

TNF- α -308 polymorphism and risk of digestive system cancers: A meta-analysis

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Abstract

AIM: To evaluate the association between the tumour necrosis factor alpha-308 (*TNF- α -308*) gene polymorphism and the risk of digestive system cancers.

METHODS: All eligible case-control studies published up to December 2012 were identified by searching PubMed, Web of Science, Embase and China National Knowledge Internet without language restrictions. The risk of digestive system cancers associated with the *TNF- α -308* polymorphism was estimated for each study using odds ratio (OR) together with its 95%CI, respectively. Cochrane Collaboration RevMan 5.1 was used to perform the analysis. A χ^2 -test-based *Q* statistic test and an *I*² test were performed to assess the between-study heterogeneity. When the *Q* test was significant (*P* < 0.05) or *I*² > 50%, the random effects model was used, otherwise the fixed effects model was used.

RESULTS: Fifty-eight studies from fifty-five publications with a total of 9986 cancer patients and 15511

healthy controls were included. Overall, a significant association was found between the *TNF- α -308* polymorphism and the risk of digestive system cancers [dominant model: OR = 1.23, 95%CI: 1.09-1.39, (G/A) vs (G/G): OR = 1.15, 95%CI: 1.02-1.28, (A/A) vs (G/G): OR = 1.44, 95%CI: 1.19-1.73, recessive model: OR = 1.38, 95%CI: 1.15-1.66]. Furthermore, when the analysis was stratified by ethnicity, similar results were observed in both the Asian and Caucasian populations, except for the dominant model and heterozygote comparisons in the Asian population [dominant model: OR = 1.24, 95%CI: 0.99-1.56, (G/A) vs (G/G): OR = 1.09, 95%CI: 0.96-1.24]. When the cancer type subgroups were examined, similar results were detected in gastric and hepatocellular carcinomas; however, no significant association was observed among other digestive system cancers.

CONCLUSION: The *TNF- α -308* gene polymorphism may be significantly associated with the risk of gastric and hepatocellular carcinomas, but not colorectal, pancreatic, or oesophageal cancer, in the Asian population.

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Key words: Tumour necrosis factor alpha; rs1800629; Polymorphism; Digestive system cancer; Meta-analysis; Association

Core tip: Genetic polymorphisms contribute to the risk of human malignant tumours. Many studies have reported the relationship between the tumour necrosis factor alpha-308 (*TNF- α -308*) gene polymorphism and risk of digestive system cancers. However, the results of these studies are inconsistent and contradictory. In this meta-analysis, our results suggest that the *TNF- α -308* polymorphism is significantly associated with the risk of gastric and hepatocellular carcinomas in the Asian

population (dominant model: 95%CI: 1.02-1.34, $P < 0.05$ and 95%CI: 1.20-2.54, $P < 0.05$, respectively). This finding indicates that certain polymorphisms and mutations at *TNF- α -308* may increase susceptibility to digestive system cancers.

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INTRODUCTION

Digestive system cancers are the most common malignant tumours worldwide, with 3.4 million new cases each year, and their mortality rates have increased gradually over the past decade^[1,2]. Molecular epidemiology has confirmed that carcinogenesis is a complex, multifactorial and multistep event, in which the interaction of environmental triggers and genetic susceptibility may play an important role. However, the exact mechanism of carcinogenesis is still not fully understood.

Tumour necrosis factor-alpha (*TNF- α*), which is mainly produced by macrophages, is a multifunctional cytokine that plays an important role in the pathogenesis of inflammatory, autoimmune, and malignant diseases^[3]. The *TNF- α* gene is located in the major histocompatibility complex class III region on the short arm of chromosome six. Several polymorphisms in the promoter region of the *TNF- α* gene have been identified and are implicated in the regulation of *TNF- α* transcription^[4-5]. The *TNF- α -308* polymorphism (rs1800629) is the most extensively studied polymorphism in digestive system cancers^[6-9]. However, the results of the studies on *TNF- α -308* have been inconclusive or inconsistent. Therefore, we conducted a meta-analysis to evaluate the association between the *TNF- α -308* polymorphism and susceptibility to digestive system cancers.

MATERIALS AND METHODS

Search strategy

A literature search was conducted using PubMed, Web of Science, Embase and CNKI for studies that were published up to December 2012 without language restrictions. The relevant studies were identified using the following terms: ["tumour necrosis factor alpha or TNF alpha or TNF- α "] AND ["genetic polymorphism or polymorphisms or variant"] AND ["digestive system cancer or gastric cancer or colorectal cancer or hepatocellular carcinoma or pancreatic cancer or oesophageal cancer"]. The search was restricted to humans. Additional studies were identified by a manual search of references of original or review articles on this topic. If more

than one cancer type was reported in one study, the data for each type was extracted separately. If data or data subsets were published in more than one article, only the publication with the largest sample size was included.

Inclusion and exclusion criteria

Studies were included if they met the following criteria: (1) studies that evaluated the association between the *TNF- α -308* polymorphism and digestive system cancer risk; (2) studies with a case-control study design; and (3) studies with detailed genotype frequencies for cases and controls or text that allowed for the calculation of these values. The major exclusion criteria were: (1) case-only studies, case reports, or review articles; (2) studies without raw data for the *TNF- α -308G/A* genotype; and (3) studies that compared the *TNF- α -308G/A* variants in precancerous lesions and other cancers.

Data extraction and quality assessment

Two investigators (Guo XF and Wang J) independently extracted the data and reached a consensus on each item. If the two investigators generated different results, they would check the data again and have a discussion to come to an agreement. If they could not reach an agreement, an expert (Dong WG) was invited to the discussion. The data extracted from the selected articles included the first author's name, year of publication, country of origin, ethnicity, cancer type, genotyping methods, and number of cases and controls. The ethnicities were categorised as Asian or Caucasian. The cancer types were categorised as gastric, colorectal, hepatocellular, pancreatic, or oesophageal.

