

Gene repertoire of the anti-poly(Glu⁶⁰Ala³⁰Tyr¹⁰) (GAT) immune response: Comparison of V_H, V_κ, and D regions used by anti-GAT antibodies and monoclonal antibodies produced after anti-idiotypic immunization

(idiotypic structure and expression/mRNA sequences)

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ABSTRACT Eight monoclonal antibodies were selected from BALB/c mice immunized with two different monoclonal anti-idiotypic antibodies recognizing two discrete idiotopes characteristic of the anti-poly(Glu⁶⁰Ala³⁰Tyr¹⁰) (GAT) antibody response. These monoclonal antibodies were previously classified as Ab₁ (anti-GAT-like) and Ab₃ (anti-anti-idiotypic) on the basis of expression of the public idiotypic specificity (*p.GAT*) studied with a xenogeneic serum, anti-GAT activity, and expression of various public idiotopes. All the heavy chain variable region (V_H) sequences from Ab₁ are nearly identical to the V_H sequences of Ab₁ anti-GAT monoclonal antibodies. The same type of results has been found with the Ab₁ κ light chain variable region (V_κ) sequences. Confirming our classification, Ab₃ V_H and V_κ sequences were found to be completely different from Ab₁ V_H and V_κ sequences. The Ab₁ diversity (*D*) regions are different from one another and different from the *D* regions found on monoclonal anti-GAT antibodies but function similarly. These *D* regions are not simply derived from already described *D* genes. Finally, our results suggest that in the anti-GAT response V_H and V_κ sequences are mainly responsible for idiotope expression.

Antigen immunization leads to the production of specific antibodies (Ab₁), which can in turn be used as immunogens to produce anti-idiotypic reagents (1–3). Immunization with these reagents (Ab₂) can also elicit an immune response (4, 5). This response consists of immunoglobulins idiotypically related to Ab₁ with or without antibody activity (Ab₁) and of anti-anti-idiotypic antibodies (Ab₃). Structural analysis of the genes involved at the different steps of this “idiotypic cascade” is a major tool in elucidating the origin of the B-cell repertoire (6). More precisely, structural studies of antibodies obtained after anti-idiotypic (anti-Id) immunization should unequivocally establish the genetic origin of Ab₃ and Ab₁ and document the relationship between Ab₁ and Ab₁.

Various systems have been analyzed at the Ab₁ level, using either myelomas or hybridomas specific for well-defined antigens. The immune response against arsonate (Ars) (7), nitrophenyl acetyl (NP) (8), phosphocholine (PC) (9), oxazolone (ox) (10), dextran (dex) (11), and poly(Glu⁶⁰Ala³⁰Tyr¹⁰) (GAT) (12) have been particularly studied. More recently several reports dealing with the study of Ab₁ and Ab₃ have been published (13, 14).

In the immune response against GAT different monoclonal anti-GAT antibodies (HP-GAT; HP, hybridoma product) have been characterized (15). Their partial amino acid sequences and complete nucleotide sequences have been re-

ported. A limited number of sequences for the heavy chain variable region (V_H) (12, 16) and κ light chain variable region (V_κ) (17–19) are used. At the idiotypic level the anti-GAT response is characterized by the expression of the public idiotypic specificity *p.GAT* defined by a xenogeneic antiserum (20, 21) and expressed in all HP-GAT (15). More recently, two discrete idiotopes characteristic of the anti-GAT responses have been identified (22–25).

In this paper we report the nucleotide sequences of two sets of monoclonal antibodies obtained from BALB/c mice immunized with two different monoclonal anti-idiotypic antibodies (HP-Id) recognizing the two idiotopes previously defined (25). The V_H, V_κ, and diversity (*D*) sequences of these monoclonals are compared to the corresponding sequences previously reported for HP-GAT. The possible contribution of V_H, V_κ, and *D* segments to idiotope expression and antibody activity is discussed.

MATERIALS AND METHODS

Detection of Idiotypic Specificities and of Anti-GAT Activity.

