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## Inhibition of Interferon-beta Responses in Multiple Sclerosis Immune Cells Associated With High-Dose Statins

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### Abstract

**Objective**—To determine whether statins affect type 1 interferon responses in relapsing-remitting multiple sclerosis (RRMS).

**Design**—Study effects of atorvastatin on type 1 interferon responses in Jurkat cells, mononuclear cells (MNCs) from therapy-naive patients with RRMS in vitro, and MNCs from interferon-treated RRMS patients in vivo in 4 conditions: no drug, statin only, interferon-beta only, and statin added on to interferon-beta therapy.

**Patients**—The study examined clinically stable patients with RRMS: 21 therapy-naive patients and 14 patients receiving interferon-beta with a statin.

**Interventions**—Statin effects on in vitro and in vivo interferon-beta–induced STAT1 transcription factor activation, expression of interferon-stimulated proteins in MNCs, and serum type 1 interferon activity.

**Results**—In vitro, atorvastatin dose dependently inhibited expression of interferon-stimulated P-Y-STAT1 by 44% ( $P < .001$ ), interferon regulatory factor 1 protein by 30% ( $P = .006$ ), and myxovirus resistance 1 protein by 32% ( $P = .004$ ) compared with no-statin control in MNCs from therapy-naive RRMS patients. In vivo, 9 of 10 patients who received high-dose statins (80 mg) had a significant reduction in interferon-beta therapy–induced serum interferon- $\alpha/\beta$  activity, whereas only 2 of 4 patients who received medium-dose statins (40 mg) had reductions. High-dose add-on statin therapy significantly blocked interferon-beta function, with less P-Y-STAT1 transcription factor activation, and reduced myxovirus resistance 1 protein and viperin protein production. Medium doses of statins did not change STAT1 activation.

**Conclusions**—High-dose add-on statin therapy significantly reduces interferon-beta function and type 1 interferon responses in RRMS patients. These data provide a putative mechanism for how statins could counteract the beneficial effects of interferon-beta and worsen disease.

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Multiple sclerosis (MS) is a chronic inflammatory disease in which autoreactive immune cells infiltrate the central nervous system (CNS), leading to demyelination and neurologic disability.<sup>1</sup> Interferon-beta ameliorates MS by altering peripheral and CNS immune responses and reducing disease activity. A total of 80% of patients with relapsing-remitting MS (RRMS) benefit from interferon-beta, but patients with progressive MS have minimal response to interferon-beta therapy.<sup>2</sup> Endogenous type 1 interferons (interferon- $\alpha$  and interferon- $\beta$ ) are important in disease progression and treatment response. Before therapy, interferon- $\alpha$ -related pathways are fundamentally dys-regulated in mononuclear cells (MNCs) from all forms of MS and are more abnormal than T<sub>H</sub>1, T<sub>H</sub>2, and other cytokine pathways.<sup>3</sup> After transition of RRMS to progressive MS, interferon- $\beta$  no longer can phosphorylate serine on STAT1 or induce certain genes in vitro.<sup>2</sup>

Statins ameliorate murine experimental autoimmune encephalomyelitis and are anti-inflammatory and neuroprotective.<sup>4,5</sup> Atorvastatin and glatiramer acetate synergize in the treatment of CNS autoimmunity,<sup>6</sup> so clinical trials in RRMS have combined interferon-beta with statins. In most placebo-controlled trials, combination therapy is safe and well tolerated but has no clinical or magnetic resonance imaging (MRI) benefit over interferon-beta monotherapy.<sup>7-9</sup> However, in a smaller, placebo-controlled trial with MS patients who were stable while taking subcutaneous interferon-beta-1a for at least 1 year prior, adding high-dose atorvastatin caused clinical and MRI exacerbations in 10 of 17 patients. The interferon-beta-only group had fewer exacerbations (1 of 10 patients;  $P=.02$ ), suggesting that statins antagonize interferon-beta therapy.<sup>10</sup> In a 307-patient, randomized, placebo-controlled, double-blind, phase 4 study, high-dose simvastatin (80 mg) added to interferon-beta-1a therapy produced no additional benefit.<sup>8</sup> There was actually a trend for higher relapse rates and disease activity in the comedication group compared with placebo, again suggesting antagonistic effects of add-on statin therapy.

How could statins impair interferon-beta therapy? Type 1 interferons bind to cell surface receptors, interferon- $\alpha$  receptor 1 and interferon- $\alpha$  receptor 2, and activate the JAK/STAT pathway, causing phosphorylation of tyrosine and serine residues on STAT1 and tyrosine on STAT2.<sup>11,12</sup> Phosphorylated STAT1-STAT2 heterodimer together with interferon-regulated factor 9 forms a complex that binds to DNA of the interferon-stimulated response element.<sup>13,14</sup> The activated P-Y-STAT1 transcription factor affects expression of 1000 genes; P-S-STAT1 enhances signaling in a subset of these genes. Interferon-beta induces myxovirus resistance 1 (MxA) and viperin proteins and endogenous interferon- $\beta$  and interferon- $\alpha$  subtypes.<sup>15,16</sup> Through this pathway, type 1 interferon alters T<sub>H</sub>1, T<sub>H</sub>2, and T<sub>H</sub>17 immunity, dendritic cell activation and maturation, cell cycle and apoptosis, and antigen presentation.<sup>17</sup>

We hypothesized that statins block the type 1 interferon pathway. We evaluated in vitro pharmacokinetic and dose effects of statins on interferon-induced phosphorylation of STAT1 and STAT2 transcription factors and downstream interferon-stimulated proteins, interferon regulatory factor 1 (IRF-1), MxA, and viperin. We also compared in vivo effects of high-dose statins plus interferon-beta therapy on interferon responses, induced proteins, and endogenous type 1 interferon activity.

