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Cysteinyl leukotrienes and their receptors: Bridging inflammation and colorectal cancer

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Core tip: Despite several advances in diagnostic and therapeutic options, colorectal cancer (CRC) continues to be a major health problem and one of the leading causes of cancer-related deaths. The inflammatory milieu has been widely recognized as one of the enabling characteristics of cancer development. Cysteinyl leukotrienes are pro-inflammatory eicosanoids implicated in chronic inflammatory bowel diseases and CRC development. Hence, targeting cysteinyl leukotrienes and their receptors could provide alternative therapeutic approaches or be used in combination with existing therapies for more efficient treatment of CRC.

Abstract

Long-standing inflammation has emerged as a hallmark of neoplastic transformation of epithelial cells and may be a limiting factor of successful conventional tumor therapies. A complex milieu composed of distinct stromal and immune cells, soluble factors and inflammatory mediators plays a crucial role in supporting and promoting various types of cancers. An augmented inflammatory response can predispose a patient to colorectal cancer (CRC). Common risk factors associated with CRC development include diet and lifestyle, altered intestinal microbiota and commensals, and chronic inflammatory bowel diseases. Cysteinyl leukotrienes are potent inflammatory metabolites synthesized from arachidonic acid and have a broad range of functions involved in the etiology of various pathologies. This review discusses the important role of cysteinyl leukotriene signaling in linking inflammation and CRC.

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INTRODUCTION

Colorectal cancer (CRC) is a global health care burden, with more than 1 million new cases diagnosed every year. It is the third most common malignancy and the fourth leading cause of cancer-related deaths worldwide^[1]. A diet high in fat but low in fiber, excessive alcohol consumption, obesity and lack of physical activity, disruption of normal gut microbiota and the presence of long-term inflammatory bowel diseases (IBDs) such as ulcerative colitis (UC) and Crohn's disease (CD) predispose to CRC. Inflammation is a host-driven response to internal and external stimuli to counter non-self or self-molecules and maintain tissue homeostasis. However, chronic inflamma-

tion can be a major health problem in allergic, cardiovascular, fibrotic, local and systemic inflammatory diseases and several cancers^[2,9]. In 1863, Rudolf Virchow was the first to speculate about the role of long-term inflammation in cancer based on his observations that cancerous tissues were frequently infiltrated by leukocytes^[10]. Current epidemiological data indicate that more than 25% of all cancers are related to long-term infections and other types of unresolved inflammation^[11-13]. Evidence from observational studies and randomized trials concerning the protective action of non-steroidal anti-inflammatory drugs (NSAIDs) such as aspirin have indicated not only a reduced long-term risk of esophageal, gastric, biliary, breast, prostate, lung, and CRC but also a lowered risk of metastasis^[14-18]. Inflammation present in the tumor microenvironment is characterized by high leukocyte infiltration, ranging in size, distribution and composition, such as tumor-associated macrophages (TAMs), mast cells, dendritic cells (DCs), natural killer cells (NKs), neutrophils, eosinophils and lymphocytes^[19-21]. These cells produce a variety of cytotoxic mediators such as reactive oxygen and nitrogen species, serine and cysteine proteases, membrane-perforating agents, matrix metalloproteinases, pro-inflammatory cytokines, interferons (IFNs) and increased levels of enzymes such as cyclooxygenase-2 (COX-2), 5-lipoxygenase (5-LOX) and phospholipase A₂ (PLA₂), hence contributing to carcinogenesis^[22-25]. This review addresses the role of cysteinyl leukotrienes in inflammation-induced colorectal carcinogenesis.

EICOSANOIDS

Eicosanoids, from the Greek word “eicosa” meaning “20,” are biologically active lipophilic molecules predominantly metabolized from arachidonic acid (AA), a 20-carbon polyunsaturated essential fatty acid, that are involved in physiological processes such as inflammation^[14]. AA belongs to the ω -6 family of polyunsaturated fatty acids and is usually found esterified at the second carbon position in the phospholipids of membranes. It serves as a precursor to several lipid pro-inflammatory mediators such as prostaglandins (PGs), prostacyclins, thromboxanes (TXAs), lipoxins and leukotrienes (LTs), which have individual as well as overlapping functions in acute and chronic inflammation^[26]. Aberrant AA metabolism is often linked to production of pro-inflammatory eicosanoids, chronic inflammatory diseases and carcinogenesis. The first eicosanoids were discovered in the 1960s, although in 1930 scientists had found that certain substances present in biological fluids such as sputum had the potential to induce contraction and relaxation in smooth muscles; they termed them “slow-reacting substances of anaphylaxis.” Despite these early observations, it was not until 1979 that “leukotrienes” were identified and defined by Samuelsson and co-workers for their biological effects in inflammatory processes^[27]. On account of their fundamental and seminal work on different eicosanoids, mainly the prostaglandins, and for their dis-

