

Efficacy of Three-Week Oxytetracycline or Rifampin Monotherapy Compared with a Combination Regimen against the Filarial Nematode *Onchocerca ochengi*

Germanus S. Bah,^{a,c} Emma L. Ward,^a Abhishek Srivastava,^{b*} Alexander J. Trees,^{a,b} Vincent N. Tanya,^c Benjamin L. Makepeace^{a,b}

Institute of Infection and Global Health, University of Liverpool, Liverpool, United Kingdom^a; Liverpool School of Tropical Medicine, Liverpool, United Kingdom^b; Institut de Recherche Agricole pour le Développement, Regional Centre of Wakwa, Ngaoundéré, Adamawa Region, Cameroon^c

Onchocerciasis (river blindness), caused by the filarial nematode *Onchocerca volvulus*, is a major cause of visual impairment and dermatitis in sub-Saharan Africa. As *O. volvulus* contains an obligatory bacterial symbiont (*Wolbachia*), it is susceptible to antibiotic chemotherapy, although current regimens are considered too prolonged for community-level control programs. The aim of this study was to compare the efficacies of oxytetracycline and rifampin, administered separately or in combination, against a close relative of *O. volvulus* (*Onchocerca ochengi*) in cattle. Six animals per group were treated with continuous or intermittent oxytetracycline regimens, and effects on adult worm viability, dermal microfilarial loads, and *Wolbachia* density in worm tissues were assessed. Subsequently, the efficacies of 3-week regimens of oxytetracycline and rifampin alone and a combination regimen were compared, and rifampin levels in plasma and skin were quantified. A 6-month regimen of oxytetracycline with monthly dosing was strongly adulticidal, while 3-week and 6-week regimens exhibited weaker adulticidal effects. However, all three regimens achieved >2-log reductions in microfilarial load. In contrast, rifampin monotherapy and oxytetracycline-rifampin duotherapy failed to induce substantive reductions in either adult worm burden or microfilarial load, although a borderline effect on *Wolbachia* density was observed following duotherapy. Dermal rifampin levels were maintained above the MIC for >24 h after a single intravenous dose. We conclude that oxytetracycline-rifampin duotherapy is less efficacious against *O. ochengi* than oxytetracycline alone. Further studies will be required to determine whether rifampin reduces oxytetracycline bioavailability in this system, as suggested by human studies using other tetracycline-rifampin combinations.

Onchocerciasis, or “river blindness,” is a vector-borne parasitic disease that continues to affect 26 million people, predominantly in sub-Saharan Africa, despite 4 decades of concerted control efforts (1). The etiological agent, *Onchocerca volvulus*, is a filarial worm that resides in subcutaneous nodules when mature and has a life span of >10 years (2). Although the nodules are generally benign, the female worm releases approximately 1,000 first-stage larvae (microfilariae [Mf]) per day (3), which accumulate in the skin and can induce severe pruritus and dermatitis. After several years, the Mf can also migrate into the eyes, where their death prompts local inflammatory reactions, visual impairment, and, ultimately, irreversible blindness. Whereas the original Onchocerciasis Control Programme was initially focused on vector control, the current African Programme for Onchocerciasis Control (APOC) is almost entirely dependent on mass drug administration (MDA) of the macrocyclic lactone ivermectin (4). This strategy has been in place in many regions of endemicity for approximately 2 decades and has been extremely successful in preventing blindness and onchodermatitis (5), since ivermectin rapidly clears Mf from the skin. Recent data have also demonstrated that ivermectin can eliminate onchocerciasis in areas of seasonal transmission (6), and this has led APOC to redefine itself as an elimination program with a target attainment date of 2025 (4). However, ivermectin MDA faces a number of challenges that may compromise the progress of APOC over the next decade. These include severe adverse events following ivermectin treatment in individuals heavily coinfecting with *Loa loa* (a species of filarial worm endemic to Central Africa) (7), continued transmission of onchocerciasis in perennial-transmission zones despite 15 to 18 years of MDA (8, 9), evidence of decreased ivermectin sus-

ceptibility in some worm populations (10, 11), variable compliance with MDA within affected communities (12), and a lack of adulticidal efficacy against the parasite (13).

Until relatively recently, no safe adulticidal drug for onchocerciasis existed. This changed with the identification of *Wolbachia* endobacteria (order *Rickettsiales*) in filariae of medical importance in the late 1990s (14, 15), which prompted a series of experiments in animal models demonstrating that antibiotics (especially the tetracyclines) not only impede the growth and embryogenesis of filarial worms (16, 17) but also can kill the adult parasites (18). Such effects were not observed at clinically relevant doses in filarial species that naturally lack *Wolbachia* symbionts (16, 19). Clinical trials of doxycycline (DOX) for human onchocerciasis were implemented rapidly, which achieved sterilization of female worms using a regimen of 200 mg/day for 4 weeks (20) or 100 mg/day for 5 weeks (21). However, significant adulticidal activity (killing of 60 to 70% of female worms) required a regimen of 200 mg/day for 6 weeks (20). This relatively protracted

Received 13 September 2013 Returned for modification 18 October 2013

Accepted 12 November 2013

Published ahead of print 18 November 2013

Address correspondence to Benjamin L. Makepeace, blm1@liv.ac.uk.

* Present address: Abhishek Srivastava, Drug Metabolism and Pharmacokinetics, Infection IMED, AstraZeneca India Pvt. Ltd., Bangalore, India.

Copyright © 2014, American Society for Microbiology. All Rights Reserved.

doi:10.1128/AAC.01995-13

The authors have paid a fee to allow immediate free access to this article.

TABLE 1 Sample sizes and treatment regimens for experiments 1 and 2

Expt	Group designation	Original group size		No. of animals lost during expt	Treatment regimen (duration) ^a
		Females	Males		
1	CON-1	6	0	1	None
1	PIR	6	0	1	OXY at 20 mg/kg i.m. once/mo (6 mo)
1	OXY3-1	6	0	0	OXY at 20 mg/kg i.m. twice/wk (3 wk)
1	OXY6	6	0	2	OXY at 20 mg/kg i.m. twice/wk (6 wk)
2	CON-2	6	0	1	DMSO at 220 mg/kg i.v. twice/wk (3 wk)
2	RIF3	5	1	0	RIF at 10 mg/kg i.v. and DMSO 220 mg/kg i.v. twice/wk (3 wk)
2	OXY3-2	6	0	0	OXY at 20 mg/kg i.m. twice/wk (3 wk)
2	COM	5	1	0	RIF at 10 mg/kg i.v., DMSO at 220 mg/kg i.v., and OXY at 20 mg/kg i.m. twice/wk (3 wk)

^a OXY, oxytetracycline; i.m., intramuscularly; DMSO, dimethyl sulfoxide; i.v., intravenously; RIF, rifampin.

course of treatment, coupled with contraindications in children below 8 years of age and in pregnant or lactating women, have prevented approval of DOX for MDA to date; this is despite the very high rates of compliance evident in a clinical trial that was conducted in a region of Cameroon where loiasis is endemic (22). Nevertheless, DOX has been applied in a small onchocerciasis focus in Venezuela to expedite elimination efforts (23).

Experiments performed *in vitro* using isolated worms or *Wolbachia*-infected cell lines (24–26), as well as *in vivo* trials in rodent models (27, 28), have indicated that rifampin (RIF) is at least as effective as the tetracyclines for symbiont depletion and, indeed, may be superior. However, two human trials of RIF on onchocerciasis failed to demonstrate that this bactericide could truncate the therapeutic duration significantly, as a 5-day regimen had no effect on either adult worms or microfilarial densities (29), whereas 2-week and 4-week regimens induced a partial embryostatic effect but were not adulticidal (30). Although these data were equivocal, there remains the possibility that a combination of a tetracycline and RIF substantially shortens the regimen required to achieve potent adulticidal effects.

A major challenge in onchocerciasis research is the inability of *Onchocerca* spp. to complete their life cycles in rodent models. However, in cattle, *Onchocerca ochengi*, which is the closest extant relative of *O. volvulus* (31), has been used extensively to investigate drug efficacy for onchocerciasis, and numerous bovine studies have displayed strong concordance with data obtained from human chemotherapeutic trials (18, 32, 33). Importantly, adult worms of *O. ochengi* reside in intradermal nodules with a histological structure highly similar to that of *O. volvulus*, and the parasites are fertile for many years (34). Here, we use this natural host-parasite system to determine whether the addition of RIF to an oxytetracycline (OXY) regimen with limited activity against adult worms can reduce the treatment duration for unequivocal parasite killing to 3 weeks.

