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## Mucosal co-infections and HIV-1 transmission and pathogenesis

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### Mucosal Barriers to Pathogen Transmission

Exogenous viral, bacterial, fungal and parasitic pathogens typically encounter several barriers to infection of their requisite host. In humans, these barriers are most commonly encountered at mucosal surfaces. These surfaces cover the eyes, lacrimal glands and tear ducts, the gastrointestinal tract, the bronchioalveolar surfaces, the mammary glands and ducts and the urogenital tracts of both men and women. When encountering transmission barriers, most pathogens must initially survive in a unique and specialized local mucosal microenvironment prior to or during transit across the mucosal epithelial barrier; others make this environment their permanent niche. During co-evolution with their host, successful pathogens have acquired mechanisms that allow survival in these microenvironments, even if only for brief periods of time. For example, the common gastrointestinal bacterium, *Helicobacter pylori*, expresses colonization factors, such as outer membrane adhesins, that allow the bacteria to overcome the inhibitory effects of gastrointestinal luminal acidity and mucus-secretion while attaching to the epithelial cells lining the mucosal surface [1]. Pulmonary anaerobes encounter relatively high oxygen content and a surfactant lipoprotein layer within the small bronchioles and pulmonary alveoli [2]. Heterosexually transmitted pathogens, including *Chlamydia trachomatis* (*C. trachomatis*), *Neisseria gonorrhoeae* (*N. gonorrhoeae*), herpes simplex viruses types 1 and 2 (HSV-1 and HSV-2) and the human immunodeficiency virus type-1 (HIV-1) are typically delivered into the female genital tract in the presence of semen. The effects of semen on sexually transmitted infections (STIs) are complex and include characteristics that enhance and that block transmission. For instance, the alkalinity of semen effectively neutralizes the relative acidity of the healthy lower genital tract of females, but semen contains antibacterial factors [3–4] and virus-enhancing [5] peptides that create mixed and pathogen-specific effects on heterosexual STI transmission. Within the healthy lower female genital tract, most surfaces are covered with exudates (vaginal) and/or layers of cervical mucus that trap and dilute pathogens, inhibiting their access to the underlying epithelial surface [6].

If a pathogen survives the intraluminal microenvironment at mucosal surfaces to reach the epithelia, most must next breach this sometimes formidable impediment to transmission. The epithelial barrier varies dramatically among mucosal sites and this variability is closely related to STI transmission rates. Beginning distally and ascending within the human female reproductive tract, the vulva and vagina and ectocervix are lined by multilayered, stratified and keratinized epithelia, the cervical transformation zone by an increasingly thin, stratified and transitional epithelium and the endocervical crypts by a single layer of non-keratinized epithelial cells [7]. Recent investigations modeling sexual transmission in non-human primate (NHP) models via atraumatic vaginal exposure to simian immunodeficiency virus (SIV) show that viral transmission occurs most readily across the single-layered endocervix [8–9]. Similar vulnerabilities to retroviral transmission are seen in another single-cell layered mucosal surface—the rectum. Studies in NHP models have confirmed

epidemiologic data demonstrating that transmission of HIV across the rectum after receptive anal intercourse is higher than after receptive vaginal intercourse [10]. Although this effect has been logically attributed to the relative vulnerability of the single-celled rectal epithelium to the micro- and macro-trauma incumbent in receptive anal intercourse, transmission characteristics in the female genital tract after atraumatic vaginal exposure (as above) suggest that other factors are likely involved. Data showing short and long-term decreases in female-to-male HIV sexual transmission with male circumcision [11–12] also point to changes in epithelial characteristics as important in HIV transmission.

