

NIH Public Access

Author Manuscript

Adv Exp Med Biol. Author manuscript; available in PMC 2014 January 08.

Adv Exp Med Biol. 2008; 614: . doi:10.1007/978-0-387-74911-2_36.

Muscle Oxygen Uptake Differs from Consumption Dynamics During Transients in Exercise

Nicola Lai^{1,3}, Nakisha Syed¹, Gerald M. Saidel^{1,3}, and Marco E. Cabrera^{1,2,3}

¹Department of Biomedical Engineering, Case Western Reserve University, Cleveland, OH 44106, USA

²Department of Pediatrics, Center for Modeling Integrated Metabolism Systems, Case Western Reserve University, Cleveland, OH 44106, USA

³Rainbow Babies Children's Hospital, Case Western Reserve University, Cleveland, OH 44106, USA

Abstract

Relating external to internal respiration during exercise requires quantitative modeling analysis for reliable inferences with respect to metabolic rate. Often, oxygen transport and metabolism based on steady-state mass balances (Fick principle) and passive diffusion between blood and tissue are applied to link pulmonary to cellular respiration. Indeed, when the work rate does not change rapidly, a quasi-steady-state analysis based on the Fick principle is sufficient to estimate the rate of O₂ consumption in working muscle. During exercise when the work rate changes quickly, however, non-invasive *in vivo* measurements to estimate muscle O₂ consumption are not sufficient to characterize cellular respiration of working muscle. To interpret transient changes of venous O₂ concentration, blood flow, and O₂ consumption in working muscle, a mathematical model of O₂ transport and consumption based on dynamic mass balances is required. In this study, a comparison is made of the differences between simulations of O₂ uptake and O₂ consumption within working skeletal muscle based on a dynamic model and quasi-steady-state approximations. The conditions are specified under which the quasi-steady-state approximation becomes invalid.

36.1 Introduction

During exercise, oxygen transport and metabolism within muscle in healthy and disease states (e.g., heart failure and diabetes) can be studied under various experimental protocols with non-invasive measurements. Measurement methods include pulmonary O_2 uptake by indirect calorimetry, muscle oxygenation by near-infrared spectroscopy and microvascular oxygenation by phosphorescence quenching. Muscle O_2 consumption (UO_{2m}) during a fast transient change (e.g., exercise) cannot be directly evaluated by measuring oxygen uptake in the lungs (VO_{2p}) . Consequently, a mathematical model is needed to relate these variables. Often, a quasi-steady-state model (e.g., Fick principle) is used to analyze dynamic responses at the onset of exercise in order to quantify relationships between the oxygen uptake, blood flow, and O2 concentration dynamics in the capillary bed (or index of extraction) within muscle [1-3]. Although this method is strictly applicable only under steady-state conditions [4], it could provide a reasonable approximation under some conditions [5]. More generally, interpretation of non-invasive measurements related to O₂ transport and metabolism in tissue can be made using dynamic mass balances if the tissue volume can be determined. Nevertheless, this has limitations also. At the microvascular level, the volume of tissue supplied with oxygen by the blood vessels is uncertain. Furthermore, in a macroscopic tissue volume, the heterogeneous spatial distribution and temporal variation of blood flow and O_2 concentration in tissue can have a significant effect on the interpretation of measurements.

In this study, simulations using a quasi-steady-state model [3] and those using a multicompartment dynamic model [6] are compared to experimental measurements during exercise. The effect of changes in muscle blood flow on the dynamic responses of venous oxygen concentration was investigated during exercise assuming specific dynamics of muscle O_2 consumption.

36.2 Methods

We consider oxygen transport and consumption in muscle to occur in a system of perfectly mixed blood and tissue compartments (Fig. 36.1) as developed previously [6]. In the blood compartment, the oxygen concentration C(t) changes with time depending on flow $Q_m(t)$ through the capillary bed of volume V_{cap} and diffusion between blood and tissue with rate coefficient *PS* according to the dynamic mass balance:

$$V_{cap} \frac{dC_{ven}^{T}}{dt} = Q_m(t) (C_{art}^{T} - C_{ven}^{T}) - PS(C_{ven}^{F} - C_{tis}^{F}) \quad (36.1)$$

where C_j^T and C_j^F represent the total and free oxygen concentrations in compartment *j*, which are related by nonlinear equations [6]. In the tissue compartment, the oxygen concentration C(t) changes with time depending on the rates of diffusion between blood and tissue cells and oxygen consumption in muscle $UO_{2m}(t)$:

$$V_{tis} \frac{dC_{tis}^{T}}{dt} = PS(C_{ven}^{F} - C_{tis}^{F}) - UO_{2m}(t) \quad (36.2)$$