covery of the role of anti-inflammatory compounds such as aspirin on prostaglandin metabolism, the scientists Bengt Samuelsson, John Vane and Sune Bergström were awarded the Nobel Prize for Physiology and Medicine in 1982.

AA, which is stored as diacylglycerol (DAG), is released from the phosphatidylinositol 4,5-bisphosphate (PIP₂) present in the outer nuclear envelope of cells, from which it is mobilized into the cytoplasm either by activation of calcium-dependent cytosolic PLA₂ or by the combined action of phospholipase C (PLC) and DAG lipase^[26,28]. Once in the cytosol, AA can be enzymatically metabolized in a three-directional manner either by cytochrome P450 or by the COX pathway into prostaglandins, prostacyclins or thromboxanes, or through the 5-LOX pathway into leukotrienes A₄ (LTA₄), B₄ (LTB₄), C₄ (LTC₄), D₄ (LTD₄) and E₄ (LTE₄) (Figure 1). The last three alternative derivatives to LTA₄ are collectively termed “cysteinyl leukotrienes” (CysLTs) owing to the presence of a cysteine residue and are structurally similar but functionally distinct. The role of these eicosanoids in maintaining intestinal epithelial cell homeostasis is well documented^[29-32]. Various epidemiological, clinical, and laboratory studies have shown that dysregulation of the COX and LOX pathways results in chronic inflammation and subsequently cancer^[14,29].

LEUKOTRIENES AND THEIR RECEPTORS

The term leukotriene is derived from the two words *leuko* for white blood cells and *trienes*, meaning three conjugated double bonds, indicating that the ability to generate LTs from AA is largely restricted to leukocytes^[28].

Synthesis of LTs is initiated by 5-LOX in concert with 5-lipoxygenase-activating protein (FLAP). The latter does not exhibit any enzymatic activity but facilitates the interaction between 5-LOX and its substrate AA. The first step in this pathway is oxygenation of AA to yield unstable 5-hydroperoxyeicosatetraenoic acid (5-HPETE), which immediately undergoes dehydration to form LTA₄. Further metabolism of LTA₄ either generates LTB₄ by the action of LTA₄ hydrolase (LTA₄-H) or leads to conjugation with glutathione in the presence of LTC₄ synthase (LTC₄-S) or glutathione S-transferase to yield LTC₄. After carrier-mediated transport of LTB₄ and LTC₄ to the extracellular milieu, LTC₄ can be further metabolized to LTD₄ through the cleavage of glutamic acid from the glutathione moiety, and additional glycine cleavage yields LTE₄^[28,33] (Figure 1).

5-LOX is expressed predominantly by neutrophils, eosinophils, monocytes, macrophages and mast cells. Although nonleukocytes express 5-LOX and FLAP to a lesser extent and are not believed to synthesize appreciable amounts of LTs, expression of LTA₄-H and/or LTC₄-S, uptake of exogenous LTA₄ and further metabolism is possible, *via* a process referred to as transcellular biosynthesis^[34].

