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MOLECULAR IMAGING REVEALS RAPID REDUCTION OF ENDOTHELIAL ACTIVATION IN EARLY ATHEROSCLEROSIS WITH APOCYNIN INDEPENDENT OF ANTI-OXIDATIVE PROPERTIES

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Abstract

OBJECTIVE—Anti-oxidative drugs continue to be developed for the treatment of atherosclerosis. Apocynin is an NADPH-oxidase-inhibitor with anti-inflammatory properties. We used contrast enhanced ultrasound (CEU) molecular imaging to assess whether short-term apocynin therapy in atherosclerosis reduces vascular oxidative stress and endothelial activation

APPROACH AND RESULTS—Genetically-modified mice with early atherosclerosis were studied at baseline and after 7 days of therapy with apocynin (4mg/kg/d I.P.) or saline. CEU molecular imaging of the aorta was performed with microbubbles targeted to vascular cell adhesion molecule 1 (VCAM-1; MB_V), to platelet GPIb α (MB_{Pl}), and control microbubbles (MB_{Cr}) . Aortic VCAM-1 was measured using Western Blot. Aortic ROS generation was measured using a lucigenin assay. Hydroethidine (HE) oxidation was used to assess aortic superoxide generation. Baseline signal for MB_V (1.3±0.3 A.U.) and MB_{Pl} (1.5±0.5 A.U.) was higher than for MB_{Ctr} (0.5±0.2 A.U., p<0.01). In saline-treated animals, signal did not significantly change for any microbubble agent whereas short-term apocynin significantly $(p<0.05)$ reduced VCAM-1 and platelet signal (MB_V: 0.3 ± 0.1 , MB_{Pl}: 0.4 ± 0.1 MB_{Ctr}: 0.3 ± 0.2 A.U., $p=0.6$ between agents). Apocynin reduced aortic VCAM-1 expression by 50% ($p<0.05$). However, apocynin therapy did not reduce either ROS content, superoxide generation, or macrophage content.

CONCLUSIONS—Short-term treatment with apocynin in atherosclerosis reduces endothelial cell adhesion molecule expression. This change in endothelial phenotype can be detected by

Disclosures: None.

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molecular imaging before any measurable decrease in macrophage content, and is not associated with a detectable change in oxidative burden.

Keywords

Apocynin; Atherosclerosis; Oxidative Stress; Microbubbles; Molecular Imaging

INTRODUCTION

Endothelial activation is a key step both in the initiation of atherosclerotic lesions as well as in their progression to a late stage, where inflammatory cell burden and susceptibility to acute atherothrombotic complications are high. Oxidative stress plays a major role in supporting and amplifying the endothelial activation in atherosclerosis¹. The family of NOX NADPH oxidase present in plaque macrophages, and in native endothelial and smooth muscle cells is a major source of reactive oxygen species (ROS) and therefore represents a potential therapeutic target².

Apocynin is a polyphenolic drug that has been isolated from plant extracts and inhibits assembly of the NOX2 isoform of the NADPH oxidase enzyme complex³. In mice with advanced atherosclerosis, long-term therapy with apocynin has been shown to reduce endothelial adhesion molecule expression, platelet adhesion, and plaque growth; while in hypercholesterolemic rabbits apocynin started at a much earlier stage of disease has been shown to prevent development of atherosclerotic lesions⁴. It is unknown whether the beneficial effects of apocynin occur early after initiation of therapy. With regards to mechanism, it is unknown whether apocynin's effects are entirely due to a reduction in oxidative stress since polyphenolic drugs such as apocynin have anti-inflammatory effects independent of their anti-oxidant properties^{5, 6}. Direct anti-inflammatory action independent of anti-oxidant properties has been substantiated by the reduced adhesion molecule expression in cultured endothelial cells exposed to apocynin^{7, 8}.

In this study we addressed many of these knowledge gaps. We performed *in vivo* ultrasound molecular imaging to test the hypothesis that short-term administration of apocynin in a model of early atherosclerosis reduces endothelial activation and platelet adhesion, two factors that are recognized to play an important role in plaque progression. Ex vivo techniques were used to evaluate whether these effects were associated with a reduction in vessel oxidative stress.

MATERIALS AND METHODS

Materials and Methods are available in the online-only Supplement.

RESULTS

Effect of apocynin on VCAM-1 expression and endothelial platelet adhesion

Apocynin treatment for 1 week reduced total aortic wall expression of VCAM-1 on Western blot by 50% (Figure 1A). On immunohistochemistry, small regions of neointimal formation were observed in the aortic root and proximal ascending portion. VCAM-1 staining was present on the endothelial lining and on macrophages in fibrofatty lesions in non-treated animals. In apocynin-treated animals, VCAM-1 expression was reduced both on endothelial cells as well as on macrophages within lesions, while the total amount of macrophages present in plaques remained unchanged (Figure 2). The reduction in endothelial VCAM-1 expression was evident both in regions overlying plaques as well as on endothelium in regions without plaques (Figure 1B–E).