MATERIALS AND METHODS

Drugs. Oxytetracycline dihydrate was obtained as a commercially available, long-acting formulation (Terramycin LA, 200 mg/ml; Pfizer Animal Health, Tadworth, United Kingdom) that is licensed for use in cattle. RIF was kindly donated by Lupin Ltd. (Mumbai, India) in solid form as the unformulated active pharmaceutical ingredient. For administration to cattle, the powder was dissolved at 1% (wt/vol) in 25 mM HEPES buffer (pH 7.0 to 7.6), 20% (vol/vol) dimethyl sulfoxide (DMSO) solvent (both cell culture grade; Sigma-Aldrich, Gillingham, United Kingdom), and sterile normal saline in intravenous bags. Prior to injection, the RIF solu-

tion was warmed to 37°C in an incubator to ensure complete dissolution and was transported to the field protected from light.

Animals, chemotherapy, and sampling strategy. Zebu cattle (Ngaoundéré Gudali breed), naturally infected with ≥ 20 palpable *O. ochengi* nodules per animal, were purchased from markets across the Adamaoua Region of Cameroon (Vina Division) and assembled at the Institut de Recherche Agricole pour le Développement (IRAD), Regional Centre of Wakwa, where transmission of *O. ochengi* is negligible. The pretreatment recording of nodule position and the randomization of animals into treatment groups were performed as previously described (35). Evaluations of antibiotic efficacy were conducted in two consecutive experiments (Table 1); the first compared continuous 3-week or 6-week OXY monotherapy (OXY3 and OXY6, respectively) with a prolonged intermittent OXY regimen (PIR) (36), whereas the second was designed to determine whether a 3-week RIF-plus-OXY combination regimen (COM) was superior to 3 weeks of RIF or OXY monotherapy. These experiments utilized different animals, with the exception of two cows from the first control (CON-1) group, which were reused in the second control (CON-2) group. Four animals from experiment 1 and one animal from experiment 2 died before the studies were completed (Tables 1 and 2) from causes unrelated to either onchocerciasis or the drug treatments.

Two nodules per animal were removed under local anesthesia in a randomized sequence at six predetermined time points, transferred to the laboratory in phosphate-buffered saline (PBS), and trimmed of excess tissue. One nodule was fixed by injection with 10% (vol/vol) neutral buffered formalin, while the diameter of the other was measured with calipers prior to parasitological analysis. Three superficial skin biopsy specimens per animal were also obtained from the ventral underside at each time point and transported to the laboratory in PBS for the determination of Mf density.

Ethical considerations. This study was approved by the Ethics Committee of the Regional Centre of Wakwa, IRAD, and authorized by the Regional Programmes Committee of IRAD. All procedures performed on cattle in Cameroon were equivalent to those previously authorized by a Home Office Project License (Animals [Scientific Procedures] Act, 1986) for experimental infections of cattle in the United Kingdom and classified under the severity limit “mild.”

Parasitology. Nodules were dissected in PBS, the male and female worms were separated, and their motility was scored on a 3-point scale after incubation at 37°C for 30 min as detailed previously (32). The Mf in skin biopsy specimens were quantified and normalized to densities per 100 mg skin as described previously (32).

Immunohistochemistry and semiquantification of *Wolbachia*. For experiment 2, the formalin-fixed nodules were embedded in paraffin and 4- μ m sections were probed with an anti-*Wolbachia* surface protein polyclonal antibody (kindly provided by M. Casiraghi, University of Milan, Italy) using the universal LSAB2 horseradish peroxidase kit (Dako United Kingdom Ltd., Ely, United Kingdom) as detailed previously (36). The

TABLE 2 Effect of oxytetracycline, rifampin, and a combination regimen on adult worms of *Onchocerca ochengi* in naturally infected cattle

Group ^a	Time (wpt)	No. of cattle	No. of nodules		No. of worms per nodule site (no. of viable worms only ^b) in:		Median motility score in ^c :	
			Intact	Degenerated ^d	Males	Females	Males	Females
CON-1	0	6	6	0	1.7	1.3	2	2
	4	6	6	0	3.3	1.3	1	2
	12	5	5	0	0.8	1.2	1.5	2
	24	5	5	0	1.4	1.0 (0.8)	2	1
	36	5	5	0	2.2 (2.0)	1.0 (0.8)	1	1
	55	5	5	0	0.8	1.0 (0.8)	2	2
PIR	0	6	6	0	4	1.2	2	2
	4	6	6	0	1.5	1.0	1	1.5
	12	5	5	0	0.8	1.0	2	1
	24	5	4	1	0.2 (1.0)	1.2	2	1
	36	5	5	0	0.4 (0.0)	1.0 (0.2)	0	0
	55 ^e	5	2	3	0	0.4 (0.2)	— ^f	1
OXY3-1	0	6	6	0	1.5	1.0	2	2
	4	6	6	0	1.5	1.0	1	2
	12	6	6	0	0.8	1.0	2	1.5
	24	6	5	1	1.3	0.8 (0.7)	1	1
	36	6	6	0	1.0 (0.3)	1.0 (0.2)	0	0
	55	6	2	4	0.2	0.3	2	1
OXY6	0	6	6	0	1.5	1.2	2	2
	4	6	6	0	1.7	1.0 (0.8)	2	1.5
	12	4	4	0	2.0	1.0	2	2
	24	4	4	0	3.0 (2.0)	1.3 (1.0)	1	1
	36	4	4	0	0.3	1.0 (0.5)	1	0.5
	55 ^e	4	2	2	0	0.5 (0.3)	— ^f	0.5
CON-2	0	6	6	0	0.8	1.0	2	2
	4	6	6	0	1.2	1.0	2	2
	12	6	6	0	1.5	1.0	2	2
	24	6	6	0	1.5	1.0 (0.3)	2	0
	36	6	5	1	1.5	0.8	2	2
	52	5	5	0	1.8	1.0	2	2
OXY3-2	0	6	6	0	1.7	1.0	2	2
	4	6	6	0	0.3	1.2 (1.0)	1.5	1
	12	6	6	0	0.3	1.0	1.5	1
	24	6	6	0	0.5	1.0 (0.7)	2	1
	36	6	6	0	0.7	1.0 (0.8)	2	1
	52 ^e	6	6	0	0.3	1.0 (0.5)	1	0.5
RIF3	0	6	6	0	1.2	1.0 (0.8)	2	2
	4	6	6	0	2.0	1.0	1	1
	12	6	6	0	1.3	1.0	2	2
	24	6	6	0	0.5	1.0 (0.8)	2	1
	36	6	6	0	0.3	1.0 (0.5)	2	0.5
	52	6	6	0	0.8 (0.5)	1.0 (0.8)	1	1
COM	0	6	6	0	0.8	1.0	2	1.5
	4	6	6	0	1.3	1.0	2	2
	12	6	6	0	0.5	1.0	2	2
	24	6	6	0	1.3	1.0 (0.8)	2	1
	36	6	5	1	1.0	0.8	1.5	2
	52	6	6	0	0.3	1.0 (0.7)	1.5	2

^a CON-1, no treatment; PIR, oxytetracycline once monthly for 6 months; OXY3-1 and OXY3-2, oxytetracycline twice weekly for 3 weeks; OXY6, oxytetracycline twice weekly for 6 weeks; CON-2, DMSO solvent only; RIF3, rifampin twice weekly for 3 weeks; COM, rifampin and oxytetracycline twice weekly for 3 weeks.

^b Worms were considered viable where the motility score was ≥ 1 . Note that *O. ochengi* nodules normally contain a single female worm and a variable number of males (0 to >10).

^c Motility was scored on a 3-point scale after incubation of worms for 30 min at 37°C, as follows: 0, no movement; 1, sluggish movement; and 2, highly active.

^d "Degenerated" refers to worm remnants only or fragmented nodules from skin biopsy specimens (in which fragmentation was too advanced to allow sexing and motility scoring).

^e The total number of viable worms recovered at the end of the experiment was significantly reduced compared with the number in control animals ($P < 0.05$).

^f Data were missing due to the absence of male worms in all nodules.

