

# Primary charge separation in bacterial photosynthesis: Oxidized chlorophylls and reduced pheophytin

(reduced bacteriopheophytin/transient electron acceptor)

J. FAJER, D. C. BRUNE, M. S. DAVIS, A. FORMAN, AND L. D. SPAULDING

Department of Applied Science and Medical Research Center, Brookhaven National Laboratory, Upton, New York 11973

Communicated by Gerhart Friedlander, October 2, 1975

**ABSTRACT** Bacteriopheophytin, the magnesium-free base of bacteriochlorophyll, undergoes reversible one-electron reduction in organic solvents to yield an anionic free radical with characteristic optical and electron spin resonance spectra. The reduction potential of bacteriopheophytin,  $E_{1/2} \approx -0.55$  V against a normal hydrogen electrode, compared to  $E_{1/2} \approx -0.85$  V for bacteriochlorophyll, renders it a likely electron acceptor in the primary charge separation of photosynthesis. Comparison of these data with picosecond optical changes recently observed upon pulsed laser excitation of bacterial reaction centers leads us to propose that bacteriopheophytin is indeed a transient electron acceptor and that the primary charge separation of bacterial photosynthesis occurs between the bacteriochlorophyll complex  $P_{870}$  and bacteriopheophytin to yield the radicals of the oxidized chlorophyll dimer cation and reduced pheophytin anion.

Light is converted by green plants and photosynthetic bacteria into chemical energy by the creation of an oxidant ( $P^+$ ) and a reductant ( $X^-$ ) which result from ejection of an electron from (bacterio)chlorophyll (P) to a nearby acceptor (X). An extensive body of evidence (for reviews, see refs. 1-5) leads to the conclusion that light impinging on a photosynthetic organism is gathered by antenna chlorophylls (and other pigments) and funneled to a special environment, the reaction center, wherein a chlorophyll "special pair" ( $P_{700}$  in plants and  $P_{870}$  in purple bacteria) undergoes one electron oxidation to yield a  $\pi$  cation radical [ $P_{700}^+$  and  $P_{870}^+$  (5)] with characteristic optical, electron spin resonance (ESR), electron-nuclear double resonance, and redox properties. Isolation of photochemically active reaction centers of purple bacteria simplifies the study of the charge separation since each reaction center contains only three different polypeptides (about 70,000 daltons), four bacteriochlorophylls, two bacteriopheophytins, one ubiquinone, and a nonheme iron, and is free of cytochromes, copper, and antenna bacteriochlorophyll. Circular dichroism (CD) spectra indicate that, within the reaction center, the bacteriochlorophylls (BChls) are strongly exciton coupled, with weaker interactions with the bacteriopheophytins (BPhs). The creation of  $P^+_{870}$ , on oxidation of the reaction center, disrupts the CD interactions and the orientation of the pigment molecules is disturbed (1-3).

The nature of the electron acceptor (X) is less clear (1-4). Midpoint potentials,  $E_m$ , for the one electron reduction of X in different species of bacteria range between 0 and -150 mv [against a normal hydrogen electrode (NHE)], but values as low as -350 mv have been reported (2, 6, 7). Removal of

ubiquinone (UQ) from *Rhodospseudomonas spheroides* reaction centers inhibits photochemical activity (8). To reconcile the UQ data with ESR evidence of an iron compound, a complex of iron-ubiquinone (Fe-UQ),  $E_m = -50$  mv, is generally considered to be a primary electron acceptor (9-11).

Recent nanosecond and picosecond laser excitations of *R. spheroides* reaction centers, poised at potentials where X(Fe-UQ) is reduced, reveal a transient state,  $P^F$ , which forms with high quantum yield.  $P^F$  exhibits optical changes that implicate both BChl and BPh and decays with an exponential time  $\tau = 150-250$  psec with the concomitant appearance of  $P^+_{870}X^-$  when X is not initially reduced. Strikingly,  $P_{870}$  oxidation seems to occur with X reduced as well (12-15). We present here evidence that equates  $P^F$  with the primary charge separation of bacterial photosynthesis and propose that bacteriopheophytin is the transient electron acceptor of the  $P_{870}$  oxidation.