## METHODS

### STUDY PARTICIPANTS

**In Vitro Experiments**—Twenty-one therapy-naive patients with RRMS, 12 women and 9 men, had a mean (SEM) age of 43.7 (2.2) years. None had been treated with immunomodulators for at least 3 months. None had ongoing infections.

**In Vivo Experiments**—Fourteen patients (4 black and 10 white; 64% female) had a mean (SEM) age of 54.2(2.6) years, an Expanded Disability Status Scale score of 4.10(0.49), an MS duration of 15.0(2.1) years, and interferon-beta treatment duration of 9.62 (1.66) years. Eleven patients were taking interferon-beta-1b, 2 patients were taking subcutaneous interferon beta-1a, and 1 patient was taking intramuscular interferon-beta-1a. There were 4 treatment groups in vivo: no drug, statin only, interferon-beta only, and statin added on to interferon-beta therapy. Serum and MNCs were obtained for all groups at various times.

Statin therapy was stopped for 5 to 7 days (>7 half-lives) to allow washout. Interferon-beta was stopped for 57 to 70 hours based on a prior study<sup>18</sup> to allow washout and to reflect basal levels of interferon-induced genes. Before phlebotomy and re-administration of interferon-beta injections, patients undergoing continuous long-term statin therapy took 40-mg (n = 4) or 80-mg (n = 10) statins to maximize statin effects at a safe dose. After washouts, blood was drawn at 8 am for baseline and then 4 hours (within 5 minutes) after the interferon-beta injection.

A total of  $5 \times 10^6$  MNCs were immediately lysed and stored for Western blotting. Another  $5 \times 10^6$  cells were cultured for 24 hours after interferon injections for ex vivo induction of MxA and viperin proteins. Serum was assayed for endogenous basal and therapy-induced type 1 interferon activity at 0 and 4 hours. All participants gave written informed consent for the University of Chicago institutional review board–approved protocol.

## DOSE RESPONSE AND KINETICS OF INTERFERONS VS STATINS

The MNCs were isolated with Ficoll-Hypaque density gradients. A total of  $4 \times 10^6$  cells/mL were cultured in RPMI with 10% fetal calf serum (GIBCO 1640; Invitrogen) at 37°C in 5% carbon dioxide. Cells were preincubated for 15 minutes to 48 hours with 1-, 5-, 10-, or 20- $\mu$ M atorvastatin (neat preparation; Anna Tallman, PharmD, Pfizer) and subsequently stimulated with interferon-beta-1b (0, 10, 20, 40, 80, 160, 320, and 500 U/mL) for 45 minutes to induce P-Y-STAT1 phosphorylation or for 24 hours to induce downstream proteins (MxA, IRF-1, and viperin; unphosphorylated STAT1 and STAT2). Stimulated cells were lysed and stored in 1 $\times$ Laemmli buffer for Western blotting.<sup>2</sup> Reversal of statin effects with 100- $\mu$ M mevalonate (Sigma Chemical Co) confirmed that the 3-hydroxy-3-methylglutaryl coenzyme A pathway affects interferon signaling.<sup>18</sup> In addition, in Jurkat T cells at  $4 \times 10^6$  cells/mL, in vitro kinetics and dose-dependent inhibition with statins combined with induction by different forms of interferon-beta were assayed with Western blots.

## SERUM INTERFERON- $\alpha/\beta$ ACTIVITY ASSAY

Serum samples from 14 RRMS patients were tested using a highly sensitive assay (limit of detection of 0.1 U/mL, well below typical 10- to 20-U/mL enzyme-linked immunosorbent assay [ELISA] thresholds). Moreover, ELISA can be less specific for serum interferon than this bioassay because ELISA detects cross-reacting but nonfunctional interferon-like proteins.<sup>19</sup> Briefly, the epithelial-derived WISH cell line (CCL-251, ATCC) was a reporter for responsiveness to interferon- $\alpha/\beta$ . Total cellular messenger RNA (mRNA) was purified, and complementary DNA was reverse transcribed and quantified by reverse transcription–polymerase chain reaction with primers for MxA-1, RNA-dependent protein kinase (protein kinase R), and interferon-induced protein with tetratricopeptide repeats 1 (IFIT-1). This bioassay was validated in large human populations and is specific for interferon- $\alpha/\beta$  activity.<sup>20–23</sup> Pretreatment of serum samples from MS patients with antibodies to interferon- $\alpha$  and interferon- $\beta$  abolishes interferon-induced gene expression in this assay.<sup>21,22,24</sup>

## WESTERN BLOT ANALYSIS

A total of  $4 \times 10^6$  MNCs/mL were induced with media alone or interferon-beta-1b at 160 U/mL for 45 minutes for assay of P-Y-STAT1 and P-Y-STAT2 or for 24 hours for STAT1, STAT2, IRF-1, MxA, and viperin.<sup>24</sup> Interferon-beta-induced MxA mRNA<sup>25</sup> is well correlated with MxA protein on Western blots.<sup>2</sup> Nonetheless, protein was used to examine interferon-beta-induced MxA responses because fluctuations are more likely to be missed with short half-life mRNA. Antibodies were goat anti-Actin (sc-1615), goat anti-P-Y701-STAT1 (sc-7988), goat anti-P-Ser-STAT1 (sc-16570-R), rabbit anti-IRF-1 (sc-20) (all Santa Cruz Biotechnology), rat anti-MxA (Stefan Lanker, PhD, Biogen), mouse anti-viperin (Peter Cresswell, PhD, Yale University), rabbit anti-STAT1 (sc-346), and rabbit anti-STAT2 (sc-476, Santa Cruz).