At steady state, these equations can be combined to yield the Fick principle for oxygen uptake of skeletal muscle:

$$VO_{2m} = Q_m (C_{art}^T - C_{ven}^T) = PS (C_{ven}^F - C_{tis}^F) = UO_{2m}$$
 (36.3)

In response to a step increase in work rate, the oxygen consumption has typically an exponential response [3]:

$$UO_{2m}(t) = UO_{2m}^{BL} + \Delta UO_{2m} \left[1 - \exp^{t_0 - t/\tau_{UO_{2m}}} \right] \quad (36.4)$$

Also, the blood flow has two phases (I, II) with different amplitudes and time constants [3]:

$$Q_m(t) = Q_m^{BL} + \Delta Q_{m,1} \left[1 - \exp^{(t_0 - t)/\tau_{Q_{m,l}}} \right] + \Delta Q_{m,2} \left[1 - \exp^{(t_0 + TD_2 - t)/\tau_{Q_{m,11}}} \right]$$
(36.5)

To simulate the oxygen concentration dynamics of skeletal muscle to a step change in work rate from baseline (*BL*), we must specify initial conditions at t_0 :

 $C_{ven}^{F}(t_0) = C_{ven}^{F,BL}; C_{tis}^{F,BL}(t_0) = C_{tis}^{F,Bl}$. Responses of venous oxygen concentration were simulated between two steady states assuming different values of muscle volume (V_m) engaged during exercise with different dynamic changes of blood flow in muscle. The differential equations of the model were solved numerically using a robust algorithm for stiff systems [7].

Page 3

For comparison with the results of Ferreira [3], we used the same initial conditions, muscle blood flows ($\tau_{Qm,I}$ and $\tau_{Qm,II}$), and oxygen consumption time constant $\tau UO_{2m} = 30$ s). Except for the parameter values in Table 36.1, values of other model parameters were obtained from previous studies [6].

Following Ferreira [3] we chose values for the arterial oxygen concentration, C_{art}^T , and free oxygen concentration $C_{ven}^{F,BL}$. The free oxygen concentration intissue was determined based on PO₂=25 mmHg. The value of *PS* was computed from Eq. (36.3) at steady state.

36.3 Results

The effects of various blood flow time profiles on venous oxygen concentration for a specific oxygen consumption dynamics are simulated.

Figure 36.2 shows dynamic changes in phase I of muscle blood flow (Fig. 36.2a) and corresponding dynamic responses of venous oxygen concentration (Fig. 36.2b). The dynamic response of the venous oxygen concentration is faster when the transient term (i.e., rate of oxygen change) is negligible ($V_m \approx 0L$) than when it is significant ($V_m = 15L$). The overshoot of the venous oxygen concentration is greater with a smaller muscle volume and with a shorter time constant of phase I ($\tau_{Qm,I}$). Even when the time constant of phase I is long, the dynamics of the venous oxygen concentration depends on the muscle volume.

Figure 36.3 shows dynamic changes in phase II of muscle blood flow (Fig. 36.3a) and corresponding dynamic responses of venous oxygen concentration (Fig. 36.3b). The dynamic response of the venous oxygen concentration is faster when the transient term is negligible ($V_m \approx 0L$) than when the transient term is significant ($V_m = 15L$). The undershoot of the venous oxygen concentration is greater with a smaller muscle volume and with a longer time constant of phase II ($\tau_{Qm,1I}$). Even when the time constant of phase II is small, the dynamics of the venous oxygen concentration depends on the muscle volume.

36.4 Discussion

Model simulations of the time response of venous oxygen concentration in working skeletal muscle were obtained with a dynamic computational model for quasi-steady-state (e.g., negligible muscle volume) and transient conditions. The simulations under quasi-steady-state conditions are equivalent to those of Ferreira [3], who applied a steady-state (Fick principle) analysis.

With a larger muscle volume, a change of muscle blood flow has less effect on the time profile of the venous oxygen concentration. In any case, an overshoot in venous oxygen concentration can occur during phase I of muscle blood flow as reported in human exercise studies [8]. Corresponding to simulations in venous oxygen concentration during phase II of muscle blood flow, experiments with rat muscle contractions show a similar undershoot response [9]. In these studies with diabetic rats where the disease induces a mismatch between oxygen delivery and oxygen consumption, an undershoot can occur in microvascular O_2 pressure at the onset of exercise.

