

REVIEW

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MicroRNA: a prognostic biomarker and a possible druggable target for circumventing multidrug resistance in cancer chemotherapy

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Abstract

Multidrug resistance (MDR) is a major obstacle to successful cancer treatment. It is often associated with an increased efflux of a variety of structurally unrelated anticancer drugs by ATP-binding cassette (ABC) transporters including P-gp, ABCG2 and MRP1. MicroRNAs (miRNAs) are small non-coding RNAs that govern posttranscriptional regulation of target genes by interacting with specific sequences in their 3' untranslated region (3'UTR), thereby promoting mRNA degradation or suppressing translation. Accumulating evidence suggests that alterations in miRNAs contribute to resistance to anticancer drugs. While miRNAs are well-known to be dysregulated in cancer, recent literature revealed that miRNA levels in biological samples may be correlated with chemotherapy response. This review summarized the coordinated network by which miRNA regulated MDR transporters. The usefulness of miRNAs as prognostic biomarkers for predicting chemotherapeutic outcome is discussed. MiRNAs may also represent druggable targets for circumvention of MDR.

Keywords: ATP-binding cassette transporters, microRNA, Multidrug resistance, Non-coding RNAs, Prognostic biomarkers, 3' untranslated region

Introduction

Resistance to anticancer drugs remains a major unresolved obstacle to successful chemotherapy. It has been estimated that most cancer deaths, if not all, are caused by chemotherapy failure because tumors quickly develop resistance after exposure to drugs [1]. In order to develop novel strategies to combat cancer drug resistance and to improve patient survival, a thorough understanding of its mechanisms is therefore badly needed. The causes of cancer drug resistance are multifactorial, including decreased accumulation/increased disposition of anticancer drugs, mutation of drug targets, enhanced cell repair and altered cell death pathways. However, the most common and extensively studied mechanism is the overexpression of the energy-dependent ATP-binding cassette (ABC) drug efflux transporters such as P-glycoprotein (MDR-1/P-gp/ABCB1), multidrug

resistance related protein (MRP-1/ABCC1), and breast cancer resistance protein (BCRP/MXR/ABCP/ABCG2) [2]. It is associated with an increased efflux of cytotoxic drugs, causing multidrug resistance (MDR) because cytotoxic drugs from different chemical structures are affected simultaneously.

MicroRNAs (miRNAs) are short endogenous non-coding RNAs that repress gene expression in a variety of eukaryotic organisms. Gene regulation by miRNAs is mediated by the formation of imperfect hybrids with the 3' untranslated region (3'UTR) sequences of the target mRNAs, leading to mRNA degradation and/or translational inhibition [3]. They play important roles in several cellular processes, such as proliferation, differentiation, apoptosis, and development, by simultaneously controlling the expression level of hundreds of genes. MiRNAs are predicted to regulate the expression of up to one third of human protein-coding genes [4-6].

Numerous recent studies have shown that miRNA expression profiles differ between normal tissues and cancerous cells derived from the same organ, and also

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between cancer types [7]. MiRNAs can act as oncogenes or tumor suppressors, contributing to different pathways in tumorigenesis [8,9]. They may be used for diagnostic and prognostic purposes and they also constitute novel targets for cancer treatment [10,11]. Recently, the evidence for the roles of miRNAs in determining drug sensitivity/resistance has been emerging. This review summarized the current understanding about the role of miRNAs in mediating cancer drug resistance. More emphasis is placed on miRNA-related regulation of the MDR transporters, though other mechanisms causing drug resistance not related to transporters will also be discussed. The possible application of miRNA-transporters regulatory network for predicting chemotherapeutic response will be highlighted. Novel strategies aiming to target miRNA-related pathways for the circumvention of multi-drug resistance will also be elaborated.

Review

Aberrant expression of miRNAs and cancer drug resistance

Evidence pointing to the role of miRNAs in determining drug sensitivity and MDR is emerging. MiRNA expression is largely dysregulated in drug-resistant cancer cells [12,13]. In a recent study on a doxorubicin-resistant breast cancer cell line MCF-7/DOX, a profound dysregulation of the miRNA profile and altered expression of two important miRNA processing enzymes Dicer and Argonaute 2 was reported [14]. The remarkable correlation between specific miRNA expression and the corresponding changes in protein levels of their specific targets having well-documented role in cancer drug resistance, may thus implies a mechanistic link between miRNAome dysregulation and the MDR phenotype. Moreover, miRNA expression patterns in the NCI-60 drug screen cell lines are significantly correlated to the sensitivity patterns of the cancer cells for a variety of anticancer drugs [15]. Furthermore, numerous miRNAs have been found to regulate drug resistance genes such as *ABCG2* [16-18], *BCL2* [19], *DHFR* [20], *MDR1* [14] and *PTEN* [21]. Importantly, modulation of miRNA expression or function can alter sensitivity of cancer cells to anticancer drugs. This could be achieved by inhibiting the function of up-regulated miRNAs or restoring the expression of down-regulated miRNAs. Together, miRNAs may represent key players in both intrinsic and acquired MDR in cancer cells [22].

The cause of cancer drug resistance is multifactorial. The role of miRNAs in mediating cancer drug resistance is separately discussed below according to whether they are regulating to (I) MDR transporters-mediated (Table 1); or (II) non-MDR transporters-mediated mechanisms (Table 2). The list is by no means exhaustive. We aim to illustrate the representative ones with potentially wider implications.

Regulation of ABC transporters-mediated MDR by miRNAs

Direct regulation by miRNAs

ABCG2 (also known as BCRP/MXR/ABCP)

ABCG2 is the first MDR transporter reported to be regulated by miRNA-mediated mechanism. It is one of the major ABC transporters contributing to the MDR phenotype. Overexpression of the *ABCG2* gene is frequently observed in cancer cell lines selected with chemotherapeutic drugs [2,45]. To date, most studies examining the regulation of *ABCG2* have focused on transcription. Gene amplification, chromosome translocation, and the use of alternative 5' promoters due to differential expression of splice variants at the 5'-untranslated region (5'UTR) of *ABCG2* mRNA have been reported to play important roles in the increased expression of *ABCG2* [46,47]. In contrast, the understanding about posttranscriptional regulation of *ABCG2* has just started to evolve.

To date, a few miRNAs (miR-520 h [36,37], -519c [16,17], -328 [18], -181a [38], & -487a [39]) have been identified by different research groups independently to regulate *ABCG2* expression by interacting directly with *ABCG2* 3'UTR and to determine the sensitivity of cancer cells to chemotherapeutic drugs (Table 1). Consistent with the hypothesis that aberrant miRNA expression can cause cancer drug resistance, low miR-328 expression was found to correlate with the overexpression of *ABCG2* in resistant MCF-7/MX100 breast cancer cells [18]. In a human retinoblastoma cell line model, it has been further demonstrated that low expressions of all three miRNAs (miR-328, -519c, & -520 h) correlate very well with high *ABCG2* expression, with concomitant expression of other stem cell markers including CD133 and ALDH1A1 [35]. On the other hand, miR-520 h has been reported to promote differentiation of hematopoietic stem cells by inhibiting *ABCG2* expression [36]. *ABCG2* has been suggested to be a survival factor for stem or cancer stem cells (CSCs) [48]. Thus these findings collectively support an important role played by miRNAs in maintaining high *ABCG2* level in CSCs, leading to drug resistance. It will be interesting to verify if the same phenomenon is also observed in tumor samples from patients not responding to cancer chemotherapy.

Drug-resistant cancer cells escape miR-519c-mediated *ABCG2* repression by shortening of *ABCG2* mRNA 3' UTR

An additional layer of complexity in miR-519c-mediated regulation of *ABCG2* has been proposed, which is associated with alternative cleavage and polyadenylation of the 3'UTR of *ABCG2* mRNA, to facilitate the drug resistance phenotype [17]. *ABCG2* mRNA was found to be more stable in drug-selected and *ABCG2*-overexpressing resistant cell lines than in their parental counterparts [16,17]. Given that the expression

Table 1 miRNAs reported to regulate MDR transporters to mediate chemoresistance

Transporter targets	MicroRNA dysregulation	Direct binding site(s) on target gene?	Prediction by miRNA database(s)	Identification method	Cancer type	Study in patient samples?	Reference
ABCB1	miR-27a↑	<i>Indirect regulation</i> -targets HIPK2 (a transcriptional corepressor that inhibits HIF-1α activity), thereby indirectly activating MDR-1/P-gp	TargetScan suggests that HIPK2 is a possible target for miR-27a	Serendipitous testing of miR-27a in the development of drug resistance in ovarian cancer cell lines	Ovarian	No	[23]
	miR-27a↑, miR-451↑	<i>Likely indirect regulation</i> (i.e. not via ABCB1 3'UTR) miR-27a & miR-451 mimics elevate ABCB1 mRNA	–	miRNA microarray profiling	Ovarian	No	[24]
	miR-122↓	NO attempt to verify binding of miR-122 on MDR-1 3'UTR	NOT performed	Serendipitous testing of downregulation of MDR related genes (MDR-1, MRP & GST-π) by miR-122	Hepatocellular carcinoma	No	[25]
	miR-296↑	<i>Likely indirect regulation</i> (i.e. not via ABCB1 3'UTR) - antagomir of miR-296 decrease ABCB1 promoter activity	–	miRNA microarray profiling	Esophageal squamous cell carcinoma	Yes	[26]
	miR-298↓ miR-1253↓	Confirmed by luciferase reporter assay / MDR-1 3'UTR miRNA site deletion / miRNA mimic / miRNA inhibitor	miRanda	miRNA microarray	Breast cancer	No	[27]
	let-7 g↓	<i>Indirect regulation</i> • targets IMP-1 (a RNA-binding protein) • IMP-1 is known to stabilize MDR1 mRNA • loss of let-7 g commonly observed in various cancers could therefore allow overexpression of IMP-1 and stability of MDR-1/P-gp to mediate drug resistance			Ovarian	Yes	[28]
ABCC1	miR-326↓	Confirmed by luciferase reporter assay / ABCC1 3'UTR miRNA site deletion / gene expression analysis after transfection with miR-326 mimic	TargetScan	miRNA microarray profiling	Breast cell line + early/advanced breast cancer tissue	Yes	[29]
	miR-1291↓	Confirmed by luciferase reporter/miRNA mimic/miRNA inhibitor	RNAhybrid, TargetScan, miRanda, PITA	Serendipitous study of the role of SNORA34 to generate miR-1291, which subsequently controls chemosensitivity	Pancreatic	No	[30]
ABCC2	miR-297↓	Luciferase reporter/miRNA mimic/miRNA inhibitor	TargetScan	miRNA microarray profiling	Colorectal	Yes	[31]
ABCC3 & ABCC6	miR-9*↓	<i>Indirect regulation</i> – • miR-9* is a SOX2-targeting miRNA	PicTar, TargetScan, miRBase, miRanda	Serendipitous study of the role of ID4 in chemoresistance of induced glioma stem cells	Glioma	Yes	[32]

Table 1 miRNAs reported to regulate MDR transporters to mediate chemoresistance (Continued)

