C. krusei* in their vaginal culture without vaginitis, who were completely asymptomatic and had no history of RVVC ($n = 7$). The control group included a mixed group of asymptomatic women who had no history of RVVC and women who had positive cultures. All participants took part in a short interview, which included questions regarding lifestyle and medical, gynecological, and sexual history. This study was reviewed and approved by the Institutional Review Board at the University of Çukurova, Adana, Turkey. The Declaration of Helsinki protocols were followed, and the patients provided written informed consent.

2.2. Identification of *C. krusei*. The *C. krusei* isolates were recovered on CHROMagar *Candida* (Becton Dickinson, Heidelberg, Germany) and appeared as dull, flat, light mauve to mauve, and colonies with a whitish border. The criteria for the identification of *C. krusei* were the absence of germ tube production in human serum at 37°C at 2 hours, the production of abundant pseudohyphae with some moderate branching on cornmeal-Tween 80 agar (Difco, Detroit, MI, USA), and weak or absent urease activity. These isolates were verified by their assimilation patterns using the API 20C AUX method (bioMérieux, Marcy l'Étoile, France) [11]. *C. krusei* ATCC 6258 was used as a positive control.

2.3. Antifungal Susceptibility Testing. Antifungal testing was conducted at the Department of Microbiology, Faculty of Medicine, Gazi University, Ankara, using a broth microdilution method and according to the guidelines of the M27-A3 document of the Clinical and Laboratory Standards Institute (CLSI). Before testing, each isolate was subcultured on Sabouraud glucose agar (SGA; Merck, Darmstadt, Germany) to ensure purity and viability. The interpretation of antifungal susceptibility was guided by criteria derived from the CLSI's M27-A3 protocol [12]. The following antifungal agents were tested: amphotericin B (0.03–16 µg/mL), 5-flucytosine (0.06–64 µg/mL), caspofungin (0.03–16 µg/mL), fluconazole (0.12–128 µg/mL), itraconazole (0.03–16 µg/mL), voriconazole (0.008–16 µg/mL), econazole (0.007–8 µg/mL), ketoconazole (0.007–8 µg/mL), miconazole (0.007–8 µg/mL), and sulconazole (0.03–16 µg/mL).

The minimal inhibitory concentrations (MICs) were determined for each antifungal agent and used to classify the susceptibility of the isolates as follows: (i) amphotericin B, MIC ≤ 1 (µg/mL), susceptible (S); (ii) 5-flucytosine, MIC ≤ 4 (µg/mL) S, MIC between 8 and 16 (µg/mL) intermediate (I), MIC ≥ 32 (µg/mL) resistant (R); (iii) caspofungin, MIC ≥ 2 (µg/mL) R; (iv) fluconazole, MIC ≤ 8 (µg/mL) S, MIC between 16 and 32 (µg/mL) susceptible dose dependent (S-DD), MIC ≥ 64 (µg/mL) R; (v) itraconazole, MIC ≤ 0.125 (µg/mL) S, MIC between 0.25 and 0.5 (µg/mL) S-DD, MIC ≥ 1 (µg/mL) R; (vi) voriconazole, ≤ 1 (µg/mL) S, MIC = 2 (µg/mL) S-DD, MIC ≥ 4 (µg/mL) R; (vii) ketoconazole, MIC ≥ 16 (µg/mL) R; and (viii) miconazole, MIC ≥ 4 (µg/mL) R [12]. Currently, there are no published criteria for defining econazole and sulconazole susceptibility [13]. These results were expressed in terms of the MIC range and the MIC₅₀, and MIC₉₀ values for each antifungal agent. All *C. krusei* isolates were declared resistant to fluconazole. *C. krusei* ATCC 6258 and *C. parapsilosis* ATCC 22019 were used as controls, as recommended by the CLSI [12, 14].