## EXPERIMENTAL

Cyclic voltammetry, controlled potential electrolysis, and coulometry were performed using a Princeton Applied Research potentiostat 173 equipped with a function generator 175 and a digital coulometer 179. Potentials were determined on  $10^{-3}$  M solutions containing 0.1 M tetrapropylammonium perchlorate against an aqueous saturated calomel electrode (= NHE -0.24 V). Optical spectra were obtained with a vacuum electrolysis cell on a Cary 17 spectrophotometer. The optical data used for the difference spectra of Figs. 3 and 4 were digitized using a Vanguard scanner and processed on a Control Data Corp. 6600 computer. ESR spectra were collected at X-band on a Varian E-12 spectrometer, equipped with a Field/Frequency Lock accessory and ramped by an SDS Sigma 2 computer. All ESR and optical spectra were obtained on samples prepared on a vacuum line using dried, distilled, and outgassed solvents with oxygen rigorously excluded. The chemical and electrochemical techniques have been described (16-18). A Coherent Radiation krypton ion laser and a quartz halogen Oriel lamp were used for the photochemical reductions. BPh *a* was prepared from BChl *a* extracted from *Chromatium vinosum* [BChls found in *C. vinosum* and *R. spheroides* have the same chemical composition (19)]. BPh was dissolved in  $CH_2Cl_2$ , which was pumped away to codistill water, and the pigment was dried under reduced pressure ( $10^{-3}$  Pa) for several hours.

## RESULTS

### The anion radical of bacteriopheophytin

BPh exhibits a reversible, one electron, halfwave reduction potential  $E_{1/2}$  of -0.58 against NHE in  $CH_2Cl_2$ . Electro-

Abbreviations: BPh, bacteriopheophytin; BPh<sup>-</sup>, reduced BPh; BChl, bacteriochlorophyll; ESR, electron spin resonance;  $E_{1/2}$ , half wave potential;  $E_m$ , midpoint potential;  $P_{870}$ , BChl special pair with 870 nm absorbance;  $P^*_{870}$ , excited singlet of  $P_{870}$ ;  $P^+_{870}$ , oxidized  $P_{870}$ ; NHE, normal hydrogen electrode; UQ, ubiquinone; X, electron acceptor, iron-ubiquinone complex.

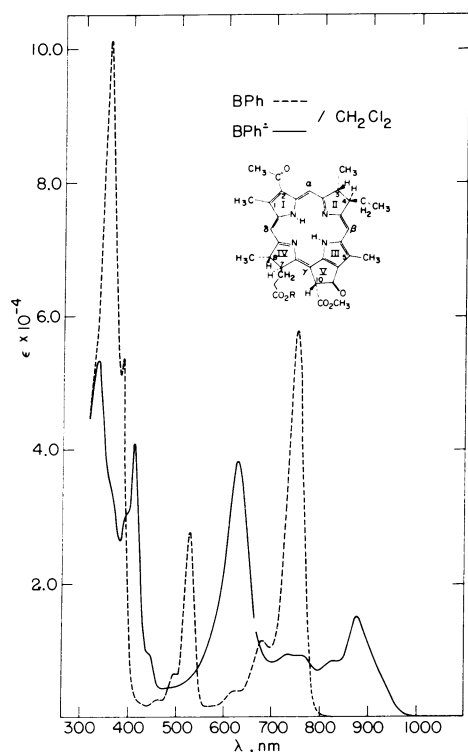


FIG. 1. Optical spectra of BPh (---) and of the anion radical BPh<sup>-</sup> (—) in CH<sub>2</sub>Cl<sub>2</sub>.

chemical reduction requires one electron and yields the optical spectra shown in Fig. 1. One electron oxidation regenerates better than 95% of the original BPh. The system can be recycled to again yield the radical spectrum. The ESR spectrum of BPh<sup>-</sup> in CH<sub>2</sub>Cl<sub>2</sub> at room temperature exhibits a *g*-value of 2.0030 ( $\pm 0.0002$ ) and a 12 line, partially resolved hyperfine structure (Fig. 2)\*. Similar spectra, with only minor variations in apparent splitting constants and line-widths are obtained by chemical reduction with an 18-crown-6 (1,4,7,10,13,16-hexaoxacyclooctadecane) ether/potassium complex in 2-methyltetrahydrofuran (20, 21) and by photochemical reduction in pyridine in the presence of Na<sub>2</sub>S (22).