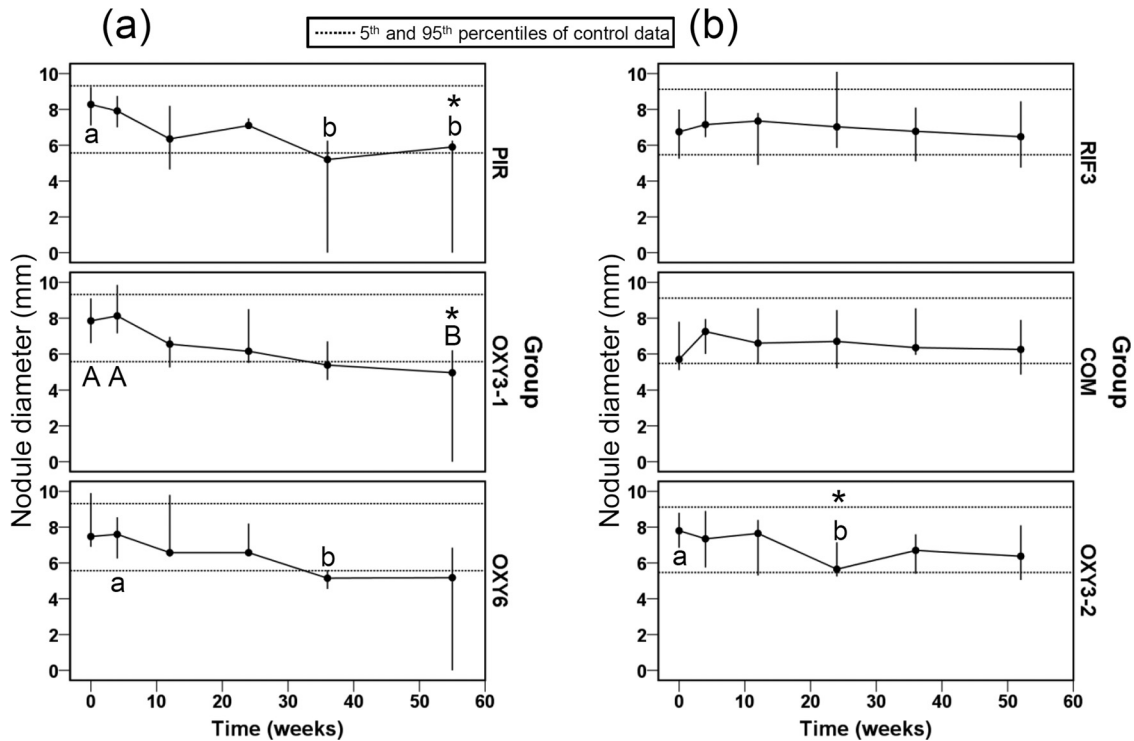


FIG 1 Nodule diameters (medians and ranges) in *Onchocerca ochengi*-infected cattle treated with oxytetracycline delivered once monthly for 6 months (PIR; $n = 5$), twice weekly for 3 weeks (OXY3-1; $n = 6$), or twice weekly for 6 weeks (OXY6; $n = 4$) (a) and with rifampin alone twice weekly for 3 weeks (RIF3; $n = 6$), rifampin and oxytetracycline twice weekly for 3 weeks (COM; $n = 6$), or oxytetracycline alone twice weekly for 3 weeks (OXY3-2; $n = 6$) (b). Within groups, medians at time points annotated with different letters are significantly different at a P of <0.05 (lowercase letters) or a P of <0.01 (uppercase letters) by Friedman's two-way ANOVA by ranks. Between groups, medians at time points annotated with an asterisk are significantly different from medians for the control group ($P < 0.05$ by the Mann-Whitney U test).

sections were counterstained with Giemsa stain, and digital images were obtained on a Microphot-FX microscope (Nikon, Tokyo, Japan) using a DCM900 microscope camera with imaging software (ScopeTek, Hangzhou, China). *Wolbachia* staining of worm sections across the entirety of each slide was semiquantified on a 3-point scale, as follows: 0, no visible bacteria; 1, sparse bacteria ($<50\%$ of worm sections were positive); and 2, profuse bacteria ($\geq 50\%$ of worm sections were positive).

Quantification of RIF in bovine plasma and skin. Four zebu cows that were extraneous to experiments 1 and 2 and did not receive other drugs were treated with a single dose of RIF at 10 mg/kg of body weight in DMSO at 220 mg/kg. A plasma sample and triplicate superficial skin biopsy specimens (mean weight, ~ 140 mg) were obtained from each animal at 0, 1, 8, 24, 48, and 72 h after treatment and were stored at -80°C . The skin biopsy specimens were thawed, combined for each animal, and macerated (protected from light) in 2 ml PBS using a T10 basic Ultra-Turrax disperser (IKA, Staufen, Germany) with an 8-mm dispersing element. To comply with the stipulations of a United Kingdom Department of Environment, Food, and Rural Affairs import license, both the skin homogenates and the plasma samples were heat inactivated at 56°C for 30 min prior to transportation to the United Kingdom on dry ice. RIF was quantified in the samples using a previously published liquid chromatographic-tandem mass spectrometric method with a lower limit of quantification of 25 ng/ml (37). Calibration standards containing known amounts of RIF (Sigma-Aldrich) were prepared separately in drug-free bovine plasma and skin homogenates obtained from United Kingdom abattoir material.

Statistical analyses. Fully quantitative time course data were analyzed by Friedman's two-way analysis of variance (ANOVA) by ranks, followed by all-pairwise comparisons (with P adjusted for multiple testing) if the significance of the omnibus test was <0.05 . Only animals with complete data sets at all six time points were included in the Friedman analyses.

Where within-group pairwise analyses were significant over time, medians between the treatment group and the respective control group at the equivalent time points were compared by the Mann-Whitney U test with exact significance. Fisher's exact test was used to analyze both the frequencies of viable and dead adult worms (both sexes combined) at the end of each experiment and the distributions of endobacterial density scores at 0, 24, and 36 weeks posttreatment start (wpt) in experiment 2. All analyses were performed with IBM SPSS Statistics v. 20 (IBM Corporation, Armonk, NY, USA), and differences were considered to be statistically significant at a critical probability of <0.05 .

RESULTS

Effects on nodule diameter and adult worm recovery. The objective of experiment 1 was to determine the simplest OXY regimen with partial adulticidal effects in order to facilitate the detection of modified efficacy when RIF was added in combination. Marked declines in nodule diameter were apparent in all three of the OXY-treated groups, although this became statistically significant at an earlier stage (36 wpt) in the PIR and OXY6 groups than in the OXY3-1 group (where 1 indicates experiment 1) (Fig. 1a). However, at the end of the experiment, only the PIR and OXY3-1 groups exhibited reduced median nodule diameters relative to those of the CON-1 group (Fig. 1a). Notably, one (OXY3-1, OXY6) or two (PIR) nodules per group had resolved completely at 55 wpt and were assigned a diameter of 0. For experiment 2, effects on nodule diameter were less pronounced than in the first experiment, with only the OXY3-2 group displaying a significant reduction in nodule diameter at 24 wpt compared with intragroup base-