Mucosal pathogens that have survived the luminal microenvironment and have breached the mucosal epithelia must surmount detection and elimination by epithelial innate and adaptive immune responses. These responses are mediated by the largest and most wide-spread component of all human lymphoid tissue, the mucosa-associated lymphoid tissues (MALT; Figure 1). MALT is comprised of intraepithelial and subepithelial immune cell populations and local, small, isolated lymph nodes. The lymphoid cell populations in MALT are sometimes organized into lymphoid follicles within the subepithelial lamina propria, most readily exemplified by the Peyer's patches of the gastrointestinal tract. MALT has been variously categorized into several regional and functional subsets, including bronchial-associated lymphoid tissues (BALT), mammary-associated lymphoid tissues, genitourinary-associated lymphoid tissues, and gut-associated lymphoid tissues (GALT). Oropharyngeal-associated lymphoid tissues (the parotid gland, parotid and salivary glands and tonsils) are part of GALT but are frequently discussed as a distinct entity. Likewise, nasopharyngeal-associated lymphoid tissues (NALT) are frequently separately acknowledged although they share anatomic and functional attributes with both oropharyngeal-associated lymphoid tissues and BALT. Detection of exogenous pathogens by MALT-associated innate immune responses involves the host complement system and host macrophages, dendritic cells, monocytes, neutrophils and eosinophils [13]. Cell-mediated innate immune pathogen recognition utilizes Toll-like receptors (TLRs), nucleotide-binding and oligomerization domain (NOD)-like receptors and helicases [14]. The mucosal bacterial pathogens *Salmonella enterica*, *Escherichia coli* and several *Brucella* species employ molecular mimicry to evade host immune signaling and the innate immune recognition and clearance mediated by these cell surface receptors [14]. Other MALT-associated immune components straddle innate and adaptive immune functionality, including natural killer (NK) and natural killer T (NKT) cells. Successful pathogens finally encounter the adaptive components of MALT, including B and T cells, which feed into and interact with systemic immunity. Several pathogens have evolved mechanisms to subvert, oftentimes only temporarily, innate and adaptive immune recognition. For example, intracellular pathogens, including several DNA viruses, can block presentation of virus-derived antigens to host immune cells via major histocompatibility (MHC) class I molecules [15–16]. Extracellular pathogens are more likely to target antigen presentation by MHC class II molecules.

Despite the extensive mechanisms employed by human pathogens to invade their host, the complementary and redundant protective mechanisms present at mucosal epithelial sites are remarkably effective. For the human immunodeficiency virus, these protective mechanisms couple with the relative fragility of the virus outside its host to keep transmission rates very low. In a study in Uganda, male-to-female HIV-1 transmission was estimated to be between 1 in 122 and 1 in 1400 coital acts, depending on stage of disease in the infected transmitting male partner [17]. A study conducted in Kenya revealed transmission rates of 1 in 159 vaginal coital acts between HIV positive women and their HIV-negative male partners [18]. Transmission rates to the receptive partner in men who have sex with men (MSM) varies along with sexual practice preferences, but was demonstrated to be approximately 1 in 70 acts of insertive anal intercourse in a study in Sydney, Australia [19]. While these numbers come from widely differing populations and transmission rates will vary by viral load and

stage of disease in the transmitter and by co-infection in each partner, the relative rates of transmission based on coital practice are informative.

## A Possible Role for Mucosal Surfaces in HIV-1 Gatekeeping

The overlapping and redundant layers of mucosal protection against HIV transmission are also likely to be responsible for the well-described finding that, while several to many HIV-1 viral variants are present in the bodily secretions of the infected transmitter, the majority of HIV-1 transmission involves only a single CCR5 tropic viral isolate [20]. The gatekeeper function of the mucosal epithelia in selecting the founder/transmitted viral variant was recently reviewed in an informed and comprehensive review by Grivel, Shattock and Margolis [21]. The authors hypothesize that multiple and redundant barriers are part of the mucosal gatekeeping function and that the efficiency of each barrier may vary among potential viral host targets. The preferential transmission of CCR5-tropic variants after direct blood inoculation suggests that post-mucosal selection also occurs.

Vulnerabilities in mucosal barrier function are exploited by HIV-1 to increase transmission efficacy. Among these is the preferential transmission across single cell-layered mucosal barriers (see above) including the endocervix [8–9] and the rectum [10]. Other vulnerabilities include physical breaks in the epithelial barrier, local inflammation and the phenotype and activation status of target cells in subepithelial sites [22]. Each of these vulnerabilities can be introduced or exacerbated by mucosal site co-infections.