The ESR spectrum in pyridine is displayed for comparison with a computer simulation (Fig. 2). The simulation assumes interaction of the unpaired electron spin density with the following nuclei: (a) 2 nitrogens (rings II and IV) with hyperfine splitting constant  $a_N = 2.3$  G ( $2.3 \times 10^{-4}$  tesla), (b) 3 protons, average  $a_H = 2.5$  G for the hydrogens of position  $\delta$ ,  $\alpha$ , and  $\beta$ , and (c) 6 protons, average  $a_{H_{CH_3}} = 3.0$  G for the methyl groups 1 and 5 on rings I and III, respectively. These assignments are deduced from ESR and electron-nuclear double resonance observations on BPh<sup>-</sup>, deuterated BPh<sup>-</sup>, and selectively deuterated BPh<sup>-</sup>†. In addition, they are in accord with the ESR spectra of the anion radical of a model compound, free base tetraphenyl bacteriochlorin, also selectively deuterated, and all the above ESR data are in general agreement with Pariser, Parr, Pople self-consistent-

\* The spectrum collapses to a singlet,  $\Delta H_{ptp} = 13$  G at 100 K.

† The different nuclear spin of deuterium,  $I = 1$  compared with  $I = \frac{1}{2}$  for hydrogen, combined with the difference in gyromagnetic ratio yield large and predictable ESR changes on substitution of deuterium for hydrogen. See ref. 33 for examples.

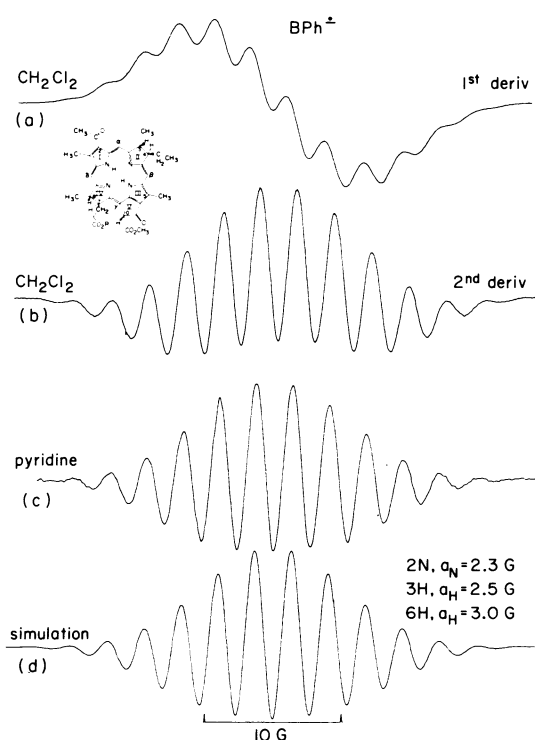


FIG. 2. (a) First and (b) second derivative ESR spectra of BPh<sup>-</sup> in CH<sub>2</sub>Cl<sub>2</sub>, (c) second derivative spectrum of BPh<sup>-</sup> in pyridine, and (d) computer simulation of (c) using the splitting constants shown.

field molecular orbital calculations by Felton‡. Thus, positions 1, 2, 5, 6,  $\delta$ ,  $\alpha$ , and  $\beta$  each carry about 0.1 unit of unpaired spin while the spin density in rings II and IV is concentrated in the nitrogens.