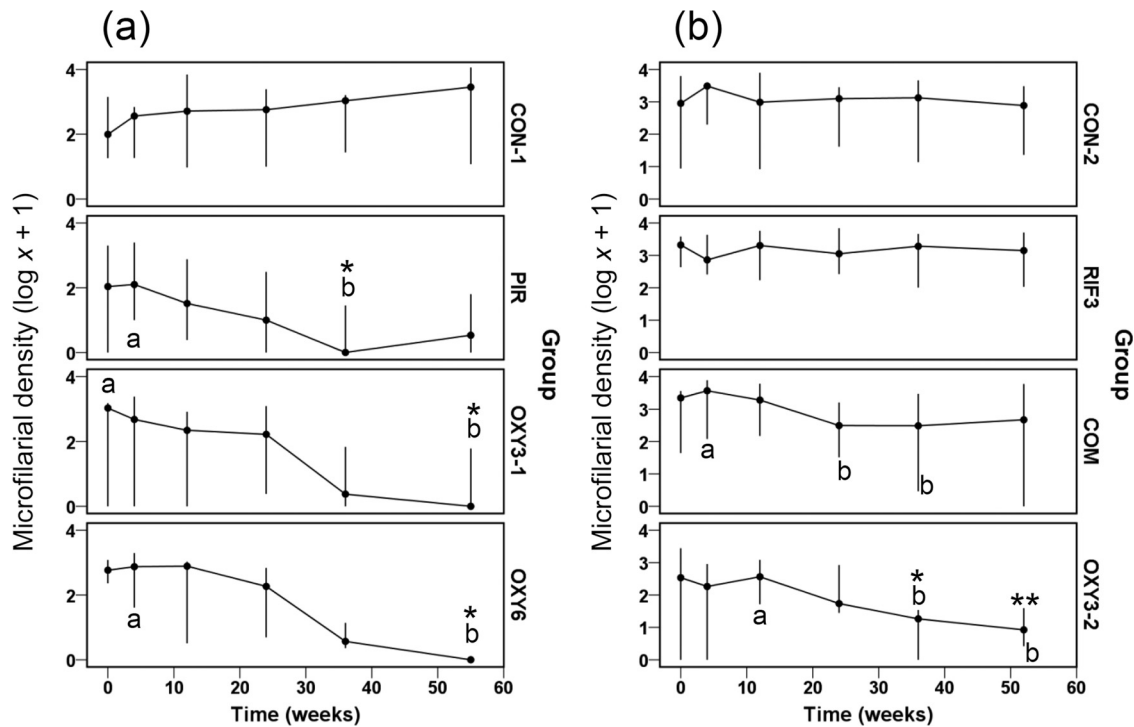


FIG 2 Microfilarial densities per 100 mg skin (medians and ranges, plotted on a $\log x + 1$ -transformed scale) in *Onchocerca ochengi*-infected cattle treated with oxytetracycline delivered once monthly for 6 months (PIR; $n = 5$), twice weekly for 3 weeks (OXY3-1; $n = 6$), or twice weekly for 6 weeks (OXY6; $n = 4$) (a) and with rifampin alone twice weekly for 3 weeks (RIF3; $n = 6$), rifampin and oxytetracycline twice weekly for 3 weeks (COM; $n = 6$), or oxytetracycline alone twice weekly for 3 weeks (OXY3-2; $n = 6$) (b). (a) Control animals (CON-1; $n = 5$) received no treatment; (b) control animals (CON-2; $n = 5$) received DMSO solvent at a dose equivalent to that of the RIF3 and COM groups. Within groups, medians at time points annotated with different letters are significantly different at a P of < 0.05 by Friedman's two-way ANOVA by ranks. Between groups, medians at time points annotated with asterisks are significantly different from those of the control group ($P < 0.05$ [*] or $P < 0.01$ [**] by the Mann-Whitney U test).

line data and CON-2 data at the equivalent time point (Fig. 1b). In addition, this reduction was not maintained at a statistically significant level at 36 and 52 wpt (Fig. 1b).

The marked adulticidal effect of the PIR treatment was confirmed by analysis of adult worm recovery and viability at the end of experiment 1 (Table 2). This regimen induced a 75% reduction in the final yield of viable adult female worms relative to that of the CON-1 group, and no adult male worms were recovered; the combined effect on both sexes was statistically significant ($P = 0.023$). For the shorter OXY regimens, the reduction in the recovery of female worms was 63% at 55 wpt, and no male worms were obtained from OXY6 nodules at this time point (Table 2). However, the decrease in total adult worm recovery was not statistically significant for the OXY3-1 and OXY6 groups, although a strong trend was apparent for the latter ($P = 0.052$). In accordance with the nodule diameter data, significant adulticidal effects were not apparent in experiment 2 for the RIF3 and COM groups (Table 2). In contrast, the OXY3-2 regimen achieved reductions in loads of viable female and male worms of 50% and 83%, respectively, which was statistically significant in aggregate ($P = 0.036$).

Effects on microfilarial density. In experiment 1, statistically significant declines in Mf density of 2 to 3 logs were observed with all three OXY regimens by 36 to 55 wpt, although complete elimination of Mf was achieved only in the OXY6 group (Fig. 2a). The reductions in Mf load were also statistically significant compared with loads in the absence of treatment (CON-1) for the PIR group at 36 wpt and for the OXY3-1 and OXY6 regimens at 55 wpt

(Fig. 2a). The lack of adulticidal effects exhibited by the RIF3 and COM groups in experiment 2 were also reflected by a limited impact on Mf, although a transient but statistically significant reduction in Mf density of ~ 1 log was apparent in the COM group at 24 and 36 wpt (Fig. 2b). Conversely, the efficacy of the OXY3-2 regimen against adult worms was mirrored by an ~ 2 -log within-group decline in Mf density at 36 and 52 wpt; moreover, this was statistically significant relative to Mf densities in the CON-2 group at these time points (Fig. 2b).

Effects on Wolbachia density in adult worms. To determine whether the poor efficacy of the RIF3 and COM regimens in experiment 2 was associated with failure to clear *Wolbachia* populations within adult worms, we stained histological sections of bovine nodules from 0, 24, and 36 wpt with an anti-*Wolbachia* surface protein antibody. In *O. ochengi* and in filarial nematodes of medical importance, including *O. volvulus*, the distribution of *Wolbachia* conforms to a highly predictable pattern, with high densities of endobacteria in the somatic hypodermal cords of both adult males and females, whereas only females contain symbionts in their gonads and, consequently, in developing embryos. Thus, sections from CON-2 nodules showed intense, punctate staining of the bacteria in the hypodermal cords of the worms and finer stippling of the symbionts in the ovaries and intrauterine embryonic stages (Fig. 3a to c). In the RIF3 and COM groups, extensive *Wolbachia* populations remained clearly visible across the time course in both the hypodermal cords and the female reproductive tracts (Fig. 3d to f and j to l, respectively); however, oocytes and