		<ul style="list-style-type: none"> • novel ID4-miR-9*-SOX2-ABCC3/ABCC6 regulatory pathway • ID4 was found to confer chemoresistance to glioma stem cells by inducing the expression of two SOX2-mediated ABC transporters (ABCC3 & ABCC6) through suppression of miR-9* 					
ABCG2	miR-212↓, miR-328↓	Effect of miR-212 & miR-328 on ABCG2 expression was evaluated by gene expression analysis and luciferase reporter gene assay	–	miRNA microarray profiling in imatinib-selected K562 cells	Leukemia (short-term /long-term imatinib treatment)	No	[33]
	miR-328↓	Confirmed by luciferase reporter assay / ABCG2 3'UTR miRNA site deletion / miRNA mimic / miRNA antagomir	PITA; TargetScan	Bioinformatic analysis	Breast	No	[18]
	miR-328↓	Confirmed by luciferase reporter assay / gene expression analysis after transfection with miRNA mimic or inhibitor	–	miRNA microarray analysis on SP [#] vs non-SP [#] cells	Colorectal cell lines + primary biopsies	Yes	[34]
	miR-328↓, miR-519c↓, miR-520 h↓	Confirmed by luciferase reporter assay with site-directed gene mutagenesis / gene expression analysis after transfection with miRNA mimics	TargetScan, PITA, MicroCosm Targets, RNA22	Bioinformatic analysis	Breast, Stem-like cells from human retinoblastoma	No	[35]
	miR-519c (shortening of ABCG2 3'UTR escape miR-519c repression)	Confirmed by luciferase reporter assay with site-directed gene mutagenesis on ABCG2 3'UTR miRNA site / gene expression analysis after transfection with miRNA mimics or inhibitor	miRBase TARGETS, RNAHybrid, UTRScan program	Bioinformatic analysis	Colon	No	[16,17]
	miR-520 h↓	Confirmed by luciferase reporter assay +/- miRNA mimic & inhibitor	PicTar, miRanda, TargetScan	miRNA microarray profiling of CD34+ hematopoietic cells	Leukemia	No	[36]
	miR-520 h↓	Confirmed by gene expression analysis after miRNA mimic transfection	miRanda, TargetScan, TarBase	Bioinformatic analysis	Pancreatic	No	[37]
	miR-181a↓	Confirmed by luciferase reporter assay and gene expression analysis +/- miRNA mimic & inhibitor	RNAhybrid	miRNA microarray analysis to compare sensitive and resistant cells	Breast	No	[38]
	miR-487a↓	Confirmed by luciferase reporter assay and gene expression analysis +/- miRNA mimic & inhibitor Confirmed by luciferase reporter assay and gene expression analysis +/- miRNA mimic & inhibitor	TargetScan, PITA, RNAhybrid	Bioinformatic analysis	Breast	No	[39]

[#] SP = side population; Non-SP = non-side population.

Table 2 miRNAs reported to regulate other mediators of drug resistance

Target gene	Biological effect of target gene	MicroRNA	Validation of miRNA binding site	Prediction by miRNA database(s)	Identification method	Cancer type/Significance/Specific type of drug resistance	Study in patient samples?	Reference
<i>BAX</i>	Pro-apoptotic	miR-296↑	Luciferase reporter assay + miRNA antagomir	–	miRNA microarray profiling	Esophageal squamous cell carcinoma	Yes	[26]
<i>BCL2</i>	Anti-apoptotic	miR-15b↓ miR-16↓	Luciferase reporter assay + site-directed mutagenesis of Bcl-2 3'UTR + miRNA mimic	miRBase & TargetScan	miRNA microarray profiling	Gastric	No	[19]
<i>BCL2</i>	Anti-apoptotic	miR-1915↓	Luciferase reporter assay + site-directed mutagenesis of Bcl-2 3'UTR + miRNA mimics & inhibitors	miRDB; TargetScan (4 possible miRNA binding sites within Bcl-2 3'UTR)	miRNA microarray profiling	Colorectal	No	[40]
<i>BCL2 & SIRT1</i>	Anti-apoptotic	miR-34a↓	<i>Direct miRNA effect</i> (on Bcl-2 & SIRT1 mRNA): Luciferase reporter +/- miRNA mimic <i>Indirect miRNA effect</i> (through HuR): Gene expression analysis +/- miRNA mimic	TargetScan	Mechanistic investigation of miR-34a/Bcl-2 and miR-34a/SIRT1 pathways in taxane-based chemotherapy	Prostate / Paclitaxel resistance	No	[41]
<i>BMI-1</i>	PcG ³ protein – transcriptional repressor	miR-200c↓	Gene expression analysis after infection with miR-200c vector	–	miRNA microarray analysis	Melanoma /miR-200c is commonly found to be downregulated in malignant melanoma that possess self-renewal cancer stem-cell like property and are more invasive. The prominent miR-200c downregulation is accompanied by Bmi-1 overexpression, which was found to cause loss of E-cadherin (thereby EMT) and overexpression of MDR transporters (including ABCG2, ABCG5 and MDR).	Yes	[42]
<i>CDH1</i>	EMT ^b transition	miR-200c↓ miR-205↓	<i>Indirect regulation:</i> The two miRNAs repress ZEB1 & ZEB2, thereby leading to loss of E-cadherin and a EMT phenotype	–	Mechanistic investigation of the role of miRNAs in EMT-linked docetaxel resistance in prostate cancer	Prostate/Docetaxel resistance/Docetaxel treatment triggers EMT to inhibit apoptosis through the proposed miR-200c/205/ZEB1/ZEB2/E-cadherin pathway	No	[43]
<i>DHFR</i>	Folate metabolism	miR-24 (DHFR SNP 829C > T makes DHFR mRNA indifferent to miR-24)	Gene expression analysis + Site-directed mutagenesis + miRNA mimics & inhibitors	MiRanda & miRBase	A 829C > T SNP identified in <i>DHFR</i> gene in Japanese population was found to be associated with increased DHFR message	Fibrosarcoma/methotrexate resistance	No	[20]
<i>HIPK2</i>	Tumor suppressor	miR-27a↑	NO attempt to verify binding of miR-27a on HIPK2 3'UTR	TargetScan	Identified during the study of indirect effect of miR-27a on MDR-1 expression	Ovarian	No	[23]

Table 2 miRNAs reported to regulate other mediators of drug resistance (Continued)

<i>PTEN</i>	Tumor suppressor	miR-214↑	Luciferase reporter assay + site-directed mutagenesis of PTEN 3'UTR + miRNA mimic	–	miRNA microarray profiling	Ovarian/cisplatin resistance	Yes	[21]
<i>TUBB3</i>	Structural protein (β-tubulin)	miR-200c↓	Gene expression analysis +/- miRNA mimic	–	miRNA microarray profiling	Resistance to microtubule –binding chemotherapeutic drugs	No	[44]

^a PcG = polycomb group protein (transcriptional repressor).

^b EMT = epithelial-to-mesenchymal transition.

level of miR-519c in the parental and resistant cells does not differ too much in the cell line model studied, the increase in mRNA stability was subsequently tied to a missing miR-519c binding site in the truncated 3'UTR of ABCG2 mRNA in drug resistant cells [17]. MiR-519c cannot bind to ABCG2 mRNA in the resistant cells because of the shorter 3'UTR, and thus miRNA-mediated mRNA degradation and/or protein translation block are relieved, contributing to ABCG2 overexpression (Figure 1).

In eukaryotes, polyadenylation is an important process that produces mature mRNA for translation. A poly(A) tail is added to 3' end of mRNA at the end of transcription, which protects the mRNA molecule from enzymatic degradation in the cytoplasm, aids in transcription termination, export of the mRNA from nucleus, and translation [49]. Importantly, alternative polyadenylation can occur when several polyA signals lie in the last exon of a protein-coding gene, generating mRNA isoforms with different 3'UTR length. While the phenomenon of alternative cleavage and polyadenylation has been known for decades, it has only been fully appreciated recently [50]. The majority of human genes contain at least two polyA sites [51], and alternative cleavage and polyadenylation has been shown to be fairly common in multiple organisms [50]. In general, proliferative cells, such as induced pluripotent stem cells and cancer cells, show a global shortening of 3'UTR as compared with their less proliferative counterparts [52-55]. As gene regulation mechanisms mediated at the 3'UTRs are mostly repressive, it is generally assumed that a truncated 3'UTR will result in higher mRNA and/or protein levels [50]. It has been reported that 3'UTR shortening of oncogene

mRNAs in cancer cells leads to increased protein abundance [54,56]. Intriguingly, the truncation of the ABCG2 3'UTR has also been reported in an undifferentiated human embryonic stem (HuES) cell line where its high ABCG2 expression was associated with the short 3'UTR variant forms [57]. In contrast, another differentiated HuES cell line with lower ABCG2 levels possesses a longer 3'UTR variant [57]. Sandberg et al. also found that rapidly proliferating cells express ABCG2 mRNA with shorter 3'UTRs, presumably to escape miRNA regulation [52].

ABCB1 (MDR-1/P-gp)

ABCB1 (MDR-1/P-gp) is the most extensively studied MDR transporter, which was discovered more than 30 years ago [58]. Overexpression of MDR-1/P-gp confers cancer cells resistance to a broad range of structurally and functionally diverse chemotherapeutic drugs [59]. It was proposed to express in more than 50% of all drug-resistant human tumors. Not long after the identification of the miRNAs regulating ABCG2, a few miRNAs directly repressing ABCB1 through binding to its 3'UTR (miR-27a, -451, -296, -298, -338, -1253) have been reported (Table 1; [14,26,27]). Downregulation of these miRNAs in resistant cancer cells leads to drug resistance.

ABCC1 (MRP-1)

Multidrug resistance-associated protein (ABCC1/MRP-1) transports a wide range of different drug classes and is also known to play a critical role in the development of MDR in cancer cells. Among the major MDR transporters, the

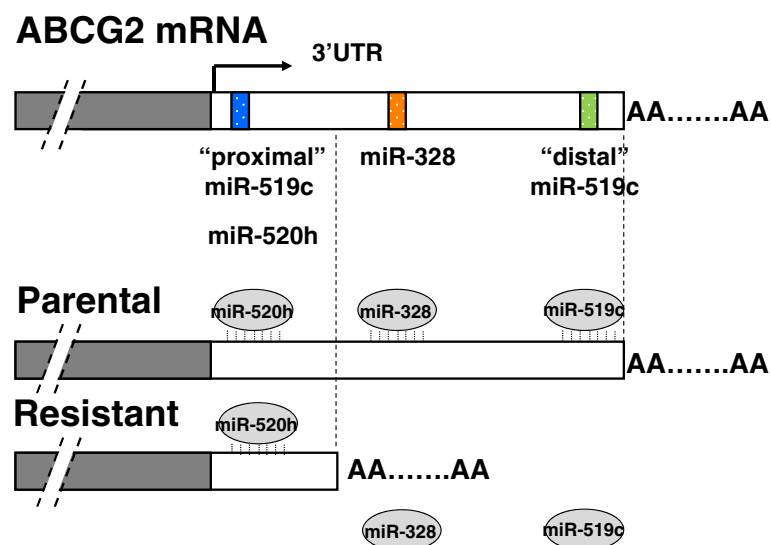


Figure 1 A proposed model for ABCG2 regulation by miR-519c. ABCG2 mRNA adopts predominantly a long form of 3'UTR in parental S1 cells but only the shorter forms in drug-resistant S1M1-80 cancer cell line [16,17]. MiR-519c (and miR-328) can only bind to the long ABCG2 3'UTR in parental cells, thus allowing ABCG2 in resistant cells to escape from their repression and thereby acquiring a higher expression.

regulation of MRP-1 by miRNAs is the least studied. To date, only miR-326 [29] and miR-1291 [30] were reported to modulate MRP-1 expression directly via interacting with its 3'UTR. In VP-16-selected MRP-1-overexpressing MCF-7 resistant cells, miR-326 was found to be downregulated and lead to MRP-1 overexpression. MiR-1291 has just been recently reported to mediate doxorubicin resistance in pancreatic cancer cells by targeting ABCC1 [30]. It was derived from a small nucleolar RNA (snoRNA: miR-1291/SNORA34), a new class of non-coding regulatory RNAs that is known to control the posttranscriptional modification of ribosomal RNAs [30]. The tissue or cell type specific processing of SNORA34 to miR-1291, and thus the overexpression of miR-1291, in pancreatic cancer may allow the development of tumor targeting therapy to combat MDR by selectively intervening the miR-1291 pathway.