The good agreement with model systems and molecular orbital calculations minimizes the possibility that the species observed is the result of a proton addition: BPh<sup>-</sup> + H<sup>+</sup> → BPhH•. Molecular orbital calculations for BPhH• indicate that large spin densities would exist at the site adjacent to the proton addition (presumably one of the *meso* positions) and would result in large couplings with the added proton‡. We observe no initial difference in the ESR spectrum of BPh<sup>-</sup> prepared photochemically in pyridine that contains small amounts of H<sub>2</sub>O or D<sub>2</sub>O, (the radical eventually undergoes isotopic exchange, *not* addition) and conversely, we detect no difference in the ESR spectra of perdeutero BPh<sup>-</sup> in perdeutero pyridine containing D<sub>2</sub>O or H<sub>2</sub>O.

In conclusion, we find an anion radical of BPh stable in several solvents with a halfwave reduction potential which varies slightly from solvent to solvent:  $E_{1/2}$  (against NHE) = -0.58 V in CH<sub>2</sub>Cl<sub>2</sub>, -0.54 in butyronitrile, -0.51 in dimethylformamide, and -0.52 in ethanol [pH = 9.4 (23)] for an averaged  $E_{1/2} \approx -0.55$  V. These compare with  $E_{1/2}$  values of +0.64 and -0.86 V for oxidation and reduction of BChl in CH<sub>2</sub>Cl<sub>2</sub>.

#### A biological role for bacteriopheophytin

We now propose that, in bacterial reaction centers, bacteriopheophytin acts as a transient electron acceptor on a picosecond time scale and that the electron donor is the P<sub>870</sub> special pair. The primary charge separation of bacterial photosyn-

‡ J. Fajer, A. Forman, M. S. Davis, L. D. Spaulding, D. C. Brune, and R. H. Felton, submitted.

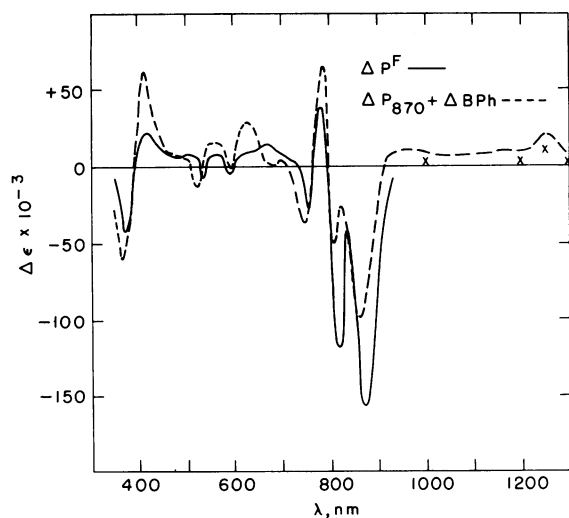


FIG. 3. Comparison of the laser induced optical changes,  $\Delta P^F$ , with the changes calculated on the assumption that  $\Delta P^F = \Delta P_{870} + \Delta BPh$ , the changes caused by the oxidation of  $P_{870}$  and the reduction of BPh.  $\Delta P^F$  observed (— and points X);  $\Delta P^F$  calculated (---).

thesis is thus the photochemical creation of the radicals of the cation  $P_{870}^+$  and the anion  $BPh^-$ . We examine here the reasons for and the consequences of this suggestion in terms of the existing experimental data and energetics involved.

The idea that the transient  $P^F$  is a pure excited singlet has been considered and discarded (13, 14) because of the order of magnitude discrepancy between the lifetime of the singlet estimated from fluorescence measurements (24, 25) and that of  $P^F$ . A triplet state has been observed *in vivo*, but only when the electron acceptor X is reduced (26–28), and it probably forms (2, 12) from  $P^F$  (see below).