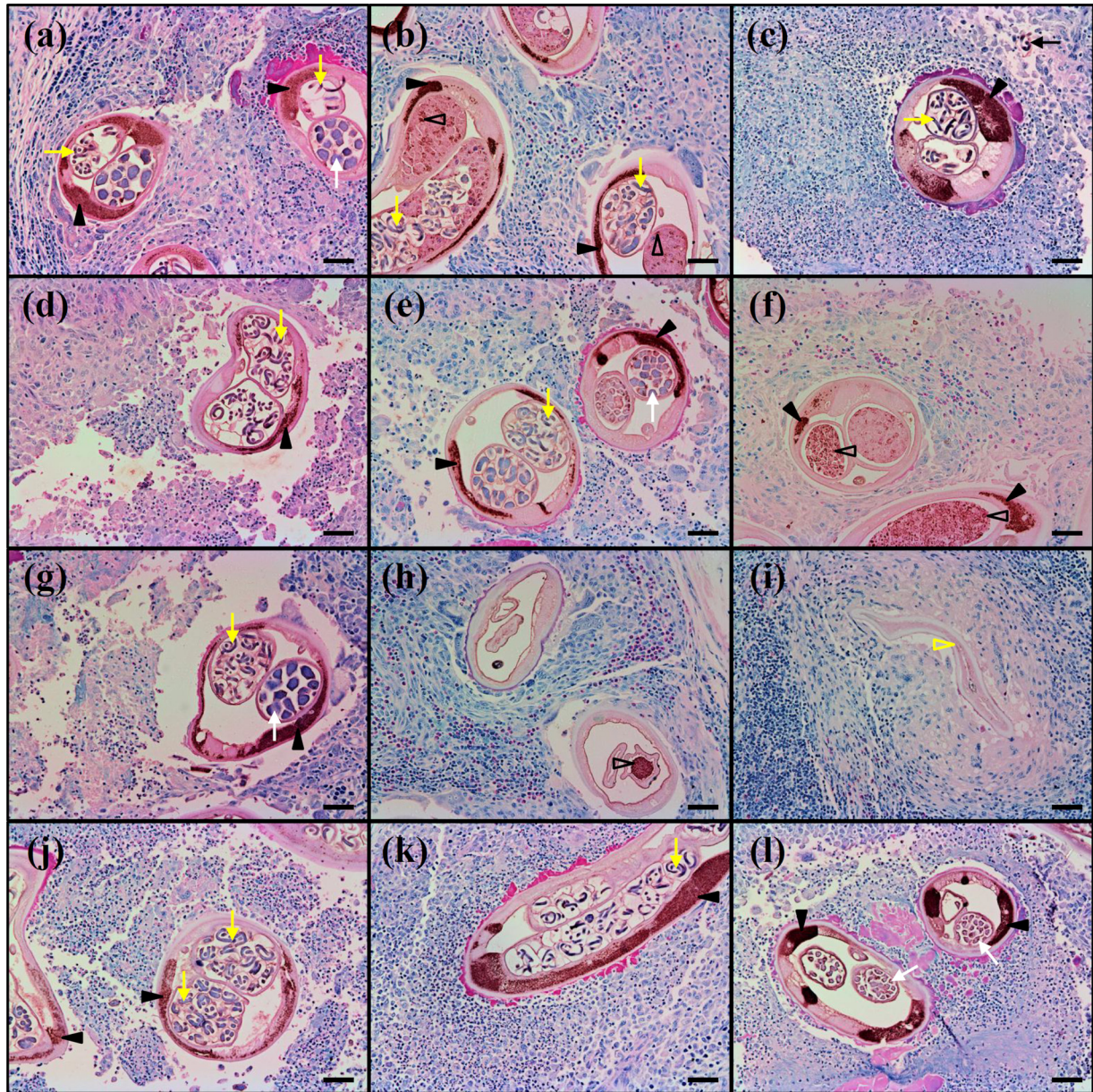


FIG 3 Localization of *Wolbachia* endobacteria in nodule sections from *Onchocerca ochengi*-infected cattle treated with DMSO solvent only (a, b, c), rifampin alone twice weekly for 3 weeks (d, e, f), oxytetracycline alone twice weekly for 3 weeks (g, h, i), or rifampin and oxytetracycline twice weekly for 3 weeks (j, k, l) at 0 (a, d, g, j), 24 (b, e, h, k), and 36 (c, f, i, l) weeks posttreatment start. Yellow arrows, intrauterine microfilariae; filled black arrowheads, *Wolbachia* organisms in the hypodermal cords; white arrows, morulae; open black arrowheads, *Wolbachia* organisms in the female reproductive tract; black arrow, migrating microfilaria; open yellow arrowhead, partially resorbed worm section. Sections were probed with an anti-*Wolbachia* surface protein polyclonal antibody, which was visualized using 3,3'-diaminobenzidine precipitate, and counterstained with Giemsa stain. Bars, 50 μ m.

morulae tended to exhibit an abnormal, granular morphology at 36 wpt (Fig. 3f and l). The OXY3-2 regimen displayed the most pronounced effects, with complete loss of *Wolbachia* staining from the hypodermal cords in most sections and a marked reduction in uterine contents at 24 wpt, although endobacteria remained detectable occasionally in degenerated oocytes (Fig. 3h). Importantly, by 36 wpt, almost-complete resolution of fragments from dead worms could be observed (Fig. 3i).

These immunohistochemical observations of *Wolbachia* density were verified using a semiquantitative scoring system, which

revealed a statistically significant reduction in numbers of nodules containing $\geq 50\%$ endobacterium-positive hypodermal cords within worm sections in the OXY3-2 group at both 24 ($P = 0.015$) and 36 ($P = 0.002$) wpt (Fig. 4a). This was not the case with either the RIF3 or the COM regimen, although a strong but nonsignificant ($P = 0.061$) pattern of reduced endobacterial density was observed at 36 wpt for the latter (Fig. 4a). In contrast, no statistically significant changes in *Wolbachia* staining within the female worm reproductive tract were apparent throughout experiment 2 (Fig. 4b).

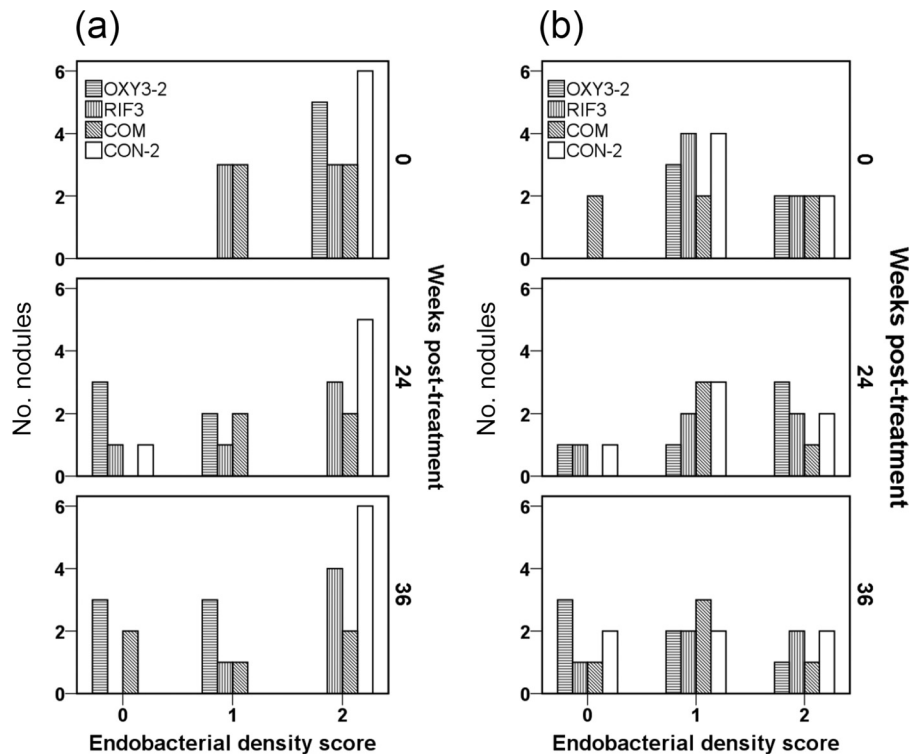


FIG 4 Endobacterial density scores in *Onchocerca ochengi* hypodermal cords (a) or *O. ochengi* female reproductive tracts (b) in nodular histological sections from infected cattle treated with DMSO solvent only (CON-2), rifampin alone twice weekly for 3 weeks (RIF3), rifampin and oxytetracycline twice weekly for 3 weeks (COM), or oxytetracycline alone twice weekly for 3 weeks (OXY3-2) at 0, 24, and 36 weeks post-treatment start. *Wolbachia* staining in worm sections across the entirety of each slide was semiquantified on a 3-point scale, as follows: 0, no visible bacteria; 1, sparse bacteria (<50% of worm sections were positive); and 2, profuse bacterial ($\geq 50\%$ of worm sections were positive). Note that data were not available for two OXY3-2 nodules, three COM nodules, and two RIF3 nodules due to complete resorption of the worms ($n = 5$) or misidentification of nodules caused by *Demodex bovis* ($n = 2$).

RIF concentrations in bovine plasma and skin. To establish if the poor efficacy of the RIF3 regimen could be explained by the pharmacokinetics of this drug in cattle, we measured RIF concentrations in the plasma and skin of four animals treated with a regimen identical to that administered to the RIF3 group. The mean maximum concentration of RIF in plasma (C_{max}) was ~ 28 $\mu\text{g/ml}$ and was maintained above the MIC for *Wolbachia* (24, 25) for at least 24 h (Fig. 5a) but was no longer detectable by 48 h. Similarly, RIF concentrations in skin, although approximately 10-fold lower than those achieved in plasma, remained above the MIC for ≥ 24 h (Fig. 5b) but were below the limit of quantification by 48 h.

DISCUSSION

The greatest barrier to the implementation of antibiotic chemotherapy for onchocerciasis has been the duration of the regimen required for permanent sterilization of female worms or potent adulticidal effects (23). Although DOX is clearly more efficacious than ivermectin, the latter can suppress Mf densities (and thus, disease manifestations) for up to 1 year following a single dose of 150 $\mu\text{g/kg}$, which makes it highly amenable to MDA programs (4). In a clinical trial in an area of Cameroon where loiasis is endemic, community-wide distribution of daily DOX therapy for 6 weeks was accompanied by very high rates of compliance (22), but to date, this has not resulted in wider application of DOX by APOC.