ABCC2 (MRP-2)

MDR-associated protein 2 (MRP-2) is a unique ABC transporter that can mediate platinum (Pt) drug resistance [60]. Pt-based anticancer drugs, including cisplatin and oxaliplatin, are the mainstay of treatment for most solid tumors. ABCC2 can recognize GSH-conjugated form of Pt drugs and effectively pump them out of the cells to confer resistance. To date, only miR-297 has been reported to be down-regulated in a oxaliplatin-resistant colon cancer cell model (HCT116/L-OHP) to cause ABCC2 overexpression and Pt drug resistance [31]. A complementary binding site for miR-297 was identified on ABCC2 3'UTR to mediate the specific gene repression.

Indirect regulation by miRNAs

MDR-1/P-gp

Besides the various miRNAs discussed above that can directly modulate MDR-1/P-gp expression by interacting with complementary sequences at its 3'UTR, indirect regulation of the MDR transporter has also been reported. Let-7 g was reported to modulate acquired resistance of ovarian cancer to taxanes via IMP-1-mediated stabilization of MDR-1 [28]. IMP-1 is an RNA binding protein that stabilizes the mRNA of a number of target genes, including MDR-1 [61]. IMP-1 was known to be a validated target for let-7 g [62]. It follows that the loss of let-7 g commonly observed in various cancers [28] could allow the overexpression of IMP-1 and thereby stabilization of MDR-1/P-gp to mediate anticancer drug resistance (Figure 2a; [28]). Moreover, a novel miR-27a/HIPK2/MDR1/P-gp pathway has been proposed that lead to paclitaxel resistance in ovarian cancer cells [23]. Homeodomain-interacting protein kinase-2 (HIPK2) was reported to inhibit HIF-1 α , thereby suppressing *MDR1* gene transcription and sensitize cancer cells to doxorubicin-induced apoptosis

[63]. Therefore, increased expression of miR-27a in resistant cells leads to downregulation of HIPK2, which indirectly allows HIF-1 α -mediated stimulation of MDR-1/P-gp and chemoresistance (Figure 2b; [23]).

Another noteworthy indirect mechanism for miRNA-mediated upregulation of MDR-1/P-gp involved the epigenetic alteration (i.e. hypomethylation) of the *MDR-1* promoter in resistant MCF-7/DOX cells (Figure 2c; [64]). The loss of cytosine methylation in the *MDR-1* promoter, which was shown to lead to P-gp overexpression and the resistance phenotype, was proposed to be mediated by the increased expression of miR-22, miR-29a, miR-132, and miR194. These miRNAs were known to target DNA methyltransferases 3A and 3B and methyl CpG binding protein 2, which mediate *MDR-1* promoter methylation [64-66]. Although the definitive proof for this hypothesis is still lacking, it has far-reaching implication in the etiology of MDR. A number of other key mediators of MDR (including ABCG2, BCL-2, PTEN, etc) are known to be repressed by DNA methylation, therefore aberrant increased expression of the aforementioned miRNAs in cancer cells may lead to derepression of these mediators to cause MDR.

ABCC3/ABCC6

Unlike most ABC transporters that are highly expressed in various anatomic regions of the normal brain, ABCC3 and ABCC6 are not detectable in normal brain tissues [67]. Surprisingly, the specific role played by these two ABC transporters in the anticancer drug resistance of glioma stem cells has been recently reported [32]. A novel regulatory pathway Inhibitor of differentiation 4 (ID4)-miR-9*-SOX2-ABCC3/ABCC6 was proposed, which induces the stemness potential of glioma stem cells and chemoresistance (Figure 3). Of note, ABCC3 and ABCC6 are not direct targets for miR-9*. However, both of these ABC transporters are transcriptionally regulated by SOX2, which is elevated in glioma stem cells by ID4-mediated suppression of miR-9*.

Regulation of MDR via non-transporter-mediated pathways by miRNAs

Alterations of several oncogenes (e.g. Bcl2, Ras, Src) and tumor suppressor genes (e.g. p53, RB, and p16) are closely associated with chemoresistance. However, the involvement of miRNA in these processes has just begun to be unraveled. A list of the most representative miRNAs regulating these non-transporter-mediated MDR pathways is summarized in Table 2. The list is by no means exhaustive but it aims to highlight a few examples according to the biological effect of the miRNA target gene (i.e. anti-apoptotic, drug metabolism, tumor suppressor, epithelial-to-mesenchymal transition (EMT)).

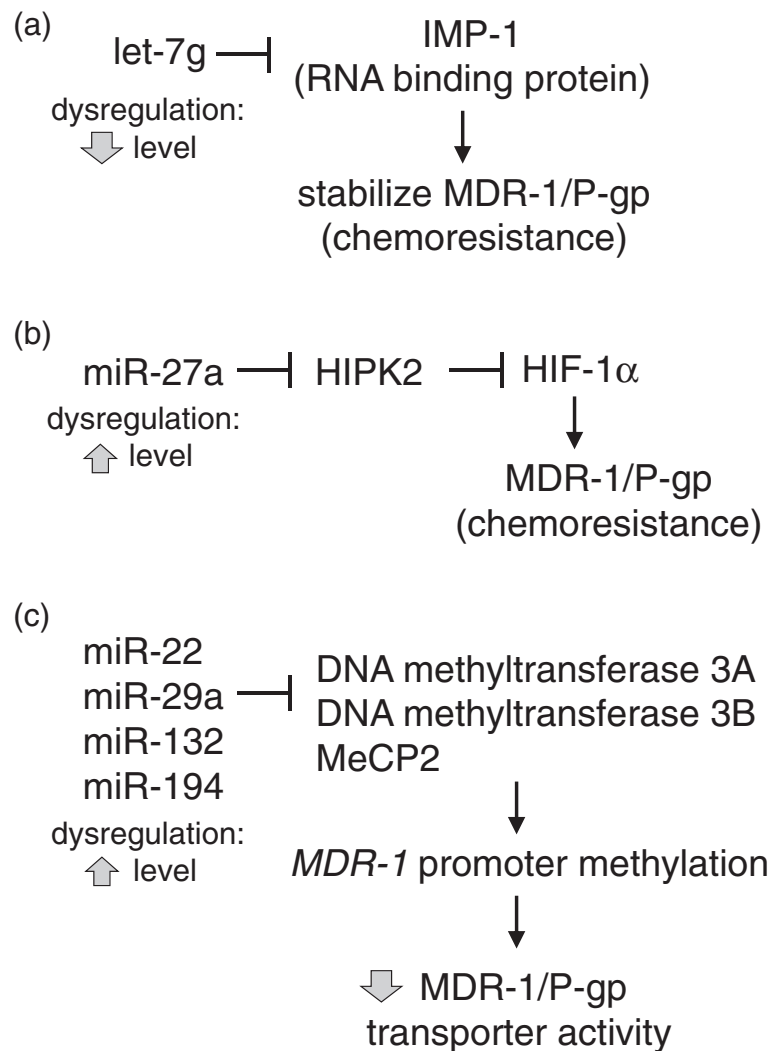


Figure 2 Indirect regulatory mechanisms of MDR-1/P-gp by miRNAs. (a) let-7 g downregulation is commonly observed in various cancers. It is known to target the RNA binding protein, IMP-1, which stabilizes MDR-1 mRNA. Therefore, let-7 g loss in resistant cells allows overexpression of IMP-1 and stability of MDR-1/P-gp to mediate drug resistance [28]. (b) Homeodomain-interacting protein kinase-2 (HIPK2) is a known target of miR-27a. HIPK2 has also been reported to inhibit HIF-1α. Increased expression of miR-27a in resistant cells leads to downregulation of HIPK2, which indirectly allows HIF-1α-mediated stimulation of MDR-1/P-gp and chemoresistance [23]. (c) A hypothetical miRNAs-DNA methylation machinery-*MDR-1* promoter methylation pathway. Increased expression of these miRNAs in resistant cells represses various DNA methylation mediators, thereby facilitating *MDR-1* promoter demethylation and increasing P-gp efflux activity to mediate chemoresistance.

(a) Anti-apoptotic

Most anticancer drugs work by induction of apoptosis. Alterations to susceptibility to apoptosis may lead to resistance to conventional cancer chemotherapy. *BCL2* (encoding the protein known as apoptosis regulator Bcl-2) is the most important pro-survival or anti-apoptotic factor often overexpressed in cancer and it is closely associated with chemotherapy resistance in various cancers. A number of miRNAs (including miR-15b, -16, -34a, -296, -1915) have been shown to modulate MDR by targeting *BCL2* [19,26,40,41]. MiR-34a is of particular interest, where both direct and indirect regulatory pathways have

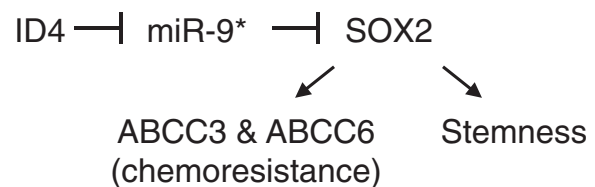


Figure 3 miR-9* is involved in an indirect regulatory mechanism of ABCC3 & ABCC6 to induce chemoresistance and stemness in glioma stem cells.

been described (Figure 4a; [41]). MiR-34a can inhibit proliferation of paclitaxel-resistant PC3PR cells by directly suppressing expression of proteins involved in cell-cycle regulation such as CDK6 and cyclin D1. On the other hand, miR-34a has also been shown to enhance apoptosis by indirectly reducing expression of SIRT1 and Bcl2 via modulating HuR.

(b) Drug metabolism

Dihydrofolate reductase (DHFR) is a key enzyme in cellular folate metabolism, which is the drug target of a widely used anticancer drug methotrexate. A naturally occurring SNP of DHFR (829C > T) has been identified near the miR-24 binding site within the 3'UTR of DHFR [20]. It interferes with miR-24 repression, thus resulting in DHFR overexpression and methotrexate resistance.

(c) Tumor suppressor

A growing list of miRNAs has been identified to regulate PTEN, an important tumor suppressor, by binding to its 3'UTR [21,68,69]. For example, miR-

214 was shown to interact with PTEN 3'UTR and inhibit PTEN translation, thereby leading to activation of the Akt pathway and cisplatin resistance [21]. The significance of this finding is that, once the miRNA-mediated regulation pathway is understood, apart from artificially altering the miRNA expression, the resistance phenotype can also be overridden by modulation of the upstream or downstream events (such as using either small-molecule Akt inhibitor or ectopic expression of PTEN cDNA lacking 3'UTR) [21].