2.4. Statistical Analysis. Data were analyzed using IBM SPSS version 19. Continuous variables, such as age and body mass index, were first divided into bins: <30, 30–39, 40–49, and >50 years of age and <25 (under or normal weight) and >25 (overweight or obese) for body mass index. Then, all categorical variables were cross classified by *C. krusei* infection or carrier status to descriptively summarize the association between the variables using the chi-squared test and to measure the degree of association using the odds ratio with a 95% confidence interval. For the multivariate analysis, logistic regression modeling of the binary data (*C. krusei* infected, carrier, or neither) was used to determine the significant predictors of *C. krusei* infection, after adjusting for other factors in the models. Factors with significance levels <0.30 were entered into the multivariate logistic model to determine the significant effect of each factor simultaneously on the prediction of *C. krusei* infection. None of the other factors contributed significantly to the prediction of *C. krusei* infection.

TABLE 1: Basic demographic characteristics of women with vaginal complaints in this study.

| Variables | <i>C. krusei</i> n (%) | | | P |
|---------------------------|------------------------|--------------|-------------|-------------|
| | Present | Absent | Total | |
| Education status | | | | |
| Illiterate | 4 (1.7) | 226 (98.3) | 230 | 0.96 |
| Primary school | 14 (1.9) | 709 (98.1) | 723 | |
| Secondary school | 2 (1.7) | 119 (98.3) | 121 | |
| High school | 5 (1.5) | 338 (98.5) | 343 | |
| College | 3 (2.4) | 122 (97.6) | 125 | |
| Marital status | | | | |
| Single | 0 (0.0) | 17 (100.0) | 17 | 0.77 |
| Married | 26 (1.8) | 1,418 (98.2) | 1,444 | |
| Widowed-Divorced | 2 (2.5) | 79 (97.5) | 81 | |
| Tobacco use | | | | |
| Yes | 6 (1.8) | 328 (98.2) | 334 | 0.6 |
| No | 22 (1.8) | 1,185 (98.2) | 1,207 | |
| Alcohol use | | | | |
| Yes | 0 (0.0) | 35 (100.0) | 35 | 0.52 |
| No | 28 (1.9) | 1,479 (98.1) | 1,507 | |
| Mean ± Standard Deviation | | | | |
| Age | 40.3 ± 10.4 | 35.3 ± 10.8 | 35.4 ± 10.8 | 0.01 |
| Gravida | 3.4 ± 1.9 | 2.9 ± 2.1 | 2.9 ± 2.0 | 0.2 |

3. Results

C. krusei isolates were recovered from non pregnant patients without diabetes mellitus ($n = 9$), pregnant patients ($n = 6$), diabetes mellitus patients ($n = 6$), and contraceptive user's ($n = 7$) with no previous history of immunodeficiency who visited the Faculty of Medicine Department of Obstetrics and Gynecology at Çukurova University from 2009 until 2012. Of the *C. krusei* isolates, 24 (85.7%) were the only species on plates and four (14.3%) were part of mixed cultures, which were always included by *C. albicans*. In our group, 24 (85.7%) women were in premenopausal and four (14.3%) in postmenopausal period who had also exposed hormone replacement therapy. The mean age of the women was 40.3 ± 10.4 years (range, 21 to 59 years old).

The basic demographic and clinical characteristics of women with vaginal *C. krusei* isolates are presented in Tables 1 and 2. Perineal laceration is significantly higher ($P = 0.006$) in the *C. krusei* group compared with the non-*C. krusei* group (Table 2). As revealed by multivariate analysis, existence of perineal laceration ($P = 0.009$) and an age of over 50 years ($P = 0.02$) were significant predictors of *C. krusei* vaginitis or carrier status (Table 3).

The MIC results for the control strains of *C. krusei* ATCC 6258 and *C. parapsilosis* ATCC 22019 were within the acceptable range. All the *C. krusei* isolates were susceptible to amphotericin B, caspofungin, ketoconazole, and miconazole, and 10 of the 28 isolates (35.7%) were defined as S-DD for 5-flucytosine. High MIC rates were observed for fluconazole, of which 42.9% of the isolates were S-DD and 57.1% were

R. Remarkably, only 42.9% and 67.9% of the isolates were susceptible to itraconazole (six S-DD and 10 R) and voriconazole (four S-DD and five R), respectively. We also observed low MIC levels for econazole and sulconazole. The antifungal susceptibilities of *C. krusei* isolated from patients with AVVC and RVVC did not differ significantly from those isolated from the group of women without symptoms of vaginitis ($P > 0.05$; Table 4).