Based on the Fick principle, the red blood cell flux (or oxygen delivery) and microvascular O_2 pressure measurements are used to compute oxygen consumption in the diabetic state [2]. This simplified analysis shows a mismatch between oxygen delivery and oxygen consumption, which accounts for the observed undershoot in microvascular O_2 pressure. Correct interpretation of this mismatch requires quantitative analysis with a more general dynamic model to determine the effect of muscle volume in the transient term of the oxygen

Adv Exp Med Biol. Author manuscript; available in PMC 2014 January 08.

balance. Furthermore, a dynamic model is essential to analyze the response of microvascular oxygen pressure in muscle at the onset of contraction in heart failure [10] and aging [11], where O_2 delivery is impaired.

Generally, venous oxygen concentration during exercise depends on the rate of oxygen mass accumulation within muscle that results from a dynamic interplay of convection, diffusion, and metabolism. Consequently, the dynamics of muscle oxygen uptake and muscle oxygen consumption differ during transient changes in exercise [6]. In this regard the extent of muscle involvement (i.e., muscle volume), which is often uncertain (especially at the microvascular level), plays a relevant role during exercise.

The more general dynamic model applied in this study consists of spatially lumped, dynamic mass-balance equations. A special case of this dynamic model is the quasi-steady-state-model, which is commonly used to analyze oxygen exchange in capillary blood and in tissue of the working muscle. Although this dynamic model of oxygen consumption in skeletalmuscle [6] is sufficient for some purposes, modifications are needed to reflect more physiological conditions. For example, the product of permeability and surface area should be a function of blood flow due to the capillary recruitment occurring at the onset of exercise [12]. Furthermore, this dynamic model assumes an exponential function to describe cellular oxygen consumption [13]. While this simple expression is sufficient to make some inferences about dynamic responses, key metabolic processes should be incorporated into the model to provide a mechanistic basis for oxygen consumption dynamics. For this purpose, future models should incorporate substrates and enzymes participating in mitochondrial oxidative phosphorylation during exercise.

The dynamics of the oxygen concentration in blood depend on the spatial distribution and temporal variation of the variables such as blood flow and hemoglobin oxygen saturation that affect convective and diffusive transport of oxygen in the micro circulation. Although these effects are not directly measurable during muscle contraction [14], more general models have been applied to account for heterogeneities of blood flow and oxygen consumption of the muscle [15–18].

In conclusion, physiological relations between oxygen transport and consumption within skeletal muscle during exercise require a model based on dynamic mass balances for oxygen in blood and tissue. Such a model can be used together with non-invasive or minimally invasive experiments to study capillary oxygen exchange during an exercise stimulus where active muscle, convection and diffusion have a significant effect. This could contribute to quantifying changes associated with aging in healthy subjects, as well as with potential pathological alterations of oxygen transport and metabolism in unhealthy subjects suffering from diabetes and heart failure [19].

Acknowledgments

Supported by grant (P50 GM-66309) from the National Institute of General Medical Sciences (NIH).

References

- Barstow TJ, Lamarra N, Whipp BJ. Modulation of muscle and pulmonary O₂ uptakes by circulatory dynamics during exercise. J. Appl. Physiol. 1990; 68:979–989. [PubMed: 2341363]
- Behnke BJ, Barstow TJ, Kindig CA, McDonough P, Musch TI, Poole DC. Dynamics of oxygen uptake following exercise onset in rat skeletal muscle. Respir. Physiol. Neurobiol. 2002b; 133:229– 239. [PubMed: 12425970]

