

Human Leukocyte Antigens and Systemic Lupus Erythematosus: A Protective Role for the HLA-DR6 Alleles *DRB1*13:02* and **14:03*

Hiroshi Furukawa^{1*}, Aya Kawasaki², Shomi Oka¹, Ikue Ito², Kota Shimada^{3,4}, Shoji Sugii³, Atsushi Hashimoto⁴, Akiko Komiya¹, Naoshi Fukui¹, Yuya Kondo⁵, Satoshi Ito⁶, Taichi Hayashi⁵, Isao Matsumoto⁵, Makio Kusaoi⁷, Hirofumi Amano⁷, Tatsuo Nagai⁸, Shunsei Hirohata⁸, Keigo Setoguchi⁹, Hajime Kono¹⁰, Akira Okamoto¹¹, Noriyuki Chiba¹², Eiichi Suematsu¹³, Masao Katayama¹⁴, Kiyoshi Migita¹⁵, Akiko Suda^{16,17}, Shigeru Ohno¹⁶, Hiroshi Hashimoto¹⁸, Yoshinari Takasaki⁷, Takayuki Sumida⁵, Shouhei Nagaoka¹⁷, Naoyuki Tsuchiya^{2,9}, Shigeto Tohma^{1,9}

1 Clinical Research Center for Allergy and Rheumatology, Sagami Hospital, National Hospital Organization, Sagami, Japan, **2** Molecular and Genetic Epidemiology Laboratory, Faculty of Medicine, University of Tsukuba, Tsukuba, Japan, **3** Department of Rheumatology, Tokyo Metropolitan Tama Medical Center, Fuchu, Japan, **4** Department of Rheumatology, Sagami Hospital, National Hospital Organization, Sagami, Japan, **5** Department of Internal Medicine, Faculty of Medicine, University of Tsukuba, Tsukuba, Japan, **6** Department of Rheumatology, Niigata Rheumatic Center, Shibata, Japan, **7** Department of Internal Medicine and Rheumatology, Juntendo University School of Medicine, Tokyo, Japan, **8** Department of Rheumatology and Infectious Disease, Kitasato University School of Medicine, Sagami, Japan, **9** Department of Allergy and Immunological Diseases, Tokyo Metropolitan Cancer and Infectious Diseases Center Komagome Hospital, Tokyo, Japan, **10** Department of Internal Medicine, Teikyo University, Tokyo, Japan, **11** Department of Rheumatology, Himeji Medical Center, National Hospital Organization, Himeji, Japan, **12** Department of Rheumatology, Morioka Hospital, National Hospital Organization, Morioka, Japan, **13** Department of Internal Medicine and Rheumatology, Clinical Research Institute, Kyushu Medical Center, National Hospital Organization, Fukuoka, Japan, **14** Department of Internal Medicine, Nagoya Medical Center, National Hospital Organization, Nagoya, Japan, **15** Clinical Research Center, Nagasaki Medical Center, National Hospital Organization, Omura, Japan, **16** Center for Rheumatic Diseases, Yokohama City University Medical Center, Yokohama, Japan, **17** Department of Rheumatology, Yokohama Minami Kyosai Hospital, Yokohama, Japan, **18** Juntendo University School of Medicine, Tokyo, Japan

Abstract

Many studies on associations between human leukocyte antigen (HLA) allele frequencies and susceptibility to systemic lupus erythematosus (SLE) have been performed. However, few protective associations with *HLA-DRB1* alleles have been reported. Here, we sought protective, as well as predispositional, alleles of *HLA-DRB1* in Japanese SLE patients. An association study was conducted for *HLA-DRB1* in Japanese SLE patients. Relative predispositional effects were analyzed by sequential elimination of carriers of each allele with the strongest association. We also explored the association of *DRB1* alleles with SLE phenotypes including the presence of autoantibody and clinical manifestations. Significantly different carrier frequencies of certain *DRB1* alleles were found to be associated with SLE as follows: increased *DRB1*15:01* ($P = 5.48 \times 10^{-10}$, corrected P (P_c) = 1.59×10^{-8} , odds ratio [OR] 2.17, 95% confidence interval [CI] 1.69–2.79), decreased *DRB1*13:02* ($P = 7.17 \times 10^{-5}$, $P_c = 0.0020$, OR 0.46, 95% CI 0.34–0.63) and decreased *DRB1*14:03* ($P = 0.0010$, $P_c = 0.0272$, OR 0.34, 95% CI 0.18–0.63). Additionally, the “**15:01/*13:02* or **14:03*” genotype tended to be negatively associated with SLE ($P = 0.4209$, OR 0.66), despite there being significant positive associations with **15:01* when present together with alleles other than **13:02* or **14:03* ($P = 1.79 \times 10^{-11}$, OR 2.39, 95% CI 1.84–3.10). This protective effect of **13:02* and **14:03* was also confirmed in SLE patients with different clinical phenotypes. To the best of our knowledge, this is the first report of a protective association between the carrier frequencies of *HLA-DRB1*13:02* and **14:03* and SLE in the Japanese population.

Citation: Furukawa H, Kawasaki A, Oka S, Ito I, Shimada K, et al. (2014) Human Leukocyte Antigens and Systemic Lupus Erythematosus: A Protective Role for the HLA-DR6 Alleles *DRB1*13:02* and **14:03*. PLoS ONE 9(2): e87792. doi:10.1371/journal.pone.0087792

Editor: Masataka Kuwana, Keio University School of Medicine, Japan

Received: October 12, 2013; **Accepted:** January 2, 2014; **Published:** February 3, 2014

Copyright: © 2014 Furukawa et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by Grants-in-Aid for Scientific Research (B, C) (22390199, 22591090), for Exploratory Research (25670458) and for Young Scientists (B) (24791018) from the Japan Society for the Promotion of Science, Health and Labour Science Research Grants from the Ministry of Health, Labour and Welfare of Japan, Grants-in-Aid for Clinical Research from National Hospital Organization, Research Grants from Daiwa Securities Health Foundation, Research Grants from Japan Research Foundation for Clinical Pharmacology, Research Grants from The Nakatomi Foundation, Research Grants from Takeda Science Foundation, Research Grants from Mitsui Sumitomo Insurance Welfare Foundation, Research Grants from SENSHIN Medical Research Foundation and research grants from pharmaceutical companies: Abbott Japan Co., Ltd., Astellas Pharma Inc., Chugai Pharmaceutical Co., Ltd., Eisai Co., Ltd., Mitsubishi Tanabe Pharma Corporation, Merck Sharp and Dohme Inc., Pfizer Japan Inc., Takeda Pharmaceutical Company Limited, Teijin Pharma Limited. The funders had no role in study design, data collection and analysis, decision to publish or preparing the manuscript.