Statistical analysis

The meta-analysis was performed using the Cochrane Collaboration RevMan 5.1 software (Copenhagen, 2008). The association between the risk of digestive system cancers and the *TNF- α -308* polymorphism was estimated for each study using the odds ratio (OR) and 95%CI. A χ^2 test-based calculation of the Q statistic was performed to assess the between-study heterogeneity^[10]. We also quantified the effect of heterogeneity with an I^2 test. When the Q test was significant ($P < 0.05$) or $I^2 > 50\%$, indicating heterogeneity across studies, the random effects model was used^[11]; otherwise, the fixed effects model was used^[12]. Before estimating the relationship between the *TNF- α -308* polymorphism and digestive system cancer risk, we tested whether the genotype frequencies of the controls were in Hardy-Weinberg equilibrium (HWE) using a χ^2 test. We first estimated this relationship with the dominant model [G/A (GA) + A/A (AA) vs G/G (GG)] and the recessive model (AA vs GA + GG) and then with the co-dominant model (GA vs GG and AA vs GG). To evaluate the ethnicity-specific and cancer type-specific effects, we performed stratification analyses with respect to ethnicity and cancer type. Sensitivity analysis was performed to evaluate the stability of the results. Funnel plots were used to evaluate publication bias.

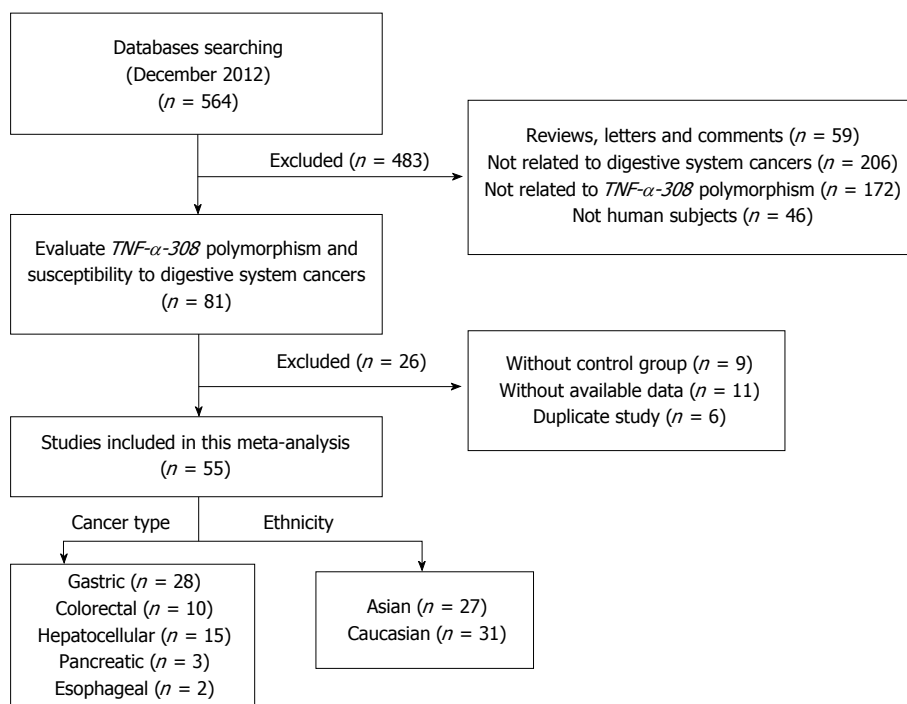


Figure 1 Flow chart showing study selection procedure. TNF- α : Tumour necrosis factor-alpha.