The detection of the *p.GAT* specificity and the percent maximal inhibition were calculated as described (20, 21). For the detection of the two idiotopes identified in the GAT response by HP-Id₂₀ and HP-Id₂₂ a solid-phase radioimmunoassay (RIA) was used (25). The idiotope binding inhibitory capacity (IBIC) of each Ab₁ was determined. IBIC was expressed as the percentage of the slope of the inhibition curves, taking the slope obtained with HP-GAT G5 as standard (IBIC = 100%).

The GAT-binding assay has been described (25). Results are expressed as the reciprocal of the dilution of supernatant giving 50% of maximal binding. Ig present in each fluid was measured and adjusted to 1 μg/ml before the assay. For the GAT-binding inhibition assay, radiolabeled G5Bb₂₋₂ (10 ng per well) was used to measure binding to GAT and various dilutions of ascitic fluids were tested for their inhibitory capacity. Results are expressed as percent of maximal inhibition obtained.

RNA Purification and Nucleotide Sequencing. The heavy- and light-chain-encoding fractions of poly(A)⁺ RNAs were prepared as described (12, 26). Nucleotide sequences were

Abbreviations: Ab₁, specific antibody; Ab₂, anti-idiotypic antibody; Ab₃, anti-anti-idiotypic antibody; Ab₁, antibody-like immunoglobulin generated after anti-idiotypic immunization; Id, idiotype; GAT, random terpolymer poly(Glu⁶⁰Ala³⁰Tyr¹⁰); V_H, variable region of immunoglobulin heavy chain; V_κ, variable region of immunoglobulin κ light chain; *D*, diversity region; *J*, joining region; HP, hybridoma product (monoclonal antibody); HP-Id, monoclonal anti-idiotypic antibody against public idiotopes; HP-GAT, monoclonal anti-GAT antibody; IBIC, idiotope binding inhibitory capacity; G5 or G5Bb₂₋₂, HP-GAT used as reference; *p.GAT*, public idiotypic specificity in the anti-GAT response; KLH, keyhole limpet hemocyanin.

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determined according to the modification of the original dideoxy method using the mRNA as template (27). Synthetic oligonucleotides, d(CAGGGGCCAGTGG) specific for the constant region of the $\gamma 1$ or $\gamma 2a$ chain, d(GCTCTCGAG-GAGAC) specific for the constant region of the μ chain, or d(TGGATGGTGGGAAGATG) specific for the constant region of the κ light chain, were synthesized according to Gait and Sheppard (28) and used as primers. Each sequence was determined with 2 μ g of enriched mRNA and 10 ng of oligonucleotide primer. The concentrations of nucleotides (unlabeled and labeled) and dideoxy analogs have already been reported (12). To minimize possible ambiguities in sequence determination on gels (27) labeling with each of the α - 32 P-labeled deoxynucleotides (dCTP, dATP, and dGTP) was performed in separate experiments and each was repeated at least twice.

RESULTS

Characteristics of Monoclonal Antibodies Derived from HP-Id-Immunized BALB/c Mice. From BALB/c mice immunized with two different HP-Ids coupled to keyhole limpet hemocyanin (KLH), two fusion experiments were performed and eight hybridomas were studied. Three hybridomas were isolated from animals immunized with HP-Id₂₀-KLH and five hybridomas were from animals stimulated with HP-Id₂₂-KLH. The characteristics of the corresponding HP have been determined (25) and are summarized in Table 1.

The expression of the *p.GAT* idiotype specificity characteristic of the anti-GAT antibody response was measured on these HPs by using the xenogeneic polyclonal antisera previously defined (20). Seven of these HPs were found to inhibit the binding of radiolabeled monoclonal anti-GAT antibody G5, which was taken as reference. One HP (22.134) was found not to inhibit in this assay. The anti-GAT activity of these reagents was tested with two different assays, and both assays gave the same results (25). Some HPs (20.8, 20.11, 20.33, and 22.186) did exhibit an anti-GAT activity comparable to G5, while others showed a low anti-GAT activity (22.162 and 22.8) or no activity (22.176, 22.134). We concluded that HP-22.134 is anti-anti-idiotypic (Ab₃), while the seven other HPs are Ab₁.