(d) Epithelial-to-mesenchymal transition (EMT)

EMT is a cellular process that describes the change of an epithelial to a motile mesenchymal phenotype. Upon EMT, primary epithelial cancer cells acquire increased invasive and migratory abilities, eventually leading to metastases. Cancer cells selected for docetaxel resistance was found to exhibit EMT-triggered E-cadherin loss and reduced apoptosis [43]. Detailed mechanistic study revealed that these

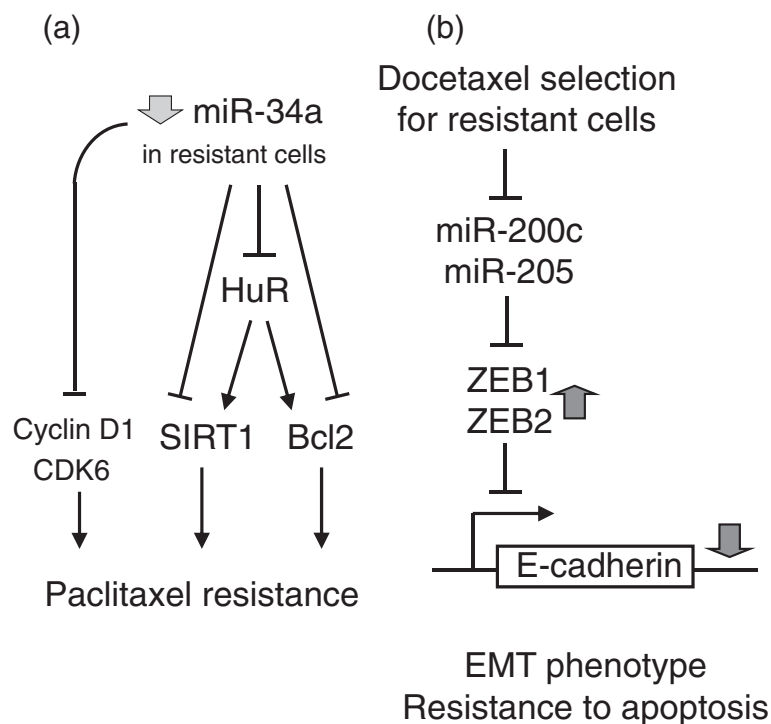


Figure 4 miR-34a mediates paclitaxel resistance via both direct and indirect mechanisms. (a) Schematic representation of direct and indirect mechanisms underlying miR-34a-mediated paclitaxel resistance. Direct pathway: miR-34a inhibit proliferation of paclitaxel-resistant PC3PR cells by directly suppressing the cell cycle regulators cyclin D1 and CDK6. Indirect pathway: miR-34a can enhance apoptosis by indirectly reducing expression of the anti-apoptotic genes (SIRT1 & BCL2) via modulating HuR [41]. **(b)** Indirect mechanism of miR-200c/-205-mediated EMT and resistance to apoptosis. Key signals that lead to EMT trigger expression of a variety of transcriptional repressors, including ZEB1, ZEB2/SIP1, Snail, and Twist. These repressors are the intracellular mediators of EMT by binding to E-box elements of genes, such as *CDH1*, that encode for the adhesion protein E-cadherin. After binding, they recruit histone deacetylases and other corepressors to facilitate transcriptional repression of E-cadherin. Reduced cellular E-cadherin expression subsequently will lead to loss of cell-cell adhesion and a series of other events, eventually leading to an invasive mesenchymal and drug resistance phenotype [43].

resistant cells have significantly reduced expression of miR-200c & -205. MiR-200c & -205 repress the transcriptional repressors, ZEB1/ZEB2, therefore reduced levels of these miRNAs allow more repression by ZEB1/ZEB2 and thereby downregulation of E-cadherin and EMT (Figure 4b; [43]). Another recent report by Liu et al. shown that miR-200c downregulation is frequently observed in metastatic melanoma, which is accompanied by overexpression of its target Bmi-1 (a Polycomb group protein class of transcriptional repressor) [42]. Upregulation of Bmi-1 was further shown to increase a number of MDR transporter genes (ABCG2, MDR1 and ABCG5) and to mediate loss of E-cadherin, collectively leading to a more resistant, malignant, and invasive EMT-like phenotype [42].

Genetic polymorphism in miRNA binding sites and anti-cancer drug resistance

Regulation by miRNAs depends on the binding to seed sequences in the 3'UTR of their target mRNAs, which subsequently leads to degradation of mRNAs and/or translation blockade. Therefore, both sequence complementarity and thermodynamics of the binding determine the likelihood of interaction between miRNAs and their targets. It follows that sequence variations such as single-nucleotide polymorphisms (SNPs) in the miRNA target site (more specifically in the seed region) could affect the miRNA-mRNA interaction and subsequently expression of the miRNA targets. To this end, a recent bioinformatics study of the entire human genome revealed a relatively high frequency of miRNA target site SNPs that disrupt target sites or create new ones [70].

The most popular SNP occurring at miRNA target site, related to anticancer drug resistance, is probably the one identified by Mishra et al. on a miR-24 binding sequence that changed cell response to methotrexate [20]. The SNP (829C → T) near the miR-24 binding site in the 3'UTR of human dihydrofolate reductase (DHFR) was found to interfere with miR-24-mediated repression, thus resulting in DHFR overexpression and methotrexate resistance. Similar to this 3'UTR polymorphism, the phenomenon of alternative transcript polyadenylation (leading to shortening of 3'UTR) described above for the regulation of ABCG2 (Figure 1) [17] is another example where miRNA-mediated gene regulation can intersect with genetic variation to mediate anticancer drug resistance.

In fact, the study of 3'UTR variations is emerging as a new research avenue that holds promise for personalized pharmacotherapy [71,72]. The idea of "miRNA-pharmacogenomics" has been proposed by Bertino et al. that study the influence of genetic polymorphisms on miRNA function as a way to predict drug behavior and to

improve drug efficacy [71]. Assessment of miRNA profile and 3'UTR polymorphisms in patients may thus allow personalized medicine [71].

Use of miRNAs as prognostic markers for predicting chemotherapy response and/or chemoresistance: a rapidly growing field

Along with strong evidence suggesting that miRNAs can be employed clinically as biomarkers for cancer classification, diagnosis, and prognosis [7,73,74], miRNAs are also rapidly gaining popularity for predicting response to chemotherapy.

(a) *In vitro* evidence from cell line studies

Blower et al. conducted one of the most systematic studies correlating global expression of 279 miRNAs and response to 3089 compounds in the NCI-60 drug screen cell line panel [15]. A good correlation between miRNA expression patterns and compound potency patterns was observed, thus strongly suggesting a prominent role played by miRNAs in mediating chemoresistance. When combined with gene expression and other biological data using multivariate analysis, miRNA expression profiles may help elucidate the complex mechanisms involved in chemosensitivity and chemoresistance.

(b) Evidence from clinical specimen analyses (miRNAs expressed in tumors)

Recent data suggest that miRNA expression in patient samples at baseline could predict chemotherapy and radiotherapy response. A summary of these miRNAs is compiled in Table 3. A few representative ones are elaborated below.

let-7i: In epithelial ovarian cancer, let-7i expression was reported to be significantly reduced in chemotherapy-resistant patients with epithelial ovarian cancer (n = 69, p = 0.003) [84]. Follow up mechanistic investigation using both gain- and loss-of-function analysis revealed that reduced let-7i expression in ovarian and breast cancer cells are linked to resistance to cisplatin chemotherapy [84]. However, a contradictory finding about the role of let-7i in drug resistance has also been reported [13]. By modulating let-7i expression with pre-miRNA or miRNA inhibitor transfection in the NCI-60 panel of human cancer cell lines, let-7i inhibition was found to enhance the sensitivity of A549 cells to one of their investigational anticancer agent NSC670550 [13].

miR-21: miR-21 dysregulation has been reported to be a predictor of tumor response in pancreatic cancer patients to conventional cytotoxic chemotherapeutic agents, including gemcitabine, docetaxel, temozolomide and 5-fluorouracil [85,86].

Table 3 Representative miRNAs from patient tumor specimens as predictive markers for treatment outcome in cancer therapy

Cancer type	miRNA dysregulation	Chemotherapeutic outcome prediction	Reference
Metastatic breast cancer	miR-26a ↑	- Multivariate analysis revealed that miR-26a and CDC2 (cell cycle regulator) are an optimal set of markers associated with favorable outcome on tamoxifen therapy, independently of traditional predictive factors (menopausal status, ER & PgR mRNA expression) - Mechanistic analysis showed that miR-26a repressed EZH2 to upregulate ER, thereby enhancing sensitivity to anti-estrogen therapy	[75]
CRC	miR-181b↓ & let-7 g↓	Associated with responsiveness to 4 th generation fluoropyrimidine-based adjuvant therapy	[76]
CRC	miR-215↑	- miR-215 level generally downregulated in clinical CRC specimen - ↑miR-215 caused chemoresistance of HCT116 to methotrexate and tomudex - No impact on treatment outcome from cisplatin and doxorubicin - High level of miR-215 was found in CRC stem cells	[77]
Metastatic CRC	miR-146b-3p↑ & miR-486-5p↑	- miRNAs level found to be more abundant in patients with mutant KRAS - Predictive of resistance to cetuximab (EGFR targeting monoclonal antibody)	[78]
Metastatic CRC	miR-200b ↑ miR-143 ↓	- In KRAS mutated tumors, ↑ miR-200b and ↓ miR-143 were associated with a good PFS in patients on cetuximab - In wild-type KRAS patients, miRNA expression did not correlate with PFS in a multivariate model	[79]
GBM	5-miRNAs signature (miR-181d, miR-518b, miR-524-5p, miR-566, miR-1227)	- Patients who had low risk scores from the 5-miRNA signature and received temozolomide treatment had better survival - Useful for identifying patients for more aggressive therapy	[80]
Hepatocellular carcinoma	miR-26↓	Lower miR-26 is associated with shorter overall survival but a better response to interferon therapy	[81]
NSCLC	miR-21 ↑	Increased miR-21 in patients not responding to platinum-based chemotherapy	[82]
Lung cancer	miR-128b LOH	Predictive of clinical response and prolonged survival following gefitinib treatment	[83]
Ovarian cancer	let-7i↓	Predictive of resistance to cisplatin	[84]
Pancreatic cancer	miR-21↑	- Predictive of resistance to gemcitabine, docetaxel, temozolomide and 5-fluorouracil - Associated with shorter overall survival in the metastatic and adjuvant setting	[85,86]
Pancreatic ductal adenocarcinoma	miR-10b ↓	Lower levels of miR-10b is associated with improved response to multimodality neoadjuvant therapy. Likelihood of surgical resection, delayed time to metastasis and increased survival	[87]

Abbreviations: CRC = colorectal cancer; EGFR = epidermal growth factor receptor; ER = estrogen receptor; GBM = glioblastoma multiforme; LOH = loss of heterozygosity; NSCLC = non-small cell lung cancer; PFS = progression-free survival; PgR = progesterone receptor.

miR-26: According to a miRNA microarray profiling study conducted in a relatively large cohort (n = 241) of patients with hepatocellular carcinoma, a list of tumor-related miRNAs closely associated with patient survival and treatment response to interferon was identified [81]. In

particular, patients whose tumors had low miR-26 expression had shorter overall survival but a better response to interferon therapy than did patients whose tumors had high expression of the miRNA.
miR-128b: In a cohort of 58 non-small-cell lung cancer patients, miR-128b loss of heterozygosity

(LOH) in the tumors was found to correlate with clinical response and prolonged survival following treatment with gefitinib [83]. Mechanistic investigation in NSCLC cell lines using miR-128b mimics or inhibitors confirmed that EGFR (the molecular target of gefitinib) is specifically regulated by the miRNA. LOH of chromosome 3p is a well-known and early genetic abnormality in lung cancer. Since miR-128b resides on the 3p22 locus, the aforementioned finding about the LOH of miR-128b could provide a functional link between a common genetic abnormality in lung cancer (i.e. loss of 3p22) and the frequent overexpression of EGFR in NSCLC patients. The clinical significance is that miR-128b may be used as a useful prognostic marker for selecting patient candidates to receive and benefit the most from EGFR tyrosine kinase inhibitors.