4. Discussion

To the best of our knowledge, this study is the largest series to exclusively investigate the prevalence of, host and risk factors for, and antifungal susceptibility of the minor isolate *C. krusei*. We also briefly summarized the baseline and demographic characteristics of women who had *C. krusei* present in their vaginal samples (Tables 1 and 2). Our data suggest that the prevalence of *C. krusei* is relatively high (5.0%) in this population, displayed no specific host preferences, and was most often associated with RVVC. Perineal laceration and increased age (>50 years) were significant predictors of *C. krusei* vaginitis (Tables 2 and 3). An important limitation of the present study is the lack of data regarding *in vivo* therapeutic drug choices and outcomes in *C. krusei* vaginitis. In addition, the number of women with *C. krusei* is very small, so, for some of the other factors we examined, the study may not have had sufficient power to detect a difference (Tables 1–3).

The presence of mixed cultures may affect the choice of treatment strategy. In our previous study, using chromogenic

TABLE 2: Basic clinical characteristics of women with vaginal complaints in the study.

| Variables | <i>C. krusei</i> n (%) | | Total | <i>P</i> |
|---|------------------------|--------------|-------|--------------|
| | Present | Absent | | |
| Diabetes mellitus | | | | |
| Absent | 21 (1.7) | 1,250 (98.3) | 1,271 | 0.21 |
| Present | 7 (2.6) | 265 (97.4) | 272 | |
| Hypothyroidism | | | | |
| Absent | 26 (1.8) | 1,452 (98.2) | 1,478 | 0.31 |
| Present | 2 (3.2) | 60 (96.8) | 62 | |
| Hyperthyroidism | | | | |
| Absent | 26 (1.7) | 1,478 (98.3) | 1,504 | 0.15 |
| Present | 2 (5.3) | 36 (94.7) | 38 | |
| Other chronic diseases | | | | |
| Absent | 19 (1.7) | 1,127 (98.3) | 1,146 | 0.28 |
| Present | 9 (2.3) | 387 (97.7) | 396 | |
| Medication other than antibiotics | | | | |
| Absent | 22 (2.0) | 1,056 (98.0) | 1,078 | 0.22 |
| Present | 6 (1.3) | 458 (98.7) | 464 | |
| Use of local steroid in the last 4 weeks | | | | |
| Absent | 27 (1.8) | 1,492 (98.2) | 1,519 | 0.35 |
| Present | 1 (4.3) | 22 (95.7) | 23 | |
| Use of systemic steroid in the last 4 weeks | | | | |
| Absent | 28 (1.9) | 1,482 (98.1) | 1,510 | 0.57 |
| Present | 0 (0.0) | 30 (100.0) | 30 | |
| Perineal laceration | | | | |
| Absent | 15 (1.3) | 1,163 (98.7) | 1,178 | 0.006 |
| Present | 13 (3.6) | 351 (96.4) | 364 | |
| Contraception | | | | |
| None | 9 (1.4) | 637 (98.6) | 646 | 0.49 |
| OC | 1 (1.0) | 103 (99.0) | 104 | |
| IUD | 8 (2.9) | 268 (97.1) | 276 | |
| Condom | 4 (1.5) | 258 (98.5) | 262 | |
| Others | 6 (2.4) | 248 (97.6) | 254 | |
| Personal allergic history | | | | |
| Absent | 24 (1.8) | 1,316 (98.2) | 1,340 | 0.51 |
| Present | 4 (2.0) | 197 (98.0) | 201 | |
| History of sexual intercourse in the last 4 weeks | | | | |
| Present | 8 (2.8) | 279 (97.2) | 287 | 0.13 |
| Absent | 20 (1.6) | 1,235 (98.4) | 1,255 | |
| Antibiotic use in the last 4 weeks | | | | |
| Absent | 27 (2.1) | 1,260 (97.9) | 1,287 | 0.07 |
| Present | 1 (0.4) | 254 (99.6) | 255 | |
| Body mass index | | | | |
| ≤19 | 1 (2.3) | 42 (97.7) | 43 | 0.4 |
| 19–24 | 4 (1.1) | 369 (98.9) | 373 | |
| 24–29 | 9 (1.6) | 561 (98.4) | 570 | |
| >29 | 14 (2.5) | 540 (97.5) | 554 | |