RESULTS

Study characteristics

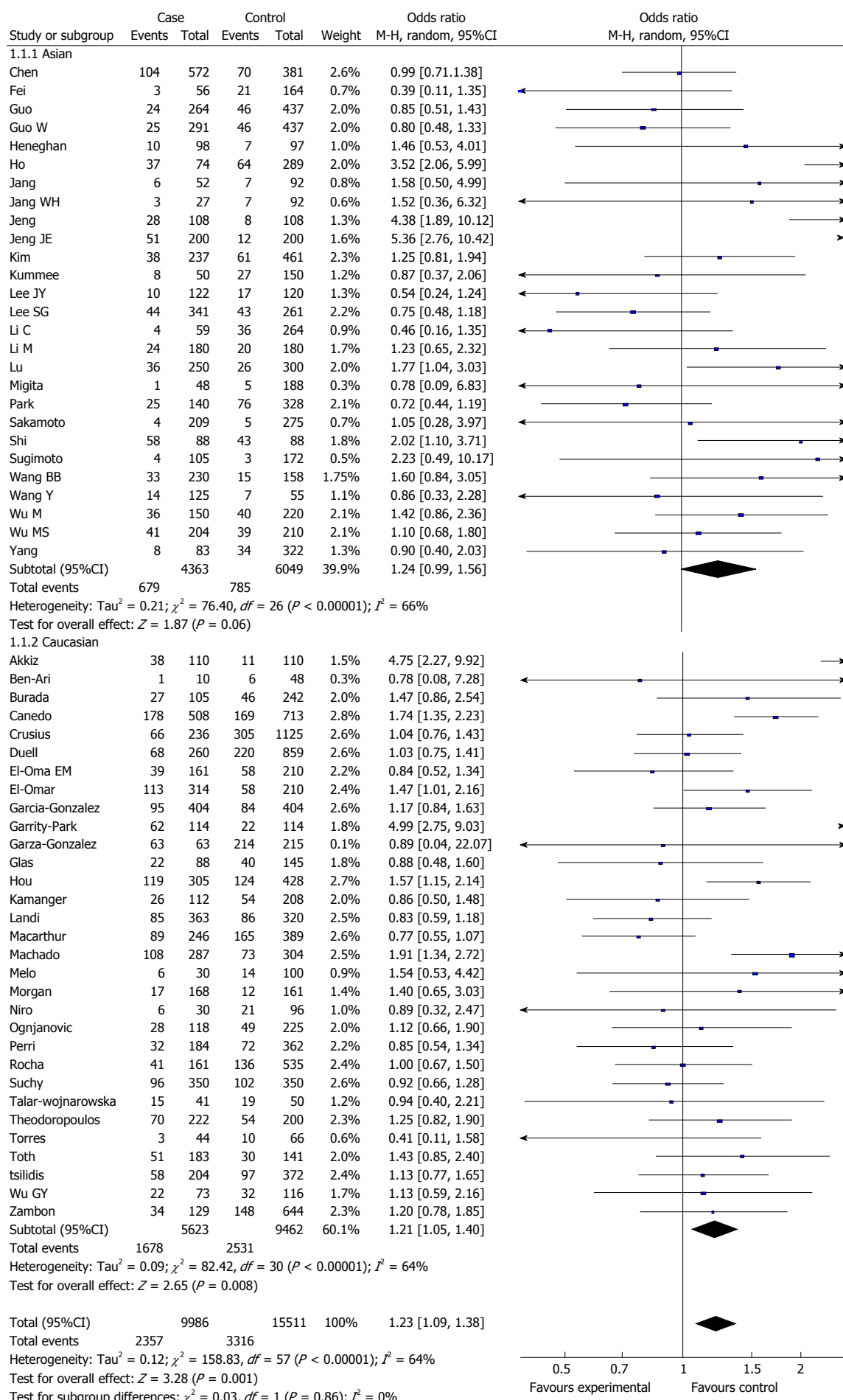
The search strategy retrieved 564 potentially relevant studies. According to the inclusion criteria, 55 studies with full-text were included in this meta-analysis and 509 studies were excluded. A flow chart of the study selection is shown in Figure 1. Because the studies of El-Omar *et al.*^[9], Guo *et al.*^[13] and Jang *et al.*^[14] each included separate analyses of two cancer types, we treated them separately in this meta-analysis^[9,13,14]. Therefore, as shown in Table 1, there were 58 case-control studies from 55 publications on the *TNF- α -308* polymorphism with a total of 9986 cancer cases and 15511 controls. Two ethnicities were addressed: 27 studies focused on Asian populations, and 31 studies focused on Caucasian populations. Five cancer types were addressed: 28 studies focused on gastric cancer^[6-9,13-36], 10 studies on colorectal cancer^[14,37-45], 15 studies on hepatocellular carcinoma^[46-60], 3 studies on pancreatic cancer^[61-63], and 2 studies on oesophageal cancer^[9,13]. The genotype distribution in the controls was consistent with HWE for all of the selected studies, except for four studies on gastric cancer^[7,13,33-34], one study on colorectal cancer^[43], six studies on hepatocellular carcinoma^[47,49,51-52,55-56], and one study on esophageal cancer^[13].