(c) Circulating miRNAs as non-invasive biomarkers for predicting chemotherapy response

In recent years, circulating miRNAs, referring to the miRNAs present in cell-free body fluids such as plasma, serum, urine and saliva, etc, have been exploited for use as biomarkers in various disease states. These circulating miRNAs are bound to proteins including argonaute2 [88] and high density lipoprotein [89], or are encapsulated in exosomes [90], which render them highly resistant to RNase activity. Therefore, they can be reliably measured by the highly sensitive and relatively inexpensive method of quantitative polymerase chain reaction (qPCR). Moreover, the ease of access, minimal invasiveness and the possibility of repeated sampling of circulating miRNAs have also made them ideal candidates for use as biomarkers. Due to space limitation, we have highlighted a few representative circulating miRNAs that have been reported as useful predictive biomarkers for chemotherapy response (Table 4).

While a number of circulating miRNAs have been identified as useful biomarkers for predicting treatment response to chemotherapy and/or surgery, the detailed mechanism is usually not determined. It remains to be determined as to whether the circulating miRNAs are actively released by surviving cancer cells or derived from the dead cancer cells. Interestingly, in prostate cancer, Lucotti et al. has nicely demonstrated that cytotoxic treatment of DU-145 prostate cancer cells by fludarabine enhanced the release of a list of exosome-associated prostate cancer secretory (PCS)-miRNAs, with the exception of miR-485-3p, which is retained by surviving cancer cells [97]. Follow-up mechanistic investigation revealed that the intracellular retention

of miR-485-3p downregulate the transcriptional repressor NF- κ B, and thus allowing the overexpression of drug resistance genes (including TOP2A, MDR1 and cyclin B2 pro-survival genes) to mediate resistance [97].

In summary, tumoral miRNA expression at diagnosis may help predict the patients' response to chemotherapy and also provide insights about mechanism of chemotherapy resistance. It may also provide guidance for rational and personalized chemotherapy selection. To this end, circulating miRNA as a novel prognostic or predictive tool is also rapidly gaining popularity [100,101] because of the non-invasive nature of the detection method. Moreover, miRNA-based profiling also has another added advantage over conventional mRNA-based methods. MiRNAs in formalin-fixed tissues, blood plasma and serum are known to be remarkably more stable than mRNAs to endogenous RNase digestion [102], thereby enabling their reliable extraction and analysis from patient specimens.

MiRNAs as druggable targets and miRNA-based therapeutics for circumvention of anticancer drug resistance

(a) MiRNA-based therapeutics

Since miRNA expression is often dysregulated in cancer cells, approaches that modulate miRNA activity could potentially produce specific anti-cancer effect. With the advancement in technology, modulation of endogenous miRNA levels can now be achieved in several ways. Oncogenic miRNAs can be targeted for downregulation using various modified antisense oligonucleotides (also known as antagomirs) to their precursor or mature forms, whereas tumor suppressive miRNAs may be directly upregulated by using synthetic miRNA mimics (including siRNA-like oligoribonucleotide duplex and chemically modified oligoribonucleotide [103]) for an anti-cancer effect. In particular, antagomirs, with 2'-O-methylation or locked nucleic acid modifications, have drawn a lot of attention. Successful *in vivo* silencing of miRNA has been achieved by their systematic administration through tail vein injections into mice [104,105]. On the other hand, a few proof-of-concept studies using artificial synthetic miRNAs have been successfully performed to target a few oncogenes and produce anticancer effect [106-108]. A list of representative miRNAs as potential molecular targets for cancer therapy is compiled in Table 5.

Table 4 Representative circulating miRNAs reported to predict response to chemotherapy and/or surgery

Cancer type	miRNA dysregulation associated with poor response	Sample type	Significance	Reference
Breast	↑ miR-125b	Serum	Increased in patients not responding to neoadjuvant chemotherapy	[91]
Breast	↑ miR-210	Plasma	Lower miR-210 plasma levels are associated with - complete response to trastuzumab (HER-2 targeted monoclonal antibody) - surgical removal of tumor - lack of tumor metastasis to lymph nodes	[92]
Colorectal (CRC)	↑ miR17-3p	Plasma	- Elevated in both CRC tissue and plasma - Lower level detected in post-operative plasma is associated with responsiveness to surgery	[93]
Colorectal (CRC)	↑ miR-29a	Serum	- Elevated in both CRC tissue and plasma - Help differentiate CRC from gastric cancer, inflammatory bowel disease and no tumor controls - Lower level detected in post-operative plasma is associated with responsiveness to surgery	[93]
Colorectal (CRC)	↑ miR-27b, miR-148a, miR-326	Plasma	Elevated in patients with metastatic CRC not responding to 5-fluorouracil and oxaliplatin-based chemotherapy	[94]
Lung	↑ miR-21	Plasma	Increased; associated with resistance to platinum-based chemotherapy	[95]
Non-Hodgkin's lymphoma (NHL)	↓ miR-92a	Plasma	- Remarkably lower in NHL patients (< 5%) than in healthy subjects - The very low plasma level of miR-92a increased in complete response phase but became lower again in the relapse phase	[96]
Prostate	↑ Prostate cancer secretory (PCS)-miRNAs	Plasma/serum	- It is not clear whether circulating miRNAs are actively released by live cancer cells or derived from dead cancer cells. - <i>In vitro</i> cytotoxic treatment of DU-145 cells enhanced the release of exosomes-associated PCS-miRNAs, with the exception of miR-485-3p, which is retained by surviving cancer cells. - The intracellular retention of miR-485-3p was shown to downregulate the transcriptional repressor NF- κ B, thus allowing the overexpression of a few drug resistance genes (including TOP2A, MDR1, and cyclin B2 pro-survival genes)	[97]
Prostate	↑ miR-21	Serum	- Increased in hormone-refractory prostate cancer - Associated with resistance to docetaxel-based chemotherapy	[98]
Advanced renal cell carcinoma	↑ miR-192 ↑ miR-193a-3p	Peripheral blood samples	- Models predicting poor and prolonged response to sunitinib were constructed - Ontology analyses revealed relevance to cancer-related pathways (angiogenesis and apoptosis) - miRNA expression signatures may be used to identify patients who may benefit the most from 1 st line therapy with sunitinib	[99]

Table 5 MiRNAs as targets for cancer therapy

Cancer type	miRNA target and its role in cancer	Delivery system to modulate the miRNA <i>in vivo</i> *	Reference
Breast cancer	miR-34a – Tumor suppressor	Cationic liposomes	[109]
Glioblastoma	miR-145 – Tumor suppressor	Adenoviruses	[110]
Glioblastoma	miR-221-222 – Oncogene	Adenoviruses	[111]
Glioblastoma Multiforme (GBM)	miR-9 – promote expression of P-gp (a multidrug resistance efflux transporter)	- Mesenchymal stem cell-derived exosomes - To deliver anti-miR-9 to temozolomide-resistant GBM to reduce P-gp expression for resistance reversal	[112]
Hepatocellular carcinoma	miR-26 – Tumor suppressor	Adenoviruses	[113]
Lung cancer	let-7 – Tumor suppressor	Adenoviruses	[114]
Lung cancer	miR-34a – Tumor suppressor	Cationic liposomes	[115]
Lymphoma	miR-155 – Oncogene	Polymer-based nanoparticles	[116]
Medulloblastoma	miR-17 ~ 92 cluster family – Sonic Hedgehog signaling	8-mer seed-targeting locked nucleic acid (LNA)-modified anti-miR oligonucleotides (nude mice)	[117]
Pancreatic cancer	miR-21 – Oncogene	Lentiviruses	[118]

* The miRNA target can be modulated by anti-miRNA oligonucleotides or miRNA-expressing constructs (delivered by viral or non-viral vectors).

(b) Potential miRNA targets for resistance circumvention

Although the use of miRNAs for cancer chemotherapy has not yet been realized in clinical trials, it has recently been demonstrated in tissue culture systems that miRNA-targeted therapy may be useful in combination with conventional chemo-radiotherapy to sensitize the cancer cells. Table 6 summarizes a few representative miRNA-modulatory approaches to circumvent anticancer drug resistance. let-7 overexpression has been shown to confer radiosensitivity in lung cancer cell lines [123]. Inhibition of miR-21 and miR-200b was reported to enhance the sensitivity of cholangiocarcinomas to gemcitabine chemotherapy [120]. MiR-21 is of particular interest, which is overexpressed in most cancer types analyzed [124].

A landmark study has been reported to illustrate the phenomenon of “oncomiR addiction” in an *in vivo* model of miR-21-induced pre-B-cell lymphoma [119]. Most intriguingly, complete tumor regression can be achieved in a few days when miR-21 was inactivated by the antisense strategy *in vivo*. Given that aberrant miR-21 expression is known to reduce sensitivity of cancer cells to a number of anticancer drugs including tamoxifen, gemcitabine, doxorubicin and docetaxel [124], inactivating miR-21 may represent a novel strategy for cancer drug resistance circumvention. Most recently, a novel approach to deliver functional anti-miR-9 by mesenchymal stem cell-derived exosomes to glioblastoma multiforme (GBM) cells has been reported to circumvent P-gp-mediated resistance to temozolomide [112]. Site-directed

Table 6 Novel approaches to circumvent chemoresistance by modulating unique miRNAs

Cancer type	miRNA targeted for inhibition	Type of resistance circumvented	Delivery system for modulation of miRNAs	Reference
B-cell lymphoma	miR-21 – oncomiR addiction	–	Antisense strategy	[119]
Cholangiocarcinomas	miR-21 & miR-200b	Gecitabine resistance mediated by PTEN-dependent activation of PI3K signaling	Transfection with miRNA-specific antisense oligonucleotides	[120]
Glioblastoma Multiforme (GBM)	miR-9 – indirectly promoting expression of the MDR transporter P-gp	P-gp-mediated resistance to temozolomide	- Mesenchymal stem cell-derived exosomes - To deliver anti-miR-9 to temozolomide-resistant GBM to reduce P-gp expression for resistance reversal	[112]
GBM	miR-21	- Temozolomide - Enhance apoptosis	Transfection with anit-miR-21 oligonucleotide	[121]
Lung cancer	miR-92b	Cisplatin resistance mediated by downregulation of the tumor suppressor gene <i>PTEN</i>	Transfection with anti-miR-92b oligonucleotide	[122]

targeted delivery of the anti-miR-9 to GBM cells was achieved because mesenchymal stem cells are able to migrate preferentially to the brain.