In vivo labeling of platelets with rhodamine-6G allowed visualization of platelet/leukocyte aggregates on the vascular endothelial surface. Platelet/leukocyte aggregates were present in regions with early plaques, but also in regions of the aortic endothelial surface with a normal appearance. Both the number of platelet/leukocyte aggregates per square millimeter $(5.0\pm0.44$ in apocynin treated animals vs 9.6 ± 0.52 in non-treated animals, p<0.01) and the percentage of endothelial surface covered by platelet/leukocyte aggregates was significantly reduced in animals that were treated with apocynin (Figure 3).

Effect of apocynin on vascular oxidative stress

Lucigenin assays showed robust NADPH oxidase activity in whole aortic rings of nontreated mice. In mice treated with apocynin, NADPH-dependent lucigenin chemiluminescence was not different compared to non-treated mice. Given the potential of lucigenin to undergo redox cycling and generate artifactual signals, high pressure liquid chromatography (HPLC) of tissue extracts after exposure of aortic rings to HE was performed to directly assess tissue superoxide content. In accordance with the results of the lucigenin assays, HE oxidation to EOH was not different between the two animal groups (Figure 4).

Molecular Imaging of VCAM-1 expression and platelet adhesion

High frequency ultrasound imaging was not of sufficient quality for evaluation in one animal in each group. In the remaining animals, there were no differences in left ventricular ejection fraction, peak aortic flow velocity or aortic diameter, indicating that apocynin treatment did not lead to hemodynamic differences that could potentially influence targeted microbubble adhesion (table).

CEU molecular imaging in the ascending aorta at baseline showed greater signal enhancement for VCAM-1-targeted and platelet-targeted microbubbles compared to control microbubbles (Figure 5). After 7 days of treatment with apocynin, signal for VCAM-1 targeted and platelet targeted microbubbles was not different from control microbubble signal (Figure 6). In contrast, in animals treated with saline injections, the signal for VCAM-1 and platelets was elevated significantly over control signal to a degree that was similar to baseline. In the subgroup of animals that were imaged before and after treatment, apocynin lead to a significant decrease in VCAM-1 targeted signal (from 1.80±0.51 to 0.30 ± 0.10 , p=0.046) and a strong trend for decrease in platelet-targeted signal $(2.21\pm0.80$ to 0.35 ± 0.07 , p=0.078). In animals imaged before and after treatment with saline injections, signal for VCAM-1 (1.44 \pm 0.55 vs. 1.00 \pm 0.23, p=1.00) and platelet-targeted signal (1.84 \pm 0.88 vs. 1.26 ± 0.40 , p=0.69) did not decrease significantly.

DISCUSSION

Endothelial activation plays a crucial role in the initiation and progression of atherosclerotic plaque formation. In this study, short term treatment with the polyphenol apocynin in a murine model of early mild atherosclerosis lead to a reduction in endothelial inflammatory phenotype and platelet adhesion. These relatively acute changes were not associated with a measurable reduction in vascular NADPH oxidase activity or superoxide content.

Endothelial adhesion molecule expression is an early and important step in the pathogenesis of atherosclerosis. Deposition of oxidized LDL in the vascular wall leads to endothelial expression of pro-inflammatory cytokines such as IL-1-β and tumor necrosis factor-α. Locally increased cytokine levels result in an upregulation of cell adhesion molecules such as VCAM-1, mediated by the transcriptional factors activated protein-1 (AP-1) and nuclear factor κB (NF-κB), this in turn promotes the recruitment of leukocytes to the vessel wall.

ROS generated by NADPH oxidases both in endothelial cells as well as in leukocytes in nascent plaques are thought to amplify vascular inflammation throughout the pathogenesis of atherosclerosis. Accordingly, mouse models with knockouts of the NADPH oxidase isoforms NOX-1 and NOX-2 or the cytosolic NADPH oxidase subunit p47phox have shown a reduction in atherosclerotic plaque formation $9-11$. In humans, functional deficiency of the GP91 $phox$ subunit is associated with smaller carotid intima-media thickness (CIMT)¹². These findings have generated interest in using inhibitors of NADPH oxidases for the treatment of atherosclerosis. Apocynin inhibits NADPH oxidase activity in leukocytes allegedly by impeding the assembly of the cytosolic subunits at the cell membrane¹³. In cell culture experiments it has been noted that the inhibitory action of apocynin occurs with a delay, suggesting that it has to undergo activation before inhibiting ROS generation. In the presence of H_2O_2 and myeloperoxidase, apocynin is converted to an apocynin radical and subsequently forms apocynin dimers, which are thought to be the active compounds that result in inhibition of NADPH oxidase activity¹⁴. Thus, the effect of apocynin on NADPH oxidase inhibition during the later stages of atherosclerosis may depend on its activation in vascular tissue and require a sufficient inflammatory cell and oxidative stress burden. Notably, however, apocynin also exerts anti-inflammatory effects that are independent of NADPH oxidase inhibition⁶. Such effects may be of importance during the early stages of atherosclerosis, when inflammatory cell load and oxidative burden are low.