OC: oral contraceptive; IUD: intrauterine device.

TABLE 3: Analysis of predictive factors for *Candida krusei* infection using univariate and multivariate logistic analyses.

| Predictores | Univariate analysis | | | Multivariate analysis | | |
|-----------------------------------|---------------------|--------------|-----------|-----------------------|--------------|-----------|
| | OR | P | %95 CI | OR | P | %95 CI |
| Age | | | | | | |
| <30 | 1.0 | — | — | — | — | — |
| 30–39 | 2.56 | 0.09 | 0.87–7.55 | 2.4 | 0.12 | 0.8–7.16 |
| 40–49 | 2.48 | 0.14 | 0.75–8 | 2.7 | 0.11 | 0.79–9.3 |
| ≥50 | 5.39 | 0.004 | 1.69–17.2 | 7.9 | 0.02 | 1.34–46.7 |
| Body mass index | 1.72 | 0.19 | 0.65–4.57 | 1.28 | 0.63 | 0.46–3.57 |
| Diabetes mellitus | 1.57 | 0.21 | 0.66–3.74 | 0.26 | 0.11 | 0.05–1.35 |
| Hyperthyroidism | 3.2 | 0.15 | 0.72–13.8 | 3.92 | 0.08 | 0.84–18.3 |
| Medication other than antibiotics | 0.63 | 0.22 | 0.25–1.56 | 0.56 | 0.23 | 0.21–1.46 |
| Use of local steroid | 2.5 | 0.35 | 0.32–19.1 | 3.11 | 0.3 | 0.38–26.2 |
| Perineal laceration | 2.87 | 0.06 | 1.25–6.1 | 3.25 | 0.009 | 1.34–7.87 |
| Antibiotics (last 4-weeks) | 0.18 | 0.07 | 0.025–1.3 | 0.19 | 0.11 | 0.03–1.48 |

media, we determined that the percentage of mixed cultures recovered from vaginal samples was as high as 14.1% in Adana, Turkey [4]. The results of this investigation (14.3%) are similar to those of our earlier study. In addition, our finding that older women (mean age, 40.3 years) are more susceptible to infection corroborates the earlier finding of Singh et al. [6] (mean age, 44 years). However, the women studied by Singh et al. [6] all had RVVC, whereas our study included not only RVVC cases but also AVVC and controls. These authors noted that *C. krusei* isolates were highly resistant to fluconazole ($MIC_{90} > 64 \mu\text{g/mL}$), consistent with our findings. In addition, these authors reported resistance to miconazole ($MIC_{90} > 4 \mu\text{g/mL}$), one of the most commonly used OTC antifungal agents, which we did not observe. However, in line with our results, clotrimazole was observed to be the most active topical imidazole against *C. krusei*. Moreover and more importantly, amphotericin B, caspofungin, itraconazole, and voriconazole were demonstrated to have favorable antifungal activity, although, in our study, several strains exhibited obvious high resistance to the latter two drugs. Of note, the authors suggested that the therapy should continue for 2–6 weeks, regardless of the agent used [6]. On the other hand, a recent study reported that fluconazole-resistant *C. albicans* appears to be emerging in clinics [15]. Therefore, antifungal susceptibility testing may assist in selecting the appropriate therapeutic drug not only for non-*C. albicans* *Candida* vaginitis but also for rare fluconazole-resistant *C. albicans* vaginitis.