Competing Interests: The following funders are supported in whole or in part by the indicated pharmaceutical companies. The Japan Research Foundation for Clinical Pharmacology is run by Daiichi Sankyo, the Takeda Science Foundation is supported by an endowment from Takeda Pharmaceutical Company and the Nakatomi Foundation was established by Hisamitsu Pharmaceutical Co., Inc. The Daiwa Securities Health Foundation was established by Daiwa Securities Group Inc. and Mitsui Sumitomo Insurance Welfare Foundation was established by Mitsui Sumitomo Insurance Co., Ltd. AH was supported by research grants from Mitsubishi Tanabe Pharma Corporation. NT is supported by SENSHIN Medical Research Foundation, which is supported by an endowment from Mitsubishi Tanabe Pharma Corporation. ST was supported by research grants from 9 pharmaceutical companies: Abbott Japan Co., Ltd., Astellas Pharma Inc., Chugai Pharmaceutical Co., Ltd., Eisai Co., Ltd., Mitsubishi Tanabe Pharma Corporation, Merck Sharp and Dohme Inc., Pfizer Japan Inc., Takeda Pharmaceutical Company Limited, Teijin Pharma Limited. The other authors declare no financial or commercial conflict of interest. This does not alter the authors' adherence to all the PLOS ONE policies on sharing data and materials.

* E-mail: h-furukawa@sagamihara-hosp.gr.jp

These authors contributed equally to this work.

Introduction

Systemic lupus erythematosus (SLE) is a prototypic autoimmune disease of unknown etiology that affects multiple organs and is associated with the production of several different autoantibodies. It is a systemic inflammatory disease susceptibility to which is associated with genetic and environmental factors [1]. Genetic risk factors for SLE include alleles in *IRF5*, *STAT4*, *BLK*, *TNFAIP3*, *TNIP1*, *FCGR2B* and others [2,3,4]; the functional role of the polymorphisms as well as the relationships with other autoimmune diseases such as rheumatoid arthritis were suggested [5,6]. Especially, altered frequencies of human leukocyte antigen (*HLA*) alleles are known to be associated with SLE. Some *HLA-DRB1* alleles are reported to be positively associated with SLE susceptibility in several ethnic groups studied: *DRB1*03:01* and **15:01* in European [7,8], **15:03* in African-American [9], **08:02* in Hispanic [10] and **15:01* and **15:02* in Asian populations [11,12,13,14]. Gene dosage effects were not noted in the associations of *HLA-DRB1* alleles with susceptibility to SLE in that homozygosity for a susceptibility allele does not confer higher disease risk than heterozygosity for that allele. However, only limited information is available concerning protective *DRB1* alleles for SLE, i.e. those with a reduced frequency in patients. Here, we sought protective, as well as predispositional, *HLA-DRB1* alleles in Japanese SLE patients. We also explored associations of *DRB1* alleles with SLE phenotypes including the presence of autoantibody and clinical manifestations of disease.

Materials and Methods

Patients and controls

A first set of 459 SLE patients (with a mean age \pm standard deviation (SD) of 48.2 ± 15.4 years) of whom 36 were men was recruited at Sagamihara Hospital, Yokohama Minami Kyosai Hospital, Tama Medical Center, Kitasato University, Komagome Hospital, Teikyo University, Himeji Medical Center, Morioka Hospital, Kyushu Medical Center and Yokohama City University Medical Center and a second set of 389 patients (41.6 ± 13.7 years of age; 31 men) at University of Tsukuba, Juntendo University and the University of Tokyo. A first set of 307 healthy controls (39.5 ± 11.1 years; 2 men) was recruited at Sagamihara Hospital or by the Pharma SNP Consortium (Tokyo, Japan) [15] and a second set of 542 healthy controls (34.0 ± 9.8 years; 245 men) at the University of Tokyo and University of Tsukuba. All patients and healthy individuals were native Japanese living in Japan. All patients with SLE fulfilled the American College of Rheumatology criteria for SLE [16]. This study was reviewed and approved by the research ethics committees of each participating institute, Sagamihara Hospital Research Ethics Committee, Nagasaki Medical Center Research Ethics Committee, Yokohama Minami Kyosai Hospital Research Ethics Committee, Tama Medical Center Research Ethics Committee, University of Tsukuba Research Ethics Committee, Kitasato University Research Ethics Committee, Komagome Hospital Research Ethics Committee, Teikyo University Research Ethics Committee, Himeji Medical Center Research Ethics Committee, Morioka Hospital Research Ethics Committee, Kyushu Medical Center Research Ethics Committee, Nagoya Medical Center Research Ethics Committee, Yokohama City University Medical Center Research Ethics Committee, the University of Tokyo Research Ethics Committee

and Juntendo University Research Ethics Committee. Written informed consent was obtained from all study participants. This study was conducted in accordance with the principles expressed in the Declaration of Helsinki.

Genotyping

Genotyping of *HLA-DRB1* and *DQB1* was performed by a polymerase chain reaction technique using sequence-specific oligonucleotide probes (WAKFlow HLA typing kits, Wakunaga, Hiroshima, Japan), using a Bio-Plex 200 system (Bio-Rad, Hercules, CA), or using MPH-2 HLA typing kits (Wakunaga). Results of *HLA-DRB1* genotyping for some of the healthy controls were reported previously [17,18]. Although a small part of second set SLE patients recruited at University of Tsukuba or Juntendo University was overlapped with that in another study which reported susceptible effects of *DRB1*09:01* [14], DNA collection and *HLA-DRB1* genotyping were independently performed for this replication study of protective association with *DRB1*13:02* and **14:03*. *DRB1-DQB1* haplotypes were elucidated by direct counting.

