

## Exon–intron organization and complete nucleotide sequence of a human major histocompatibility antigen DC $\beta$ gene

(major histocompatibility complex/class II antigen)

DAN LARHAMMAR, JENS JØRGEN HYLDIG-NIELSEN, BO SERVENIUS, GÖRAN ANDERSSON, LARS RASK, AND PER A. PETERSON

Department of Cell Research, The Wallenberg Laboratory, University of Uppsala, Box 562, S-751 22 Uppsala, Sweden

Communicated by Jean Dausset, August 15, 1983

**ABSTRACT** We have determined the complete nucleotide sequence of a human class II histocompatibility antigen DC $\beta$  gene. The gene spans more than 7 kilobases and contains five exons corresponding to the different domains of the DC $\beta$  polypeptide. The exon–intron organization is thus analogous to that of class II antigen  $\alpha$ -chain genes, class I antigen heavy chain genes, and the constant parts of immunoglobulin genes, emphasizing further the evolutionary relationship among these molecules. The mature polypeptide deduced from the DC $\beta$  gene shows 93% and 88% homology, respectively, to sequences derived from two DC $\beta$  cDNA clones of other haplotypes. The allelic polymorphism of DC $\beta$  chains resides predominantly in the first extracellular domain, whereas the rest of the polypeptide is virtually constant. The exons of the DC $\beta$  gene display high homology to the corresponding exons of a murine I-A $\beta$  gene. Also, the introns show significant homology. The DC $\beta$  chains lack eight amino acids in the cytoplasmic tail, as compared to DR and I-A  $\beta$  chains. This is probably due to a non-functional splice junction of DC $\beta$  genes, causing a separate cytoplasmic exon to be nonexpressed.

The D region of the human major histocompatibility complex (MHC) harbors genes involved in the regulation of the immune response (1). The known products of this region, the class II antigens, are polymorphic glycoproteins composed of two dissimilar subunits: an  $\alpha$  chain of approximately 34,000 daltons and a  $\beta$  chain of approximately 28,000 daltons (2). The polymorphism is primarily carried by the  $\beta$  chains (3). Class II antigens are expressed on the surface of cells involved in various aspects of an immune response such as B lymphocytes, subpopulations of T lymphocytes, and macrophages (4).

Three different class II heterodimers, denoted DR (2), DC (5), and SB (6), have been identified by immunochemical techniques and primed lymphocyte typing. Also a fourth antigen called BR has been proposed (7). Southern blot analyses with fragments derived from  $\beta$ -chain cDNA clones provide evidence that the class II family contains more than three  $\beta$ -chain genes (8). The exact function of the different class II antigens in cell interactions is as yet unknown.

Through cloning of cDNAs corresponding to class II antigen  $\alpha$  and  $\beta$  chains, we have recently shown that the two chains display structural homology to each other as well as to class I antigens and immunoglobulins (9–11), suggesting that these immunologically important proteins have evolved from a common ancestor by gene duplications.

As a step towards the understanding of the evolution and mechanisms for generation and maintenance of the polymorphism of class II antigens, we present here the nucleotide sequence of a human class II antigen  $\beta$ -chain gene.

The publication costs of this article were defrayed in part by page charge payment. This article must therefore be hereby marked "advertisement" in accordance with 18 U.S.C. §1734 solely to indicate this fact.

### MATERIALS AND METHODS

**Isolation of Genomic Clone.** A genomic library was constructed from DNA donated by an HLA-homozygous human individual typed to be Dw4/DR4 (unpublished data) in the cosmid vector pHEP (12). The library was screened with a 627-base-pair (bp) *Ava* I fragment containing almost the entire coding part of the DC $\beta$  cDNA clone pII- $\beta$ -1 [previously denoted pDR- $\beta$ -1 (9)]. Screening of the cosmid library and growth and analysis of cosmid clones were performed as described (13), as were Southern blot analyses (14). DNA probes were labeled by nick-translation (15).

**DNA Sequence Determination.** Nucleotide sequences were determined with the chemical degradation procedure (16) and a modification of the dideoxy chain termination method (17) using exonuclease III and synthetic oligonucleotide primers (unpublished results).

### RESULTS AND DISCUSSION

**Isolation of a Cosmid Containing a DC $\beta$  Gene.** Genomic DNA from a panel of DR-homozygous human individuals has been analyzed in Southern blotting experiments using a fragment from a DC $\beta$  cDNA clone as probe (8). All blots with DNA digested with different enzymes show one strong constant band and one or two strong polymorphic bands. The  $\beta$ -chain gene of cosmid clone cosII-102 accounts for the strong polymorphic DC $\beta$  bands of the genomic donor DNA. Thus, this clone was chosen for further characterization.