In this investigation, amphotericin B, caspofungin, ketoconazole, and miconazole were observed to be active against all *C. krusei* isolates (Table 4). In contrast to the findings of Singh et al. [6], but in line with those of Richter et al. [16], itraconazole exhibited high S-DD and R rates, 35.7% and 21.4%, respectively. Although the new broad-spectrum oral antifungal voriconazole is rarely used in patients with VVC, we observed that 67.9% of the isolates were susceptible to

voriconazole. Pfaller et al. [8] reported a higher rate, stating that 81.5% of 426 genital *C. krusei* isolates were susceptible to voriconazole using the CLSI M44-A disk diffusion method. In contrast to our findings, Lyon et al. [17] reported that fluconazole resistance rates were highly predictive of resistance to voriconazole. Although specific clinical cutoff points have not yet been assigned for econazole and sulconazole susceptibility, we observed low MIC values for both drugs. Nystatin suppositories and boric acid could be therapies of choice for *C. krusei* vaginitis [6].

This study is the largest to date to investigate the antifungal drug-resistance profile of *C. krusei* vaginal isolates and the epidemiologic risk factors of infection. In this investigation, perineal laceration and increasing age (>50 years) were important predictive factors for *C. krusei* vaginitis or carrier status (Table 3). This study also revealed that the topical imidazoles (ketoconazole and miconazole), which can be prescribed safely in routine practice, were effective against all *C. krusei* isolates. In addition, the vaginal *C. krusei* isolates were less susceptible to itraconazole (42.9%) and voriconazole (67.9%) than to other antifungal therapeutics. These findings may have implications for the *in vivo* therapeutic treatment of *C. krusei* vaginitis (Table 4). Thus, the identification of *C. krusei* in vaginal samples and *in vitro* antifungal testing will assist in the selection of appropriate antifungal agents and therapy duration. Future clinical trials to determine the *in vivo* efficacy of the current drugs for women with *C. krusei* vaginitis are required.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

TABLE 4: Antifungal susceptibility of 28 vaginal *Candida krusei* isolates stratified according to clinical forms.