## A Variety of Interactions between Mucosal Co-infections and HIV-1 transmission and Disease

In this special hot topics issue of *Current HIV Research* the invited contributors were tasked with discussing the effects of specific mucosal co-infections on HIV pathogenesis. An emphasis on HIV transmission was suggested. Contributors were requested to submit primary, original data, topic reviews or a combination of approaches. Each contributing group brought its individual expertise to these submissions. All contributors demonstrate that associations between HIV-1 and other mucosal infections are the result of more than just common risk factors.

## Bacterial vaginosis and *Trichomonas vaginalis*

Mirmonsef, et al., have addressed the interactions between HIV-1 and two common inflammatory conditions of the lower female genital tract. One involves infection with the vaginal and ectocervical pathogen, *Trichomonas vaginalis* (*T. vaginalis*); the other involves complex alterations in the normal vaginal flora. The latter has been variably labeled, but is most usually called bacterial vaginosis (BV). Each of these conditions is frequently asymptomatic or minimally symptomatic. For BV, the authors review epidemiologic data linking the presence of BV to HIV-1 acquisition in and transmission by women. They highlight possible mechanisms for this association, including altered vaginal acidity secondary to decreases in hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>)-producing lactobacilli, the creation of a pro-inflammatory local microenvironment involving IL-8, IL-1 $\alpha$  and -1 $\beta$ , RANTES and IL2, bacteria-associated peptidoglycan stimulation of epithelial TLR2, and direct damage to the vaginal mucosa. The bacteria associated with BV can produce small chain fatty acids, including butyric acid, and sialidases, glycosidases and proteases that can stimulate inflammatory cascades and cell migration or inhibit innate immunity respectively. The authors discuss the intriguing finding that the normal vaginal flora of healthy non-human primates resembles that of women with BV. In their discussion of *T. vaginalis* as a co-factor in HIV-1 transmission, Mirmonsef, et al., review epidemiologic data linking this relatively

asymptomatic infection to increased HIV-1 seroconversion rates in affected women and increased viral shedding in those already infected with HIV-1. They review data mechanistically linking these effects to ectocervical epithelial disruption, to the breakdown of cervical mucus and to pro-inflammatory changes, including recruitment of CD4+ T cell targets and the secretion of IL-1 $\beta$ , IL8 and TNF $\alpha$  into the local microenvironment. Women with *T. vaginalis* also commonly experience BV, possibly secondary to direct effects of *T. vaginalis* on H<sub>2</sub>O<sub>2</sub>-producing lactobacilli.

## Neisseria gonorrhoea

Jarvis and Chang have reviewed the very complex data linking infection with *Neisseria gonorrhoea* (*N. gonorrhoea*) with HIV transmission and HIV-specific immune responses. Unlike BV, *T. vaginalis*, and *Chlamydia trachomatis* (*C. trachomatis*), infection with *N. gonorrhoeae* is oftentimes quite symptomatic; like *C. trachomatis*, *N. gonorrhoea* is an ascending infection that can spread to the upper female genital tract and cause pelvic inflammatory disease. Most *N. gonorrhoea* infections are highly inflammatory, with adhesion proteins, including pili, opacity-associated (Opa) proteins, lactosyl lipooligosaccharide (LOS) and porin proteins causing cytokine/chemokine secretion via TLR activation and related activities. As seen in *T. vaginalis*, CD4+ T cell targets are recruited to the endocervices of women infected with *N. gonorrhoeae*. The authors review data demonstrating that infection with *N. gonorrhoea* increases viremia in women chronically infected with HIV-1, but advise that population based investigations are conflicting as to the effects of *N. gonorrhoea* on systemic immunity in these women. Further controversies are discussed in terms of the effects of *N. gonorrhoea* on HIV-1 infection of target cells (T cells, dendritic cells and macrophages) *in vitro*. Here *N. gonorrhoeae* exerts cell-type dependent enhancing or inhibiting effects suggesting to the authors that these interactions may be better studied at the tissue level. Gavin and Chang finish their contribution with an exciting review of their own work and that of others on the effects of *N. gonorrhoea* on epithelial cell defensin-mediated enhancement of HIV-1 infectivity and link these finding to the gatekeeping function of mucosal epithelia.