Statistical analysis

Differences of allele carrier frequencies, genotype frequencies, haplotype carrier frequencies or amino acid residue carrier frequencies were analyzed by Fisher's exact test using 2×2 contingency tables. Adjustment for multiple comparisons was performed using the Bonferroni method. Corrected *P* (*P_c*) values were calculated by multiplying the *P* value by the number of alleles, haplotypes or amino acid residues tested. Relative predispositional effects (RPE) were analyzed by sequential elimination of carriers of each allele with the strongest association [19]. Correction for multiple testing of SLE with different clinical and phenotypic manifestations was performed by calculating false discovery rate *Q*-value [20].

Results

Association analysis of the first set of SLE patients and healthy controls

HLA-DRB1 and *DQB1* genotyping was performed in 459 SLE patients and 307 healthy controls in the first set to compare carrier frequencies of each allele or haplotype (Table 1, left column, Table S1). Although the positive association between the carrier frequency of *DRB1*15:01* and SLE failed to reach significance in this set (*P_c* = 0.0705, odds ratio [OR] 1.76, 95% confidence interval [CI] 1.22–2.53, Table 1), a significant protective association was found for *DRB1*13:02* and SLE (*P_c* = 0.0123, OR 0.42, 95% CI 0.26–0.68). Additionally, the carrier frequency of the *DRB1*14:03* allele was lower in SLE, although this difference was not statistically significant (*P_c* = 0.0812, OR 0.30, 95% CI 0.14–0.68). The DR6 (*DRB1*13* and **14*) allele carrier frequency was significantly lower in SLE (*P* = 4.34×10^{-5} , OR 0.51, 95% CI 0.37–0.70), but DR2 (*DRB1*15* and **16*) was not different (*P* = 0.1743, OR 1.23). A significant negative association was found for *DQB1*06:04* and SLE (*P_c* = 0.0499, OR 0.46, 95% CI 0.28–0.77, Table S1). The *DRB1*13:02-DQB1*06:04* haplotype was negatively associated with SLE (*P_c* = 0.0414, OR 0.30, 95% CI 0.26–0.72). Thus, we detected certain *DRB1* and *DQB1*

Table 1. HLA-DRB1 allele carrier frequency in the SLE patients and controls.

	1 st Set					2 nd set					combined						
	Case (n = 459)	Control (n = 307)	OR	Pc	P	Case (n = 389)	Control (n = 542)	OR	Pc	P	Case (n = 848)	Control (n = 849)	OR	Pc	P	95%CI	P (RPE)
DRB1*01:01	41 (8.9)	27 (8.8)	1.0000	1.02	NS	34 (8.7)	65 (12.0)	0.70	NS	0.1312	75 (8.8)	92 (10.8)	0.80	NS	0.1921		
DRB1*04:01	19 (4.1)	4 (1.3)	0.0292	3.27	0.8189	15 (3.9)	15 (2.8)	1.41	NS	0.3546	34 (4.0)	19 (2.2)	1.82	NS	0.0371	(1.03–3.23)	
DRB1*04:03	15 (3.3)	15 (4.9)	0.2612	0.66	NS	16 (4.1)	29 (5.4)	0.76	NS	0.4403	31 (3.7)	44 (5.2)	0.69	NS	0.1560		0.0117
DRB1*04:05	94 (20.5)	68 (22.1)	0.5890	0.91	NS	79 (20.3)	125 (23.1)	0.85	NS	0.3357	173 (20.4)	193 (22.7)	0.87	NS	0.2621		
DRB1*04:06	17 (3.7)	26 (8.5)	0.0062	0.42	0.1738	10 (2.6)	36 (6.6)	0.37	0.1533	0.0053	27 (3.2)	62 (7.3)	0.42	0.0052	0.0002	(0.26–0.66)	0.0020
DRB1*04:10	15 (3.3)	10 (3.3)	1.0000	1.00	NS	10 (2.6)	15 (2.8)	0.93	NS	1.0000	25 (2.9)	25 (2.9)	1.00	NS	1.0000		
DRB1*08:02	59 (12.9)	31 (10.1)	0.2549	1.31	NS	40 (10.3)	32 (5.9)	1.83	0.5114	0.0176	99 (11.7)	63 (7.4)	1.65	0.0848	0.0029	(1.18–2.30)	0.0082
DRB1*08:03	93 (20.3)	50 (16.3)	0.1855	1.31	NS	61 (15.7)	76 (14.0)	1.14	NS	0.5119	154 (18.2)	126 (14.8)	1.27	NS	0.0673		0.0151
DRB1*09:01	139 (30.3)	72 (23.5)	0.0394	1.42	NS	118 (30.3)	151 (27.9)	1.13	NS	0.4208	257 (30.3)	223 (26.3)	1.22	NS	0.0670		
DRB1*11:01	11 (2.4)	18 (5.9)	0.0192	0.39	0.5378	15 (3.9)	16 (3.0)	1.32	NS	0.4639	26 (3.1)	34 (4.0)	0.76	NS	0.3576		
DRB1*12:01	37 (8.1)	20 (6.5)	0.4836	1.26	NS	36 (9.3)	43 (7.9)	1.18	NS	0.4771	73 (8.6)	63 (7.4)	1.18	NS	0.3730		
DRB1*12:02	12 (2.6)	6 (2.0)	0.6330	1.35	NS	14 (3.6)	24 (4.4)	0.81	NS	0.6156	26 (3.1)	30 (3.5)	0.86	NS	0.6839		
DRB1*13:02	30 (6.5)	44 (14.3)	0.0004	0.42	0.0123	39 (10.0)	93 (17.2)	0.54	0.0646	0.0022	69 (8.1)	137 (16.1)	0.46	1.51 × 10 ⁻⁵	5.21 × 10 ⁻⁷	(0.36–0.80)	7.17 × 10 ⁻⁵
DRB1*14:03	9 (2.0)	19 (6.2)	0.0029	0.30	0.0812	5 (1.3)	21 (3.9)	0.32	0.7156	0.0247	14 (1.7)	40 (4.7)	0.34	0.0127	0.0004	(0.18–0.63)	0.0010
DRB1*14:05	14 (3.1)	11 (3.6)	0.6836	0.85	NS	10 (2.6)	28 (5.2)	0.48	NS	0.0635	24 (2.8)	39 (4.6)	0.60	NS	0.0715		
DRB1*14:06	9 (2.0)	12 (3.9)	0.1176	0.49	NS	4 (1.0)	11 (2.0)	0.50	NS	0.2962	13 (1.5)	23 (2.7)	0.56	NS	0.1284		
DRB1*14:54	33 (7.2)	20 (6.5)	0.7727	1.11	NS	18 (4.6)	29 (5.4)	0.86	NS	0.6521	51 (6.0)	49 (5.8)	1.04	NS	0.8375		
DRB1*15:01	119 (25.9)	51 (16.6)	0.0025	1.76	0.0705	98 (25.2)	65 (12.0)	2.47	8.90 × 10 ⁻⁶	3.07 × 10 ⁻⁷	217 (25.6)	116 (13.7)	2.17	1.59 × 10 ⁻⁸	5.48 × 10 ⁻¹⁰	(1.69–2.79)	5.48 × 10 ⁻¹⁰
DRB1*15:02	69 (15.0)	63 (20.5)	0.0514	0.69	NS	76 (19.5)	116 (21.4)	0.89	NS	0.5118	145 (17.1)	179 (21.1)	0.77	NS	0.0414	(0.61–0.98)	
DR2 (DRB1*15:01)	187 (40.7)	110 (35.8)	0.1743	1.23	NS	170 (43.7)	183 (33.8)	1.52	NS	0.0026	357 (42.1)	293 (34.5)	1.38	NS	0.0014	(1.13–1.68)	
DR6 (DRB1*13:02 and *14:03)	98 (21.4)	107 (34.9)	4.34 × 10 ⁻⁵	0.51	0.0004	83 (21.3)	180 (33.2)	0.55	6.68 × 10 ⁻⁵	6.68 × 10 ⁻⁵	181 (21.3)	287 (33.8)	0.53	NS	1.06 × 10 ⁻⁸	(0.43–0.66)	