We examined the effects of treatment with apocynin in a murine model of early atherosclerosis in mice with fibrofatty lesions. Assessing treatment effects of potential drug candidates during the early stages of atherosclerosis is of clinical significance, since interventions that are started early during the pathogenesis of atherosclerosis are thought to afford a larger risk reduction for cardiovascular events than interventions that are initiated when clinical atherosclerotic disease is established¹⁵. Our data indicate that treatment with apocynin results in a rapid decrease in endothelial expression of VCAM-1. These results are in line with observations in cell culture showing a decrease in VCAM-1 expression in response to apocynin⁸, and extend observations made in advanced atherosclerosis to earlier stages of plaque development⁴. The decrease in signal from VCAM-1 targeted microbubbles was more pronounced than differences in VCAM-1 expression determined with Western blot. We believe this to be due to the capability of ultrasound molecular imaging to specifically assess endothelial inflammatory phenotype¹⁶ and thus to detect a more pronounced decrease in endothelial VCAM-1 compared to Western blot which assesses expression in the whole vascular wall. In addition, the observation that the decrease in vascular inflammation was not associated with a reduction of vascular NADPH oxidase activity or tissue superoxide content indicates that the antiinflammatory effect of apocynin observed in our study was probably mediated through a ROS-independent mechanism in the very early stages of atherosclerotic plaque development, possibly mediated instead by its effects on cytochrome P450 pathways¹⁷.

In addition to endothelial cell inflammatory activation, platelet-endothelial interactions play a role in vascular inflammation and the pathogenesis of atherosclerosis. Platelet-endothelial interactions mediated by P-Selectin and von Willebrand factor- GPIbα ligation in the absence of plaque rupture accelerate plaque formation in murine atherosclerosis $18-20$. The interaction of activated platelets with the endothelial surface results in the secretion of proinflammatory cytokines CD40L and IL-1β, as well as of chemokines like RANTES (regulated on activation, normal T cell expressed and secreted) and platelet factor 4 (PF4) to the vascular wall, all of which facilitate increased monocyte recruitment 2^{1-23} . Platelets from patients with functional deficiency of GP91*phox* subunit of NADPH oxidase or control subject platelets treated with apocynin have reduced *in vitro* platelet recruitment and aggregation, indicating a direct role of NADPH oxidase in platelet reactivity²⁴. Our molecular imaging results indicate a decrease in platelet-endothelial interactions after

treatment with apocynin. While we did not specifically investigate the pathways responsible for the action of apocynin in platelets, previous data indicate that apocynin influences platelet aggregation by mechanisms that are dependent on NADPH oxidase activity²⁵ but also on changes in arachidonic acid metabolism with a decrease in thromboxane A2 formation⁵.

Several limitations of our study deserve attention. First, as the aim of our study was to assess the acute effects of apocynin treatment on the endothelial inflammatory phenotype, we did not expect an influence of treatment on plaque size and thus did not perform histological analysis. However, long-term treatment with apocynin has been shown to reduce plaque formation in our mouse model⁵. Also, the dose used in our study was in the low range of doses used in published animal studies, however, there is no established optimal dose, and our data demonstrate an effect of the treatment on both endothelial inflammatory phenotype and platelet adhesion. Furthermore, while we applied well-established techniques to measure NADPH oxidase activity and superoxide content without finding an effect of apocynin therapy, we cannot exclude that locally restricted (e.g. endothelial) and/or changes in other ROS species contributed to the observed effect. Finally, our methods for evaluating platelet adhesion did not allow differentiation of direct endothelial attachment and platelet-monocyte complexes.

In summary, we show that in a murine model of early atherosclerosis, treatment with apocynin leads to a rapid decrease in endothelial inflammation and platelet adhesion, which are detectable using ultrasound molecular imaging. Our data indicate that these effects of apocynin are not associated with a measurable decrease in ROS generation.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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ABBREVIATIONS

VWF Von Willebrand Factor

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SIGNIFICANCE

Anti-oxidative drugs continue to be developed for the treatment of atherosclerosis. In this study, we assessed whether short-term therapy with the NADPH oxidase inhibitor apocynin reduces vascular oxidative stress, endothelial inflammatory activation and platelet adhesion in a murine model of early atherosclerosis, and whether molecular imaging is capable of detecting these changes. Assessing the treatment effects of potential drug candidates during the early stages of atherosclerosis is of clinical significance, since interventions that are started early during the pathogenesis of atherosclerosis are thought to afford a larger risk reduction for cardiovascular events than interventions that are initiated late. We show that ultrasound molecular imaging is capable of detecting reductions in endothelial inflammatory activation and in platelet adhesion at a stage before any measurable, treatment associated decrease in macrophage content occurs. These phenotypic changes are not associated with measurable changes in oxidative stress.