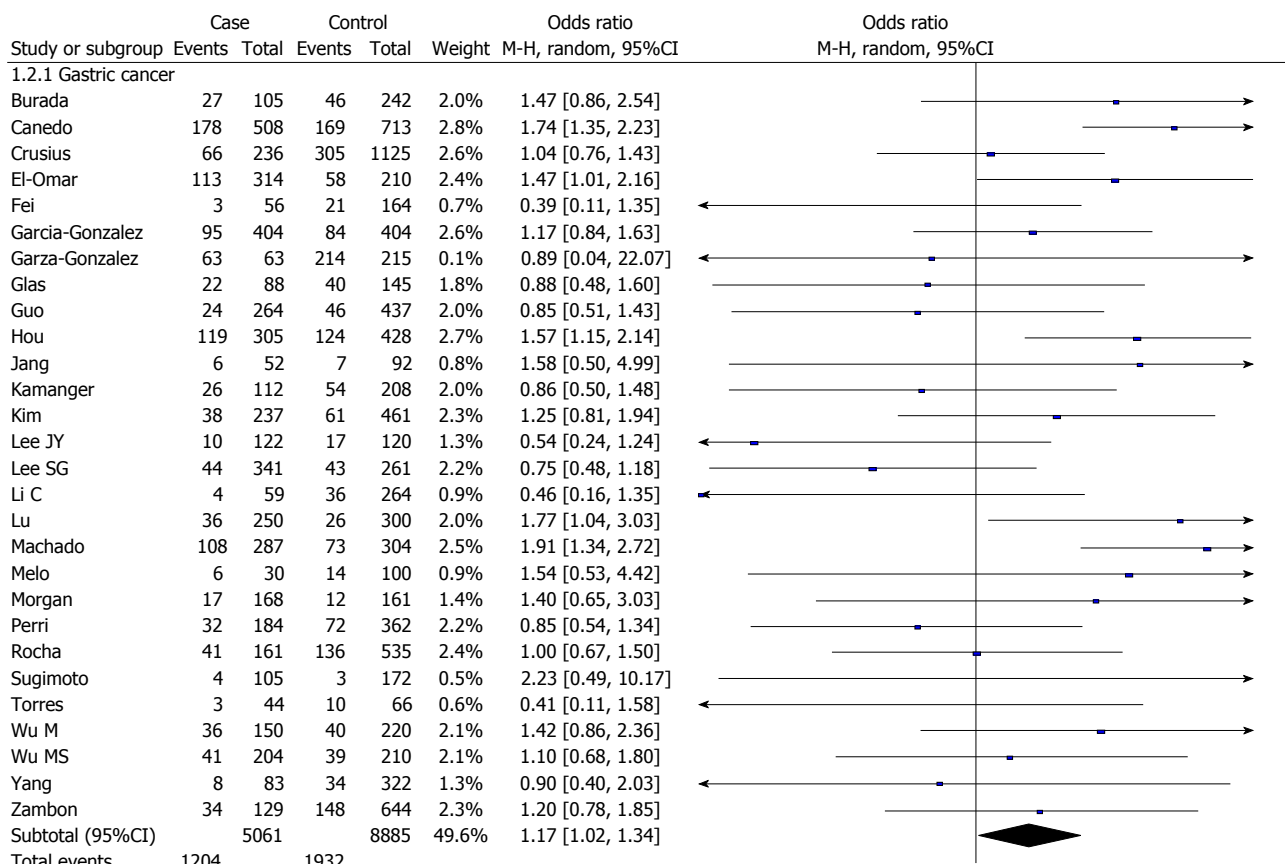
Quantitative data synthesis

Overall, there was a significant difference in the *TNF- α -308G/A* genotype distribution between the digestive system cancer patients and the controls (dominant model: OR = 1.23, 95%CI: 1.09-1.39, $P < 0.00001$; GA *vs* GG: OR = 1.15, 95%CI: 1.02-1.28, $P < 0.0001$; AA *vs* GG: OR = 1.44, 95%CI: 1.19-1.73, $P = 0.23$; recessive model:

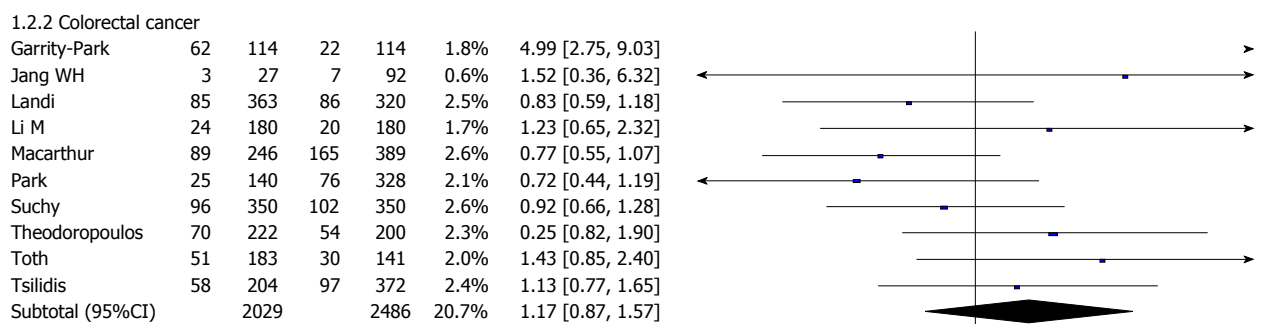
OR = 1.38, 95%CI: 1.15-1.66, $P = 0.50$) (Table 2, Figure 2). In the analysis of the ethnic subgroups, similar results were observed in the Caucasian population; but in the Asian population, we found that there was no significant association between the *TNF- α -308* polymorphism and the risk of digestive system cancers in the dominant model and heterozygote comparisons (GA + AA *vs* GG: OR = 1.24, 95%CI: 0.99-1.56, GA *vs* GG: OR = 1.09, 95%CI: 0.96-1.24) (Table 2, Figure 2). When stratified by cancer type, similar results were detected for gastric and hepatocellular carcinomas; however, no significant association was observed among the other digestive system cancer types (Table 2, Figure 3). Furthermore, we found that there was significant heterogeneity for the dominant model and heterozygote comparisons both overall and in the stratified analyses: $I^2 = 64\%$ and 52% in the overall population, $I^2 = 66\%$ and 45% ($P = 0.008$) in the Asian population, $I^2 = 64\%$ and 58% in the Caucasian population, $I^2 = 76\%$ and 70% in colorectal cancer, and $I^2 = 73\%$ and 66% in hepatocellular carcinoma. In addition, there was evidence of heterogeneity in gastric cancer (dominant model: $P = 0.009$). Thus, the random effects model was employed in the OR calculations. Then, sensitivity analyses were conducted to determine whether modification of the inclusion criteria of the meta-analysis affected the final results. We examined the influence of these studies on the pooled OR by repeating the meta-analysis while excluding the study that was not in HWE. The estimated pooled OR did not show a significant change (Table 2), indicating that our results are statistically robust. The shapes of the funnel plots did not reveal any evidence of asymmetry, suggesting that there was no publication bias among the studies (Figure 4).

Figure 2
Subgroup analysis of tumour necrosis factor α -308 polymorphism by ethnicity (dominant model).