SLE: systemic lupus erythematosus, OR: odds ratio, CI: confidence interval, Pc: corrected P value, NS: not significant, RPE: relative predispositional effects. Allele carrier frequencies are shown in parenthesis (%). Alleles with more than 1% of the frequency in controls are shown. Association was tested by Fisher's exact test using 2 × 2 contingency tables. RPE were tested by sequential elimination of carriers of each of the alleles DRB1*15:01, *13:02, *14:03, *04:06, *08:02, *08:03 and *04:03.
doi:10.1371/journal.pone.0087792.t001

alleles which either conferred protection against or susceptibility to SLE.

Replication in the second set and combined analysis

We then sought to confirm these findings in an independent set of SLE patients and healthy controls by *DRB1* genotyping the second set of 389 SLE patients and 542 healthy controls (Table 1, central column). In this case, a significant positive association was found for *DRB1*15:01* and SLE ($P_c = 8.90 \times 10^{-6}$, OR 2.47, 95% CI 1.75–3.49) and it was the protective association with *DRB1*13:02* which just failed to achieve significance ($P_c = 0.0646$, OR 0.54, 95% CI 0.36–0.80). As in the first set, the protective association with *DRB1*14:03* also failed to reach significance ($P_c = 0.7156$, OR 0.32, 95% CI 0.12–0.86). Carrier frequencies of DR6 ($P = 6.68 \times 10^{-5}$, OR 0.55, 95% CI 0.40–0.74) and DR2 ($P = 0.0026$, OR 1.52, 95% CI 1.16–1.99) alleles were significantly lower and higher, respectively, in SLE.

We further explored associations between these *DRB1* alleles and SLE in a combined analysis, using RPE testing [19]. RPE were analyzed by sequential elimination of carriers of each allele with the strongest association (Table 1, right column). The strongest association was between *DRB1*15:01* and SLE, confirmed in this combined analysis ($P = 5.48 \times 10^{-10}$, $P_c = 1.59 \times 10^{-8}$, OR 2.17, 95% CI 1.69–2.79). The second round of comparisons was conducted after the elimination of *DRB1*15:01* carriers, revealing the next strongest association to be between *DRB1*13:02* and SLE ($P = 7.17 \times 10^{-5}$, $P_c = 0.0020$). The third round was after the elimination of both *DRB1*15:01* or **13:02* carriers, now showing the strongest association to be between *DRB1*14:03* and SLE ($P = 0.0010$, $P_c = 0.0272$). Further rounds after elimination of *DRB1*15:01*, **13:02* or **14:03* carriers revealed only tentative associations between the remaining *DRB1* alleles and SLE, particularly for *DRB1*04:06* ($P = 0.0020$, $P_c = 0.0509$), **08:02* ($P = 0.0082$, $P_c = 0.2040$), **08:03* ($P = 0.0151$, $P_c = 0.3615$) and **04:03* ($P = 0.0117$, $P_c = 0.2683$). We therefore focused on the *DRB1* alleles with the strongest SLE associations, namely *DRB1*15:01*, **13:02* and **14:03*.

Genotype analysis of HLA-DRB1*15:01, *13:02 and *14:03

We next compared genotype frequencies of *HLA-DRB1*15:01*, **13:02* and **14:03* to seek associations with SLE (Table 2). A significant positive association was found for “*DRB1*15:01/allele other than *15:01*” ($P = 4.83 \times 10^{-9}$, OR 2.14, 95% CI 1.65–2.76). On the other hand, “*DRB1*13:02/allele other than *13:02*” and the “**13:02/*13:02*” genotypes were both negatively associated with SLE ($P = 6.47 \times 10^{-6}$, OR 0.49, 95% CI 0.36–0.67 and $P = 0.0211$, OR 0.11, 95% CI 0.01–0.87, respectively). A negative association of the “**13:02 or *14:03/alleles other than *13:02 or *14:03*” genotype with SLE was also observed ($P = 5.53 \times 10^{-9}$, OR 0.43, 95% CI 0.33–0.58), again although a significant positive association for the genotype “alleles other than **13:02* or **14:03/alleles other than *13:02 or *14:03*” was present ($P = 3.69 \times 10^{-10}$, OR 2.41, 95% CI 1.82–3.19). A significant positive association was found with “**15:01/alleles other than *13:02 or *14:03*” ($P = 1.79 \times 10^{-11}$, OR 2.39, 95% CI 1.84–3.10). Thus, protective effects of **13:02* and **14:03* are dominant over the predisposing effects of **15:01* in SLE.