In addition, we have tested the idiotope expression by these HPs, using two HP-Ids (HP-Id₂₀ and HP-Id₂₂) that recognize the two idiotopes characteristic of the anti-GAT response (Table 1). The seven Ab₁s were able to inhibit the binding of G5 to these HP-Id. When compared to HP-GAT G5, the idiotopes expressed by these different Ab₁s are not identical, since the inhibition curves were found to have different slopes. However, all the Ab₁s isolated expressed the

same characteristic idiotopes as all the BALB/c HP-GATs studied (22). As expected, Ab₃ inhibited only the binding of HP-Id₂₂ to G5 (25).

Primary Structure of V_H Regions of Ab₁ and Ab₃ Monoclonal Antibody. The nucleotide sequence of seven heavy chain regions, encompassing most of the V_H, the entire D, and the beginning of the joining (J_H) segments was determined (Fig. 1). Within the limits of the analysis (four residues were undetermined), each of the five HPs has exactly the same V_H sequence as monoclonal anti-GAT G5. One (20.11) has only four substitutions, at position 56, 59, 67, and 68. These four changes do not lead to a net charge difference between the molecules (Gly → Val, Lys → Arg, Lys → Arg, and Ala → Gly). No silent substitution was identified.

While Ab₁ anti-GAT D regions usually code for five amino acids, the D regions of Ab₁ are very heterogeneous. None of these D segments is identical to D segments of monoclonal anti-GAT antibodies, which were found to derive either from DSP-2 or from DFL-16 (12). The D-J border is very heterogeneous, and two Ab₁ D segments are very short and encode only three amino acids. The seven J_H segments sequenced are J_{H-4}, which is also used by most monoclonal anti-GATs (G5, G8Ca₁₋₇, and G7Ab₂₋₉). At the D-V_H borders HP 20.33 has lost one triplet, while in all the other Ab₁s there is a silent substitution.

HP 22.134 uses a completely different V_H chain; this is consistent with the fact that it has been identified as an Ab₃. However, it belongs to the same V_H subgroup as the Ab₁ (29) and also uses J_{H-4}.

Primary Structure of Ab₁ and Ab₃ V_κ Regions. Three types of sequences were already defined in the V_κ involved in the anti-GAT response. One was identified on HP-GAT G5 and on several other monoclonal anti-GAT antibodies derived from BALB/c mice. The other sequences were obtained from DBA/2 and (DBA/2 × BALB/c)F₁ hybridomas (H-56 and H-51 fusion experiments). V_κ chains from HPs H-51 and H-56 vary from the V_κ chain of G5 at seven positions (17).

The partial sequences of six V_κ chains from Ab₁ are shown in Fig. 2. Strikingly, they are all identical and show the same substitutions as H-51 and H-56, when compared to the G5 sequence: at positions 50 (Arg → Lys), 83 (Met → Leu), 89 (Phe → Ser), and 91 (Gly → Ser). However, in HP 22.176 one substitution has been found at position 40 (Pro → Gln). The substitutions expressed only in H-51 and not in H-56 at positions 81, 87, 92, and 94 are not recovered on the Ab₁ V_κ chains.