A lack of contraindication in children, coupled with RIF's bactericidal mode of action, has led it to be mooted as a more favor-

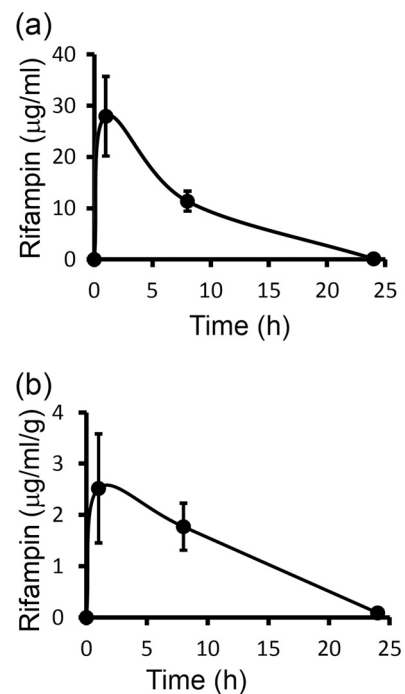


FIG 5 Rifampin concentration-time profiles (means \pm standard deviations; $n = 4$) in bovine plasma (a) and bovine skin (b) following a single intravenous injection at 10 mg/kg.

able alternative to DOX in this context. However, RIF alone has not proved to be strongly adulticidal in human trials, even if administered daily for 4 weeks (30). In the current study, we reasoned that a combination of a tetracycline and RIF might be more efficacious than either drug alone, as inhibition of both translation (by the former) and RNA synthesis (by the latter) is expected to induce synergistic effects. In addition, an earlier *in vivo* trial in the rodent filarial model *Litomosoides sigmodontis* revealed enhanced efficacy (prophylaxis, growth retardation, and embryostasis) for DOX and RIF duotherapy for 14 days compared with the efficacy of DOX monotherapy for 21 days (27). Moreover, an ~50% adulticidal effect of 14-day duotherapy was observed in a human clinical trial against *Wuchereria bancrofti* (38), a pathogen responsible for lymphatic filariasis which also harbors *Wolbachia*. Surprisingly, not only did our data fail to support the superiority of duotherapy, they also consistently reflected an impaired efficacy of the combination relative to that of OXY alone. Thus, in experiment 2, only OXY monotherapy attained significant reductions in Mf loads, *Wolbachia* densities in the hypodermal cords, and the recovery of viable adult worms at the end of the trial.

Unfortunately, no commercial formulation of RIF licensed for administration to ruminants is available worldwide. For welfare reasons, we were unable to deliver RIF on a daily schedule due to the large volumes of intravenous solution required, and accordingly, drug concentrations fell below the MIC for *Wolbachia* (0.0625 mg/liter [24, 25]) between 24 and 48 h. As RIF levels are reduced by 20 to 25% by heat inactivation of plasma (37), the concentrations determined in our bovine samples may have been underestimated; nevertheless, it is likely that our regimen achieved a therapeutic dose in skin for only two periods slightly in excess of 24 h each week. Although this drug is generally considered to have good penetrance and residual activity in tissue, we are not aware of other studies that have quantified RIF in skin. However, in a case of human RIF overdose, discoloration of skin resolved after 24 h, while urine and tears remained red-orange for several days (39). Possibly, the subcutaneous location of human nodules renders *O. volvulus* more susceptible to RIF than is *O. ochengi*, especially as the drug is photosensitive (40). Conversely, the long-acting formulation of OXY is known to achieve therapeutic levels for 3 to 5 days following a single intramuscular injection (41); hence, a twice-weekly schedule is effectively a continuous regimen. While these considerations provide a superficial explanation for the inefficacy of RIF monotherapy relative to OXY monotherapy, intermittent regimens of OXY are generally superior to continuous administration in this system, as observed in the current study with PIR and in several previous trials (18, 36, 42). Moreover, the relatively rapid clearance of RIF in cattle does not explain why RIF-OXY duotherapy is apparently less efficacious than OXY alone.

In monogastric species, daily therapy with RIF is generally well tolerated, and *in vivo* trials against *L. sigmodontis* and *Brugia pahangi* in rodents used doses of 50 mg/kg (27) and up to 100 mg/kg (28), respectively. However, even clinical trials of “higher-dose” RIF for tuberculosis in humans have not exceeded regimens of 20 mg/kg daily (43), which raises questions regarding the interpretation of the rodent studies. Notably, 20 doses of RIF monotherapy at 100 mg/kg failed to achieve a significant adulticidal effect on *B. pahangi* transplanted into jirds (28), whereas the potent antifilarial properties of a 14-day combination regimen of RIF and DOX against *L. sigmodontis* in mice were manifested when che-

motherapy was initiated simultaneously with infection using third-stage larvae (27). Thus, these effects against *L. sigmodontis* were not genuinely adulticidal. Combinations of RIF and DOX have rarely been used in human clinical trials for other conditions, although in the case of scrub typhus (caused by another member of the *Rickettsiales*, *Orientia tsutsugamushi*), duotherapy was ineffective compared to either drug alone (44). The reasons for this failure were not determined, but in duotherapy for brucellosis, pharmacokinetic interference between DOX and RIF has been reported. Hence, plasma levels of DOX were reduced by 50% in brucellosis patients exhibiting metabolic induction of hepatic microsomal enzymes (45), while in a larger study, a statistically significant inverse correlation between DOX and RIF plasma concentrations was apparent, which resulted in two cases of therapeutic failure or relapse (46). Moreover, there was a trend for DOX levels to be lower in brucellosis patients also receiving RIF who had a “rapid acetylator” *N*-acetyltransferase-2 genotype (46). Although, to the best of our knowledge, no studies on liver function following RIF therapy have been performed on ruminants, a detrimental effect on OXY pharmacokinetics caused by metabolic induction is a plausible explanation for the poor efficacy of duotherapy in our study.

In conclusion, despite promising early indications *in vitro* and in small-animal studies *in vivo*, RIF (whether alone or in combination with a tetracycline) may not represent the breakthrough in adulticidal therapy for onchocerciasis that was hoped. In addition to having potentially unfavorable pharmacokinetic interactions with tetracyclines, RIF remains of critical importance in tuberculosis control in the countries where onchocerciasis is endemic (47), rendering RIF MDA for onchocerciasis extremely unlikely. In this context, the recent discovery that *Wolbachia* is highly susceptible to another RNA polymerase inhibitor, coralopyronin A, which lacks activity against mycobacteria, is timely (48). Moreover, the successful chemical synthesis of this natural product (49) provides an avenue for its clinical development and ultimate application to human filarial diseases (50).

ACKNOWLEDGMENTS

We are very grateful for the technical assistance of David Ekale and Henrietta Nganyung at IRAD (Ngaoundéré, Cameroon) and of Catherine Hartley at the University of Liverpool. We also thank Shrikant Kulkarni and Mukul Jerath of Lupin Ltd. (Mumbai, India) for the kind donation of rifampin.

This study was supported by the European Commission (contracts INCO-CT-2006-032321 and HEALTH-F3-2010-242131).

We declare that we have no conflict of interest.

REFERENCES

1. Anonymous. 2010. Onchocerciasis (river blindness), p 123–128. In Crompton DWT, Peters P (ed), Working to overcome the global impact of neglected tropical diseases: first WHO report on neglected tropical diseases. World Health Organization, Geneva, Switzerland.
2. Plaisier AP, Van Oortmarsen GJ, Remme J, Habbema JD. 1991. The reproductive lifespan of *Onchocerca volvulus* in West African savanna. *Acta Trop* 48:271–284. [http://dx.doi.org/10.1016/0001-706X\(91\)90015-C](http://dx.doi.org/10.1016/0001-706X(91)90015-C).
3. Schulz-Key H. 1990. Observations on the reproductive biology of *Onchocerca volvulus*. *Acta Leiden* 59:27–44.
4. Crump A, Morel CM, Omura S. 2012. The onchocerciasis chronicle: from the beginning to the end? *Trends Parasitol* 28:280–288. <http://dx.doi.org/10.1016/j.pt.2012.04.005>.
5. Tielsch JM, Beeche A. 2004. Impact of ivermectin on illness and disability associated with onchocerciasis. *Trop. Med. Int. Health* 9:A45–A56. <http://dx.doi.org/10.1111/j.1365-3156.2004.01213.x>.