Associations of DRB1 with SLE of different clinical and phenotypic manifestations

We analyzed the associations of genotype frequencies of **15:01*, **13:02* and **14:03* separately in SLE patients with different clinical and autoantibody phenotypes to confirm the protective

effects of **13:02* and **14:03* in SLE patients with different manifestations. The significant positive association of the genotype “**15:01/alleles other than *13:02 or *14:03*” with SLE of a certain phenotype was confirmed for almost all factors assessed (Table 3, left column). The same was true for the negative association of the “**15:01/*13:02 or *14:03*” genotype with SLE regardless of phenotype (Table 3, right column). Thus, the protective effects of **13:02* and **14:03* were confirmed in all the different phenotypic manifestations of SLE.

Amino acid residues in the DRβ chain are associated with SLE

Amino acid residues in the HLA-DRβ chain were also analyzed for associations with SLE. The amino acid residue 13S in the DRβ chain showed strong protective associations with SLE ($P = 2.24 \times 10^{-8}$, $P_c = 7.63 \times 10^{-7}$, OR 0.55, 95% CI 0.45–0.68, Figure 1) and is shared by **13:02* and **14:03*, whereas the amino acid residues 32H ($P = 9.76 \times 10^{-5}$, $P_c = 0.0033$, OR 0.67, 95% CI 0.55–0.82), 67L ($P = 0.0003$, $P_c = 0.0118$, OR 0.70, 95% CI 0.58–0.85), and 71E ($P = 1.84 \times 10^{-5}$, $P_c = 0.0006$, OR 0.53, 95% CI 0.40–0.71) showed only moderate protective associations and are possessed by **13:02* and **14:03*, **14:03*, and **13:02*, respectively. Thus, association analysis suggested roles for certain defined amino acid residues in the DRβ chain in SLE susceptibility.

Discussion

Several studies have shown that certain *HLA-DR* alleles are positively associated with SLE. However, few studies have focused on negative associations of *HLA* alleles with SLE. To the best of our knowledge, this is the first report of a negative association of the HLA-DR6 alleles *DRB1*13:02* and **14:03* with Japanese SLE, although a lower frequency of DR5 [21] or DR6 [22] alleles in Asian patients with SLE has been reported before. Several studies have noted positive associations of *DRB1*15:01* [11,21,23], **15:02* [12,13] or **09:01* [3,14,24,25] alleles with SLE in Asians. However, here we only confirmed the association with **15:01*, but not **15:02* or **09:01* in Japanese SLE patients in general (Table 1). The association of **15:02* with SLE has never been reported in a Japanese population, suggesting that this allele may not be the primary genetic factor in itself, but a marker for a nearby gene. Since the allele frequencies between Thai and Japanese are comparable [13,18], the differences in allele frequencies could not explain the reason of the lack of association of **15:02* in Japanese SLE. Because HLA is the strongest genetic factor for SLE, it is quite difficult to explain the reason by other genetic backgrounds than *HLA* region. Since the amino acid sequences of **15:01* and **15:02* are almost same, presented peptide will be same. Environmental background could not explain the reason. HLA region is in strong linkage disequilibrium. Therefore, we cannot rule out the possibility that another causative genes, namely *DRB5* or *DQA1* genes, might exist in the *HLA* region in linkage disequilibrium with the culprit gene in the *DRB1* locus.

In the genotype analysis of *HLA-DRB1*15:01*, **13:02* and **14:03*, a nominally lower frequency of the “**15:01/*13:02 or *14:03*” genotype in SLE was observed, although a positive association was revealed for the genotype of “**15:01/alleles other than *13:02 or *14:03*” (Table 2). The protective effects of **13:02* or **14:03* thus seem to overcome the predispositional effects of **15:01* in SLE. *DRB1*13:02* commonly belongs to the haplotype *DRB1*13:02-DQB1*06:04-DPBI*04:01* positively selected in Japanese in recent history [26]. The *DRB1*13:02* allele is also a protective allele for cervical cancer caused by human papilloma

Table 2. HLA-DRB1 genotype frequency in the SLE patients and controls.

	Case (n = 848)	Control (n = 849)	P	OR	95%CI
*15:01/alleles other than *15:01	198 (23.3)	106 (12.5)	4.83 × 10 ⁻⁹	2.14	(1.65–2.76)
*15:01/*15:01	19 (2.2)	10 (1.2)	0.0958	1.92	
*13:02/alleles other than *13:02	68 (8.0)	128 (15.1)	6.47 × 10 ⁻⁶	0.49	(0.36–0.67)
*13:02/*13:02	1 (0.1)	9 (1.1)	0.0211	0.11	(0.01–0.87)
*15:01/*13:02	8 (0.9)	12 (1.4)	0.5009	0.66	
*15:01/alleles other than *15:01 or *13:02	190 (22.4)	94 (11.1)	3.30 × 10 ⁻¹⁰	2.32	(1.77–3.03)
*14:03/alleles other than *14:03	13 (1.5)	40 (4.7)	0.0002	0.31	(0.17–0.59)
*14:03/*14:03	1 (0.1)	0 (0.0)	0.4997	3.01	
*15:01/*14:03	2 (0.2)	3 (0.4)	1.0000	0.67	
*13:02/*14:03	0 (0.0)	1 (0.1)	1.0000	0.33	
*15:01/alleles other than *15:01, *13:02 or *14:03	188 (22.2)	91 (10.7)	1.58 × 10 ⁻¹⁰	2.37	(1.81–3.11)
*13:02 or *14:03/any alleles	83 (9.8)	176 (20.7)	3.69 × 10 ⁻¹⁰	0.41	(0.31–0.55)
*13:02 or *14:03/alleles other than *13:02 or *14:03	81 (9.6)	166 (19.6)	5.53 × 10 ⁻⁹	0.43	(0.33–0.58)
*13:02 or *14:03/*13:02 or *14:03	2 (0.2)	10 (1.2)	0.0379	0.20	(0.04–0.91)
alleles other than *13:02 or *14:03/alleles other than *13:02 or *14:03	765 (90.2)	673 (79.3)	3.69 × 10 ⁻¹⁰	2.41	(1.82–3.19)
*15:01/*13:02 or *14:03	10 (1.2)	15 (1.8)	0.4209	0.66	
*15:01/alleles other than *13:02 or *14:03	207 (24.4)	101 (11.9)	1.79 × 10 ⁻¹¹	2.39	(1.84–3.10)