## Chlamydia trachomatis

Schust, et. al., have also taken a largely mechanistic approach to their contribution on the effects of *C. trachomatis* infection on HIV-1 transmission across the female genital tract. Although *C. trachomatis* is an ascending infection frequently associated with the development of PID, these authors have concentrated their efforts on the cells present at the likely site of atraumatic transmission of both HIV-1 and *C. trachomatis*—the single-layered endocervical epithelia. Female genital tract epithelial cells have been implicated in HIV-1 transcytosis and CCR5 preferential HIV-1 gatekeeping [23]. The authors have used a newly-described [24–25], primary-like endocervical epithelial cell line to show that infections with non-disseminating genital serovars of *C. trachomatis* increases the cell surface expression of the alternative HIV-1 receptor, GalCer and of the HIV-1 co-receptors CXCR4 and CCR5 on epithelial cells. They provide primary *in vitro* data demonstrating that *C. trachomatis* infection increases the binding of HIV-1 to endocervical epithelial cells and subsequent infection of co-cultured, susceptible MT4-R5 T cells and primary clinical data indicating that CD4+, CXCR4+ and/or CCR5+ T cell targets are recruited to the endocervical mucosae of women with *C. trachomatis* cervicitis.

## Herpes Simplex Virus

Barnabas and Celum have approached their discussion of the effects of co-infection with herpes simplex virus (HSV) on HIV-1 transmission from a largely epidemiologic viewpoint

with a specific focus on HSV type 2 (HSV-2). HSV-2 typically enters its host via breaks in the epidermis or across mucosal epithelia. Typical disease manifestations include ulcerative lesions at the site of infection and cycles of latency in the dorsal root ganglia alternating with repeat local lesions and or asymptomatic viral shedding. The authors emphasize the dramatic variations in the worldwide disease prevalence and summarize fairly large data sets showing that HIV-1 transmission and acquisition are increased in HSV-2 infected women and men and in HSV-2 infected men who have sex with men. While dermal disruption with ulcerative lesions is an obvious mechanism by which HSV-2 co-infection may increase HIV-1 transmission, the contributors review other potential mechanisms, including local recruitment of HIV-target cells to sites of HSV-2 infection, HSV-induced transactivation of HIV-1 long-terminal repeats, increased HIV rna production and the surprising long-term local persistence of HIV-1 target cells at the sites of treated and healed HSV-2 lesions. The contributors then use the co-infection paradigm to discuss lessons learned about HSV-2 pathogenesis from HIV-1 intervention studies. They review several population-based investigations that demonstrate the treatment HSV-2 in HIV-1 infected individuals has little effect on HIV-1 viral load and transmission. These results demonstrate that, despite standard antiviral therapies, HSV-2 infected individuals are in an almost continual phase of disease reactivation and viral shedding. The authors complete their review with a discussion of the promising effects of high dose HSV-2 suppressive therapies on HIV-1 disease progression in co-infected populations.

## Mammary and Bronchioalveolar Mucosal Co-infections

This special hot topics issue of Current HIV Research does not contain a contribution on pulmonary co-infections and HIV-1 transmission nor on mammary co-infections and HIV-1 transmission, although each could be the focus of its own special issue. HIV transmission across the pulmonary tract is exceedingly rare, but there is a large body of literature available on the interplay between HIV-1 and pulmonary pathogens, including *Mycobacterium tuberculosis* (TB) and *Pneumocystis carinii* (*P carinii*). One investigator has shown a lung-specific expansion of CD4+ CCR5+ T cells among patients with active TB that is not found in matched peripheral blood nor in patients with non-TB pulmonary disease and suggested that the TB-infected lower respiratory tract and that could serve as a potential site for transmucosal HIV-1 transmission [26]. Others have linked *M. tuberculosis* infection to HIV-1 reactivation from latency (reviewed in [27]; see below for similar effects by periodontal pathogens). The nearly exclusive place of the mammary gland in HIV-1 transmission is in mother-to-neonate transmission during breastfeeding. Among HIV-positive women that breastfeed, 16–35% will transmit HIV-1 to their infant in the absence of antiretroviral therapy (ART) [28–30]. Breast milk contains both cell-free and cell-associated virus, and higher levels of both have been associated with transmission [31–32]. Generalized breast infection and inflammation (mastitis) are associated with an elevated risk for maternal-to-fetal HIV-1 transmission when compared to HIV-infected mothers without breast complaints; the risk is even higher in the presence of severe and localized breast infections (abscesses) [33]. The discrete point of viral entry in the infant who obtains HIV-1 through breastfeeding is not conclusively known, and infection may occur via the oral mucosa, tonsils, and/or the neonatal gastrointestinal tract. Recent studies using fetal organ tissue models have shown that both cell-free and cell-associated virus can transmigrate across fetal epithelial cells from the oral cavity and the intestines, and thus demonstrate a potential mechanism for transmission of HIV-1 contained in breast milk [34–35]. Studies in primates [36] and mice [37] have shown that cells present in breast milk can survive their transit through the stomach, traverse the intestinal tract and remain functional in the nursing neonate. Other major risk factors for HIV-1 transmission across the oropharyngeal and gastrointestinal tracts include orogenital sexual contact and receptive anal intercourse.