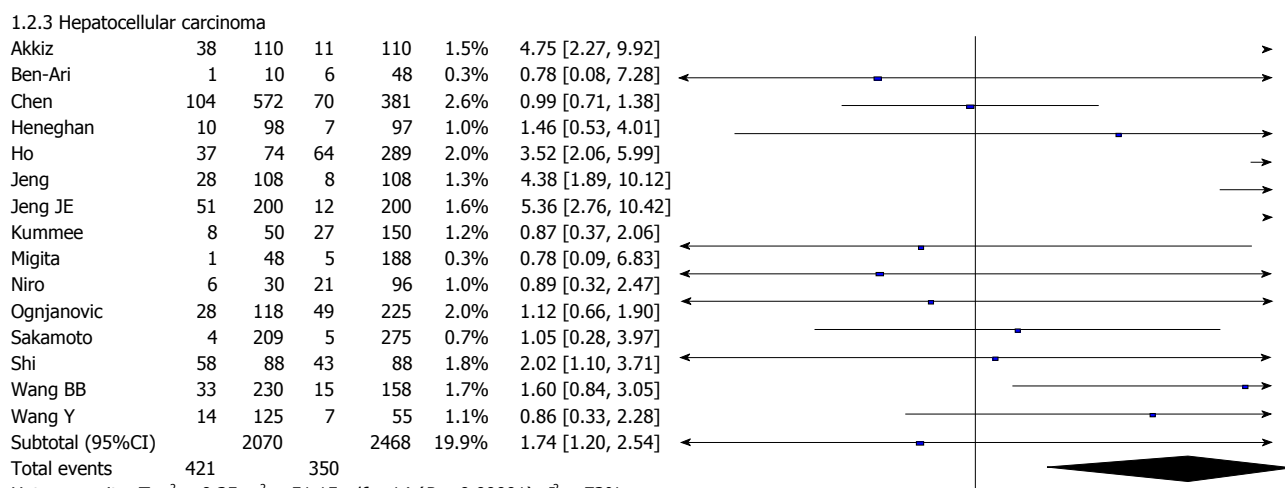




Heterogeneity: $\tau^2 = 0.05$; $\chi^2 = 47.60$, $df = 27$ ($P = 0.009$); $I^2 = 43\%$
 Test for overall effect: $Z = 2.32$ ($P = 0.02$)



Heterogeneity: $\tau^2 = 0.16$; $\chi^2 = 37.06$, $df = 9$ ($P < 0.00001$); $I^2 = 76\%$
 Test for overall effect: $Z = 1.02$ ($P = 0.31$)



Heterogeneity: $\tau^2 = 0.35$; $\chi^2 = 51.15$, $df = 14$ ($P < 0.00001$); $I^2 = 73\%$
 Test for overall effect: $Z = 2.89$ ($P = 0.004$)

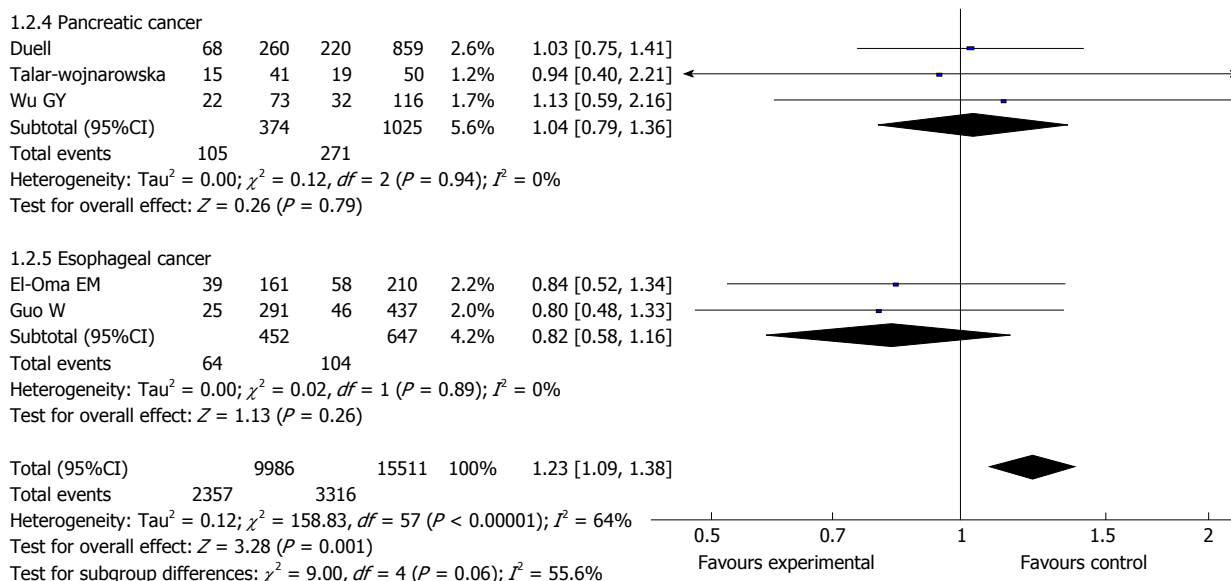


Figure 3 Subgroup analysis of tumor necrosis factor α -308 polymorphism by cancer type (dominant model).

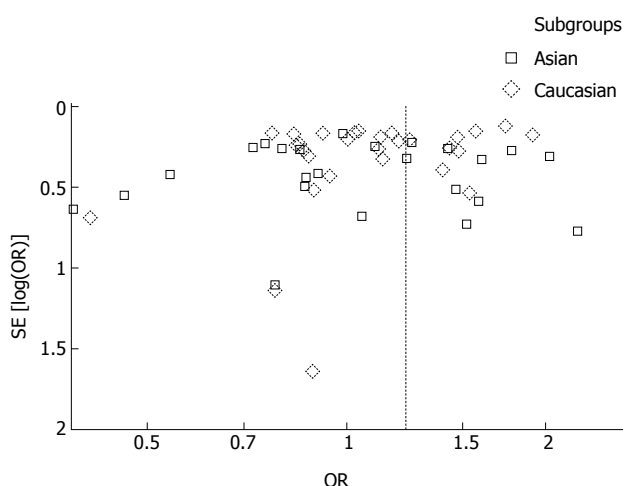


Figure 4 Funnel plots analysis to detect publication bias. Each point represents an independent study for the indicated association.