CysLT signaling is initiated upon binding of a li-

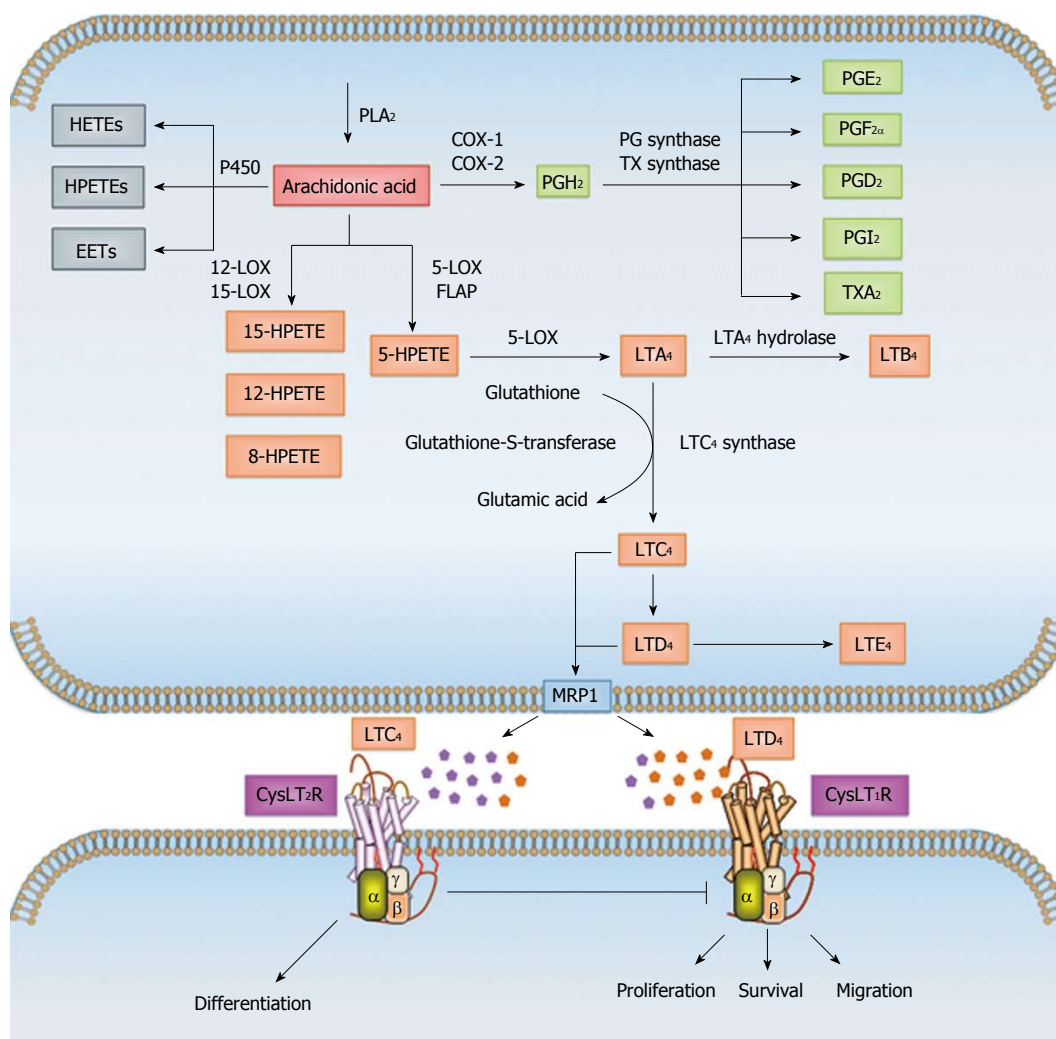


Figure 1 Overview of arachidonic acid metabolism. Arachidonic acid (AA) is a polyunsaturated fatty acid found in the phospholipids of cell membranes. AA is mobilized into the cytoplasm mostly by the activation of calcium-dependent cytosolic phospholipase A₂ (cPLA₂). Free AA in the cytosol can be enzymatically metabolized to eicosanoids through three major pathways: the cytochrome P450, cyclooxygenase (COX) and/or 5-lipoxygenase (5-LOX) pathways. In the P450 pathway, AA is metabolized to epoxyeicosatrienoic acids (EETs), hydroxyeicosatetraenoic acids (HETEs) and hydroperoxyeicosatetraenoic acids (HPETEs). In the COX pathway, AA is enzymatically converted to the intermediate prostaglandin H₂ (PGH₂), which is then sequentially metabolized to prostanoids, including prostaglandins (PGs), such as PGE₂, PGF₂, PGD₂ and PGI₂, and thromboxanes (TXs) such as TXA₂ by specific prostaglandin and thromboxane synthases. In the LOX pathway, AA is metabolized by 12- and 15-LOX to 8-, 12- and 15-HPETE or by 5-LOX and 5-lipoxygenase activating protein (FLAP) to intermediary 5-HPETE. 5-HPETE is further processed to form leukotrienes (LTs), the first of which is the unstable leukotriene A₄ (LTA₄). LTA₄ is subsequently converted to leukotriene B₄ (LTB₄) by LTA₄ hydrolase or together with glutathione to leukotriene C₄ (LTC₄) by LTC₄ synthase and glutathione-S-transferase. LTC₄ is converted by ubiquitous enzymes to form leukotriene D₄ (LTD₄) and leukotriene E₄ (LTE₄). The members of the multidrug resistance-associated protein (MRP) family are efflux transporters for both PGs and LTs. The cysteinyl leukotrienes (CysLTs) LTC₄, LTD₄ and LTE₄ act *via* G protein-coupled receptors CysLT₁R and CysLT₂R at the cell surface and induce different signaling mechanisms.