## Periodontal pathogens

There is a growing body of literature posing credible mechanisms by which co-infection with the periodontal pathogen *Porphyromonas gingivalis* (*P. gingivalis*) might increase transmission of HIV-1 across the oral mucosa. Some of these mechanisms, including *P. gingivalis*-associated increases in the expression of CCR5 on oral keratinocytes [38], are reminiscent of the effects of *C. trachomatis* on HIV-1 receptor and co-receptor expression on genital epithelial cells (Schust, et. al., this issue). Also similar are data demonstrating that oral epithelial cells can maintain non-replicative HIV-1 infection [39] and that *P. gingivalis* infection and its resultant CCR5 induction promotes transmission of CCR5-tropic HIV-1 variants across human oral epithelial cells (gingival and tonsil) to susceptible T cells [40].

The authors of our final contribution in this special Hot Topics issue have also addressed interactions between the periodontal pathogen, *P. gingivalis*, and HIV-1, but have focused their efforts on their own work on the effects of *P. gingivalis* on AIDS progression in co-infected individuals. *P. gingivalis* expresses several virulence factors, including small chain fatty acids (SCFAs), that Imai, et al., have studied in the context of HIV-1 infections [41–42]. The authors review their own published data outlining the specific effects of the SCFA, butyric acid, on the reactivation of integrated HIV-1 from latency [41–42]. They provide evidence supporting a mechanism involving histone deacetylation [42] and primary data demonstrating that these effects are not generalizable to all oral bacteria, including *Escherichia coli* (*E. coli*) and *Prevotella nigrescens* (*P. nigrescens*), and that local cytokine secretion in the presence of *P. gingivalis*-induced inflammation may also be involved. Imai, et al., conclude with the intriguing suggestion that HIV-1 reactivation from latency could be similarly induced by anerobic, butyric-acid producing bacteria known to be present at other mucosal sites.

## Concluding Remarks

As we continue to increase our knowledge about HIV-1 transmission and pathogenesis, it has become increasingly clear that mucosal co-infections play an important role in both. This will have particular ramifications to strategies aimed at preventing transmission in both resource-poor and more developed settings where many of these co-infections are very common and are inadequately treated. The contributors to this special Hot Topics issue of Current HIV Research have outlined many of the mechanisms involved in mucosal pathogen-associated vulnerabilities to HIV-1 transmission across the otherwise formidable barriers to infection present at most mucosal epithelial sites. These and other experts in infectious disease and mucosal immunology are certain to be at the forefront when advances are made in combating mucosal HIV-1 transmission.