(c) Therapeutic drugs altering miRNA profile in cancer cells

Apart from exhibiting aberrant expression of a few miRNAs, human cancers are in fact characterized by impaired miRNA processing and global miRNA dysregulation [125]. It has been recently shown that miRNA expression can be differentially altered by xenobiotic drugs in different human cell lines [126]. The drugs identified are not necessarily anticancer drugs. The practical implication is that they could be safely administered with other conventional anticancer drugs in an attempt to reverse miRNA-mediated drug resistance. Along this line of investigation, the fluoroquinolone class of antibiotics has been shown to enhance RNA interference and promote miRNA processing [127,128]. This may represent a novel approach to modulate multiple miRNAs simultaneously or to restore the global miRNA expression (i.e. miRNAome) to provide a cancer-specific growth inhibitory effect. It has also been proposed that specific class of drugs might be screened for their effects on shifting the miRNA expression profile of a cancer cell towards that of a normal tissue [129]. To this end, the SM2miR database has been established to provide a fairly comprehensive repository about the influences of small molecules on miRNA expression, which could promote the further development of miRNA-targeting therapeutics [130]. Calin et al. has proposed structure-based approaches (such as molecular docking) to identify compounds that may target specific miRNAs [131]. However, three-dimensional structure prediction of miRNA still remains a challenge. There has also been attempts to identify small molecules targeting specific miRNAs in human cancers based on transcriptional responses [132]. As more is discovered in this research area, the specific modulation of miRNAs by therapeutic drugs may become feasible in the future.

(d) Off-target effects

Each miRNA typically targets up to hundreds of transcripts directly or indirectly, and multiple miRNAs can target a given gene. It follows that miRNAs are also tied to some tightly regulated gene expression networks in normal cells. The therapeutic outcome of a miRNA-targeted chemotherapy or resistance reversal regimen may thus depend on the number of miRNA targets and the affinities for each of these targets that are expressed in a given tumor microenvironment. It

could be difficult to rule out the by-stander off-target effects, if any. Moreover, an appropriate method to deliver the effective miRNA mimic/antagomir to the right cell type must also be considered in order to prevent unwanted side effects. Indeed, a fatal side effect as a result of saturation of miRNA pathways has been reported in an animal study [133]. It appears that these hurdles have to be overcome before an effective miRNA-targeted strategy can be realized for circumvention of anticancer drug resistance in cancer patients.

Conclusions

The emerging role of miRNAs as regulators of gene expression, and their dysregulation in human cancer has provided opportunities for their therapeutic application in the capacity of cancer detection, diagnosis and prognosis prediction. Selective targeting of some miRNAs may be useful in enhancing chemosensitivity, and may have future applications in modulating therapeutic response to molecular targeted chemotherapeutics in selected cancer subtypes. A better understanding about the complex regulatory pathways that control miRNAs function and their tumor-specific effect will be needed in order to fully realize the promise of miRNAs in cancer diagnosis, chemotherapy and drug resistance circumvention.

Abbreviations

ABCG2/BCRP: Breast cancer resistance protein; ABC transporters: ATP-binding cassette transporters; CSC: Cancer stem cell; DHFR: Dihydrofolate reductase; EGFR: Epidermal growth factor receptor; EMT: Epithelial-to-mesenchymal transition; GSH: Glutathione; HuES: Human embryonic stem cell; LOH: Loss of heterozygosity; MDR-1/ABCB1: P-glycoprotein; MeCP2: Methyl CpG binding protein 2; miRNA/miR: microRNA; MRP-1/ABCC1: Multidrug resistance associated protein 1; PcG: Polycomb group protein; Pt: Platinum; PTEN: Phosphatase and tensin homolog; SnoRNA: Small nucleolar RNA; SP: Side population; 3'UTR: 3'untranslated region.

Competing interests

The author declares that he has no competing interests.

Author's contributions

KKT completed the final draft and approved the manuscript.

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References

1. Goldman B: Multidrug resistance: can new drugs help chemotherapy score against cancer? *J Natl Cancer Inst* 2003, **95**:255-257.
2. Gottesman MM, Fojo T, Bates SE: Multidrug resistance in cancer: role of ATP-dependent transporters. *Nat Rev Cancer* 2002, **2**:48-58.
3. Bartel DP: MicroRNAs: target recognition and regulatory functions. *Cell* 2009, **136**:215-233.

4. Lim LP, Glasner ME, Yekta S, Burge CB, Bartel DP: Vertebrate microRNA genes. *Science* 2003, **299**:1540.
5. Lewis BP, Burge CB, Bartel DP: Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets. *Cell* 2005, **120**:15–20.
6. Xie X, Lu J, Kulbokas EJ, et al: Systematic discovery of regulation motifs in human promoters and 3'UTRs by comparison of several mammals. *Nature* 2005, **434**:338–345.
7. Lu J, Getz G, Miska EA, et al: MicroRNA expression profiles classify human cancers. *Nature* 2005, **435**:834–838.
8. Esquela-Kerscher A, Slack FJ: Oncomirs – microRNAs with a role in cancer. *Nat Rev Cancer* 2006, **6**:259–269.
9. Hammond SM: MicroRNAs as tumor suppressors. *Nat Genet* 2007, **39**:582–583.
10. Tricoli JV, Jacobson JW: MicroRNA: potential for cancer detection, diagnosis, and prognosis. *Cancer Res* 2007, **67**:4553–4555.
11. Blenkiron C, Miska EA: miRNAs in cancer: approaches, aetiology, diagnostics and therapy. *Hum Mol Genet* 2007, **16**:R106–R113.
12. Fojo T: Multiple paths to a drug resistance phenotype: mutations, translocations, deletions and amplification of coding genes or promoter regions, epigenetic changes and microRNAs. *Drug Resist Update* 2007, **10**:59–67.
13. Blower PE, Chung JH, Verducci JS, et al: MicroRNAs modulate the chemosensitivity of tumor cells. *Mol Cancer Ther* 2008, **7**:1–9.
14. Kovalchuk O, Fikowski J, Meservy J, et al: Involvement of microRNA-451 in resistance of the MCF-7 breast cancer cells to chemotherapeutic drug doxorubicin. *Mol Cancer Ther* 2008, **7**:2152–2159.
15. Blower PE, Verducci JS, Lin S, et al: MicroRNA expression profiles for the NCI-60 cancer cell line panel. *Mol Cancer Ther* 2007, **6**:1483–1491.
16. To KK, Zhan Z, Litman T, Bates SE: Regulation of ABCG2 expression at the 3'untranslated region of its mRNA through modulation of transcript stability and protein translation by a putative microRNA in the S1 colon cancer cell line. *Mol Cell Biol* 2008, **28**:5147–5161.
17. To KK, Robey RW, Knutsen T, Zhan Z, Ried T, Bates SE: Escape from hsa-miR-519c enables drug-resistant cells to maintain high expression of ABCG2. *Mol Cancer Ther* 2009, **8**:2959–2968.
18. Pan YZ, Morris ME, Yu AM: MicroRNA-328 negatively regulates the expression of breast cancer resistance protein (BCRP/ABCG2) in human cancer cells. *Mol Pharmacol* 2009, **75**:1374–1379.
19. Xia L, Zhang D, Du R, et al: MiR-15b and miR-16 modulate multidrug resistance by targeting BCL2 in human gastric cancer cells. *Int J Cancer* 2008, **123**:372–379.
20. Mishra PJ, Humeniuk R, Longo Sorbello GS, Banerjee D, Bertino JR: A miR-24 microRNA binding-site polymorphism in dihydrofolate reductase gene leads to methotrexate resistance. *Proc Natl Acad Sci USA* 2007, **104**:13513–13518.
21. Yang H, Kong W, He L, et al: MicroRNA expression profiling in human ovarian cancer: miR-214 induces cell survival and cisplatin resistance by targeting PTEN. *Cancer Res* 2008, **68**:425–433.
22. Allen KE, Weiss GJ: Resistance may not be futile: microRNA biomarkers for chemoresistance and potential therapeutics. *Mol Cancer Ther* 2010, **9**:3126–3136.
23. Li Z, Hu S, Wang J, et al: MiR-27a modulates MDR1/P-glycoprotein expression by targeting HIPK2 in human ovarian cancer cells. *Gynecol Oncol* 2010, **119**:125–130.
24. Zhu H, Wu H, Liu X, et al: Role of microRNA miR-27a and miR-451 in the regulation of MDR1/P-glycoprotein expression in human cancer cells. *Biochem Pharmacol* 2008, **76**:582–588.
25. Xu Y, Xia F, Ma L, et al: MicroRNA-122 sensitizes HCC cancer cells to adriamycin and vincristine through modulating expression of MDR and inducing cell cycle arrest. *Cancer Lett* 2011, **310**:160–169.
26. Hong L, Han Y, Zhang H, et al: The prognostic and chemotherapeutic value of miR-296 in esophageal squamous cell carcinoma. *Ann Surg* 2010, **251**:1056–1063.
27. Bao L, Hazari S, Mehra S, Kaushal D, Moroz K, Dash S: Increased expression of P-glycoprotein and doxorubicin chemoresistance of metastatic breast cancer is regulated by miR-298. *Am J Pathol* 2012, **180**:2490–2503.
28. Boyerinas B, Park SM, Murmann AE, et al: Let-7 modulates required resistance of ovarian cancer to Taxanes via IMP-1-mediated stabilization of multidrug resistance 1. *Int J Cancer* 2012, **130**:1787–1797.
29. Liang Z, Wu H, Xia J, et al: Involvement of miR-326 in chemotherapy resistance of breast cancer through modulating expression of multidrug resistance-associated protein 1. *Biochem Pharmacol* 2010, **79**:817–824.
30. Pan YZ, Zhou A, Hu Z, Yu AM: Small nucleolar RNA-derived microRNA hsa-miR-1291 modulates cellular drug disposition through direct targeting of ABC transporter ABCC1. *Drug Metab Dispos* 2013; doi: 10.1124/dmd.113.052092.
31. Xu K, Liang X, Shen K, et al: MiR-297 modulates multidrug resistance in human colorectal carcinoma by down-regulating MRP-2. *Biochem J* 2012, **446**:291–300.
32. Jeon HM, Sohn YW, Oh SY, et al: ID4 imparts chemoresistance and cancer stemness to glioma cells by derepressing miR-9^a-mediated suppression of SOX2. *Cancer Res* 2011, **71**:3410–3421.
33. Turrini E, Haensch S, Laechelt S, Diewock T, Bruhn O, Cascorbi I: MicroRNA profiling in K-562 cells under imatinib treatment: influence of miR-212 and miR-328 on ABCG2 expression. *Pharmacogenet Genomics* 2012, **22**:198–205.
34. Xu XT, Xu Q, Tong JL, et al: MicroRNA expression profiling identifies miR-328 regulates cancer stem cell-like SP cells in colorectal cancer. *Br J Cancer* 2012, **106**:1320–1330.
35. Pan YZ, Seigel GM, Hu ZH, Huang M, Yu AM: Breast cancer resistance protein BCRP/ABCG2 regulatory microRNAs (hsa-miR-328, -519c, -520 h) and their differential expression in stem-like ABCG2+ cancer cells. *Biochem Pharmacol* 2011, **81**:783–792.
36. Liao R, Sun J, Zhang L, et al: MicroRNAs play a role in the development of human hematopoietic stem cells. *J Cell Biochem* 2008, **104**:805–817.
37. Wang F, Xue X, Wei J, et al: hsa-miR-520 h downregulates ABCG2 in pancreatic cancer cells to inhibit migration, invasion, and side populations. *Br J Cancer* 2010, **103**:567–574.
38. Jiao X, Zhao L, Ma M, Bai X, He M, Yan Y, Wang Y, Chen Q, Zhao X, Zhou M, Cui Z, Zheng Z, Wang E, Wei M: MiR-181a enhances drug sensitivity in mitoxantrone-resistant breast cancer cells by targeting breast cancer resistance protein (BCRP/ABCG2). *Breast Cancer Res Treat* 2013, **139**:717–730.
39. Ma MT, He M, Wang Y, Jiao XY, Zhao L, Bai XF, Yu ZJ, Wu HZ, Sun ML, Song ZG, Wei MJ: MiR-487a resensitizes mitoxantrone (MX)-resistant breast cancer cells (MCF-7/MX) to MX by targeting breast cancer resistance protein (BCRP/ABCG2). *Cancer Lett* 2013, **339**:107–115.
40. Xu K, Liang X, Cui D, Wu Y, Shi W, Liu J: miR-1915 inhibits Bcl-2 to moderate multidrug resistance by increasing drug-sensitivity in human colorectal carcinoma cells. *Mol Carcinog* 2013, **52**:70–78.
41. Kojima K, Fujita Y, Nozawa Y, Deguchi T, Ito M: MiR-34a attenuates paclitaxel-resistance of hormone-refractory prostate cancer PC3 cells through direct and indirect mechanisms. *Prostate* 2010, **70**:1501–1512.
42. Liu S, Tetzlaff MT, Cui R, Xu X: miR-200c inhibits melanoma progression and drug resistance through down-regulation of Bmi-1. *Am J Pathol* 2012, **181**:1823–1835.
43. Pühr M, Hoefler J, Schafer G, et al: Epithelial-to-mesenchymal transition leads to docetaxel resistance in prostate cancer and is mediated by reduced expression of miR-200c and miR-205. *Am J Pathol* 2012, **181**:2188–2201.
44. Cochrane DR, Spoelstra NS, Howe EN, Nordeen SK, Richer JK: MicroRNA-200c mitigates invasiveness and restores sensitivity to microtubule-targeting chemotherapeutic agents. *Mol Cancer Ther* 2009, **8**:1055–1066.
45. Robey RW, Polgar O, Deeken J, To KW, Bates SE: determining its relevance in clinical drug resistance. *Cancer Metastasis Rev* 2007, **26**:39–57.
46. Knutsen T, Rao VK, Ried T, et al: Amplification of 4q21-q22 and the MXR gene in independently derived mitoxantrone-resistant cell lines. *Genes Chromosomes Cancer* 2000, **27**:110–116.
47. Nakanishi T, Bailey-Dell KJ, Hassel BA, et al: Novel 5'untranslated region variants of BCRP mRNA are differentially expressed in drug-selected cancer cells and in normal human tissues: implications for drug resistance, tissue-specific expression, and alternative promoter usage. *Cancer Res* 2006, **66**:5007–5011.
48. Krishnamurthy P, Ross DD, Nakanishi T, et al: The stem cell marker Bcrp/ABCG2 enhances hypoxic cell survival through interactions with heme. *J Biol Chem* 2004, **279**:24218–24225.
49. Guhaniyogi J, Brewer G: Regulation of mRNA stability in mammalian cells. *Gene* 2001, **265**:11–23.
50. Di Giannardino DC, Nishida K, Manley JL: Mechanisms and consequences of alternative polyadenylation. *Mol Cell* 2011, **43**:853–866.