gand to one of the two G-protein-coupled receptors (GPCRs), CysLT₁R and CysLT₂R located at the plasma membrane^[35,36], although the presence of other CysLT receptors such as GPR17, P2Y₁₂, and CysLT₁R have also been suggested^[37-39]. Both CysLT₁R and CysLT₂R can also be localized to the nuclear membrane, since CysLT₁R has a bipartite nuclear localization sequence and CysLT₂R possesses an interferon regulatory 7 (IRF7) site, which in turn carries a nuclear localization sequence domain^[40-42]. While the affinity of CysLT₁R for LTD₄ is high, the CysLT₂R has a low but an equal affinity for LTD₄ and LTC₄^[35,36]. Functionally, CysLTs induce smooth muscle contraction, vascular leakage, eosinophil recruitment in inflammatory diseases, mucus production and chemo-

taxis^[43-46].

LTB₄ also plays a pivotal role in inflammatory processes such as leukocyte chemoattraction, particularly of granulocytes and T cells, induction of rapid invasion and recruitment of these cells to the plasma membrane of endothelial cells, production of reactive oxygen species, and induction of gene expression^[47,48]. LTB₄ mediates its signaling *via* two GPCRs: BLT₁ and BLT₂^[49,50]. BLT₁ binds to LTB₄ with an affinity higher than that of the BLT₂ receptor. The tissue distribution of the two receptors is quite different. Whereas BLT₁ expression in both mice and humans has been reported to be predominantly restricted to peripheral leukocytes, BLT₂ expression in humans appears to be fairly ubiquitous, with the highest

level observed in the spleen, liver, and lymphocytes^[51].

CYSTEINYL LEUKOTRIENES AND THEIR RECEPTORS IN COLORECTAL CANCER

IBD and colorectal cancer

Inflammation and CRC initiation and dissemination go hand in hand^[10,52]. The most well-established connection exists between IBD-both UC and CD- and CRC^[53-55]. “IBD” is a name given to a group of prolonged inflammatory disorders of the intestinal tract associated with debilitating symptoms and epithelial damage. The risk of developing CRC is 30%-50% higher in patients with IBD^[56,57]. IBDs are characterized by increased leukocyte infiltration into the intestinal wall, where they can induce non-specific inflammation through activation and production of AA-derived pro-inflammatory metabolites such as LTs and PGs and subsequent tissue injury. Thus, the gastrointestinal tract is richly supplied with these eicosanoids that mediate several gastrointestinal diseases, including cancers. High levels of LTs such as LTE₄ have been detected in the urine of patients with UC and CD^[58,59]. Among CysLTs, the presence of LTD₄ at an IBD site increases the risk of consequential cancer development, and specific LTD₄ antagonists have been shown to reduce colonic inflammation^[60]. Although UC is fundamentally similar to CD, a few differences exist, primarily the presentation of a cytokine profile with a T helper 2 (Th2) antibody-mediated response^[61]. CD is an autoimmune disease associated with T helper 1 (Th1)-mediated cytokines such as interleukin-12 (IL-12), IFN- γ and tumor necrosis factor-alpha (TNF- α)^[61,62].

Colitis-associated cancer (CAC) is known to be highly infiltrated by several cells of the innate immune system, including neutrophils, mast cells, NKs, DCs and TAMs^[63]. Moreover, recent evidence supports the concept that malignant tumors also recruit a specific subpopulation of myeloid cells called myeloid-derived suppressor cells^[64]. These cells share some characteristics with monocytes, macrophages, neutrophils, and DCs and help suppress any potential anti-tumor immune response and tumor angiogenesis. As in several cancers, including CRC, in which the major inflammatory cellular components are macrophages, TAMs contribute immensely to cancer growth and expansion. TAMs are macrophages that display an M2 type (alternatively activated phenotype) and secrete high levels of Th2 cytokines, growth factors and inflammatory mediators that promote tumor growth, angiogenesis, and metastasis^[65,66]. We have observed a high intra-tumoral density of TAMs in colon cancer tissue compared with the adjacent normal tissue, and M2 macrophages were required for effective colon cancer cell migration *via* factors derived from M2 macrophages and their association with signal regulatory protein alpha (SIRP- α) through CD47^[67].