## STATISTICAL ANALYSIS

Values from washout vs treated experiments were compared with unpaired *t* tests. Baseline vs drug-induced values were analyzed with paired *t* tests in the same MS patients tested at all conditions.

## RESULTS

### INHIBITION OF TYPE 1 INTERFERON SIGNALING IN VITRO

Optimal conditions for interferon-stimulated STAT activation and downstream protein expression (MxA and IRF-1) were determined with different doses (0, 1, 5, 10, and 20  $\mu$ M) and kinetics (0 and 15 minutes and 1, 3, and 24 hours) of atorvastatin before treatment in Jurkat T cells and RRMS MNCs.

Three interferon-beta forms (interferon-beta-1a for intramuscular and subcutaneous use, and interferon-beta-1b for subcutaneous use but tested in vitro here) induced tyrosine phosphorylation of STAT1 in Jurkat cells after 45 minutes in vitro (eFigure 1A; <http://www.archneuro.com>). Preincubation with atorvastatin for 24 hours inhibited interferon-beta-stimulated P-Y-STAT1 and MxA and IRF-1 protein expression in a dose-dependent manner for all 3 interferon-beta forms (160 U/mL) in Jurkat T cells in vitro (eFigures 1B). In human U937 monocytoid cells, all 3 interferon-beta forms exhibited similar dose responses and inhibition by statin before incubation (data not shown). Statins inhibited interferon-induced tyrosine phosphorylation on STAT1 but not on STAT2. P-S-STAT1 and nonphosphorylated STAT1 and STAT2 levels did not change (data not shown), indicating that atorvastatin specifically targets P-Y-STAT1 in Jurkat cells.

Blockade of interferon-stimulated P-Y-STAT1 began at 15 minutes and was maximal after 24 hours before incubation with high-dose atorvastatin (eFigure 2A). A total of 10  $\mu$ M of atorvastatin markedly decreased interferon-beta-1b-stimulated MxA and IRF-1 production; lower statin doses were less inhibitory. High-dose atorvastatin inhibition of interferon-beta-1b-induced P-Y-STAT1 in MNCs from therapy-naive RRMS patients was confirmed with blockade of interferon  $\gamma$ , a strong inducer of P-Y-STAT1 (eFigure 2B).

STAT1 and STAT2 must be phosphorylated for interferon-stimulated gene expression. After pretreatment for 24 hours with 10  $\mu$ M of atorvastatin, MNCs from 21 therapy-naive RRMS patients were stimulated with 160 U/mL of interferon-beta-1b for 45 minutes. Atorvastatin reduced interferon-stimulated P-Y-STAT1 by 44% compared with no-statin control ( $P < .001$ ) (Figure 1). One hour of 100- $\mu$ M mevalonate before incubation reversed statin inhibition,<sup>26</sup> indicating specificity of atorvastatin in inhibiting P-Y-STAT1. Atorvastatin did not block induction of type 1 interferon-stimulated P-S-STAT1 or unphosphorylated STAT1 and STAT2 in MNCs (Figure 1).

Pretreatment atorvastatin reduced downstream interferon-beta-stimulated IRF-1 (30% reduction,  $P=.006$ ) and MxA protein (32% reduction,  $P=.004$ ) compared with no-statin control in paired MNCs from the same therapy-naive RRMS patients (Figure 1). Pretreatment simvastatin (10  $\mu\text{M}$ ) also significantly inhibited type 1 interferon responses in MNCs from RRMS patients (data not shown).

## HIGH-DOSE STATIN ADD-ON THERAPY AND IN VIVO INTERFERON- $\beta$ SIGNALING IN RRMS PATIENTS

To determine whether statin add-on therapy impairs interferon-beta therapy induction of endogenous serum interferon- $\beta$  activity, we measured serum type 1 interferon activity and interferon-induced proteins in 14 interferon-beta-treated RRMS patients under 4 different conditions (Figure 2). To determine the optimal time for measuring serum interferon activity, we first performed kinetics in stable RRMS patients receiving interferon-beta therapy but no statins. Blood was drawn at baseline and periodically from 10 minutes to 27 hours, and serum interferon activity was analyzed with a highly sensitive assay. Figure 3A shows representative kinetics from a stable RRMS patient given interferon-beta-1a (44  $\mu\text{g}$ , 9 MU subcutaneously) after a 3-day interferon washout. Serum type 1 interferon activity was elevated by 30 minutes after interferon-beta injection and remained high until 6 hours later, then declined by 27 hours. Interferon- $\beta$  averaged a 3-fold induction of P-Y-STAT1 from baseline. This interferon activity initially reflects administered interferon-beta and later is from therapy-induced endogenous interferon- $\beta$  and interferon- $\alpha$ , based on blocking experiments with specific anti-interferon- $\alpha$  and anti-interferon- $\beta$  antibodies. We used the 4-hour point for in vivo interferon- $\beta$  stimulation because interferon- $\alpha/\beta$  induction was still high 4 hours after interferon-beta injection, and many interferon-stimulated genes are induced within 4 hours.<sup>27</sup>

Statin add-on therapy significantly reduced serum type 1 interferon activity compared with interferon-beta monotherapy (Figure 3B). Nine of 10 patients who received high-dose statins (80 mg of atorvastatin or simvastatin) had significant reduction in serum interferon- $\alpha/\beta$  activity. Two of these patients had undetectable levels of serum interferon activity at all conditions even with this sensitive bioassay (Figure 3C). However, 2 of 4 patients who received medium-dose statins (40 mg) had no reduction in serum interferon- $\alpha/\beta$  activity. These data indicate that high-dose statin add-on therapy inhibits interferon- $\beta$  activity in most patients, whereas moderate doses have lesser inhibitory effects.