Figure 1.

Assessment of vascular cell adhesion molecule 1 (VCAM-1) expression after 7 days of saline or apocynin treatment. **A**, VCAM-1 protein expression in the ascending aorta assessed by Western blot in nontreated (lanes 1–4) vs apocynin-treated (lanes 5–8) animals, n=4 per group, **P*<0.05 vs nontreated animals. Representative examples of fluorescent immunohistochemistry images of the base of the aorta demonstrating endothelial VCAM-1 expression (red fluorescence) in a nontreated animal at 10-fold magnification (**B**), in the same animal at 40-fold magnification (**C**), and reduced VCAM-1 expression in an apocynintreated animal (**D** and **E**). Autofluorescence delineating vessel anatomy is shown in green, 4 ´,6-diamidino-2-phenylindole staining of the nuclei in blue.

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Figure 2.

Assessment of plaque macrophage content after 7 days of saline or apocynin treatment. **A**, Percentage of the plaque area covered with macrophages (Mac-2 staining) at the base of the aorta and in the ascending aorta (n=8 in each group; *P*=nonsignificant vs non-treated). **B**, Example of trans-illumination image used for plaque delineation. **C**, Example of Mac-2 staining used for quantification of macrophage content in the plaque.

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Figure 3.

Assessment of platelet adhesion on the aortic endothelial surface after 7 days of saline or apocynin treatment. **A**, Percentage of the endothelial surface of the ascending aorta covered with platelet/leukocyte aggregates (n=5 in each group), $*P<0.05$ vs nontreated animals. Examples of en face fluorescence microscopy demonstrating 2 platelet/leukocyte aggregates on normal appearing endothelial surface in a nontreated animal (**B**) and absence of platelet/ leukocyte aggregates in an apocynin-treated animal (**C**). Scale bar, 25 µm.

Figure 4.

Assessment of reactive oxygen species generation after 7 days of saline or apocynin treatment. **A**, Superoxide-generating activity of whole aortic rings after the addition of 100 µmol/L NADPH at 4 minutes. Measurements represent relative light units (RLU).*P*=nonsignificant (ns) between saline-treated and apocynin-treated animals (n=10 in each group). **B**, High pressure liquid chromatography analysis of 2-hydroxyethidium generated in vascular rings exposed to 50 µmol/L hydroethidine. *P*=ns between salinetreated and apocynin-treated animals (n=8 in each group).

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Figure 5.

Molecular imaging of the ascending aorta before the start of treatment.**A**, Mean±SEM background-subtracted signal intensity for microbubbles targeted to vascular cell adhesion molecule 1 (VCAM-1; MB_{VCAM}), to glycoprotein Iba on activated thrombocytes (MB_{Pl}), and control microbubbles (MB_{Ctr}). **P*<0.01 vs MB_{ctr}, #*P*<0.05 vs MB_{ctr}(n=12). Examples of color-coded contrast-enhanced ultrasound (CEU) images after injection of MB_{Ctr} (**B**), of MB_{VCAM} (**C**), and of MB_{PI} (**D**). The color scale for the CEU images is shown at the bottom of each frame.

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Figure 6.

Molecular imaging of the ascending aorta after 7 days of saline or apocynin treatment. **A**, Mean \pm SEM background-subtracted signal intensity in saline-treated (n=9) and apocynintreated (n=10) animals for microbubbles targeted to vascular cell adhesion molecule 1 (VCAM-1; MB_{VCAM}), to glycoprotein Ib α on activated thrombocytes (MB_{Pl}), and control microbubbles (MB_{Ctr}). **P*<0.01 vs MB_{ctr}, #*P*<0.01 vs MB_{ctr}. \mathbb{P} *P*<0.05 vs the same microbubble in apocynin-treated animals. Examples of color-coded contrast-enhanced ultrasound (CEU) images after injection of $MB_{\text{Ctr}}(B)$, of $MB_{\text{VCAM}}(C)$, and of $MB_{\text{Pl}}(D)$ in saline-treated animals. In the same order, examples of color-coded CEU images after injection of MB_{Ctr} (**E**), of MB_{VCAM} (**F**), and of MB_{Pl} (**G**) in apocynin-treated animals are shown.

Table

Echocardiographic data (mean ± 1 SD)