Eicosanoids and colorectal cancer

Apart from its role in inflammation-associated dis-

eases such as asthma, psoriasis, rheumatoid arthritis and IBD^[68], LTB₄ has pro-tumorigenic effects in breast cancer, melanoma, lymphoma, and head and neck carcinoma^[69-72]. Increased expression of LTB₄ and its receptor BLT₁ have been demonstrated in human CRC tissue^[73]. Ihara *et al.*^[73] demonstrated significant expression of BLT₁ in the colon cancer cell lines Caco-2 and HT-29. Using both the 5-LOX inhibitor AA-861 and selective BLT₁ antagonist U75302 in these cell lines, the authors showed induction of apoptosis and reduced proliferation^[73]. LTB₄-stimulated extracellular signal-regulated kinase (Erk) activation in these cancer cells was also abrogated by U75302. A subsequent study investigated the effectiveness of another LTB₄ receptor antagonist (LY293111) in combination with gemcitabine, an anti-tumor adjuvant and radiosensitizer, on the proliferation rate of human colon cancer cell lines LoVo and HT-29 in an athymic heterotrophic xenograft mouse model and found a significant reduction in tumor growth due to apoptosis *via* the mitochondrial pathway^[74]. The findings from these and several other studies emphasize the role of LTB₄ signaling in colon cancer cells and warrant the use of specific LTB₄ receptor antagonists to suppress CRC expansion.

Among the eicosanoids derived from the COX-pathway, PGE₂ is the most abundant and extensively studied in cancer, including CRC^[29]. In both the spontaneous adenomatous polyposis coli (Apc)^{Min/+} mouse model of intestinal cancer and the azoxymethane (AOM)-induced CRC mouse model, PGE₂ has been shown to increase the tumor burden^[75,76]. Selective inhibition of PGE₂ synthesis, through genetic deletion of microsomal PGES (mPGES-1), significantly reduced tumor formation in an Apc^{Min/+} and AOM-induced mouse model of intestinal and CRC, respectively, and further established the role of PGE₂ in tumorigenesis^[77,78].

Increased expression of the enzymes responsible for production of PGs and LTs-COX-2 and 5-LOX, respectively has been documented in human colorectal adenocarcinomas compared with adjacent normal mucosa^[79-81]. Various clinical trials over the past two decades have highlighted the use of eicosanoid-depressing and anti-inflammatory drugs in the prevention and treatment of CRC. Two groups of compounds have shown promising results: aspirin (NSAID) and celecoxib (COX-2 selective inhibitor)^[82].

Cysteinyl leukotrienes, their receptors and colorectal cancer

Upregulated expression of CysLT₁R has been observed in several human cancers, including transitional cell carcinoma (TCC) in the bladder, neuroblastomas, and brain, prostate, breast, and CRCs^[6,80,83-86]. We have shown that high CysLT₁R tumor expression is associated with a poor survival prognosis in breast and CRC patients^[80,86], whereas concomitant low CysLT₁R and high CysLT₂R expression indicate a good prognosis in CRC patients^[87]. We have also demonstrated high CysLT₁R expression in established colon cancer cell lines^[42,80]. The CysLT₁R and