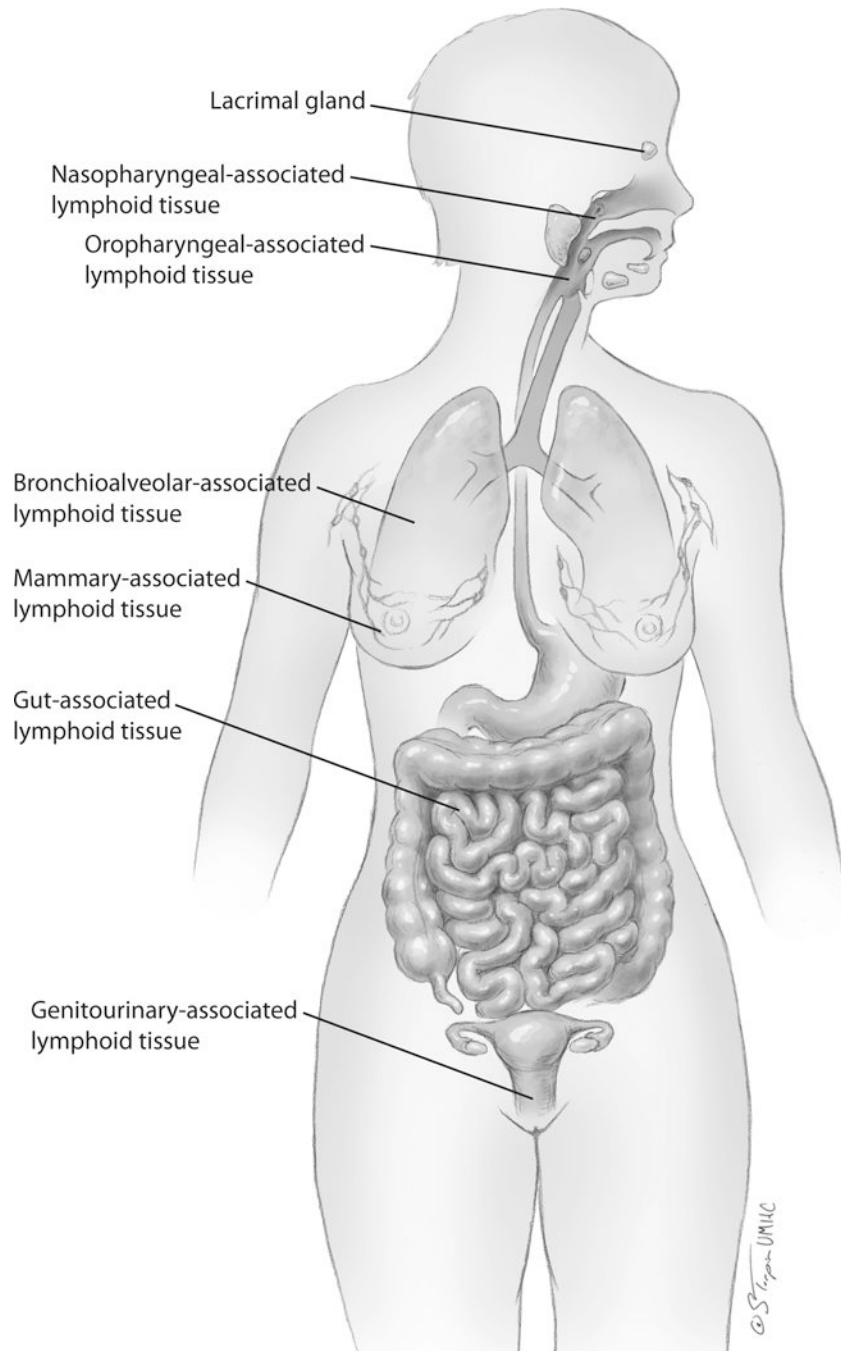
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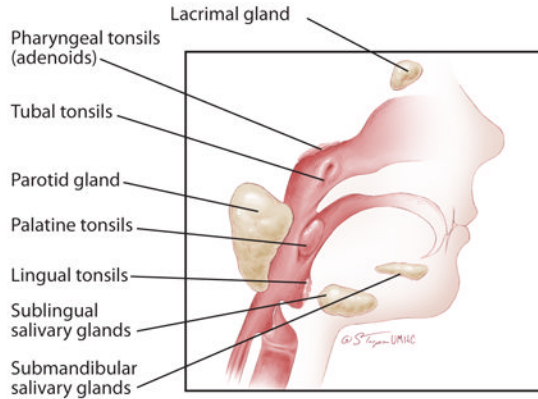




**Figure 1. Mucosal Immune Sites**

The immune cells populating mucosal sites create the largest immune barrier in the human body. Innate immune responses are particularly prevalent at mucosal immune sites although adaptive responses are also represented. The vast majority of exogenous pathogens first encounter host immune defenses at mucosal sites. Mucosal immune tissues are anatomically subdivided. One paradigm divides these tissues into: 1) nasopharyngeal-associated lymphoid tissues (NALT), 2) oropharyngeal-associated lymphoid tissues, 3) bronchioalveolar-associated lymphoid tissues, 4) mammary-associated lymphoid tissues, 5) gut (gastrointestinal)-associated lymphoid tissues and 6) genitourinary-associated lymphoid tissues. The lacrimal gland and eye also exert mucosa-associated immune reactivities.