SLE: systemic lupus erythematosus, OR: odds ratio, CI: confidence interval. Genotype frequencies are shown in parenthesis (%). Association was tested by Fisher's exact test using 2 × 2 contingency tables.

doi:10.1371/journal.pone.0087792.t002

virus infection [27] and *DPBI*04:01* is protective for hepatitis B infection [28].

The *DRBI* and *DQBI* alleles which showed significant associations are in strong linkage disequilibrium. In order to elucidate which of the *DRBI* and *DQBI* genes was responsible for

Table 3. HLA-DRB1 genotype frequency in the SLE patients and controls relative to SLE phenotype.

	n	*15:01/alleles other than *13:02, or *14:03	P	OR	Q	95%CI	*15:01/*13:02, or *14:03	P	OR	Q	95%CI
age of onset < 20	144	36 (25.0)	7.58 × 10 ⁻⁵	2.47	0.0001	(1.60–3.80)	1 (0.7)	0.4920	0.39	0.8806	
anti-Ro/SS-A antibodies (+)	326	80 (24.5)	2.13 × 10 ⁻⁷	2.41	6.04 × 10 ⁻⁷	(1.74–3.34)	4 (1.2)	0.6138	0.69	0.8806	
anti-La/SS-B antibodies (+)	70	14 (20.0)	0.0591	1.85	0.0628		1 (1.4)	1.0000	0.81	1.0000	
anti-RNP antibodies (+)	240	60 (25.0)	1.62 × 10 ⁻⁶	2.47	3.07 × 10 ⁻⁶	(1.72–3.53)	1 (0.4)	0.2192	0.23	0.8806	
anti-Sm antibodies (+)	220	51 (23.2)	5.14 × 10 ⁻⁵	2.23	7.49 × 10 ⁻⁵	(1.53–3.25)	2 (0.9)	0.5478	0.51	0.8806	
anti-dsDNA antibodies (+)	603	148 (24.5)	5.53 × 10 ⁻¹⁰	2.41	3.13 × 10 ⁻⁹	(1.82–3.18)	7 (1.2)	0.3916	0.65	0.8806	
antiphospholipid syndrome (+)	165	37 (22.4)	0.0007	2.14	0.0009	(1.41–3.26)	1 (0.6)	0.4925	0.34	0.8806	
malar rash (+)	301	90 (25.4)	2.08 × 10 ⁻⁸	2.52	8.85 × 10 ⁻⁸	(1.83–3.45)	2 (0.7)	0.2655	0.37	0.8806	
discoid rash (+)	116	84 (27.9)	5.28 × 10 ⁻¹⁰	2.87	3.13 × 10 ⁻⁹	(2.07–3.97)	2 (1.7)	1.0000	0.98	1.0000	
photosensitivity (+)	238	28 (24.1)	0.0007	2.36	0.0009	(1.47–3.78)	3 (1.3)	0.7770	0.71	0.8806	
arthritis (+)	410	60 (25.2)	1.44 × 10 ⁻⁶	2.50	3.06 × 10 ⁻⁶	(1.74–3.57)	4 (1.0)	0.3336	0.55	0.8806	
serositis (+)	161	96 (23.4)	3.46 × 10 ⁻⁷	2.26	8.41 × 10 ⁻⁷	(1.66–3.08)	0 (0.0)	0.1477	0.17	0.8806	
renal disorder (+)	380	35 (21.7)	0.0015	2.06	0.0017	(1.34–3.16)	5 (1.3)	0.6354	0.74	0.8806	
neurologic disorder (+)	130	93 (24.5)	7.01 × 10 ⁻⁸	2.40	2.38 × 10 ⁻⁷	(1.76–3.28)	3 (2.3)	0.7219	1.31	0.8806	
hemolytic anemia (+)	95	35 (26.9)	1.84 × 10 ⁻⁵	2.73	3.14 × 10 ⁻⁵	(1.76–4.24)	2 (2.1)	0.6857	1.20	0.8806	
lymphopenia (+)	459	18 (18.9)	0.0708	1.73	0.0708		4 (0.9)	0.2335	0.49	0.8806	
thrombocytopenia (+)	180	122 (26.6)	5.58 × 10 ⁻¹¹	2.68	9.49 × 10 ⁻¹⁰	(2.00–3.60)	2 (1.1)	0.7513	0.62	0.8806	
Control	849	101 (11.9)					15 (1.8)				

SLE: systemic lupus erythematosus, OR: odds ratio, CI: confidence interval. Genotype frequencies are shown in parenthesis (%). Associations were tested by Fisher's exact test using 2 × 2 contingency tables. To correct for multiple testing, the false discovery rate Q-value was calculated.