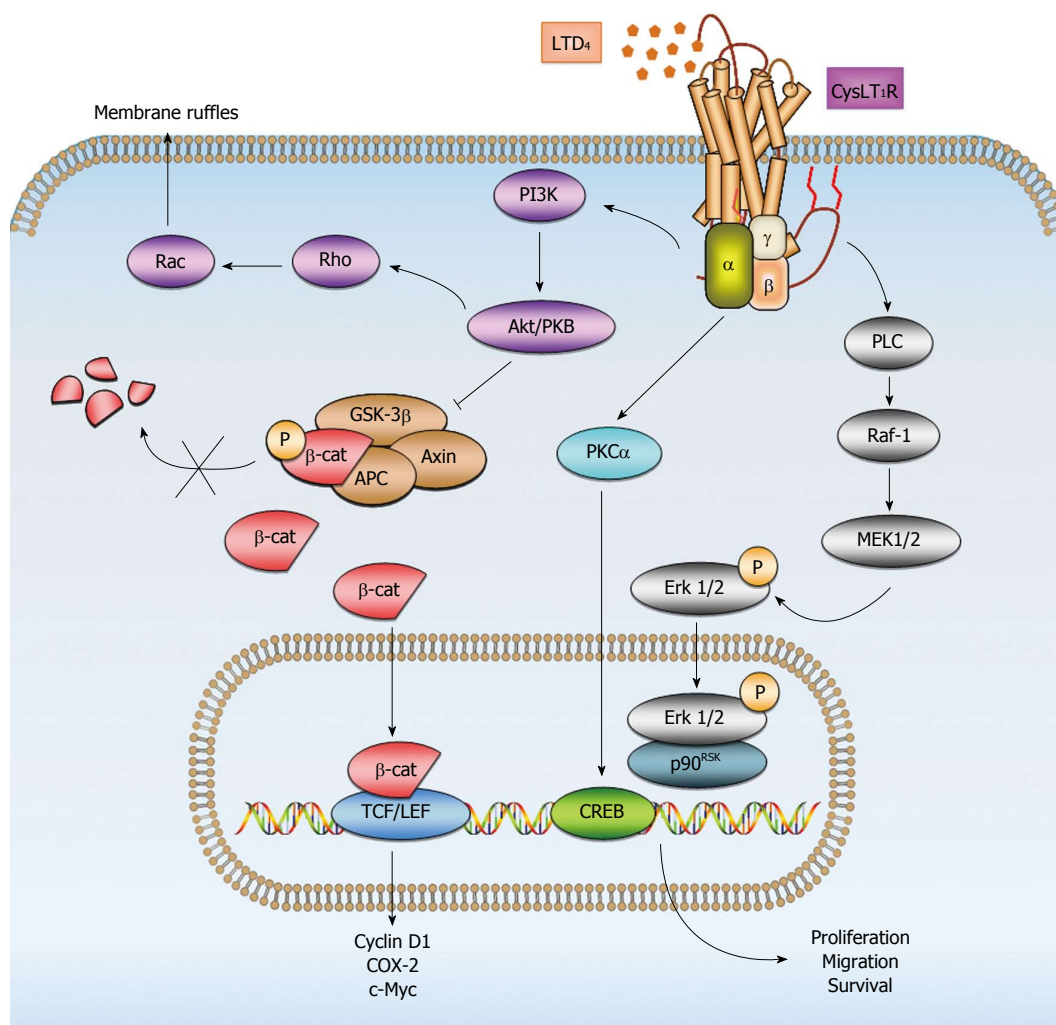


Figure 2 Signaling pathways of LTD₄ and CysLT₁R. Upon stimulation with LTD₄, various downstream signaling pathways become activated. LTD₄ induces cell membrane ruffles through phosphoinositide 3-kinase (PI3K) signaling, which in turn stimulates protein kinase Akt/PKB and the small GTPases Rho and Rac. Akt/PKB can inhibit glycogen synthase kinase-3 β (GSK-3β), which comprises the destruction complex for cytosolic β-catenin together with Axin and adenomatous polyposis coli (APC). Inhibition of the destruction complex leads to the accumulation of β-catenin in the cytosol and translocation to the nucleus. In the nucleus, β-catenin interacts primarily with members of the T-cell factor/lymphoid enhancer factor (TCF/LEF) family of transcription factors to activate target genes, leading to the expression of various proteins, such as cyclin D1, COX-2 and c-Myc, which contribute to diverse cellular processes, including proliferation and migration. LTD₄-CysLT₁R signaling also regulates cell proliferation via mitogen-activated protein kinase (MAPK)/extracellular signal-regulated kinase (Erk) pathways. LTD₄-CysLT₁R in turn activates phospholipase C (PLC), Raf-1 and mitogen-activated protein kinase kinase (MEK-1/2). This activation leads to the translocation of p-Erk from the cytosol to the nucleus and interaction with p90^{RSK}, which can activate various transcription factors. Conversely, LTD₄-CysLT₁R mediates survival through protein kinase Cα (PKCα) and the transcription factor cAMP response element-binding protein (CREB). β-cat: β-catenin.