## Oropharyngeal and Nasopharyngeal-associated Lymphoid Tissue

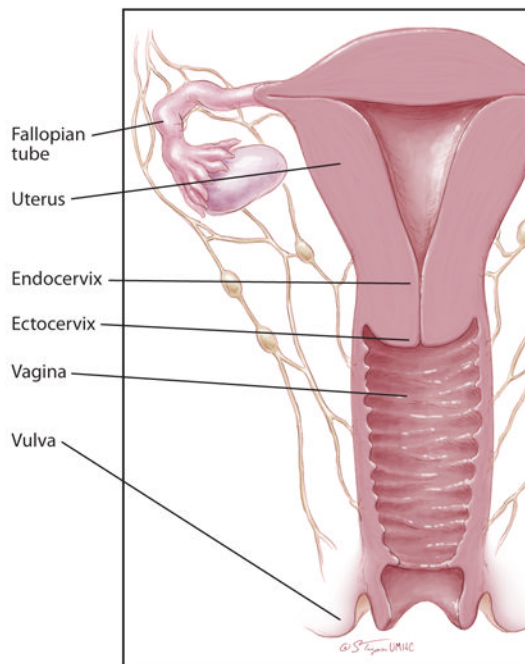


### Periodontal Pathogens

(gingiva, tonsils)

- Increased epithelial expression of HIV-1 receptors and co-receptors
- Increased HIV-1 reactivation
- Increased epithelial binding and transmission of HIV-1
- Local proinflammatory cytokine alterations
- SCFA (butyric acid)-induced histone deacetylation

## Genital-associated Lymphoid Tissue



### N. gonorrhoea

(vaginal, cervical and ascending infection)

- Adhesion-induced local inflammation
- Endocervical CD4+ T cell recruitment
- Increased viremia in chronically-infected women
- TLR activation
- Increased local proviral defensin production

### C. trachomatis

(endocervical and ascending infection)

- Increased epithelial expression of HIV-1 receptors and co-receptors
- Increased epithelial binding of HIV-1
- Increased epithelial transmission of HIV-1
- Endocervical CD4+ T cell recruitment

### Bacterial vaginosis

(vaginal)

- Direct mucosal damage
- Altered vaginal acidity
- Proinflammatory cytokine alterations
- TLR-2 stimulation
- SCFA (butyric acid)-induced inflammation

### T. vaginalis

(vaginal and cervical)

- Ectocervical disruption
- Increased viral shedding
- Proinflammatory cytokine alterations
- Mucus breakdown
- CD4+ T cell recruitment
- BV coinfection

### HSV-2

(vulvar, vaginal and ectocervical)

- Epithelial breakdown
- Increased HIV-1 rna production
- Local target cell recruitment and long-term retention

### Figure 2. Mechanisms by which co-infections increase HIV-1 transmission and/or pathogenesis at mucosal immune sites

Periodontal pathogens, including *P. gingivalis*, can infect tissues of the gingival and tonsils. These pathogens have been shown to exert direct effects on the mucosal epithelial cell and indirect effects on the local mucosal immune environment that can aid in HIV-1 transmission and spread after oral-genital exposures. The sexually transmitted infections, *C. trachomatis*, *N. gonorrhoea* and HSV can also infect oropharyngeal tissues but investigations on their effects on HIV-1 transmission at these sites has not been well-described. Sexually transmitted co-infections with *N. gonorrhoea*, *C. trachomatis*, *T. vaginalis* and *HSV-2* and the presence of bacterial vaginosis are associated with increased transmission of HIV-1 across

the female genital tract and/or increased HIV-1 viremia. The mechanisms causing these increases are variable and pathogen specific, although common mechanisms include epithelial disruption and local recruitment of target immune cells to the subepithelial tissues.