**Exon–Intron Organization of the DC $\beta$  Gene.** Two overlapping fragments containing the DC $\beta$  gene [a 7.7-kilobase (kb) *Eco*RI fragment and an 11-kb *Bam*HI fragment] were subcloned in pUC9 to facilitate sequence determination (see Fig. 1). A restriction map of the DC $\beta$  gene and the sequencing strategy are shown in Fig. 1. Exons were localized by comparison with the DC $\beta$  cDNA clone pII- $\beta$ -2 (unpublished results). The nucleotide sequence and the translated amino acid sequence are shown in Fig. 2. The DC $\beta$  gene encompasses more than 7 kb and contains five exons correlating with the different domains of the DC $\beta$  polypeptide (see Fig. 3). The first exon corresponds to the 5' untranslated region, the signal sequence, and four amino acids of the first domain. The remainder of the first domain and the second domain are encoded by exons 2 and 3, respectively. Exon 4 encodes the connecting peptide, the membrane-spanning segment, and six amino acids of the cytoplasmic tail. The last four amino acids of the cytoplasmic tail are encoded by a separate exon also containing the 3' untranslated region. All splice junctions conform to the G-T—A-G rule (18). Overall, the exon–intron organization is analogous to that of genes for class II antigen  $\alpha$  chains (refs. 19 and 20; unpub-

Abbreviation: bp, base pair(s).





homology, respectively. Thus, within one subset of class II  $\beta$  chains, the first extracellular domain carries most of the allelic polymorphisms, whereas the second domain appears virtually constant (25). In the nucleotide sequences encoding the first domains of the three DC $\beta$  chains, mutations in the first and second positions of codons are at least as prevalent as third-base mutations. This pattern of nucleotide substitutions is also found in the variable domain exons of immunoglobulins (26–28), particularly in the codons of the hypervariable regions (29). Models involving either positive selection favoring substitutions leading to amino acid replacements (26) or suppression of silent mutations (28) have been proposed. Whereas the class I antigen polymorphism may be due partly to gene conversion events (30, 31), the DC $\beta$  pattern of nucleotide substitutions is consistent with multiple independent mutational events, as has been suggested for the immunoglobulins (27). In the nucleotide stretches encoding the DC $\beta$  second domain, the pattern of substitutions is as expected for a conserved region—i.e., silent third-base substitutions in the codons are more frequent than first- and second-base substitutions. At only 4 of 34 variant positions do all three DC $\beta$  chains have unique amino acids. This low number may be due to structural constraints, allowing only a few different amino acids at a certain position.

Exon 4, encoding the connecting peptide, the membrane-spanning segment, and the first six amino acids of the cytoplasmic tail, is highly conserved. Its translated amino acid sequence is identical to that of the cDNA clone pII- $\beta$ -1 and differs from that deduced from pII- $\beta$ -2 at only four positions.

The 3' untranslated region of the DC $\beta$  gene in cosII-102 shows 98% and 92% homology to pII- $\beta$ -1 and pII- $\beta$ -2, respectively. The polyadenylation signal A-T-T-A-A-A (ref. 9; see also ref. 32 and references therein) is present 330 bp downstream of the termination codon. Although the sequence determination was extended more than 100 bp 3' of the region, no additional polyadenylation signal was found.

In all four exons available for comparison, the DC $\beta$  gene shows greater homology to pII- $\beta$ -1 than to pII- $\beta$ -2. The mature DC $\beta$  chain deduced from the gene displays an overall homology of 93% and 88%, respectively, to the amino acid sequences deduced from pII- $\beta$ -1 and pII- $\beta$ -2. The DC $\beta$  chain encoded by cosII-102 thus appears to be more closely related to that deduced from pII- $\beta$ -1 than to that of pII- $\beta$ -2.