Ex vivo MNCs were studied in 4 different treatment conditions to confirm the in vitro effects of statins on interferon- $\beta$  responses. There was a significant in vivo reduction in tyrosine phosphorylation of STAT1 in the high-dose statin add-on group compared with interferon-beta monotherapy, whereas medium-dose statins did not affect STAT1 phosphorylation (Figure 4A). In addition, 6 of 14 RRMS patients receiving combination therapy had significant reduction in interferon-stimulated MxA and viperin proteins compared with interferon-beta monotherapy (Figure 4B). These results demonstrate that high-dose statin add-on therapy blocks interferon- $\beta$  responses in vivo.

Together, our data from cell culture, in vitro studies in therapy-naive RRMS patients, and in vivo studies in interferon-beta-treated RRMS patients receiving statin add-on therapy reveal that high-dose statins inhibit interferon- $\alpha/\beta$  activity by blocking tyrosine phosphorylation on STAT1 and preventing interferon responses.

## COMMENT

We demonstrate that high-dose statins inhibit interferon signaling. Atorvastatin dose dependently inhibits interferon- $\beta$  induction of P-Y-STAT1 and downstream proteins.



Preincubation in vitro with statins in Jurkat T cells and MNCs blocked interferon responses within 15 minutes and reached maximal inhibition at 24 hours (eFigures 1 and 2). This finding is consistent with other dose and pharmacokinetic studies.<sup>26,28</sup> Statins inhibit cholesterol synthesis but are also anti-inflammatory and thus are a potential therapy for MS and other neuroinflammatory diseases.<sup>29–32</sup>

Statins suppress proinflammatory T<sub>H</sub>1 and T<sub>H</sub>17 responses in experimental autoimmune encephalomyelitis and MS lymphocytes.<sup>28,33–35</sup> In 30 RRMS patients, monotherapy with 80 mg of simvastatin appeared to reduce the volume and number of gadolinium-positive MRI lesions by 44% from baseline in patients with active disease.<sup>36</sup> Treatment with high-dose atorvastatin for 9 months reduced MRI contrast-enhancing lesions (CELS).<sup>37</sup> In these uncontrolled studies with significant baseline MRI activity, the decrease in activity could have arisen from regression to the mean.<sup>38</sup>

Potential mechanisms of statin benefit in MS include (1) regulating extracellular kinase ERK and p38 phosphorylation through Rac and Rho pathways, which would block T<sub>H</sub>1 activation and induce a T<sub>H</sub>2 shift<sup>39</sup>; (2) impairing activation of Ras superfamily GTPases to inhibit the major histocompatibility class II antigen presentation pathway<sup>40</sup>; (3) blocking STAT activation to inhibit interleukin 17 production<sup>34</sup>; and (4) disturbing formation of cholesterol-containing microdomains (lipid rafts), thereby inhibiting function of the T-cell receptor and major histocompatibility class I and II.<sup>41–44</sup> However, MRI and clinical effects may be complex in humans because simvastatin inhibits CNS remyelination by blocking oligodendrocyte progenitor differentiation,<sup>45</sup> and atorvastatin promotes some proinflammatory T<sub>H</sub>1 responses by raising interleukin 12p70.<sup>46</sup>

High-dose atorvastatin in vitro specifically blocks formation of P-Y-STAT1 but not P-S-STAT1 or P-Y-STAT2 in MNCs from therapy-naive RRMS patients (Figure 2). High-dose statins in vivo also block interferon-beta-induced transcription factor activation and expression of interferon-induced proteins in RRMS; moderate-dose statins were less inhibitory (Figures 3 and 4). Our results may explain why some clinical studies with high-dose statins (80 mg/d) added to interferon-beta therapy found loss of clinical benefit or worsening of MRI, whereas studies with relatively low-dose statins (20 mg/d) are more variable.<sup>7,8,10,37,47,48</sup>

In the interferon-beta-only group of the SENTINEL trial (intramuscular interferon-beta-1a with or without natalizumab), a subgroup of 40 RRMS patients with ongoing disease activity while taking interferon-beta received low to high doses of various statins. No differences were found in clinical activity, CELs, or new T2 lesions.<sup>7</sup> In another study,<sup>47</sup> the total relapse rate was lower with 40 mg of simvastatin added on to intramuscular interferon-beta-1a, but the MRI results did not favor simvastatin. With low-dose atorvastatin added on (20 mg/d) to patients with active disease while receiving subcutaneous interferon-beta-1a therapy, CELs and relapses were reduced compared with baseline in the combination group vs the interferon monotherapy group.<sup>48</sup> In 16 RRMS patients with consistent baseline MRI activity, 80 mg of atorvastatin added on to 22 μg of interferon-beta-1a or to interferon-beta-1b therapy nonsignificantly reduced the number and volume of CELs vs baseline but increased T2 lesions for 9 months.<sup>38</sup> A parallel atorvastatin-only group showed similar effects, so regression to the mean is possible. A large phase 4 study (307 RRMS patients) demonstrated that 80 mg of simvastatin added on to weekly interferon-beta-1a did not benefit clinical and MRI activity and suggested that simvastatin should not be added as treatment for RRMS.<sup>8</sup> Simvastatin (80 mg) (n = 21) and placebo (n = 16) groups had no difference in expression of interferon-β-inducible genes *IL10*, *TNFSF10*, *MX1*, and *IRF7* in PAX gene-collected whole blood, appropriately obtained 9 to 12 hours after injection of interferon-beta-1a intramuscularly.