The oxidation of  $P_{870}$  is well characterized by steady-state photochemical and chemical techniques and leads to typical difference spectra (1, 2). These reflect the disruption of the complex of two sets of BChls that give rise to the 800- and 870-nm bands and the appearance of the  $\pi$  cation radical (30–32) of the bacteriochlorophyll pair,  $P_{870}^+$ . Of all the radicals possibly involved in photosynthesis, i.e.,  $BChl^+$ ,  $BChl^-$ ,  $BPh^+$ ,  $BPh^-$ , and  $P_{870}^+$ , that have been observed optically (17, 33–35), *only* the dimer absorbs in the red region between 1100 and 1300 nm ( $\lambda_{max} = 1250$  nm). The most recent psec data indicate (15) that the 1250-nm band appears immediately upon excitation. The rapid bleaching of the 870-nm band ( $\leq 7$  psec) and the quantum yield of about 1 found for the bleaching of the 870-nm band upon 880-nm excitation, even at 4 K (29; R. K. Clayton, private communication) further suggest that oxidation of the  $P_{870}$  complex is a primary step which proceeds from the excited singlet.

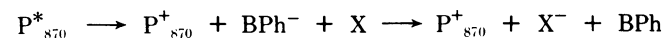
If we thus accept that oxidized  $P_{870}$  is the electron donor observed immediately upon psec excitation and if X (the UQ–Fe complex) is already reduced, then the choice of primary electron acceptor is limited to either BChl or BPh.

If it is BPh, then  $P^F$  represents the sum of the oxidized  $P_{870}$  and the reduced BPh, and the optical changes reported for the appearance of  $P^F$ ,  $\Delta P^F$ , are given by the spectral differences  $\Delta P_{870} + \Delta BPh$  where  $\Delta P_{870} = P_{870}^+ - P_{870}$  and  $\Delta BPh = BPh^- - BPh$ .  $\Delta P_{870}$  has been determined both by

photo- and chemically-induced oxidation. The data of Reed (34) have been used to calculate Fig. 3. [The absorption due to X and  $X^-$  is small (35, 36) relative to the other changes in  $P_{870}$ , so we feel justified in ignoring it. Use of a chemically oxidized  $\Delta P_{870}$  spectrum (35) yields results similar to those displayed in Fig. 3.]  $\Delta P^F$  is a composite of the data reported by Parson *et al.* for nsec excitation with X reduced (Fig. 2A of ref. 13) for the region 300–700 nm and the psec data with X not reduced, for the 700- to 900-nm region (Fig. 1 of ref. 13). The first part of the spectrum is consistent with the sparser data obtained after 20 psec in the 400- to 700-nm region with X not reduced (13). Kaufmann *et al.* (14) reported similar results. The points X shown in the 1000- to 1300-nm region are the data of Dutton *et al.* (15), which were obtained under nonsaturating light intensities.

The discrepancy near 800 nm in the calculated spectrum is due in part to the bleaching of that band as it undergoes kinetics of its own with  $\tau \approx 50$  psec (13). These changes may well reflect (13) the disruption of the reaction center when  $P_{870}^+$  is formed and would be consistent with circular dichroism data (3). At large changes in absorbance ( $\Delta A$ ), the psec data also contain apparent artifacts such as  $\Delta A = 0.16$  for a solution that has a nominal absorbance  $A = 0.12$  at 870 nm (see Fig. 1 of ref. 13). We emphasize the good agreement between practically all maxima and minima in the calculated and experimental spectra over the entire 350- to 1300-nm region.

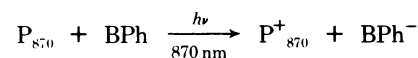
According to the mechanism we propose, the optical changes should first reflect the appearance of  $P_{870}^+$  and  $BPh^-$ . Then, if X was not originally reduced, the disappearance of  $BPh^-$  should reveal the spectrum of  $P_{870}^+$  and  $X^-$ , i.e., the standard  $P^+X^- - PX$  difference spectrum:



Experimental data at several wavelengths typical of both  $P_{870}$  and BPh confirm this suggestion (12–15). Furthermore, since  $BPh^-$  does not absorb above 950 nm (see Fig. 1), the infrared bands characteristic of  $P_{870}^+$  should appear immediately and undergo no further kinetics until  $P_{870}^+$  is reduced again. This is also observed (15). The experimental data, kinetic as well as optical, thus support our scheme.

Note that if X is initially reduced,  $P^F$  can decay via a back reaction to yield  $P_{870}^*$  and fluorescence or a triplet of  $P_{870}$ . Increased fluorescence (37) and triplets consonant with the  $P_{870}$  pair but with unusual spin polarization have been observed (26, 28, 38, 40) when X is reduced<sup>†</sup>. The triplet is probably the state  $P^R$  detected by Parson *et al.* (12).