| Antifungals | Acute VVC (<i>n</i> = 7) | Recurrent VVC (<i>n</i> = 13) | Control (<i>n</i> = 8) |
|--|---------------------------|--------------------------------|-------------------------|
| Amphotericin B ($\mu\text{g/mL}$) | | | |
| MIC range | 0.03–0.25 | 0.03–0.5 | 0.06–0.5 |
| MIC ₅₀ | 0.03 | 0.25 | 0.25 |
| MIC ₉₀ | 0.25 | 0.25 | 0.25 |
| <i>R</i> , $\geq 2 \mu\text{g/mL}$ | | | |
| <i>n</i> (%) | — | — | — |
| 5-Flucytosine ($\mu\text{g/mL}$) | | | |
| MIC range | 0.125–8 | 0.125–8 | 0.125–16 |
| MIC ₅₀ | 2 | 2 | 8 |
| MIC ₉₀ | 8 | 8 | 16 |
| <i>R</i> , $\geq 32 \mu\text{g/mL}$ | | | |
| <i>n</i> (%) | — | — | — |
| Caspofungin ($\mu\text{g/mL}$) | | | |
| MIC range | 0.03–0.06 | 0.03–0.06 | 0.03–0.06 |
| MIC ₅₀ | 0.03 | 0.03 | 0.03 |
| MIC ₉₀ | 0.06 | 0.03 | 0.06 |
| <i>R</i> , $\geq 2 \mu\text{g/mL}$ | | | |
| <i>n</i> (%) | — | — | — |
| Fluconazole ($\mu\text{g/mL}$)[#] | | | |
| MIC range | 16–128 | 16–128 | 16–128 |
| MIC ₅₀ | 128 | 64 | 32 |
| MIC ₉₀ | 128 | 128 | 128 |
| <i>R</i> , $\geq 64 \mu\text{g/mL}$ | | | |
| <i>n</i> (%) | 4 | 9 | 3 |
| S-DD, 16–32 $\mu\text{g/mL}$ | | | |
| <i>n</i> (%) | 3 | 4 | 5 |
| Itraconazole ($\mu\text{g/mL}$) | | | |
| MIC range | 0.125–16 | 0.125–16 | 0.125–8 |
| MIC ₅₀ | 0.25 | 0.25 | 0.125 |
| MIC ₉₀ | 4 | 4 | 2 |
| <i>R</i> , $\geq 1 \mu\text{g/mL}$ | | | |
| <i>n</i> (%) | 3 | 4 | 3 |
| Voriconazole ($\mu\text{g/mL}$) | | | |
| MIC range | 0.125–8 | 0.125–16 | 0.25–8 |
| MIC ₅₀ | 1 | 0.5 | 0.5 |
| MIC ₉₀ | 4 | 4 | 2 |
| <i>R</i> , $\geq 4 \mu\text{g/mL}$ | | | |
| <i>n</i> (%) | 2 | 2 | 1 |
| Ketoconazole ($\mu\text{g/mL}$) | | | |
| MIC range | 0.125–8 | 0.25–8 | 0.25–8 |
| MIC ₅₀ | 0.25 | 4 | 0.25 |
| MIC ₉₀ | 2 | 8 | 8 |
| <i>R</i> , $\geq 16 \mu\text{g/mL}$ | | | |
| <i>n</i> (%) | — | — | — |
| Econazole ($\mu\text{g/mL}$) | | | |
| MIC range | 0.5–1 | 0.125–2 | 0.5–1 |
| MIC ₅₀ | 1 | 1 | 1 |
| MIC ₉₀ | 1 | 2 | 1 |
| <i>R</i> , ND | | | |
| <i>n</i> (%) | — | — | — |

TABLE 4: Continued.

| Antifungals | Acute VVC (n = 7) | Recurrent VVC (n = 13) | Control (n = 8) |
|----------------------------------|-------------------|------------------------|-----------------|
| Miconazole ($\mu\text{g/mL}$) | | | |
| MIC range | 0.25–0.5 | 0.06–1 | 0.25–0.5 |
| MIC ₅₀ | 0.25 | 0.25 | 0.25 |
| MIC ₉₀ | 0.5 | 1 | 0.5 |
| R, $\geq 4 \mu\text{g/mL}$ | | | |
| n (%) | — | — | — |
| Sulconazole ($\mu\text{g/mL}$) | | | |
| MIC range | 1–4 | 0.06–4 | 1–2 |
| MIC ₅₀ | 2 | 1 | 1 |
| MIC ₉₀ | 4 | 4 | 2 |
| R, ND | | | |
| n (%) | — | — | — |

VVC: vulvovaginal candidiasis; R: resistance; S-DD: susceptible dose dependent; ND: not determined. * All isolates were declared resistant to fluconazole.