Our *in vivo* study design differed from other studies<sup>7,8</sup> that found no changes in interferon responses. We used statistically powerful, paired, within-subject analysis to minimize variability between patients receiving or not receiving statin therapy vs cross-sectional comparisons between placebo and statin groups. We measured more stable protein production instead of mRNA and used MNCs instead of whole blood to eliminate the up to 15-fold higher signals from polymorphonuclear leukocytes and reticulocytes in whole blood.<sup>27</sup> Moreover, our serum interferon activity assay is much more sensitive than ELISA.<sup>24</sup>

We tested only 14 RRMS patients, but statistical significance was found for multiple measures. We did not study long-term statin effects on clinical and MRI activity in these 14 RRMS patients because prolonged block of interferon therapy could allow recurrence of clinical activity.<sup>10</sup> Different statins may have various effects on interferon-beta therapy based on their half-lives, pharmacokinetics, and blood-brain barrier penetration based on hydrophobicity vs hydrophilicity.<sup>28,49,50</sup> Divergent results among clinical studies could be due to various doses and forms of statins,<sup>51</sup> weekly vs every-other-day interferon-beta, effects on oligodendroglia and immune cells,<sup>9</sup> and wide pharmacogenomic divergence in response to statins.<sup>52</sup>

In conclusion, high-dose statin add-on therapy impaired the ability of interferon-beta to activate STAT1 and, in turn, to induce IRF-1, serum type 1 interferons, and MxA and viperin proteins. More important, subtle shifts in immune cell activation or expression of regulatory proteins can disproportionately increase an ordinarily small percentage of autoreactive cells.<sup>53</sup> This study provides evidence that high-dose statins (80 mg/d) inhibit interferon effects by targeting STAT1 activation *in vitro* and during interferon-beta therapy. This finding suggests that MS patients who have high cholesterol levels should be cautious when combining high-dose statin therapy with interferon-beta.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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## References

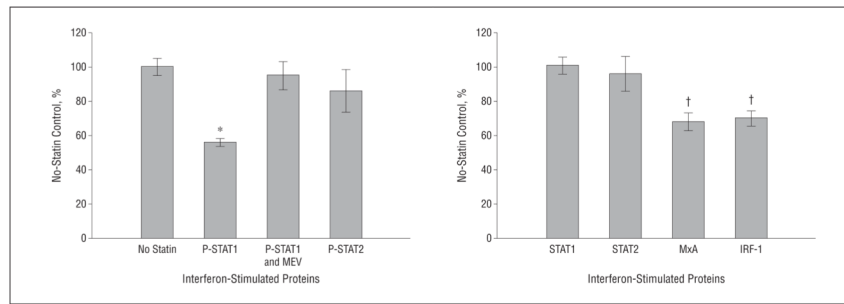
1. Lassmann H, Brück W, Lucchinetti CF. The immunopathology of multiple sclerosis: an overview. *Brain Pathol.* 2007; 17(2):210–218. [PubMed: 17388952]
2. Feng X, Petraglia AL, Chen M, Byskosh PV, Boos MD, Reder AT. Low expression of interferon-stimulated genes in active multiple sclerosis is linked to subnormal phosphorylation of STAT1. *J Neuroimmunol.* 2002; 129(1–2):205–215. [PubMed: 12161037]
3. Yamaguchi KD, Ruderman DL, Croze E, et al. IFN- $\beta$ -regulated genes are abnormally expressed in therapy-naïve MS mononuclear cells: unbiased gene expression analysis parallels literature on signaling pathways. *J Neuroimmunol.* 2008; 195:116–120. [PubMed: 18279974]
4. Youssef S, Stüve O, Patarroyo JC, et al. The HMG-CoA reductase inhibitor, atorvastatin, promotes a Th2 bias and reverses paralysis in central nervous system autoimmune disease. *Nature.* 2002; 420(6911):78–84. [PubMed: 12422218]
5. Greenwood J, Steinman L, Zamvil SS. Statin therapy and autoimmune disease: from protein prenylation to immunomodulation. *Nat Rev Immunol.* 2006; 6 (5):358–370. [PubMed: 16639429]