doi:10.1371/journal.pone.0087792.t003

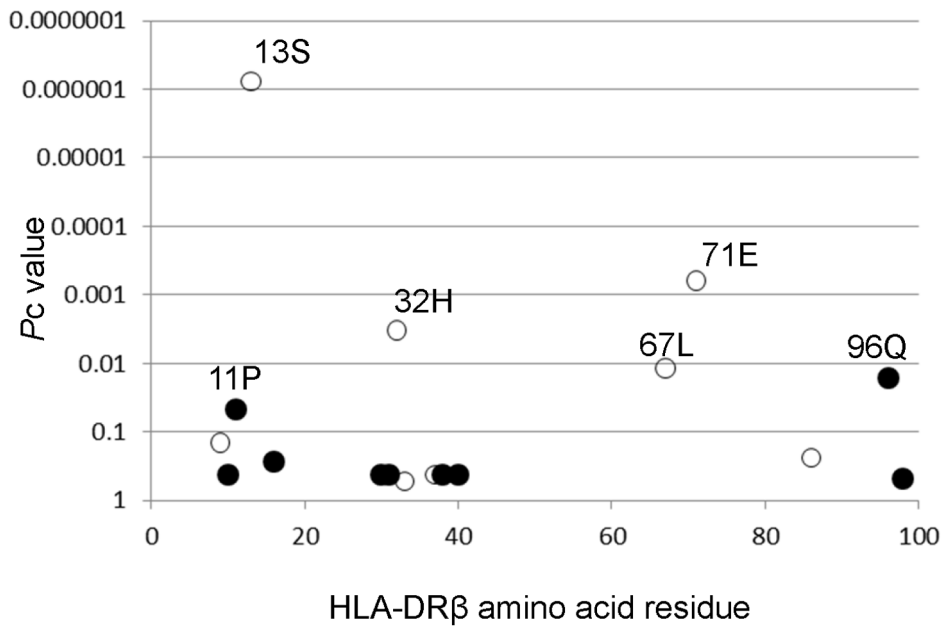


Figure 1. Associations of amino acid residues in the DR β chain with SLE. Corrected P (P_c) values were calculated by multiplying the P value by the number of amino acid residues tested. Associations were established by Fisher's exact test using 2×2 contingency tables. Positive associations were indicated in filled circles and negative in open circles. doi:10.1371/journal.pone.0087792.g001

primary association, haplotype analysis of *DRB1-DQB1* was performed. A significant negative association was found for the *DQB1*06:04* allele and the *DRB1*13:02-DQB1*06:04* haplotype with SLE in the first set comparison (Table S1). The primary role of *DRB1* or *DQB1* was not elucidated, because the strong linkage disequilibrium between *DRB1*13:02* and *DQB1*06:04* results in a low frequency of *DRB1*13:02* in patients without *DQB1*06:04* and similarly a low frequency of *DQB1*06:04* in patients without *DRB1*13:02*.

It was reported that anti-Ro/SS-A-positive rheumatoid arthritis patients were more frequently *DRB1*08:03*-positive and an association of *DRB1*15:01* and anti-La/SS-B antibodies has been reported in Japanese rheumatoid arthritis patients [29]. In the present study, a significant positive association of *DRB1*04:05* with the presence of anti-Ro/SS-A antibodies in SLE patients was found; in contrast, the *DRB1*12:02* allele was associated with the presence of anti-La/SS-B antibodies. These four *DRB1* alleles are in linkage disequilibrium with *DPB1*05:01* in the Japanese population [30], suggesting a role for *DPB1* in the production of autoantibodies to ribonucleoprotein. Susceptibility alleles for background diseases, *DRB1*15:01* in SLE or *DRB1*04:05* in rheumatoid arthritis, might not be easily detected as autoantibody-associated alleles in a comparison of antibody-positive and -negative patients. Alternatively, these findings could be explained by differences in the pathogenesis of rheumatoid arthritis and SLE.

Amino acid residues 13, 32, 67 and 71 of the HLA-DR β chain were found to be associated with SLE (Figure 1). Residues 13, 32, 67 and 71 form the HLA-DR peptide-binding groove [31]. These data suggest the involvement of peptide antigens bound to specific HLA molecules in controlling the development of SLE.

References

- Rahman A, Isenberg DA (2008) Systemic lupus erythematosus. *N Engl J Med* 358: 929–939.
- Koga M, Kawasaki A, Ito I, Furuya T, Ohashi J, et al. (2011) Cumulative association of eight susceptibility genes with systemic lupus erythematosus in a Japanese female population. *J Hum Genet* 2011: 12.

The negative association with *HLA-DR6* alleles needs to be confirmed in future independent studies. Because the allelic distribution of *HLA* in other ethnic populations is different from the Japanese, the protective role of some *DRB1* alleles in SLE in other populations should be determined.

This is the first identification of a negative association of *HLA-DRB1*13:02* and **14:03* with SLE. Our findings support the dominantly protective role of *HLA-DR6* alleles in the pathogenesis of SLE.

Supporting Information

Table S1 HLA-DQB1 allele and DRB1-DQB1 haplotype carrier frequency in the 1st set of SLE patients and controls.
(PDF)

Acknowledgments

We thank Ms. Mayumi Yokoyama (Sagamihara Hospital) for secretarial assistance.

Author Contributions

Conceived and designed the experiments: HF NT ST. Performed the experiments: HF A. Kawasaki S. Oka II. Analyzed the data: HF. Contributed reagents/materials/analysis tools: HF K. Shimada SS AH A. Komiya NF YK SI TH IM M. Kusaoi HA TN SH K. Setoguchi HK AO NC ES M. Katayama KM AS S. Ohno HH YT TS SN NT ST. Wrote the paper: HF NT ST. Collected clinical information: HF K. Shimada SS AH A. Komiya NF YK SI TH IM M. Kusaoi HA TN SH K. Setoguchi HK AO NC ES M. Katayama KM AS S. Ohno HH YT TS SN NT ST.