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References

- [1] J. D. Sobel, "Vulvovaginal candidosis," *The Lancet*, vol. 369, no. 9577, pp. 1961–1971, 2007.
- [2] M. Ilkit and A. B. Guzel, "The epidemiology, pathogenesis, and diagnosis of vulvovaginal candidosis: a mycological perspective," *Critical Reviews in Microbiology*, vol. 37, no. 3, pp. 250–261, 2011.
- [3] F. Parazzini, E. Di Cintio, V. Chiantera, and S. Guaschino, "Determinants of different *Candida* species infections of the genital tract in women," *European Journal of Obstetrics Gynecology & Reproductive Biology*, vol. 93, no. 2, pp. 141–145, 2000.
- [4] A. B. Guzel, M. Ilkit, T. Akar, R. Burgut, and S. C. Demir, "Evaluation of risk factors in patients with vulvovaginal candidiasis and the value of chromID *Candida* agar versus CHROMagar *Candida* for recovery and presumptive identification of vaginal yeast species," *Medical Mycology*, vol. 49, no. 1, pp. 16–25, 2010.
- [5] P. Nyirjesy, S. M. Seeney, M. H. T. Grody, C. A. Jordan, and H. R. Buckley, "Chronic fungal vaginitis: the value of cultures," *American Journal of Obstetrics and Gynecology*, vol. 173, no. 3, pp. 820–823, 1995.
- [6] S. Singh, J. D. Sobel, P. Bhargava, D. Boikov, and J. A. Vazquez, "Vaginitis due to *Candida krusei*: epidemiology, clinical aspects, and therapy," *Clinical Infectious Diseases*, vol. 35, no. 9, pp. 1066–1070, 2002.
- [7] P. Nyirjesy, A. B. Alexander, and M. V. Weitz, "Vaginal *Candida parapsilosis*: pathogen or bystander?" *Infectious Disease in Obstetrics and Gynecology*, vol. 13, no. 1, pp. 37–41, 2005.
- [8] M. A. Pfaller, D. J. Diekema, D. L. Gibbs et al., "*Candida krusei*, a multidrug-resistant opportunistic fungal pathogen: geographic and temporal trends from the ARTEMIS DISK Antifungal Surveillance Program, 2001 to 2005," *Journal of Clinical Microbiology*, vol. 46, no. 2, pp. 515–521, 2008.
- [9] J. D. Sobel, M. Zervos, B. D. Reed et al., "Fluconazole susceptibility of vaginal isolates obtained from women with complicated *Candida* vaginitis: clinical implications," *Antimicrobial Agents and Chemotherapy*, vol. 47, no. 1, pp. 34–38, 2003.
- [10] L. Agatensi, F. Franchi, V. Mondello et al., "Vaginopathic and proteolytic *Candida* species in outpatients attending a gynaecology clinic," *Journal of Clinical Pathology*, vol. 44, no. 10, pp. 826–830, 1991.
- [11] H. R. Buckley, "Identification of yeasts," in *Medical Mycology: A Practical Approach*, E. G. V. Evans and M. D. Richardson, Eds., pp. 97–109, IRL Press, Oxford, UK, 1989.
- [12] Clinical Laboratory Standards Institution, *Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts. Approved Standard CLSI Document M27-A3*, Wayne, Pa, USA, 2008.
- [13] A. Kalkanci, A. B. Güzel, I. I. J. Khalil, M. Aydin, M. Ilkit, and S. Kustimur, "Yeast vaginitis during pregnancy: susceptibility testing of 13 antifungal drugs and boric acid and the detection of four virulence factors," *Medical Mycology*, vol. 50, no. 6, pp. 585–593, 2012.
- [14] M. A. Pfaller, D. Andes, D. J. Diekema, A. Espinel-Ingroff, and D. Sheehan, "Wild-type MIC distributions, epidemiological cutoff values and species-specific clinical breakpoints for fluconazole and *Candida*: time for harmonization of CLSI and EUCAST broth microdilution methods," *Drug Resistance Updates*, vol. 13, no. 6, pp. 180–195, 2010.
- [15] D. Marchaim, L. Lemanek, S. Bheemreddy, K. S. Kaye, and J. D. Sobel, "Fluconazole-resistant *Candida albicans* vulvovaginitis," *Obstetrics & Gynecology*, vol. 120, no. 6, pp. 1407–1414, 2012.
- [16] S. S. Richter, R. P. Galask, S. A. Messer, R. J. Hollis, D. J. Diekema, and M. A. Pfaller, "Antifungal susceptibilities of *Candida* species causing vulvovaginitis and epidemiology of recurrent cases," *Journal of Clinical Microbiology*, vol. 43, no. 5, pp. 2155–2162, 2005.
- [17] G. M. Lyon, S. Karatela, S. Sunay, and Y. Adiri, "Antifungal susceptibility testing of *Candida* isolates from the *Candida* surveillance study," *Journal of Clinical Microbiology*, vol. 48, no. 4, pp. 1270–1275, 2010.