6. Traore MO, Sarr MD, Badji A, Bissan Y, Diawara L, Doumbia K, Goita SF, Konate L, Mounkoro K, Seck AF, Toe L, Toure S, Remme JH. 2012. Proof-of-principle of onchocerciasis elimination with ivermectin treatment in endemic foci in Africa: final results of a study in Mali and Senegal. *PLoS Negl. Trop. Dis.* 6:e1825. <http://dx.doi.org/10.1371/journal.pntd.0001825>.
7. Gardon J, Gardon-Wendel N, Demanga N, Kamgno J, Chippaux JP, Boussinesq M. 1997. Serious reactions after mass treatment of onchocerciasis with ivermectin in an area endemic for *Loa loa* infection. *Lancet* 350:18–22. [http://dx.doi.org/10.1016/S0140-6736\(96\)11094-1](http://dx.doi.org/10.1016/S0140-6736(96)11094-1).
8. Katarbarwa MN, Lakwo T, Habomugisha P, Agunyo S, Byamukama E, Oguttu D, Tukesiga E, Unoba D, Dramuke P, Onapa A, Tukahebwa EM, Lwamafa D, Walsh F, Unnasch TR. 2013. Transmission of *Onchocerca volvulus* continues in Nyagak-Bondo focus of northwestern Uganda after 18 years of a single dose of annual treatment with ivermectin. *Am. J. Trop. Med. Hyg.* 89:293–300. <http://dx.doi.org/10.4269/ajtmh.13-0037>.
9. Katarbarwa MN, Eyamba A, Nwane P, Enyong P, Kamgno J, Kuete T, Yaya S, Aboutou R, Mukenge L, Kafando C, Siaka C, Mkpouweiko S, Ngangue D, Biholong BD, Andze GO. 2013. Fifteen years of annual mass treatment of onchocerciasis with ivermectin have not interrupted transmission in the west region of Cameroon. *J. Parasitol. Res.* 2013:420928. <http://dx.doi.org/10.1155/2013/420928>.
10. Osei-Atweneboana MY, Eng Boakye JKDA, Gyapong JO, Prichard RK. 2007. Prevalence and intensity of *Onchocerca volvulus* infection and efficacy of ivermectin in endemic communities in Ghana: a two-phase epidemiological study. *Lancet* 369:2021–2029. [http://dx.doi.org/10.1016/S0140-6736\(07\)60942-8](http://dx.doi.org/10.1016/S0140-6736(07)60942-8).
11. Osei-Atweneboana MY, Awadzi K, Attah SK, Boakye DA, Gyapong JO, Prichard RK. 2011. Phenotypic evidence of emerging ivermectin resistance in *Onchocerca volvulus*. *PLoS Negl. Trop. Dis.* 5:e998. <http://dx.doi.org/10.1371/journal.pntd.0000998>.
12. Brieger WR, Okeibunor JC, Abiose AO, Ndyomugenyi R, Wanji S, Elhassan E, Amazigo UV. 2012. Characteristics of persons who complied with and failed to comply with annual ivermectin treatment. *Trop. Med. Int. Health* 17:920–930. <http://dx.doi.org/10.1111/j.1365-3156.2012.03007.x>.
13. Awadzi K, Attah SK, Addy ET, Opoku NO, Quartey BT. 1999. The effects of high-dose ivermectin regimens on *Onchocerca volvulus* in onchocerciasis patients. *Trans. R. Soc. Trop. Med. Hyg.* 93:189–194. [http://dx.doi.org/10.1016/S0035-9203\(99\)90305-X](http://dx.doi.org/10.1016/S0035-9203(99)90305-X).
14. Taylor MJ, Bilo K, Sood HF, Archer JP, Underwood AP. 1999. 16S rDNA phylogeny and ultrastructural characterization of *Wolbachia* intracellular bacteria of the filarial nematodes *Brugia malayi*, *B. pahangi*, and *Wuchereria bancrofti*. *Exp. Parasitol.* 91:356–361. <http://dx.doi.org/10.1006/expr.1998.4383>.
15. Henkle-Dührsen K, Eckelt VH, Wildenburg G, Blaxter M, Walter RD. 1998. Gene structure, activity and localization of a catalase from intracellular bacteria in *Onchocerca volvulus*. *Mol. Biochem. Parasitol.* 96:69–81. [http://dx.doi.org/10.1016/S0166-6851\(98\)00109-1](http://dx.doi.org/10.1016/S0166-6851(98)00109-1).
16. Hoerauf A, Nissen-Pähle K, Schmetz C, Henkle-Dührsen K, Blaxter ML, Büttner DW, Gallin MY, Al-Qaoud KM, Lucius R, Fleischer B. 1999. Tetracycline therapy targets intracellular bacteria in the filarial nematode *Litomosoides sigmodontis* and results in filarial infertility. *J. Clin. Invest.* 103:11–18. <http://dx.doi.org/10.1172/JCI4768>.
17. Bandi C, McCall JW, Genchi C, Corona S, Venco L, Sacchi L. 1999. Effects of tetracycline on the filarial worms *Brugia pahangi* and *Dirofilaria immitis* and their bacterial endosymbionts *Wolbachia*. *Int. J. Parasitol.* 29:357–364. [http://dx.doi.org/10.1016/S0020-7519\(98\)00200-8](http://dx.doi.org/10.1016/S0020-7519(98)00200-8).
18. Langworthy NG, Renz A, Mackenstedt U, Henkle-Dührsen K, de Bronsvort MB, Tanya VN, Donnelly MJ, Trees AJ. 2000. Macrofilariocidal activity of tetracycline against the filarial nematode *Onchocerca ochengi*: elimination of *Wolbachia* precedes worm death and suggests a dependent relationship. *Proc. R. Soc. B Biol. Sci.* 267:1063–1069. <http://dx.doi.org/10.1098/rspb.2000.1110>.
19. Brouqui P, Fournier PE, Raoult D. 2001. Doxycycline and eradication of microfilaremia in patients with loiasis. *Emerg. Infect. Dis.* 7:604–605. <http://dx.doi.org/10.3201/eid0707.017747>.
20. Hoerauf A, Specht S, Buttner M, Pfarr K, Mand S, Fimmers R, Marfo-Debrekyei Y, Konadu P, Debrah AY, Bandi C, Brattig N, Albers A, Larbi J, Batsa L, Taylor MJ, Adjei O, Buttner DW. 2008. *Wolbachia* endobacteria depletion by doxycycline as antifilarial therapy has macrofilaricidal activity in onchocerciasis: a randomized placebo-controlled study. *Med. Microbiol. Immunol.* 197:295–311. <http://dx.doi.org/10.1007/s00430-007-0062-1>.
21. Hoerauf A, Specht S, Marfo-Debrekyei Y, Buttner M, Debrah AY, Mand S, Batsa L, Brattig N, Konadu P, Bandi C, Fimmers R, Adjei O, Buttner DW. 2009. Efficacy of 5-week doxycycline treatment on adult *Onchocerca volvulus*. *Parasitol. Res.* 104:437–447. <http://dx.doi.org/10.1007/s00436-008-1217-8>.
22. Wanji S, Tendongfor N, Nji T, Esum M, Che JN, Nkwescheu A, Alassa F, Kamnang G, Enyong PA, Taylor MJ, Hoerauf A, Taylor DW. 2009. Community-directed delivery of doxycycline for the treatment of onchocerciasis in areas of co-endemicity with loiasis in Cameroon. *Parasit. Vectors* 2:39. <http://dx.doi.org/10.1186/1756-3305-2-39>.
23. Taylor MJ, Hoerauf A, Townson S, Slatko BE, Ward SA. 18 July 2013. Anti-*Wolbachia* drug discovery and development: safe macrofilaricides for onchocerciasis and lymphatic filariasis. *Parasitology* <http://dx.doi.org/10.1017/S0031182013001108>.
24. Hermans PG, Hart CA, Trees AJ. 2001. *In vitro* activity of antimicrobial agents against the endosymbiont *Wolbachia pipientis*. *J. Antimicrob. Chemother.* 47:659–663. <http://dx.doi.org/10.1093/jac/47.5.659>.
25. Fenollar F, Maurin M, Raoult D. 2003. *Wolbachia pipientis* growth kinetics and susceptibilities to 13 antibiotics determined by immunofluorescence staining and real-time PCR. *Antimicrob. Agents Chemother.* 47:1665–1671. <http://dx.doi.org/10.1128/AAC.47.5.1665-1671.2003>.
26. Townson S, Tagboto S, McGarry HF, Egerton GL, Taylor MJ. 2006. *Onchocerca* parasites and *Wolbachia* endosymbionts: evaluation of a spectrum of antibiotic types for activity against *Onchocerca gutturosa* *in vitro*. *Filaria J.* 5:4. <http://dx.doi.org/10.1186/1475-2883-5-4>.
27. Volkmann L, Fischer K, Taylor M, Hoerauf A. 2003. Antibiotic therapy in murine filariasis (*Litomosoides sigmodontis*): comparative effects of doxycycline and rifampicin on *Wolbachia* and filarial viability. *Trop. Med. Int. Health* 8:392–401. <http://dx.doi.org/10.1046/j.1365-3156.2003.01040.x>.
28. Townson S, Hutton D, Siemienka J, Hollick L, Scanlon T, Tagboto SK, Taylor MJ. 2000. Antibiotics and *Wolbachia* in filarial nematodes: antifilarial activity of rifampicin, oxytetracycline and chloramphenicol against *Onchocerca gutturosa*, *Onchocerca lienalis* and *Brugia pahangi*. *Ann. Trop. Med. Parasitol.* 94:801–816. <http://dx.doi.org/10.1080/00034980020027988>.
29. Richards FO, Jr, Amann J, Arana B, Punkosdy G, Klein R, Blanco C, Lopez B, Mendoza C, Dominguez A, Guarner J, Maguire JH, Eberhard M. 2007. No depletion of *Wolbachia* from *Onchocerca volvulus* after a short course of rifampin and/or azithromycin. *Am. J. Trop. Med. Hyg.* 77:878–882.
30. Specht S, Mand S, Marfo-Debrekyei Y, Debrah AY, Konadu P, Adjei O, Buttner DW, Hoerauf A. 2008. Efficacy of 2- and 4-week rifampicin treatment on the *Wolbachia* of *Onchocerca volvulus*. *Parasitol. Res.* 103:1303–1309. <http://dx.doi.org/10.1007/s00436-008-1133-y>.
31. Morales-Hojas R, Cheke RA, Post RJ. 2006. Molecular systematics of five *Onchocerca* species (Nematoda: Filarioidea) including the human parasite, *O. volvulus*, suggest sympatric speciation. *J. Helminthol.* 80:281–290. <http://dx.doi.org/10.1079/JOH2006331>.
32. Renz A, Trees AJ, Achukwi D, Edwards G, Wahl G. 1995. Evaluation of suramin, ivermectin and CGP 20376 in a new macrofilaricidal drug screen, *Onchocerca ochengi* in African cattle. *Trop. Med. Parasitol.* 46:31–37.
33. Bronsvort BM, Renz A, Tchakouté V, Tanya VN, Ekale DD, Trees AJ. 2005. Repeated high doses of avermectins cause prolonged sterilisation, but do not kill *Onchocerca ochengi* adult worms in African cattle. *Filaria J.* 4:8. <http://dx.doi.org/10.1186/1475-2883-4-8>.
34. Trees AJ, Wahl G, Klager S, Renz A. 1992. Age-related differences in parasitosis may indicate acquired immunity against microfilariae in cattle naturally infected with *Onchocerca ochengi*. *Parasitology* 104:247–252. <http://dx.doi.org/10.1017/S0031182000061680>.
35. Hansen RD, Trees AJ, Bah GS, Hetzel U, Martin C, Bain O, Tanya VN, Makepeace BL. 2011. A worm's best friend: recruitment of neutrophils by *Wolbachia* confounds eosinophil degradation against the filarial nematode *Onchocerca ochengi*. *Proc. R. Soc. B Biol. Sci.* 278:2293–2302. <http://dx.doi.org/10.1098/rspb.2010.2367>.
36. Gilbert J, Nfon CK, Makepeace BL, Njongmeta LM, Hastings IM, Pfarr KM, Renz A, Tanya VN, Trees AJ. 2005. Antibiotic chemotherapy of onchocerciasis: in a bovine model, killing of adult parasites requires a sustained depletion of endosymbiotic bacteria (*Wolbachia* species). *J. Infect. Dis.* 192:1483–1493. <http://dx.doi.org/10.1086/462426>.
37. Srivastava A, Waterhouse D, Ardrey A, Ward SA. 2012. Quantification of rifampicin in human plasma and cerebrospinal fluid by a highly sensitive and rapid liquid chromatographic-tandem mass spectrometric