All the J_κ segment sequences are J_{κ2}, as in G5, H-51, and H-56. However, other J_κ segments are used in the anti-GAT response (17). Concerning the V_κ-J_κ junction, the same

Table 1. Characteristics of monoclonal antibodies derived from HP-Id-immunized BALB/c mice

Immunizing HP-Id	HP	<i>p.GAT</i> expression,* %	Type of HP	Idiotype expression,† %		Anti-GAT activity‡	Isotypes
				Group 20	Group 22		
HP-Id ₂₀ -KLH	20.8	75	Ab ₁	130	120	+	$\gamma 1$ κ
	20.11	75	Ab ₁	100	100	+	$\gamma 1$ κ
	20.33	80	Ab ₁	90	90	+	μ κ
HP-Id ₂₂ -KLH	22.186	80	Ab ₁	120	100	+	μ κ
	22.162	80	Ab ₁	40	40	+/-	$\gamma 1$ κ
	22.8	90	Ab ₁	25	30	+/-	μ κ
	22.176	80	Ab ₁	50	40	-	μ κ
HP-Id ₂₂ -KLH	22.134	0	Ab ₃	—	—	-	$\gamma 2a$ κ

*Percent binding inhibition by hybridoma supernatants of rabbit anti-idiotypic antiserum to 5 ng of radiolabeled HP-GAT G5. G5 is taken as a probe of the public idiotype specificity in the GAT response (*p.GAT*).

†IBIC determined by the inhibition of the binding of radiolabeled G5 to purified HP-Id₂₀- or HP-Id₂₂-coated plates.

‡Anti-GAT activity was determined by two independent assays. See Table 2.

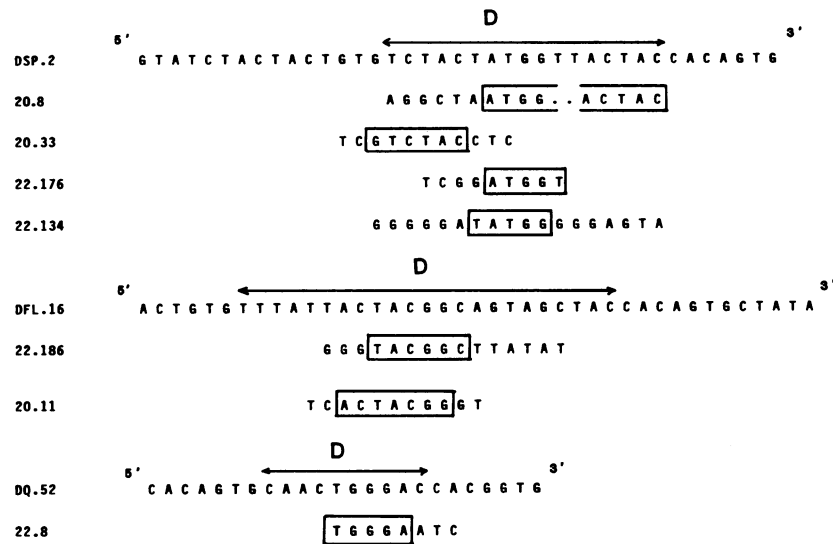


FIG. 3. Genetic origin of the Ab₁ and Ab₃ D regions. Ab₁ and Ab₃ D segments were compared with D germ-line sequences (30). Arrows indicate germ-line D coding sequences; boxes enclose nucleotides in common with germ-line sequences; and dots indicate hypothetical deleted nucleotides.

preferential substrate of terminal transferase (31). Alternatively, since the homologous regions between the Ab₁ D segment and the germ-line D segments considered are very short, one may consider our D segments as the products of non-identified D genes.

Anti-idiotypic immunization has been used as a tool to derive monoclonal reagents after a selective pressure only on idiotype expression. These reagents have been characterized at the serological level (25) and their primary structures have been determined. These reagents have also permitted the further analysis of the relationship between V_H, V_κ, J_H, and D segments in the expression of idiotypic determinants and antibody activity. The germ-line V_H and V_κ are critical for determining conformations leading to idiotype expression, while in the context of these defined V_H and V_κ segments, D segments appear to be more involved in determining the antibody activity. In general terms the repertoire expressed after anti-Id immunization is comparable to the repertoire expressed after antigen stimulation. These results suggest that immunization with monoclonal anti-Id antibodies may be used for vaccinations in situations in which appropriate HP-Ids are available (32). HP-Ids recognizing idiotopes expressed by protective antibodies should be good candidates.

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