- Ferreira LF, Poole DC, Barstow TJ. Muscle blood flow-O₂ uptake interaction and their relation to on-exercise dynamics of O₂ exchange. Respir. Physiol. Neurobiol. 2005; 147:91–103. [PubMed: 15848127]
- Stringer WW, Whipp BJ, Wasserman K, Porszasz J, Christenson P, French WJ. Non-linear cardiac output dynamics during ramp-incremental cycle ergometry. Eur. J. Appl. Physiol. 2005; 93:634– 639. [PubMed: 15578205]
- 5. Kemp G. Kinetics of muscle oxygen use, oxygen content and blood flow during exercise. J. Applied Physiology. 2005; 99:2463–2469.
- Lai N, Dash RK, Nasca MM, Saidel GM, Cabrera ME. Relating pulmonary oxygen uptake to muscle oxygen consumption at exercise onset: in vivo and in silico studies. Eur. J. Appl. Physiol. 2006; 97(4):380–394. [PubMed: 16636861]
- 7. Hindmarsh AC. A systematized collection of ode solvers. Scientific computing. 1983:55–64. DLSODE, http://www.netlib.org/odepack/.
- Grassi B, Poole DC, Richardson RS, Knight DR, Erickson BK, Wagner PD. Muscle O₂ uptake kinetics in humans: implications for metabolic control. J. Appl. Physiol. 1996; 80:988–998. [PubMed: 8964765]
- Behnke BJ, Kindig CA, McDonough P, Poole DC, Sexton WL. Dynamics of microvascular oxygen pressure during rest-contraction transition in skeletal muscle of diabetic rats. Am. J. Physiol. Heart Circ. Physiol. 2002a; 283:H926–H932. [PubMed: 12181120]
- Diederich ER, Behnke BJ, McDonough P, Kindig CA, Barstow TJ, Poole DC, Musch TI. Dynamics of microvascular oxygen partial pressure in contracting skeletal muscle of rats with chronic heart failure. Cardiovasc. Res. 2002; 56:479–486. [PubMed: 12445889]
- Behnke BJ, Delp MD, Dougherty PJ, Musch TI, Poole DC. Effects of aging on microvascular oxygen pressures in rat skeletal muscle. Respir. Physiol. Neurobiol. 2005; 146:259–268. [PubMed: 15766914]
- Caldwell JH, Martin GV, Raymond GM, Bassingthwaighte JB. Regional myocardial flow and capillary permeability-surface area products are nearly proportional. Am. J. Physiol. Heart Circ. Physiol. 1994; 267:H654–H666.
- Binzoni T, Colier W, Hiltbrand E, Hoofd L, Cerretelli P. Muscle O₂ consumption by NIRS: a theoretical model. J. Appl. Physiol. 1999; 87:683–688. [PubMed: 10444628]
- Pittman RN. Oxygen supply to contracting skeletal muscle at the microcirculatory level: diffusion vs. convection. Acta Physiol. Scand. 2000; 168:593–602. [PubMed: 10759595]
- 15. Beard DA. Computational framework for generating transport models from databases of microvascular anatomy. Ann. Biomed. Eng. 2001; 29:837–843. [PubMed: 11764314]
- Beard DA, Schenkman KA, Feigl EO. Myocardial oxygenation in isolated hearts predicted by an anatomically realistic microvascular transport model. Am. J. Physiol. Heart Circ. Physiol. 2003; 285:H1826–H1836. [PubMed: 12869375]
- Dash RK, Bassingthwaighte JB. Simultaneous blood–tissue exchange of oxygen, carbon dioxide, bicarbonate and hydrogen Ion. Ann. Biomed. Eng. 2006; 34:1129–1148. [PubMed: 16775761]
- Popel AS. Theory of oxygen transport to tissue. Crit. Rev. Biomed. Eng. 1989; 17:257–321. [PubMed: 2673661]
- Poole DC, Behnke BJ, Padilla DJ. Dynamics of muscle microcirculatory oxygen exchange. Med. Sci. Sports Exerc. 2005; 37:1559–1566. [PubMed: 16177609]

Lai et al.



Fig. 36.1.

Oxygen consumption and transport in skeletal muscle.

Adv Exp Med Biol. Author manuscript; available in PMC 2014 January 08.

Lai et al.



Fig. 36.2.

(a) Blood flow dynamic for different time constants $\tau_{Qm,I}$, ($\tau_{Qm,II} = 30$ s); (b) Effect of blood flow dynamic on the dynamic response of the venous O₂ concentration obtained with quasi-steady-state and dynamic model.

Lai et al.



Fig. 36.3.

(a) Blood flow dynamic for different time constants $\tau_{Qm,II}$, ($\tau_{Qm,I} = 4$ s); (b) Effect of blood flow dynamic on the dynamic response of the venous O₂ concentration obtained with quasi-steady-state and dynamic model.

Table 36.1

Initial conditions and model parameters

$C_{ven}^{F,BL}$ [mM]	$C_{tis}^{F,BL}$ [mM]	$C_{art}^{F,BL}$ [mM]	<i>PS</i> [L min ⁻¹]
3.48 .10-2	3.37 · 10-2	$1.05 \cdot 10^{-1}$	2026