We consider now the energetics of the reaction we postulated:



The midpoint potential for the one electron oxidation of  $P_{870}$  is +0.45 V against NHE (2, 3, 41).  $E_{1/2}$  of a reversible, one-electron process virtually equals the thermodynamic potential, and therefore the  $E_{1/2}$  values found for the BPh reduction can be used to estimate the free energy gained by the photochemically-induced charge separation.  $E_{1/2} \approx$

<sup>†</sup> Recombination of the transient radicals of  $P^F$  with a dynamic electron polarization mechanism could induce the unusual spin population of the triplet (39). The  $BPh^- - P_{870}^+$  annihilation was recently considered by Thurnauer *et al.* (40). See also Dutton *et al.* (15).

<sup>§</sup> BPh presumably occurs in intact organisms as well as in isolated reaction centers.



and to Drs. M. C. Thurnauer, J. R. Norris, J. J. Katz, R. E. Connors, and R. H. Clarke for preprints of their triplet work (38, 40). This work was performed under the auspices of the U.S. Energy Research and Development Administration.

1. Clayton, R. K. (1973) *Annu. Rev. Biophys. Bioeng.* **2**, 131-136.
2. Parson, W. W. & Cogdell, R. J. (1975) *Biochim. Biophys. Acta* **416**, 105-149.
3. Sauer, K. (1975) in *Bioenergetics of Photosynthesis*, ed. Govindjee (Academic Press, New York), pp. 115-181.
4. Frenkel, A. W. (1970) *Biol. Revs.* **45**, 595-616.
5. Katz, J. J. & Norris, J. R. (1973) in *Current Topics in Bioenergetics*, eds. Sanadi, D. R. & Packer, L. (Academic Press, New York), Vol. 5, pp. 41-75.
6. Loach, P. A., Kung, M. & Hales, B. J. (1975) *Ann. N.Y. Acad. Sci.* **244**, 297-318.
7. Govindjee, R., Smith, W. R., Jr. & Govindjee (1974) *Photochem. Photobiol.* **20**, 191-199.
8. Cogdell, R. J., Brune, D. C. & Clayton, R. K. (1974) *FEBS Lett.* **45**, 344-347.
9. Bolton, J. R. & Cost, K. (1973) *Photochem. Photobiol.* **18**, 417-421.
10. Dutton, P. L., Leigh, J. S. & Reed, D. W. (1973) *Biochim. Biophys. Acta* **292**, 654-664.
11. Feher, G., Isaacson, R. A., McElroy, J. D., Ackerson, L. C. & Okamura, M. W. (1974) *Biochim. Biophys. Acta* **368**, 135-139.
12. Parson, W. W., Clayton, R. K. & Cogdell, R. J. (1975) *Biochim. Biophys. Acta* **387**, 268-278.
13. Rockley, M. G., Windsor, M. W., Cogdell, R. J. & Parson, W. W. (1975) *Proc. Nat. Acad. Sci. USA* **72**, 2251-2255.
14. Kaufmann, K. J., Dutton, P. L., Netzel, T. L., Leigh, J. S. & Rentzepis, P. M. (1975) *Science* **188**, 1301-1304.
15. Dutton, P. L., Kaufmann, K. J., Chance, B. & Rentzepis, P. M. (1975) *FEBS Lett.*, in press.
16. Fajer, J., Borg, D. C., Forman, A., Dolphin, D. & Felton, R. H. (1970) *J. Am. Chem. Soc.*, **92**, 3451-3459.
17. Fajer, J., Borg, D. C., Forman, A., Dolphin, D. & Felton, R. H. (1973) *J. Am. Chem. Soc.* **95**, 2739-2741.