51. Tian B, Hu J, Zhang H, Lutz CS: A large-scale analysis of mRNA polyadenylation of human and mouse genes. *Nucleic Acids Res* 2005, **33**:201–212.
52. Sandberg R, Neilson JR, Sarma A, Sharp PA, Burge CB: Proliferating cells express mRNAs with shortened 3'untranslated regions and fewer microRNA target sites. *Science* 2008, **320**:1643–1647.
53. Ji Z, Tian B: Reprogramming of 3'untranslated regions of mRNAs by alternative polyadenylation in generation of pluripotent stem cells from different cell types. *PLoS One* 2009, **4**:e8419.
54. Mayr C, Bartel DP: Widespread shortening of 3'UTR by alternative cleavage and polyadenylation activates oncogenes in cancer cells. *Cell* 2009, **138**:673–684.
55. Shepard PJ, Choi EA, Lu J, Flanagan LA, Hertel KJ, Shi Y: Complex and dynamic landscape of RNA polyadenylation revealed by PAS-Seq. *RNA* 2011, **17**:761–772.
56. Legendre M, Ritchie W, Lopez F, Gautheret D: Differential repression of alternative transcripts: a screen for miRNA targets. *PLoS Comput Biol* 2006, **2**:e43.
57. Apati A, Orban TI, Varga N, et al: High level functional expression of the ABCG2 multidrug transporter in undifferentiated human embryonic stem cells. *Biochim Biophys Acta* 2008, **1778**:2700–2709.
58. Juliano RL, Ling V: A surface glycoprotein modulating drug permeability in Chinese hamster ovary cell mutants. *Biochim Biophys Acta* 1976, **455**:152–162.
59. Ambudkar SV, Kimchi-Sarfaty C, Sauna ZE, Gottesman MM: P-glycoprotein: from genomics to mechanism. *Oncogene* 2003, **22**:7468–7485.
60. Zhou SF, Wang LL, Di YM, et al: Substrates and inhibitors of human multidrug resistance associated proteins and the implications in drug development. *Curr Med Chem* 2008, **15**:1981–2039.
61. Sparanese D, Lee CH: CRD-BP shields c-myc and MDR-1 RNA from endonucleolytic attack by a mammalian endoribonuclease. *Nucleic Acids Res* 2007, **35**:1209–1221.
62. Boyerinas B, Park SM, Shomron N, et al: Identification of let-7-regulated oncofetal genes. *Cancer Res* 2008, **68**:2587–2591.
63. Nardinocchi L, Puca R, Sacchi A, D'Orazi G: Inhibition of HIF-1alpha activity by homeodomain-interacting protein kinase-2 correlates with sensitization of chemoresistant cells to undergo apoptosis. *Mol Cancer* 2009, **8**:1.
64. Chekhun VF, Lukyanova NY, Kovalchuk O, Tryndyak VP, Pogribny IP: Epigenetic profiling of multidrug-resistant human MCF-7 breast adenocarcinoma cells reveals novel hyper- and hypomethylated targets. *Mol Cancer Ther* 2007, **6**:1089–1098.
65. Fabbri M, Garzon R, Cimmino A, et al: MicroRNA-29 family reverts aberrant methylation in lung cancer by targeting DNA methyltransferases 3A and 3B. *Proc Natl Acad Sci USA* 2007, **104**:15805–15810.
66. Klein ME, Liyo DT, Ma L, Lmpey S, Mandel G, Goodman RH: Homeostatic regulation of MeCP2 expression by a CREB-induced microRNA. *Nat Neurosci* 2007, **10**:1513–1514.
67. Nies AT, Jedlitschky G, Konig J, et al: Expression and immunolocalization of the multidrug resistance proteins, MRP1-MRP6 (ABCC1-ABCC6), in human brain. *Neuroscience* 2004, **129**:349–360.
68. Garofalo M, Di Leva G, Romano G, et al: miR-221&222 regulate TRAIL resistance and enhance tumorigenicity through PTEN and TIMP3 downregulation. *Cancer Cell* 2009, **16**:498–509.
69. Vinciguerra M, Sgroi A, Veyrat-Durebex C, Rubbia-Brandt L, Buhler LH, Foti M: Unsaturated fatty acids inhibit the expression of tumor suppressor phosphatase and tensin homolog (PTEN) via microRNA-21 up-regulation in hepatocytes. *Hepatology* 2009, **49**:1176–1184.
70. Saunders MA, Liang H, Li WH: Human polymorphism at microRNAs and microRNA target sites. *Proc Natl Acad Sci USA* 2007, **104**:3300–3305.
71. Bertino JR, Banerjee D, Mishra PJ: Pharmacogenomics of microRNA: a miRSNP towards individualized therapy. *Pharmacogenomics* 2007, **8**:1625–1627.
72. Mishra PJ, Banerjee D, Bertino JR: MiRSNPs or MiR-polymorphisms, new players in microRNA mediated regulation of the cell. *Cell Cycle* 2008, **7**:853–858.
73. Yanaihara N, Caplen N, Bowman E, et al: Unique microRNA molecular profiles in lung cancer diagnosis and prognosis. *Cancer Cell* 2006, **9**:189–198.
74. Mitchell PS, Parkin RK, Kroh EM, et al: Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci USA* 2008, **105**:10513–10518.
75. Jansen MP, Reijm EA, Sieuwerts AM, Ruigrok-Ritstier K, Look MP, Rodriguez-Gonzalez FG, Heine AA, Martens JW, et al: High miR-26a and low CDC2 levels associate with decreased EZH2 expression and with favorable outcome on tamoxifen in metastatic breast cancer. *Breast Cancer Res Treat* 2012, **133**:937–947.
76. Nakajima G, Hayashi K, Xi Y, Kudo K, Uchida K, Takasaki K, Yamamoto M, Ju J: Non-coding microRNAs hsa-let-7 g and hsa-miR-181b are associated with chemoresponse to S-1 in colon cancer. *Cancer Genomics Proteomics* 2006, **3**:317–324.
77. Song B, Wang Y, Titmus MA, Botchkina G, Formentini A, Kormann M, Ju J: Molecular mechanism of chemoresistance by miR-215 in osteosarcoma and colon cancer cells. *Mol Cancer* 2010, **9**:96.
78. Ragusa M, Majorana A, Statello L, Maugeri M, Salito L, Barbagallo D, Guglielmino MR, Duro LR, Angelica R, Caltabiano R, Biondi A, Di Vita M, Privitera G, et al: Specific alterations of microRNA transcriptome and global network structure in colorectal carcinoma after cetuximab treatment. *Mol Cancer Ther* 2010, **9**:3396–3409.
79. Mekenkamp LJ, Tol J, Dijkstra JR, de Krijger I, et al: Beyond KRAS mutation status: influence of KRAS copy number status and microRNAs on clinical outcome to cetuximab in metastatic colorectal cancer patients. *BMC Cancer* 2012, **12**:292.
80. Zhang W, Zhang J, Yan W, You G, Bao Z, Li S, et al: Whole-genome microRNA expression profiling identifies a 5-microRNA signature as a prognostic biomarker in Chinese patients with primary glioblastoma multiforme. *Cancer* 2013, **119**:814–824.
81. Ji J, Shi J, Budhu A, et al: MicroRNA expression, survival, and response to interferon in liver cancer. *N Eng J Med* 2009, **361**:1437–1447.
82. Gao W, Lu X, Liu L, Xu J, Feng D, Shu Y: MiRNA-21: a biomarker predictive of platinum-based adjuvant chemotherapy response in patients with non-small cell lung cancer. *Cancer Biol Ther* 2012, **13**:330–340.
83. Weiss GJ, Bemis LT, Nakajima E, et al: EGFR regulation by microRNA in lung cancer: correlation with clinical response and survival to gefitinib and EGFR expression in cell lines. *Ann Oncol* 2008, **19**:1053–1059.
84. Yang N, Kaur S, Volinia S, et al: MicroRNA microarray identifies let-7i as a novel biomarker and therapeutic target in human epithelial ovarian cancer. *Cancer Res* 2008, **68**:10307–10314.
85. Hwang JH, Voortman J, Giovannetti E, et al: Identification of microRNA-21 as a biomarker for chemoresistance and clinical outcome following adjuvant therapy in resectable pancreatic cancer. *PLoS One* 2010, **5**:e10630.
86. Giovannetti E, Funel N, Peters GJ, et al: MicroRNA-21 in pancreatic cancer: correlation with clinical outcome and pharmacologic aspects underlying its role in the modulation of gemcitabine activity. *Cancer Res* 2010, **70**:4528–4538.
87. Preis M, Gardner TB, Gordon SR, Pipas JM, et al: MicroRNA-10b expression correlates with response to neoadjuvant therapy and survival in pancreatic ductal adenocarcinoma. *Clin Cancer Res* 2011, **17**:5812–5821.
88. Arroyo JD, Chevillet JR, Kroh EM, et al: Argonaute2 complexes carry a population of circulating microRNAs independent of vesicles in human plasma. *Proc Natl Acad Sci USA* 2011, **108**:5003–5008.
89. Vickers KC, Palmisano BT, Shoucri BM, Shamburek RD, Remaley AT: MicroRNAs are transported in plasma and delivered to recipient cells by high-density lipoproteins. *Nat Cell Biol* 2011, **13**:423–433.
90. Gallo A, Tandon M, Alevizos I, Illei GG: The majority of microRNAs detectable in serum and saliva is concentrated in exosomes. *PLoS ONE* 2012, **7**:e30679.
91. Wang H, Tan G, Dong L, Cheng L, Li K, Wang Z, et al: Circulating miR-125b as a marker predicting chemoresistance in breast cancer. *PLoS ONE* 2012, **7**:e34210.
92. Jung EJ, Santarpia L, Kim J, Esteva FJ, Moretti E, Buzdar AU, Di Leo A, Le XF, Bast RC Jr, Park ST, Pusztai L, Calin GA: Plasma microRNA 210 levels correlate with sensitivity to trastuzumab and tumor presence in breast cancer patients. *Cancer* 2012, **118**:2603–2614.
93. Ng EK, Chong WW, Jin H, Lam EK, Shin VY, Yu J, et al: Differential expression of microRNAs in plasma of patients with colorectal cancer: a potential marker for colorectal cancer screening. *Gut* 2009, **58**:1375–1381.
94. Kjersem JB, Ikdahl T, Lingjaerde OC, Guren T, Tveit KM, Kure EH: Plasma microRNAs predicting clinical outcome in metastatic colorectal cancer patients receiving first line oxaliplatin-based treatment. *Mol Oncol* 2013. doi:10.1016/j.molonc.2013.09.001.