6. Stüve O, Youssef S, Weber MS, et al. Immunomodulatory synergy by combination of atorvastatin and glatiramer acetate in treatment of CNS autoimmunity. *J Clin Invest*. 2006; 116(4):1037–1044. [PubMed: 16543951]
7. Rudick RA, Pace A, Rani MR, et al. Effect of statins on clinical and molecular responses to intramuscular interferon beta-1a. *Neurology*. 2009; 72(23):1989–1993. [PubMed: 19506220]
8. Sorensen PS, Lycke J, Erälinna JP, et al. SIMCOMBIN study investigators. Simvastatin as add-on therapy to interferon  $\beta$ -1a for relapsing-remitting multiple sclerosis (SIMCOMBIN study): a placebo-controlled randomised phase 4 trial. *Lancet Neurol*. 2011; 10(8):691–701. [PubMed: 21742556]
9. Sellner J, Weber MS, Vollmar P, Mattle HP, Hemmer B, Stüve O. The combination of interferon-beta and HMG-CoA reductase inhibition in multiple sclerosis: enthusiasm lost too soon? *CNS Neurosci Ther*. 2010; 16(6):362–373. [PubMed: 20626428]
10. Birnbaum G, Cree B, Altafullah I, Zinser M, Reder AT. Combining beta interferon and atorvastatin may increase disease activity in multiple sclerosis. *Neurology*. 2008; 71(18):1390–1395. [PubMed: 18525027]
11. Stark GR, Kerr IM, Williams BR, Silverman RH, Schreiber RD. How cells respond to interferons. *Annu Rev Biochem*. 1998; 67:227–264. [PubMed: 9759489]
12. Takaoka A, Yanai H. Interferon signalling network in innate defence. *Cell Microbiol*. 2006; 8(6):907–922. [PubMed: 16681834]
13. Ghislain JJ, Fish EN. Application of genomic DNA affinity chromatography identifies multiple interferon- $\alpha$ -regulated Stat2 complexes. *J Biol Chem*. 1996; 271(21):12408–12413. [PubMed: 8647845]
14. Li X, Leung S, Qureshi S, Darnell JE Jr, Stark GR. Formation of STAT1-STAT2 heterodimers and their role in the activation of IRF-1 gene transcription by interferon- $\alpha$ . *J Biol Chem*. 1996; 271(10):5790–5794. [PubMed: 8621447]
15. Reder AT. MxA: a biomarker for predicting multiple sclerosis disease activity. *Neurology*. 2010; 75(14):1222–1223. [PubMed: 20921508]
16. Hinson ER, Joshi NS, Chen JH, et al. Viperin is highly induced in neutrophils and macrophages during acute and chronic lymphocytic choriomeningitis virus infection. *J Immunol*. 2010; 184(10):5723–5731. [PubMed: 20410488]
17. Vosslamber S, van Baarsen LG, Verweij CL. Pharmacogenomics of IFN- $\beta$  in multiple sclerosis: towards a personalized medicine approach. *Pharmacogenomics*. 2009; 10(1):97–108. [PubMed: 19102719]
18. Ghittoni R, Patrussi L, Pirozzi K, et al. Simvastatin inhibits T-cell activation by selectively impairing the function of Ras superfamily GTPases. *FASEB J*. 2005; 19(6):605–607. [PubMed: 15677697]
19. Jabs WJ, Hennig, Zawatzky R, Kirchner H. Failure to detect antiviral activity in serum and plasma of healthy individuals displaying high activity in ELISA for IFN-alpha and IFN-beta. *J Interferon Cytokine Res*. 1999; 19(5):463–469. [PubMed: 10386858]
20. Niewold TB, Kelly JA, Flesch MH, Espinoza LR, Harley JB, Crow MK. Association of the IRF5 risk haplotype with high serum interferon- $\alpha$  activity in systemic lupus erythematosus patients. *Arthritis Rheum*. 2008; 58(8):2481–2487. [PubMed: 18668568]
21. Hua J, Kirou K, Lee C, Crow MK. Functional assay of type I interferon in systemic lupus erythematosus plasma and association with anti-RNA binding protein autoantibodies. *Arthritis Rheum*. 2006; 54(6):1906–1916. [PubMed: 16736505]
22. Niewold TB, Hua J, Lehman TJ, Harley JB, Crow MK. High serum IFN- $\alpha$  activity is a heritable risk factor for systemic lupus erythematosus. *Genes Immun*. 2007; 8(6):492–502. [PubMed: 17581626]
23. Niewold TB, Kariuki SN, Morgan GA, Shrestha S, Pachman LM. Elevated serum interferon- $\alpha$  activity in juvenile dermatomyositis: associations with disease activity at diagnosis and after thirty-six months of therapy. *Arthritis Rheum*. 2009; 60(6):1815–1824. [PubMed: 19479879]
24. Feng X, Reder NP, Yanamandala M, et al. Type I interferon signature is high in lupus and neuromyelitis optica but low in multiple sclerosis. *J Neurol Sci*. 2012; 313(1–2):48–53. [PubMed: 22036215]