DISCUSSION

TNF, an important pro-inflammatory cytokine, plays an important role in the regulation of cell differentiation, proliferation and death as well as in inflammation and the innate and adaptive immune response. TNF has also been implicated in a wide variety of human diseases. The presence of DNA sequence variations in the regulatory region might interfere with transcription of the *TNF* gene, influencing the circulating level of TNF and thus increasing susceptibility to human diseases, such as cancer^[64]. The TNF enhancer polymorphism has been implicated in several diseases, and the *TNF- α -308* polymorphism has been described as the most important TNF polymorphism in human disease susceptibility. The significance of these polymorphisms reflects their possible influence on the transcription of the *TNF* gene. However, the results of studies in this area are inconsistent. Canedo *et al*^[7] found

that the *TNF- α -308G/A* polymorphism increases the risk of gastric carcinoma. However, some studies have reported that no statistically significant association exists between the *TNF- α -308G/A* polymorphism and cancer risk^[14,20].

The current meta-analysis, which included 58 case-control studies and 25497 subjects, was conducted to explore the association of the *TNF- α -308* polymorphism with digestive system cancer risk. Overall, a significant association was identified between the *TNF- α -308* polymorphism and the risk of digestive system cancers. When the analysis was stratified by ethnicity, we found a statistically significant association between this polymorphism and the risk of these cancers in the Caucasian population. However, no significant association was observed in the dominant model and heterozygote comparisons in the Asian population, which could be due to ethnic differences. When the analysis was stratified by cancer type, we found a significant association between this polymorphism and gastric and hepatocellular carcinoma risk under all four genetic models, but no significant association was observed among colorectal, pancreatic or oesophageal cancer.

Heterogeneity is a potential problem when interpreting the results of meta-analyses. In this meta-analysis, heterogeneity was found in the dominant model and heterozygote comparisons in both the overall and subgroup analyses; thus, the random effects model was used. Sensitivity analyses were also conducted by excluding the study that was not in HWE. With this exclusion, the estimated pooled OR did not change significantly, strengthening our confidence in our results. This finding suggests that the population selection and the study that was not in HWE were not sources of heterogeneity. Alternatively, lifestyle, environment and other unknown factors may be sources of heterogeneity. Moreover, no publication bias was shown, suggesting that our results