- method. *J. Pharm. Biomed. Anal.* 70:523–528. <http://dx.doi.org/10.1016/j.jpba.2012.05.028>.
38. Debrah AY, Mand S, Marfo-Debrekeye Y, Batsa L, Albers A, Specht S, Klarmann U, Pfarr K, Adjei O, Hoerauf A. 2011. Macroparasitocidal activity in *Wuchereria bancrofti* after 2 weeks treatment with a combination of rifampicin plus doxycycline. *J. Parasitol. Res.* 2011:201617. <http://dx.doi.org/10.1155/2011/201617>.
 39. Gross DJ, Dellinger RP. 1988. Red/orange person syndrome. *Cutis* 42: 175–177.
 40. Ali J, Ali N, Sultana Y, Baboota S, Faiyaz S. 2007. Development and validation of a stability-indicating HPTLC method for analysis of antitubercular drugs. *Acta Chromatogr.* 18:168–179.
 41. Davey LA, Ferber MT, Kaye B. 1985. Comparison of the serum pharmacokinetics of a long acting and a conventional oxytetracycline injection. *Vet. Rec.* 117:426–429. <http://dx.doi.org/10.1136/vr.117.17.426>.
 42. Nfon CK, Makepeace BL, Njongmeta LM, Tanya VN, Trees AJ. 2007. Lack of resistance after re-exposure of cattle cured of *Onchocerca ochengi* infection with oxytetracycline. *Am. J. Trop. Med. Hyg.* 76:67–72.
 43. Diacon AH, Patientia RF, Venter A, van Helden PD, Smith PJ, McIl- leron H, Maritz JS, Donald PR. 2007. Early bactericidal activity of high-dose rifampin in patients with pulmonary tuberculosis evidenced by positive sputum smears. *Antimicrob. Agents Chemother.* 51:2994–2996. <http://dx.doi.org/10.1128/AAC.01474-06>.
 44. Watt G, Kantipong P, Jongsakul K, Watcharapichat P, Phulsuksombati D, Strickman D. 2000. Doxycycline and rifampicin for mild scrub-typhus infections in northern Thailand: a randomised trial. *Lancet* 356:1057–1061. [http://dx.doi.org/10.1016/S0140-6736\(00\)02728-8](http://dx.doi.org/10.1016/S0140-6736(00)02728-8).
 45. Garraffo A, Dellamonica P, Fournier JP, Lapalus P, Bernard E. 1988. The effect of rifampicin on the pharmacokinetics of doxycycline. *Infection* 16:297–298. <http://dx.doi.org/10.1007/BF01645076>.
 46. Colmenero JD, Fernandez-Gallardo LC, Agundez JA, Sedeno J, Benitez J, Valverde E. 1994. Possible implications of doxycycline-rifampin inter- action for treatment of brucellosis. *Antimicrob. Agents Chemother.* 38: 2798–2802. <http://dx.doi.org/10.1128/AAC.38.12.2798>.
 47. Aristoff PA, Garcia GA, Kirchhoff PD, Hollis Showalter HD. 2010. Rifamycins—obstacles and opportunities. *Tuberculosis (Edinb.)* 90:94–118. <http://dx.doi.org/10.1016/j.tube.2010.02.001>.
 48. Schiefer A, Schmitz A, Schaberle TF, Specht S, Lammer C, Johnston KL, Vassilyev DG, Konig GM, Hoerauf A, Pfarr K. 2012. Corallopyronin A specifically targets and depletes essential obligate *Wolbachia* endobacteria from filarial nematodes *in vivo*. *J. Infect. Dis.* 206:249–257. <http://dx.doi.org/10.1093/infdis/jis341>.
 49. Rentsch A, Kalesse M. 2012. The total synthesis of corallopyronin A and myxopyronin B. *Angew. Chem. Int. Ed. Engl.* 51:11381–11384. <http://dx.doi.org/10.1002/anie.201206560>.
 50. Schaberle TF, Schiefer A, Schmitz A, Konig GM, Hoerauf A, Pfarr K. 2013. Corallopyronin A—a promising antibiotic for treatment of filaria- sis. *Int. J. Med. Microbiol.* <http://dx.doi.org/10.1016/j.ijmm.2013.08.010>.