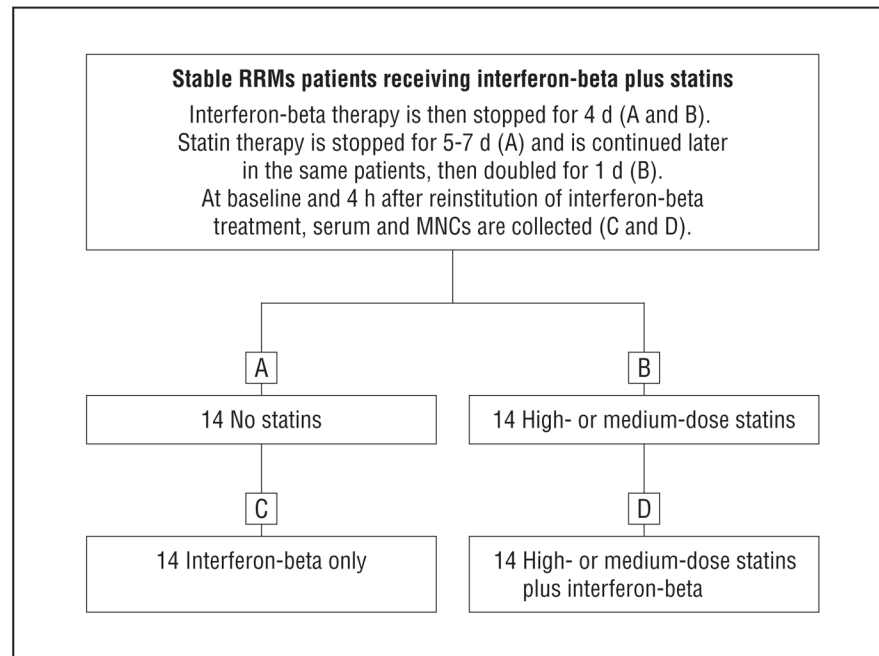
25. van der Voort LF, Vennegoor A, Visser A, et al. Spontaneous MxA mRNA level predicts relapses in patients with recently diagnosed MS. *Neurology*. 2010; 75(14):1228–1233. [PubMed: 20921509]
26. Kwak B, Mulhaupt F, Myit S, Mach F. Statins as a newly recognized type of immunomodulator. *Nat Med*. 2000; 6(12):1399–1402. [PubMed: 11100127]
27. Reder AT, Velichko S, Yamaguchi KD, et al. IFN- $\beta$ 1b induces transient and variable gene expression in relapsing-remitting multiple sclerosis patients independent of neutralizing antibodies or changes in IFN receptor RNA expression. *J Interferon Cytokine Res*. 2008; 28(5):317–331. [PubMed: 18547162]
28. Youssef S, Stüve O, Patarroyo JC, et al. The HMG-CoA reductase inhibitor, atorvastatin, promotes a Th2 bias and reverses paralysis in central nervous system autoimmune disease. *Nature*. 2002; 420(6911):78–84. [PubMed: 12422218]
29. Zamvil SS, Steinman L. Cholesterol-lowering statins possess anti-inflammatory activity that might be useful for treatment of MS. *Neurology*. 2002; 59(7):970–971. [PubMed: 12370448]
30. Stüve O, Prod'homme T, Slavin A, et al. Statins and their potential targets in multiple sclerosis therapy. *Expert Opin Ther Targets*. 2003; 7(5):613–622. [PubMed: 14498824]
31. Neuhaus O, Stüve O, Archelos JJ, Hartung HP. Putative mechanisms of action of statins in multiple sclerosis: comparison to interferon- $\beta$  and glatiramer acetate. *J Neurol Sci*. 2005; 233(1–2):173–177. [PubMed: 15949504]
32. Peng X, Jin J, Giri S, et al. Immunomodulatory effects of 3-hydroxy-3-methylglutaryl coenzyme-A reductase inhibitors, potential therapy for relapsing remitting multiple sclerosis. *J Neuroimmunol*. 2006; 178(1–2):130–139. [PubMed: 16870268]
33. Aktas O, Waiczies S, Smorodchenko A, et al. Treatment of relapsing paralysis in experimental encephalomyelitis by targeting Th1 cells through atorvastatin. *J Exp Med*. 2003; 197(6):725–733. [PubMed: 12629065]
34. Zhang X, Jin J, Peng X, Ramgolam VS, Markovic-Plese S. Simvastatin inhibits IL-17 secretion by targeting multiple IL-17-regulatory cytokines and by inhibiting the expression of IL-17 transcription factor RORC in CD4<sup>+</sup> lymphocytes. *J Immunol*. 2008; 180(10):6988–6996. [PubMed: 18453621]
35. Zhang X, Tao Y, Troiani L, Markovic-Plese S. Simvastatin inhibits IFN regulatory factor 4 expression and Th17 cell differentiation in CD4<sup>+</sup> T cells derived from patients with multiple sclerosis. *J Immunol*. 2011; 187(6):3431–3437. [PubMed: 21856936]
36. Vollmer T, Key L, Durkalski V, et al. Oral simvastatin treatment in relapsing-remitting multiple sclerosis. *Lancet*. 2004; 363(9421):1607–1608. [PubMed: 15145635]
37. Paul F, Waiczies S, Wuerfel J, et al. Oral high-dose atorvastatin treatment in relapsing-remitting multiple sclerosis. *PLoS One*. 2008; 3(4):e1928. [PubMed: 18398457]
38. Zhao Y, Traboulsee A, Petkau AJ, Li D. Regression of new gadolinium enhancing lesion activity in relapsing-remitting multiple sclerosis. *Neurology*. 2008; 70(13 pt 2):1092–1097. [PubMed: 18003938]
39. Dunn SE, Youssef S, Goldstein MJ, et al. Isoprenoids determine Th1/Th2 fate in pathogenic T cells, providing a mechanism of modulation of autoimmunity by atorvastatin. *J Exp Med*. 2006; 203(2):401–412. [PubMed: 16476765]
40. Ghittoni R, Napolitani G, Benati D, et al. Simvastatin inhibits the MHC class II pathway of antigen presentation by impairing Ras superfamily GTPases [published correction appears in *Eur J Immunol*. 2006;36(12):3381]. *Eur J Immunol*. 2006; 36(11):2885–2893. [PubMed: 17048274]
41. Feng X, Heyden NV, Ratner L. Alpha interferon inhibits human T-cell leukemia virus type 1 assembly by preventing Gag interaction with rafts. *J Virol*. 2003; 77(24):13389–13395. [PubMed: 14645593]
42. Kuipers HF, Biesta PJ, Groothuis TA, Neeffjes JJ, Mommaas AM, van den Elsen PJ. Statins affect cell-surface expression of major histocompatibility complex class II molecules by disrupting cholesterol-containing microdomains. *Hum Immunol*. 2005; 66(6):653–665. [PubMed: 15993711]
43. Ehrenstein MR, Jury EC, Mauri C. Statins for atherosclerosis: as good as it gets? *N Engl J Med*. 2005; 352(1):73–75. [PubMed: 15635116]