Table 1 Characteristics of studies included in the meta-analysis

Ref.	Year	Country	Ethnicity	Cancer type	Genotyping method	Case				Control				P
						Total	GG	GA	AA	Total	GG	GA	AA	
Burada <i>et al</i> ^[6]	2012	Romania	Caucasian	Gastric	TaqMan	105	78	26	1	242	196	44	2	0.78
Canedo <i>et al</i> ^[7]	2008	Portugal	Caucasian	Gastric	TaqMan	508	330	178 ¹		713	544	169 ¹		NA
Crusius <i>et al</i> ^[8]	2008	Spain	Caucasian	Gastric	Real-time PCR	236	170	64	2	1125	820	274	31	0.17
El-Omar <i>et al</i> ^[9]	2003	United States	Caucasian	Gastric	TaqMan	314	201	87	26	210	152	52	6	0.55
Guo <i>et al</i> ^[13]	2005	China	Asian	Gastric	PCR-RFLP	264	240	20	4	437	391	40	6	<0.01
Jang <i>et al</i> ^[14]	2001	South Korea	Asian	Gastric	PCR-RFLP	52	46	4	2	92	85	7	0	0.70
Fei <i>et al</i> ^[15]	2004	China	Asian	Gastric	PCR	56	53	3	0	164	143	20	1	0.74
Garcia-Gonzalez <i>et al</i> ^[16]	2007	Spain	Caucasian	Gastric	TaqMan	404	309	84	11	404	320	77	7	0.35
Garza-Gonzalez <i>et al</i> ^[17]	2005	Mexico	Caucasian	Gastric	PCR-RFLP	63	0	8	55	215	1	35	179	0.61
Glas <i>et al</i> ^[18]	2004	Germany	Caucasian	Gastric	PCR-RFLP	88	66	19	3	145	105	36	4	0.67
Hou <i>et al</i> ^[19]	2007	Poland	Caucasian	Gastric	TaqMan	305	186	98	21	428	304	109	15	0.19
Kamangar <i>et al</i> ^[20]	2006	Finland	Caucasian	Gastric	TaqMan	112	86	23	3	208	154	52	2	0.29
Kim <i>et al</i> ^[21]	2006	South Korea	Asian	Gastric	PCR-RFLP	237	199	34	4	461	400	59	2	0.91
Lee <i>et al</i> ^[22]	2004	South Korea	Asian	Gastric	PCR	341	297	43	1	261	218	42	1	0.49
Lee <i>et al</i> ^[23]	2005	South Korea	Asian	Gastric	PCR-RFLP	122	112	10	0	120	103	17	0	0.40
Li <i>et al</i> ^[24]	2005	China	Asian	Gastric	PCR-RFLP	59	55	4	0	264	228	34	2	0.56
Lu <i>et al</i> ^[25]	2005	China	Asian	Gastric	PCR-DHPLC	250	214	36	0	300	274	24	2	0.08
Machado <i>et al</i> ^[26]	2003	Portugal	Caucasian	Gastric	PCR-SSCP	287	179	105	3	304	231	69	4	0.65
Melo <i>et al</i> ^[27]	2009	Brazil	Caucasian	Gastric	PCR-RFLP	30	24	5	1	100	86	13	1	0.53
Morgan <i>et al</i> ^[28]	2006	Honduras	Caucasian	Gastric	TaqMan	168	151	17	0	161	149	12	0	0.62
Perri <i>et al</i> ^[29]	2005	Italy	Caucasian	Gastric	PCR-RFLP	184	152	30	2	362	290	65	7	0.15
Rocha <i>et al</i> ^[30]	2005	Brazil	Caucasian	Gastric	PCR-RFLP	161	120	37	4	535	399	123	13	0.34
Sugimoto <i>et al</i> ^[31]	2007	Japan	Asian	Gastric	PCR-RFLP	105	101	4	0	172	169	3	0	0.91
Torres <i>et al</i> ^[32]	2004	Colombia	Caucasian	Gastric	PCR	44	41	3	0	66	56	10	0	0.51
Wu <i>et al</i> ^[33]	2002	China	Asian	Gastric	Direct sequencing	150	114	27	9	220	180	27	13	<0.01
Wu <i>et al</i> ^[34]	2004	China	Asian	Gastric	Direct sequencing	204	163	29	12	210	171	26	13	<0.01
Yang <i>et al</i> ^[35]	2009	South Korea	Asian	Gastric	SNaPshot	83	75	8	0	322	288	34	0	0.32
Zamboni <i>et al</i> ^[36]	2005	Italy	Caucasian	Gastric	TaqMan	129	95	31	3	644	496	138	10	0.91
Garrity-Park <i>et al</i> ^[37]	2008	Ireland	Caucasian	Colorectal	PCR, sequencing	114	52	49	13	114	92	20	2	0.46
Jang <i>et al</i> ^[14]	2001	South Korea	Asian	Colorectal	PCR-RFLP	27	24	3	0	92	85	7	0	0.70
Landi <i>et al</i> ^[38]	2003	Spain	Caucasian	Colorectal	TaqMan	363	278	80	5	320	234	76	10	0.22
Li M <i>et al</i> ^[39]	2011	China	Asian	Colorectal	PCR-RFLP	180	156	15	9	180	160	19	1	0.60
Macarthur <i>et al</i> ^[40]	2005	Scotland	Caucasian	Colorectal	TaqMan	246	157	74	15	389	224	145	20	0.58
Park <i>et al</i> ^[41]	1998	South Korea	Asian	Colorectal	PCR-RFLP	140	115	24	1	328	252	72	4	0.65
Suchy <i>et al</i> ^[42]	2008	Poland	Caucasian	Colorectal	PCR-RFLP	350	254	87	9	350	248	95	7	0.55
Theodoropoulos <i>et al</i> ^[43]	2006	Greece	Caucasian	Colorectal	PCR-RFLP	222	152	56	14	200	146	44	10	0.01
Toth <i>et al</i> ^[44]	2007	Hungary	Caucasian	Colorectal	PCR-SSP	183	132	48	3	141	111	30	0	0.16
Tsilidis <i>et al</i> ^[45]	2009	United States	Caucasian	Colorectal	TaqMan	204	146	55	3	372	275	90	7	0.91
Akkiz <i>et al</i> ^[46]	2009	Turkey	Caucasian	Hepatocellular	PCR-RFLP	110	72	35	3	110	99	11	0	0.58
Ben-Ari <i>et al</i> ^[47]	2003	United States	Caucasian	Hepatocellular	PCR-SSP	10	9	1 ¹		48	42	6 ¹		NA
Chen <i>et al</i> ^[48]	2005	China	Asian	Hepatocellular	TaqMan	572	468	95	9	381	311	67	3	0.77
Heneghan <i>et al</i> ^[49]	2003	China	Asian	Hepatocellular	ASO-PCR	98	88	10	0	97	90	6	1	0.03
Ho <i>et al</i> ^[50]	2004	China	Asian	Hepatocellular	PCR-RFLP	74	37	34	3	289	225	62	2	0.30
Jeng <i>et al</i> ^[51]	2007	China	Asian	Hepatocellular	PCR-SSO	108	80	28 ¹		108	100	8 ¹		NA
Jeng JE <i>et al</i> ^[52]	2009	China	Asian	Hepatocellular	PCR-SSO	200	149	51 ¹		200	188	12 ¹		NA
Kummee <i>et al</i> ^[53]	2007	Thailand	Asian	Hepatocellular	PCR-RFLP	50	42	8	0	150	123	26	1	0.77
Migita <i>et al</i> ^[54]	2005	Japan	Asian	Hepatocellular	PCR-SSP	48	47	1	0	188	183	5	0	0.85
Niro <i>et al</i> ^[55]	2005	Italy	Caucasian	Hepatocellular	Direct sequencing	30	24	6 ¹		96	75	21 ¹		NA
Ognjanovic <i>et al</i> ^[56]	2009	United States	Caucasian	Hepatocellular	TaqMan	118	90	28 ¹		225	176	49 ¹		NA
Sakamoto <i>et al</i> ^[57]	2008	Japan	Asian	Hepatocellular	PCR-RFLP	209	205	4	0	275	270	5	0	0.88
Shi <i>et al</i> ^[58]	2011	China	Asian	Hepatocellular	PCR-RFLP	88	30	43	15	88	45	35	8	0.75
Wang <i>et al</i> ^[59]	2003	Japan	Asian	Hepatocellular	Direct sequencing	125	111	13	1	55	48	6	1	0.16
Wang <i>et al</i> ^[60]	2010	China	Asian	Hepatocellular	PCR-SSO	230	197	30	3	158	143	15	0	0.53
Duell <i>et al</i> ^[61]	2006	United States	Caucasian	Pancreatic	PCR-RFLP	260	192	63	5	859	639	198	22	0.16
Talor-wojnarowska <i>et al</i> ^[62]	2009	Poland	Caucasian	Pancreatic	PCR-RFLP	41	26	12	3	50	31	17	2	0.86
Wu GY <i>et al</i> ^[63]	2010	Germany	Caucasian	Pancreatic	PCR-RFLP	73	51	20	2	116	84	30	2	0.72
El-Omar <i>et al</i> ^[9]	2003	United States	Caucasian	Esophageal	TaqMan	161	122	34	5	210	152	52	6	0.55
Guo <i>et al</i> ^[13]	2005	China	Asian	Esophageal	PCR-RFLP	291	266	21	4	437	391	40	6	<0.01

¹Numbers of GA+AA. P_{HWE} was calculated by goodness-of fit χ^2 -test, and $P_{HWE} < 0.05$ was considered statistically significant. PCR-DHPLC: Polymerase chain reaction-based denaturing high-performance liquid chromatography; HWE: Hardy-Weinberg equilibrium; NA: Not available; GG: Guanine/Guanine; GA: Guanine/Adenine; AA: Adenine/Adenine; PCR-RFLP: Polymerase chain reaction-restriction fragment length polymorphism.

are accurate.