3. Kim I, Kim YJ, Kim K, Kang C, Choi CB, et al. (2009) Genetic studies of systemic lupus erythematosus in Asia: where are we now? *Genes Immun* 10: 421–432.
4. Furukawa H, Kawasaki A, Oka S, Shimada K, Matsui T, et al. (2013) Association of a single nucleotide polymorphism in the SH2D1A intronic region with systemic lupus erythematosus. *Lupus* 22: 497–503.
5. Deng FY, Lei SF, Zhang YH, Zhang ZL, Guo YF (2013) Functional relevance for associations between genetic variants and systemic lupus erythematosus. *PLoS One* 8: e53037.
6. Perricone C, Ceccarelli F, Valesini G (2011) An overview on the genetic of rheumatoid arthritis: a never-ending story. *Autoimmun Rev* 10: 599–608.
7. Black CM, Welsh KI, Fielder A, Hughes GR, Batchelor JR (1982) HLA antigens and Bf allotypes in SLE: evidence for the association being with specific haplotypes. *Tissue Antigens* 19: 115–120.
8. Tsuchiya N, Kawasaki A, Tsao BP, Komata T, Grossman JM, et al. (2001) Analysis of the association of HLA-DRB1, TNF α promoter and TNFR2 (TNFRSF1B) polymorphisms with SLE using transmission disequilibrium test. *Genes Immun* 2: 317–322.
9. Suggs MJ, Majithia V, Lewis RE, Cruse JM (2011) HLA DRB1*1503 allelic haplotype predominance and associated immunodysregulation in systemic lupus erythematosus. *Exp Mol Pathol* 91: 548–562.
10. Reveille JD, Moulds JM, Ahn C, Friedman AW, Baethge B, et al. (1998) Systemic lupus erythematosus in three ethnic groups: I. The effects of HLA class II, C4, and CR1 alleles, socioeconomic factors, and ethnicity at disease onset. LUMINA Study Group. *Lupus in minority populations, nature versus nurture. Arthritis Rheum* 41: 1161–1172.
11. Hashimoto H, Nishimura Y, Dong RP, Kimura A, Sasazuki T, et al. (1994) HLA antigens in Japanese patients with systemic lupus erythematosus. *Scand J Rheumatol* 23: 191–196.
12. Lu LY, Ding WZ, Fici D, Deulofeut R, Cheng HH, et al. (1997) Molecular analysis of major histocompatibility complex allelic associations with systemic lupus erythematosus in Taiwan. *Arthritis Rheum* 40: 1138–1145.
13. Sirikong M, Tsuchiya N, Chandanayingyong D, Bejrachandra S, Suthipinittharm P, et al. (2002) Association of HLA-DRB1*1502-DQB1*0501 haplotype with susceptibility to systemic lupus erythematosus in Thais. *Tissue Antigens* 59: 113–117.
14. Shimane K, Kochi Y, Suzuki A, Okada Y, Ishii T, et al. (2013) An association analysis of HLA-DRB1 with systemic lupus erythematosus and rheumatoid arthritis in a Japanese population: effects of *09:01 allele on disease phenotypes. *Rheumatology (Oxford)* 52: 1172–1182.
15. Kamatani N, Kawamoto M, Kitamura Y, Harigai M, Okumoto T, et al. (2004) Establishment of B-cell lines derived from 996 Japanese individuals. *Tissue Culture Res Commun* 23: 71–80.
16. Hochberg M (1997) Updating the American College of Rheumatology revised criteria for the classification of systemic lupus erythematosus. *Arthritis Rheum* 40: 1725.
17. Shibue T, Tsuchiya N, Komata T, Matsushita M, Shiota M, et al. (2000) Tumor necrosis factor alpha 5'-flanking region, tumor necrosis factor receptor II, and HLA-DRB1 polymorphisms in Japanese patients with rheumatoid arthritis. *Arthritis Rheum* 43: 753–757.
18. Furukawa H, Oka S, Shimada K, Sugii S, Ohashi J, et al. (2012) Association of human leukocyte antigen with interstitial lung disease in rheumatoid arthritis: A protective role for shared epitope. *PLoS ONE* 7: e33133.
19. Payami H, Joe S, Farid NR, Stenszky V, Chan SH, et al. (1989) Relative predispositional effects (RPEs) of marker alleles with disease: HLA-DR alleles and Graves disease. *Am J Hum Genet* 45: 541–546.
20. Benjamini Y, Hochberg Y (1995) Controlling the false discovery rate: a practical and powerful approach to multiple testing. *J R Statist Soc B* 57: 289–300.
21. Zhang J, Ai R, Chow F (1997) The polymorphisms of HLA-DR and TNF B loci in northern Chinese Han nationality and susceptibility to systemic lupus erythematosus. *Chin Med Sci J* 12: 107–110.
22. Doherty DG, Ireland R, Demaine AG, Wang F, Veerapan K, et al. (1992) Major histocompatibility complex genes and susceptibility to systemic lupus erythematosus in southern Chinese. *Arthritis Rheum* 35: 641–646.
23. Lee HS, Chung YH, Kim TG, Kim TH, Jun JB, et al. (2003) Independent association of HLA-DR and FCgamma receptor polymorphisms in Korean patients with systemic lupus erythematosus. *Rheumatology (Oxford)* 42: 1501–1507.
24. Hong GH, Kim HY, Takeuchi F, Nakano K, Yamada H, et al. (1994) Association of complement C4 and HLA-DR alleles with systemic lupus erythematosus in Koreans. *J Rheumatol* 21: 442–447.
25. Li CF, He XH, Teng Q, Jiang ZF (2003) Association of HLA-A, B, and DR haplotypes with genotype in Chinese children with systemic lupus erythematosus. *Zhonghua Er Ke Za Zhi* 41: 422–425.
26. Kawashima M, Ohashi J, Nishida N, Tokunaga K (2012) Evolutionary analysis of classical HLA class I and II genes suggests that recent positive selection acted on DPB1*04:01 in Japanese population. *PLoS One* 7: e46806.
27. Madeleine MM, Johnson LG, Smith AG, Hansen JA, Nisperos BB, et al. (2008) Comprehensive analysis of HLA-A, HLA-B, HLA-C, HLA-DRB1, and HLA-DQB1 loci and squamous cell cervical cancer risk. *Cancer Res* 68: 3532–3539.
28. Kamatani Y, Wattanapokayakit S, Ochi H, Kawaguchi T, Takahashi A, et al. (2009) A genome-wide association study identifies variants in the HLA-DP locus associated with chronic hepatitis B in Asians. *Nat Genet* 41: 591–595.
29. Furukawa H, Oka S, Shimada K, Sugii S, Hashimoto A, et al. (2013) Association of increased frequencies of HLA-DPB1*05:01 with the presence of anti-Ro/SS-A and anti-La/SS-B antibodies in Japanese rheumatoid arthritis and systemic lupus erythematosus patients. *PLoS ONE* 8: e53910.
30. Nakajima F, Nakamura J, Yokota T (2001) Analysis of HLA haplotypes in Japanese, using high resolution allele typing. *MHC* 8: 1–32.
31. Jardtzyk TS, Brown JH, Gorga JC, Stern IJ, Urban RG, et al. (1994) Three-dimensional structure of a human class II histocompatibility molecule complexed with superantigen. *Nature* 368: 711–718.