44. Fessler MB, Parks JS. Intracellular lipid flux and membrane microdomains as organizing principles in inflammatory cell signaling. *J Immunol.* 2011; 187(4):1529–1535. [PubMed: 21810617]
45. Miron VE, Zehntner SP, Kuhlmann T, et al. Statin therapy inhibits remyelination in the central nervous system. *Am J Pathol.* 2009; 174(5):1880–1890. [PubMed: 19349355]
46. Sellner J, Greeve I, Findling O, et al. Effect of interferon-beta and atorvastatin on Th1/Th2 cytokines in multiple sclerosis. *Neurochem Int.* 2008; 53(1–2):17–21. [PubMed: 18524417]
47. Togha M, Karvigh SA, Nabavi M, et al. Simvastatin treatment in patients with relapsing-remitting multiple sclerosis receiving interferon beta 1a: a double-blind randomized controlled trial. *Mult Scler.* 2010; 16(7):848–854. [PubMed: 20488825]
48. Lanzillo R, Orefice G, Quarantelli M, et al. Atorvastatin Combined to Interferon to Verify the Efficacy (ACTIVE) in relapsing-remitting active multiple sclerosis patients: a longitudinal controlled trial of combination therapy. *Mult Scler.* 2010; 16(4):450–454. [PubMed: 20150398]
49. Corsini A, Bellosta S, Baetta R, Fumagalli R, Paoletti R, Bernini F. New insights into the pharmacodynamic and pharmacokinetic properties of statins. *Pharmacol Ther.* 1999; 84(3):413–428. [PubMed: 10665838]
50. Butterfield DA, Barone E, Mancuso C. Cholesterol-independent neuroprotective and neurotoxic activities of statins: perspectives for statin use in Alzheimer disease and other age-related neurodegenerative disorders. *Pharmacol Res.* 2011; 64(3):180–186. [PubMed: 21536132]
51. Neuvonen PJ, Niemi M, Backman JT. Drug interactions with lipid-lowering drugs: mechanisms and clinical relevance. *Clin Pharmacol Ther.* 2006; 80(6):565–581. [PubMed: 17178259]
52. Kajinami K, Akao H, Polisecki E, Schaefer EJ. Pharmacogenomics of statin responsiveness. *Am J Cardiol.* 2005; 96(9A):65K–70K.
53. Germain RN. The art of the probable: system control in the adaptive immune system. *Science.* 2001; 293(5528):240–245. [PubMed: 11452112]

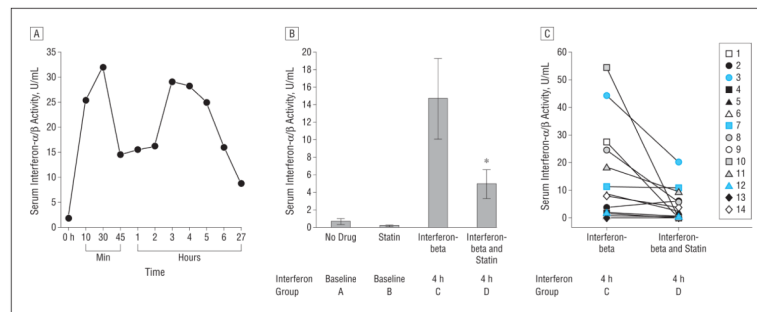


**Figure 1.**

In vitro atorvastatin reduces interferon-beta effects. Mononuclear cells from 21 therapy-naive patients with relapsing-remitting multiple sclerosis were pretreated at 24 hours with 10- $\mu$ M atorvastatin, then induced with 160 U/mL of interferon-beta-1b for 45 minutes (phosphorylated/activated STAT transcription factors) and 24 hours (downstream proteins, STAT1, STAT2, interferon regulatory factor 1 [IRF-1], and myxovirus resistance 1 [MxA]). Proteins were quantified with Western blots and normalized with actin. \* $P < .001$  vs no-statin control, † $P < .05$ . MEV indicates 100- $\mu$ M mevalonate. Error bars indicate SEM.

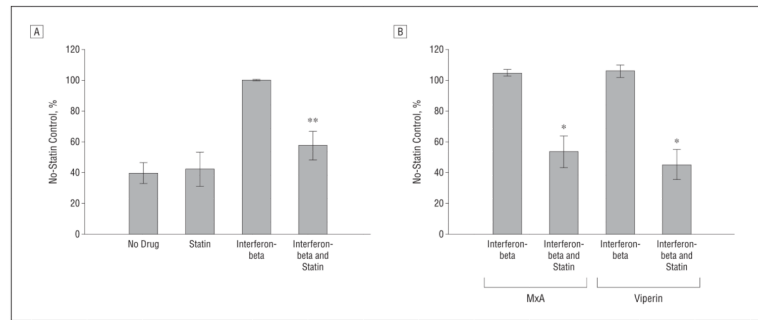


**Figure 2.** Fourteen clinically stable patients with relapsing-remitting multiple sclerosis (RRMS) receiving interferon-beta plus statin therapy stopped interferon-beta therapy and stopped (A and C) or continued medium- or high-dose (B and D) statin therapy. Serum type 1 interferon activity and Western blots of STAT1 and STAT2 phosphorylation were performed at 0 and 4 hours after interferon-beta injection; *in vivo*-induced myxovirus resistance 1 and viperin proteins were measured with Western blots at 24 hours. MNCs indicates mononuclear cells.



**Figure 3.** Statins reduce interferon-beta therapy induction of serum type 1 interferon activity in 14 stable patients with relapsing-remitting multiple sclerosis. A, In vivo Rebif kinetics after a 3-day washout. B and C, Statin add-on therapy blocks interferon-beta therapy induction of serum interferon- $\alpha/\beta$  activity in 14 patients with relapsing-remitting multiple sclerosis. Serum samples were obtained at 8 am after statin washout or long-term statin alone and then exactly 4 hours after interferon-beta injections or high-dose statins plus 4 hours of interferon-beta therapy. \* $P < .001$  vs interferon alone (paired  $t$  test). Error bars indicate SEM.





**Figure 4.** Comparison of interferon and interferon with statin therapy. The addition of a statin blocks interferon-beta therapy induction of P-Y-STAT1 (A) and myxovirus resistance 1 (MxA) and viperin proteins (B) in 14 patients with relapsing-remitting multiple sclerosis. Mononuclear cell lysates from no drug, statin alone, interferon-beta alone at 4 hours, and 4 hours of interferon-beta plus statin conditions are shown. Proteins were quantified with Western blots and normalized with actin. \* $P < .001$  vs interferon-beta (paired  $t$  test). Error bars indicate SEM.