95. Wei J, Gao W, Zhu C, Liu YQ, Mei Z, Cheng T, et al: **Identification of plasma microRNA-21 as a biomarker for early detection and chemosensitivity of non-small cell lung cancer.** *Chin J Cancer* 2011, **30**:407–414.
96. Ohyashiki K, Umetsu T, Yoshizawa S, Ito Y, Ohyashiki M, Kawashima H, et al: **Clinical impact of down-regulated plasma miR-92a levels in non-Hodgkin's lymphoma.** *PLoS ONE* 2011, **6**:e16408.
97. Lucotti S, Rainaldi G, Evangelista M, Rizzo M: **Fludarabine treatment favors the retention of miR-485-3p by prostate cancer cells: implications for survival.** *Mol Cancer* 2013, **12**:52.
98. Zhang HL, Yang LF, Zhu Y, Yao XD, Zhang SL, Dai B, et al: **Serum miRNA-21: elevated levels in patients with metastatic hormone-refractory prostate cancer and potential predictive factor for the efficacy of docetaxel-based chemotherapy.** *Prostate* 2011, **71**:326–331.
99. Gamez-Pozo A, Anton-Aparicio LM, Bayona C, Borrega P, Gallegos Sancho MI, Garcia-Dominguez R, et al: **MicroRNA expression profiling of peripheral blood samples predicts resistance to first-line sunitinib in advanced renal cell carcinoma patients.** *Neoplasia* 2012, **14**:1144–1152.
100. Madhavan D, Zucknick M, Wallwiener M, et al: **Circulating miRNAs as surrogate markers for circulating tumor cells and prognostic markers in metastatic breast cancer.** *Clin Cancer Res* 2012, **18**:5972–5982.
101. Hu Z, Chen X, Zhao Y, et al: **Serum microRNA signatures identified in a genome-wide serum microRNA expression profiling predict survival of non-small-cell lung cancer.** *J Clin Oncol* 2010, **28**:1721–1726.
102. Xi Y, Nakajima G, Gavin E, et al: **Systematic analysis of microRNA expression of RNA extracted from fresh frozen and formalin-fixed paraffin-embedded samples.** *RNA* 2007, **13**:1668–1674.
103. Hutvagner G, Zamore PD: **RNAi: nature abhors a double-strand.** *Curr Opin Genet Dev* 2002, **12**:225–232.
104. Krutzfeldt J, Rajewsky N, Braich R, et al: **Silencing of microRNAs in vivo with 'antagomirs'.** *Nature* 2005, **438**:685–689.
105. Elmen J, Lindow M, Silahatoglu A, et al: **Antagonism of microRNA-122 in mice by systematically administered LNA-antimiR leads to up-regulation of a large set of predicted target mRNAs in the liver.** *Nucleic Acids Res* 2008, **36**:1153–1162.
106. Dickins RA, Hamann MT, Zilfou JT, Simpson DR, Ibarra I, Hannon GJ: **Probing tumor phenotypes using stable and regulated synthetic microRNA precursors.** *Nat Genet* 2005, **37**:1289–1295.
107. Tsuda N, Kawano K, Efferson CL, Ioannides CG: **Synthetic microRNA and double-strand RNA targeting the 3'-untranslated region of HER-2/neu mRNA inhibit HER-2 protein expression in ovarian cancer cells.** *Int J Oncol* 2005, **27**:1299–1306.
108. Liang Z, Wu H, Reddy S, Zhu A, Wang S, Blevins D: **Blockade of invasion and metastasis of breast cancer cells via targeting CXCR4 with an artificial microRNA.** *Biochem Biophys Res Commun* 2007, **363**:542–546.
109. Li L, Xie X, Luo J, Liu M, Xi S, Guo J, Kong Y, Wu M, Gao J, Xie Z, et al: **Targeted expression of mir-34a using the t-*visa* system suppresses breast cancer cell growth and invasion.** *Mol Ther* 2012, **20**:2326–2334.
110. Lee SJ, Kim SJ, Seo HH, Shin SP, Kim D, Park CS, Kim KT, Kim YH, Jeong JS, Kim IH: **Over-expression of mir-145 enhances the effectiveness of hsvtk gene therapy for malignant glioma.** *Cancer Lett* 2012, **320**:72–80.
111. Wang X, Han L, Zhang A, Wang G, Jia Z, Yang Y, Yue X, Pu P, Shen C, Kang C: **Adenovirus-mediated shRNAs for co-repression of mir-221 and mir-222 expression and function in glioblastoma cells.** *Oncol Rep* 2011, **25**:97–105.
112. Munoz JL, Bliss SA, Greco SJ, Ramkissoon SH, Ligon KL, Rameshwar P: **Delivery of functional anti-miR-9 by mesenchymal stem cell-derived exosomes to glioblastoma multiforma cells conferred chemosensitivity.** *Mol Ther Nucleic Acids* 2013, **2**:e126.
113. Kota J, Chivukula RR, O'Donnell KA, Wentzel EA, Montgomery CL, Hwang HW, Chang TC, Vivekanandan P, Torbenson M, Clark KR, et al: **Therapeutic microRNA delivery suppresses tumorigenesis in a murine liver cancer model.** *Cell* 2009, **137**:1005–1017.
114. Trang P, Medina PP, Wiggins JF, Ruffino L, Kelnar K, Omotola M, Homer R, Brown D, Bader AG, Weidhaas JB, et al: **Regression of murine lung tumors by the let-7 microRNA.** *Oncogene* 2010, **29**:1580–1587.
115. Trang P, Wiggins JF, Daige CL, Cho C, Omotola M, Brown D, Weidhaas JB, Bader AG, Slack FJ: **Systemic delivery of tumor suppressor microRNA mimics using a neutral lipid emulsion inhibits lung tumors in mice.** *Mol Ther* 2011, **19**:1116–1122.
116. Babar IA, Cheng CJ, Booth CJ, Liang X, Weidhaas JB, Saltzman WM, Slack FJ: **Nanoparticle-based therapy in an in vivo microRNA-155 (mir-155)-dependent mouse model of lymphoma.** *Proc Natl Acad Sci USA* 2012, **109**:E1695–E1704.
117. Murphy BL, Obad S, Bihannic L, Ayrault O, Zindy F, Kauppinen S, Roussel MF: **Silencing of the miR-17 92 cluster family inhibits medulloblastoma progression.** *Cancer Res* 2013, **73**:7068–7078.
118. Sicard F, Gayral M, Lulka H, Buscail L, Cordelier P: **Targeting miR-21 for the therapy of pancreatic cancer.** *Mol Ther* 2013, **21**:986–994.
119. Medina PP, Nolde M, Slack FJ: **OncomiR addiction in an in vivo model of microRNA-21-induced pre-B-cell lymphoma.** *Nature* 2010, **467**:86–90.
120. Meng F, Henson R, Lang M, et al: **Involvement of human micro-RNA in growth and response to chemotherapy in human cholangiocarcinoma cell lines.** *Gastroenterology* 2006, **130**:2113–2129.
121. Wong ST, Zhang XQ, Zhuang JT, Chan HL, Li CH, Leung GK: **MicroRNA-21 inhibition enhances in vitro chemosensitivity of temozolomide-resistant glioblastoma cells.** *Anticancer Res* 2012, **32**:2835–2841.
122. Li Y, Li L, Guan Y, Liu X, Meng Q, Guo Q: **Mir-92b regulates the cell growth, cisplatin chemosensitivity of A549 non small cell lung cancer cell line and target PTEN.** *Biochem Biophys Res Commun* 2013, **440**:604–610.
123. Weidhaas JB, Babar I, Nallur SM, et al: **MicroRNAs as potential agents to alter resistance to cytotoxic anticancer therapy.** *Cancer Res* 2007, **67**:11111–11116.
124. Pan X, Wang ZX, Wang R: **MicroRNA-21: a novel therapeutic target in human cancer.** *Cancer Biol Ther* 2011, **10**:1224–1232.
125. Iorio MV, Croce CM: **Causes and consequences of microRNA dysregulation.** *Cancer J* 2012, **18**:215–222.
126. Rodrigues AC, Li X, Radecki L, et al: **MicroRNA expression is differentially altered by xenobiotic drugs in different human cell lines.** *Biopharm Drug Dispos* 2011, **32**:355–367.
127. Shan G, Li Y, Zhang J, et al: **A small molecule enhances RNA interference and promotes microRNA processing.** *Nature Biotech* 2008, **26**:933–940.
128. Melo S, Villanueva A, Moutinho C, et al: **The small molecule enoxacin is a cancer-specific growth inhibitor that acts by enhancing TAR RNA-binding protein 2-mediated microRNA processing.** *Proc Natl Acad Sci USA* 2011, **108**:4394–4399.
129. Scott GK, Mattie MD, Berger CE, Benz SC, Benz CC: **Rapid alteration of microRNA levels by histone deacetylase inhibition.** *Cancer Res* 2006, **66**:1277–1281.
130. Liu X, Wang S, Meng F, et al: **SM2miR: a database of the experimentally validated small molecules' effects on microRNA expression.** *Bioinformatics* 2013, **29**:409–411.
131. Zhang S, Chen L, Jung EJ, Calin GA: **Targeting microRNAs with small molecules: from dream to reality.** *Clin Pharmacol Ther* 2010, **87**:754–758.
132. Jiang W, Chen Z, Liao M, Li W, et al: **Identification of links between small molecules and miRNAs in human cancers based on transcriptional responses.** *Sci Rep* 2012, **2**:282.
133. Grimm D, Streetz KL, Jopling CL, et al: **Fatality in mice due to oversaturation of cellular microRNA/short hairpin RNA pathways.** *Nature* 2006, **441**:537–541.

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