**DC $\beta$  Genes Lack a Cytoplasmic Exon.** The cytoplasmic tail of DC $\beta$  chains is eight amino acids shorter than that of DR $\beta$  chains (11). The nucleotide sequence of the murine I-A $\beta$  gene (33), the structural homologue of DC $\beta$ , has provided evidence that a separate exon of 24 nucleotides accounts for this difference. A nucleotide stretch of 24 bp is indeed present in the DC $\beta$  gene, displaying clear homology to the corresponding segments of the DR $\beta$  cDNA clones and the A $\beta$  gene. The absence of this segment in the two DC $\beta$  cDNA clones whose sequences have been determined to date is probably due to an altered 5' splice junction of this exon. Instead of the preferred sequence A-G<sup>+</sup>G (18), A-A<sup>+</sup>G is found. Whether the difference in cytoplasmic tails between DC $\beta$  and DR $\beta$  chains has any functional significance remains to be investigated. Although this exon is not expressed in the DC $\beta$  genes, it remains well conserved at the genomic level (21) of the 24 nucleotides are identical with the A $\beta$  cytoplasmic exon. However, the entire intron between exons 4 and 5 of the DC $\beta$  gene is highly homologous to the corresponding stretch of the A $\beta$  gene, indicating that there could be a general conservative pressure acting on the nucleotides of this part of the DC $\beta$  gene. Also, the third DC $\beta$  intron shows clear homology to the corresponding A $\beta$  intron. Significant homology can also be detected in parts of intron 1 and intron 2 by using a computer alignment program (34).

**Promoter Region.** The transcriptionally important DNA sequences G-G<sup>+</sup>C-A-A-T-C-T (CAT) and TATA are expected at about 70 bp and 30 bp upstream from the cap site of transcribed genes, respectively (18). Because no full-length DC $\beta$  cDNA clone is available that can define the cap site, assignment of these elements can only be tentative. A computer comparison of the 5' part of the DC $\beta$  gene with the corresponding region of the A $\beta$  gene aligns a TATA-like sequence and a possible cap sequence with the proposed A $\beta$  TATA and cap sequences, respectively (33). These putative TATA and cap elements, as well as a CAT-like sequence, are underlined in Fig. 2. Provided the proposed cap site is correct, the DC $\beta$  mRNA will have a 5' untranslated region of 329 nucleotides, unless an intron occurs in this region. The homology between the DC $\beta$  gene and the A $\beta$  gene in the region extending from the putative cap sites to the initiation codons is about 60%, counting insertions as mismatches.

The nucleotide stretches flanking the first domain exon are extremely rich in G+C, a feature shared with the nucleotide stretches flanking the first domain exon of the A $\beta$  gene (33) as well as the first and second domain exons of class I heavy chain genes in man (21) and mouse (22), but not of the DR $\alpha$  (19), I-E $\alpha$  (ref. 20; unpublished data), and  $\beta_2$ -microglobulin (35) genes. The significance of the G+C-rich stretches is unclear. However, it is interesting to note that they flank exons encoding polymorphic domains.

**Concluding Remarks.** The DC $\beta$ -chain gene reported here corresponds to the highly polymorphic DC $\beta$  gene detected in Southern blotting analyses of genomic DNA. The gene displays all the characteristics of a functional gene. The polymorphic region of DC $\beta$  chains is predominantly located in the first extracellular domain. The exon-intron organization of the DC $\beta$  gene is similar to that of class II antigen  $\alpha$ -chain genes, class I antigen heavy chain genes, and the constant parts of immunoglobulin genes; the similarity supports previous evidence for an evolutionary relationship among these molecules.

We thank Ms. E. Rossi for help with computer analyses, Ms. P. Ågren for technical assistance, and Dr. P. Lind and Mr. P. Wenkler for preparation of oligonucleotide primers. This work was supported by grants from the Swedish Cancer Society, King Gustav V's 80-Years Fund, and Marcus Borgström's Fund.

1. Benacerraf, B. (1981) *Science* **212**, 1229–1238.
2. Klareskog, L., Sandberg-Trägårdh, L., Rask, L., Lindblom, J. B., Curman, B. & Peterson, P. A. (1977) *Nature (London)* **265**, 248–251.
3. Charron, D. J. & McDevitt, H. O. (1979) *Proc. Natl. Acad. Sci. USA* **76**, 6567–6571.
4. Hämmerling, G. J. (1976) *Transplant. Rev.* **40**, 64–82.
5. Tosi, R., Tanigaki, N., Centis, D., Ferrara, G. B. & Pressman, D. (1978) *J. Exp. Med.* **148**, 1592–1611.
6. Shaw, S., Johnson, A. H. & Shearer, G. M. (1980) *J. Exp. Med.* **152**, 565–580.
7. Tanigaki, N. & Tosi, R. (1982) *Immunol. Rev.* **66**, 5–37.
8. Böhme, J., Owerbach, D., Denaro, M., Lernmark, Å., Peterson, P. A. & Rask, L. (1983) *Nature (London)* **301**, 82–84.
9. Larhammar, D., Schenning, L., Gustafsson, K., Wiman, K., Claesson, L., Rask, L. & Peterson, P. A. (1982) *Proc. Natl. Acad. Sci. USA* **79**, 3687–3691.
10. Larhammar, D., Gustafsson, K., Claesson, L., Bill, P., Wiman, K., Schenning, L., Sundelin, J., Widmark, E., Peterson, P. A. & Rask, L. (1982) *Cell* **30**, 153–161.
11. Larhammar, D., Andersson, G., Andersson, M., Bill, P., Böhme, J., Claesson, L., Denaro, M., Emmoth, E., Gustafsson, K., Hammerling, U., Helden, E., Hyldig-Nielsen, J. J., Lind, P., Schenning, L., Servenius, B., Widmark, E., Rask, L. & Peterson, P. A. (1983) *Hum. Immunol.*, in press.
12. Grosveld, F. G., Lund, T., Murray, E. J., Mellor, A. L., Dahl, H. H. M. & Flavell, R. A. (1982) *Nucleic Acids Res.* **10**, 6715–6732.