Some limitations of this meta-analysis should be ad-

ressed. First, the number of published studies, especially for oesophageal and pancreatic cancers, was not suf-

Table 2 Stratified analysis of the tumor necrosis factor alpha polymorphism and digestive system cancers risk

Group	GA + AA vs GG			GA vs GG			AA vs GG			AA vs GA + GG		
	n	OR (95%CI)	P ¹	n	OR (95%CI)	P ¹	n	OR (95%CI)	P ¹	n	OR (95%CI)	P ¹
Overall	58	1.23 (1.09, 1.38) ²	< 0.00001	52	1.14 (1.01, 1.28) ²	< 0.00001	44	1.43 (1.19, 1.73)	0.26	44	1.38 (1.15, 1.66)	0.55
Studies with HWE	46	1.18 (1.03, 1.34) ²	< 0.00001	46	1.14 (1.00, 1.29) ²	< 0.00001	38	1.54 (1.25, 1.90)	0.15	38	1.48 (1.20, 1.81)	0.40
Cancer type												
Gastric	28	1.23 (1.12, 1.34) ²	0.009	27	1.15 (1.04, 1.27)	0.07	22	1.38 (1.06, 1.80)	0.63	22	1.33 (1.03, 1.72)	0.67
Colorectal	10	1.17 (0.87, 1.57) ²	< 0.0001	10	1.10 (0.83, 1.45) ²	0.0004	9	1.45 (0.76, 2.75) ²	0.02	9	1.40 (0.99, 2.00)	0.07
Hepatocellular	15	1.74 (1.20, 2.54) ²	< 0.00001	10	1.58 (1.05, 2.39) ²	0.002	8	2.55 (1.38, 4.70)	0.49	8	2.15 (1.19, 3.90)	0.66
Pancreatic	3	1.04 (0.79, 1.36)	0.94	3	1.04 (0.79, 1.38)	0.88	3	0.99 (0.46, 2.14)	0.63	3	0.99 (0.46, 2.13)	0.60
Esophageal	2	0.82 (0.58, 1.16)	0.89	2	0.80 (0.55, 1.15)	0.89	2	1.01 (0.42, 2.43)	0.95	2	1.05 (0.44, 2.51)	0.92
Ethnicity												
Asian	27	1.24 (0.99, 1.56) ²	< 0.00001	25	1.07 (0.94, 1.22) ²	0.008	19	1.55 (1.11, 2.17)	0.43	19	1.47 (1.05, 2.06)	0.60
Caucasian	31	1.21 (1.05, 1.40) ²	< 0.00001	27	1.17 (1.01, 1.35) ²	< 0.0001	25	1.38 (1.10, 1.74)	0.18	25	1.34 (1.08, 1.67)	0.39

¹Test for heterogeneity; ²Random-effects model was used when the P for heterogeneity test was < 0.05. GG: Guanine/Guanine; GA: Guanine/Adenine; AA: Adenine/Adenine; HWE: Hardy-Weinberg equilibrium.

ficiently large for a comprehensive analysis, and some studies with small sample sizes may not have enough statistical power to prove authentic associations. Therefore, our analysis should be interpreted with caution, and more studies are needed. Second, our results were based on unadjusted estimates, and lack of information for the data analysis may cause serious confounding bias. Third, significant heterogeneity was found in some models, which may lead to failure to confirm marginal associations. In spite of these limitations, our meta-analysis had several advantages. First, a substantial number of cases and controls were pooled from different studies, which significantly increased the statistical power of the analysis. Second, the quality of the case-control studies included in the current meta-analysis was satisfactory and met our inclusion criteria. Third, we did not detect any publication bias, suggesting that the whole pooled result is unbiased.

In summary, this meta-analysis suggests that the *TNF- α -308* polymorphism increases susceptibility to digestive system cancers in the Caucasian population. The *TNF- α -308 AA* genotype is closely related to the risk of digestive system cancers in people of Asian descent. The *TNF- α -308* polymorphism may be significantly associated with the risk of gastric and hepatocellular carcinomas, but not colorectal, pancreatic, or oesophageal cancer. Future studies should use standardised unbiased genotyping methods, examine homogeneous cancer patients and well-matched controls, and include multiethnic groups.

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COMMENTS

Background

Digestive system cancers are the most common malignant tumors worldwide. Tumor necrosis factor alpha-308 (*TNF- α -308*) polymorphism (rs1800629) is the most extensively studied polymorphism in digestive system cancers. However,

the results are different or even inconsistent.

Research frontiers

Molecular epidemiology has confirmed that carcinogenesis is a complex, multifactorial, and multistep event, and genetic mutation play an important role in the process. Many studies have reported the association between the *TNF- α -308* polymorphism and human malignant tumors, but no agreements have been reached till now.

Innovations and breakthroughs

This meta-analysis systemically assessed the association between *TNF- α -308* polymorphism and risk of digestive system cancers. Results show that *TNF- α -308* polymorphism may be significantly associated with the risk of gastric and hepatocellular carcinomas in Asians.

Applications

This study results indicate that *TNF- α -308* polymorphism may be used as a detectable biomarker for gastric and hepatocellular carcinoma patients.

Peer review

The authors present a meta-analysis study over the influence of a polymorphism of *TNF- α* on digestive system cancers. The manuscript is well written and interesting, especially because it is the first meta-analysis study on the subject.

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