18. Fajer, J., Bielski, B. H. J. & Felton, R. H. (1968) *J. Phys. Chem.* **72**, 1281-1288.
19. Katz, J. J., Strain, H. H., Harkness, A. L., Studier, M. H., Svec, W. A., Janson, T. R. & Cope, B. T. (1972) *J. Am. Chem. Soc.* **94**, 7938-7939.
20. Grokel, G. W., Cram, D. J., Liotta, C. L., Harris, H. P. & Cook, F. L. (1974) *J. Org. Chem.* **39**, 2445-2446.
21. Komarynsky, M. A. & Weissman, S. I. (1975) *J. Am. Chem. Soc.* **97**, 1589.
22. Pakshina, Ye. V. & Krasnovskii, A. A. (1974) *Biofizika* **19**, 238-243.
23. Gilman, S. (1957) Ph.D. Dissertation, Syracuse University.
24. Zankel, K. L., Reed, D. L. & Clayton, R. K. (1968) *Proc. Nat. Acad. Sci. USA* **61**, 1243-1249.
25. Slooten, L. (1972) *Biochim. Biophys. Acta* **256**, 452-466.
26. Leigh, J. S. & Dutton, P. L. (1974) *Biochim. Biophys. Acta* **357**, 67-77, and references therein.
27. Wraight, C. A., Leigh, J. S., Dutton, P. L. & Clayton, R. K. (1974) *Biochim. Biophys. Acta* **333**, 401-403.
28. Uphaus, R. A., Norris, J. R. & Katz, J. J. (1974) *Biochem. Biophys. Res. Commun.* **61**, 1057-1063.
29. Wraight, C. A. & Clayton, R. K. (1974) *Biochim. Biophys. Acta* **333**, 246-260.
30. Feher, G., Hoff, A. J., Isaacson, R. A. & Ackerson, L. C. (1975) *Ann. N.Y. Acad. Sci.* **244**, 239-259.
31. Norris, J. R., Scheer, H. & Katz, J. J. (1975) *Ann. N.Y. Acad. Sci.* **244**, 260-280.
32. Borg, D. C. (1975) in *Free Radicals in Biology*, ed. Pryor, W. A. (Academic Press, New York), pp. 69-147.
33. Fajer, J., Borg, D. C., Forman, A., Felton, R. H., Dolphin, D. & Vegh, L. (1974) *Proc. Nat. Acad. Sci. USA* **71**, 994-998.
34. Reed, D. W. (1969) *J. Biol. Chem.* **244**, 4936-4941.
35. Clayton, R. K. & Straley, S. C. (1972) *Biophys. J.* **12**, 1221-1234.
36. Slooten, L. (1972) *Biochim. Biophys. Acta* **275**, 208-218.
37. Clayton, R. K., Fleming, H. & Szuts, E. A. (1972) *Biophys. J.* **12**, 46-63.
38. Clarke, R. H., Connors, R. E., Norris, J. R. & Thurnauer, M. C. (1975) *J. Am. Chem. Soc.*, in press.
39. Wan, J. K. S., Wong, S. K. & Hutchinson, D. A. (1974) *Acc. Chem. Res.* **7**, 58-64.
40. Thurnauer, M. C., Katz, J. & Norris, J. R. (1975) *Proc. Nat. Acad. Sci. USA* **72**, 3270-3274.
41. Kuntz, I. D., Loach, P. A. & Calvin, M. (1964) *Biophys. J.* **4**, 227-249.
42. Duysens, L. N. M. (1958) *Brookhaven Symp. Biol.* **11**, 10-23.
43. Ross, R. T. & Calvin, M. (1967) *Biophys. J.* **7**, 595-614.
44. Knox, R. S. (1969) *Biophys. J.* **9**, 1351-1362, and references therein.
45. Prince, R. C. & Dutton, P. L. (1975) *Biochim. Biophys. Acta* **387**, 609-613.
46. Dutton, P. L. & Wilson, D. F. (1974) *Biochim. Biophys. Acta* **346**, 165-212.
47. Ke, B., Chaney, T. H. & Reed, D. W. (1970) *Biochim. Biophys. Acta* **216**, 373-383.
48. Clayton, R. K. & Yau, H. F. (1972) *Biophys. J.* **12**, 867-881.