13. Grosveld, F. G., Dahl, H.-H. M., de Boer, E. & Flavell, R. A. (1981) *Gene* 13, 227-237.
14. Southern, E. M. (1975) *J. Mol. Biol.* 98, 503-517.
15. Rigby, P. W. J., Dieckmann, M., Rhodes, C. & Berg, P. (1977) *J. Mol. Biol.* 113, 237-251.
16. Maxam, A. M. & Gilbert, W. (1980) *Methods Enzymol.* 65, 499-560.
17. Sanger, F., Nicklen, S. & Coulson, A. R. (1977) *Proc. Natl. Acad. Sci. USA* 74, 5463-5467.
18. Breathnach, R. & Chambon, P. (1981) *Annu. Rev. Biochem.* 50, 349-383.
19. Das, H. K., Lawrence, S. K. & Weissman, S. M. (1983) *Proc. Natl. Acad. Sci. USA* 80, 3543-3547.
20. Mathis, D. J., Benoist, C. O., Williams, V. E., II, Kanter, M. R. & McDevitt, H. O. (1983) *Cell* 32, 745-754.
21. Malissen, M., Malissen, B. & Jordan, B. R. (1982) *Proc. Natl. Acad. Sci. USA* 79, 893-897.
22. Steinmetz, M., Moore, K. W., Frelinger, J. G., Sher, B. T., Shen, F.-W., Boyse, E. A. & Hood, L. (1981) *Cell* 25, 683-692.
23. Gough, N. (1981) *Trends Biochem. Sci.* 6, 203-205.
24. Kreil, G. (1981) *Annu. Rev. Biochem.* 50, 317-348.
25. Kaufman, J. F. & Strominger, J. L. (1982) *Nature (London)* 297, 694-697.
26. Baltimore, D. (1981) *Cell* 24, 592-594.
27. Bothwell, A. L. M., Paskind, M., Reth, M., Imanishi-Kari, T., Rajewsky, K. & Baltimore, D. (1981) *Cell* 24, 625-637.
28. Selsing, E., Miller, J., Wilson, R. & Storb, U. (1982) *Proc. Natl. Acad. Sci. USA* 79, 4681-4685.
29. Rechavi, G., Bienz, B., Ram, D., Ben-Neriah, Y., Cohen, J. B., Zakut, R. & Givol, D. (1982) *Proc. Natl. Acad. Sci. USA* 79, 4405-4409.
30. Boss, J. M., Gillam, S., Zitomer, R. S. & Smith, M. (1981) *J. Biol. Chem.* 256, 12958-12961.
31. Pease, L. R., Schulze, D. H., Pfaffenbach, G. M. & Nathenson, S. G. (1983) *Proc. Natl. Acad. Sci. USA* 80, 242-246.
32. Weiss, E., Golden, L., Zakut, R., Mellor, A., Fahrner, K., Kvist, S. & Flavell, R. A. (1983) *EMBO J.* 2, 453-462.
33. Larhammar, D., Hammerling, U., Denaro, M., Lund, T., Flavell, R. A., Rask, L. & Peterson, P. A. (1983) *Cell* 34, 179-188.
34. Orcutt, B. C., Dayhoff, M. O. & Barker, W. C. (1982) *ALIGN: Alignment Score Program*, NBR Report 820501-08710 (National Biomedical Research Foundation, Washington, DC).
35. Parnes, J. R. & Seidman, J. G. (1982) *Cell* 29, 661-669.