CysLT₂R expression ratio seems to be important in the disease etiology of CRC. Accordingly, we have shown that these two receptors are co-localized and form hetero- and homodimers in a human intestinal epithelial cell line and that LTC₄ stimulation of CysLT₂R negatively regulates the plasma membrane expression of CysLT₁R by inducing internalization of the receptor heterodimer complex^[88]. The expression of CysLT₁R has also been positively correlated with the cell survival factors COX-2 and Bcl-xL in tumor specimens from patients with CRC^[80].

We have previously observed that LTD₄, via CysLT₁R, induces upregulation of proteins associated with CRC - such as COX-2, β-catenin, and Bcl-2 - in intestinal epithelial cells^[89]. Upregulation of β-catenin expression was shown to be dependent on phosphoinositide 3 kinase

(PI3K)-glycogen synthase kinase 3β (GSK-3β) signaling^[30]. Moreover, our previous work has shown that LTD₄-induced CysLT₁R signaling results in cell proliferation, survival, and migration through distinct signaling pathways. LTD₄-induced CysLT₁R-signaling through cAMP response element-binding protein (CREB) and p90 ribosomal s6 kinase (p90^{RSK}) was shown to induce survival and proliferation, respectively, while inducing migration via the PI3K-Rac signaling pathway^[90,91] (Figure 2).

COX-2 expression has also been detected in various colon cancer cells^[92], and we have shown that LTD₄ via CysLT₁R enhances survival by activating the mitogen-activated protein kinase kinase (Mek)/Erk signaling pathway and increasing COX-2 and subsequent Bcl-2 expression in the colon cancer cell line Caco-2^[93]. Furthermore, we have demonstrated that LTD₄ via CysLT₁R induces

increased proliferation and migration in the colon cancer cell line HCT-116, probably *via* the GSK-3 β / β -catenin pathway with subsequent increased transcription of the target genes *MYC* and *CCD1*^[94]. By contrast, decreased expression of CysLT₂R has been observed in different colon cancer cell lines (Caco-2 and SW480) compared with an epithelial intestinal cell line, and LTC₄ stimulation of CysLT₂R has been shown to induce differentiation as demonstrated by increased intestinal alkaline phosphatase activity in Caco-2 cells^[42]. In the same colon cancer cell line, anti-tumorigenic IFN- α was shown to induce CysLT₂R promoter activity and expression, whereas mitogenic epidermal growth factor (EGF) displayed the opposite effect, suppressing CysLT₂R promoter activity and expression. LTC₄-mediated CysLT₂R signaling suppressed EGF-induced cell migration, and IFN- α induced expression of the differentiation marker mucin-2 and alkaline phosphatase activity^[95].

These results indicate potential pro- and anti-tumorigenic properties conveyed by CysLT₁R and CysLT₂R, respectively, in CRC.

LEUKOTRIENE RECEPTOR ANTAGONISTS AND LEUKOTRIENE SYNTHESIS INHIBITORS

CysLT₁R antagonists have been used in studies of inflammatory diseases such as rheumatoid arthritis and atherosclerosis^[96,97]. The CysLT₁R antagonists pranlukast, zafirlukast and montelukast are commercially available and are currently in clinical use to treat asthmatic patients^[98]. Emerging data suggest that the pro-inflammatory CysLTs might have an important role in solid tumors.

CysLT₁R antagonist treatment has been shown to inhibit tumor growth by inducing apoptosis in a variety of human urological cancer cell lines (*e.g.*, renal cell carcinoma, bladder cancer, prostate cancer, and testicular cancer)^[99]. Montelukast has been shown to induce early apoptosis in a bladder transitional cell carcinoma (TCC) cell line, as well as in three different prostate cancer cell lines^[6,83]. In addition, montelukast has been shown to induce the intrinsic apoptotic pathway, resulting in cleavage of caspases 3 and 9, and cell cycle arrest in neuroblastoma cell lines^[84].

Studies in CysLT₁R-deficient mice have revealed its role in enhanced vascular permeability during an acute inflammatory response^[100]. Pranlukast and montelukast have been shown to reduce vascular permeability by regulating vascular endothelial growth factor (VEGF) expression in allergen-induced asthmatic lungs of mice^[101]. Furthermore, the two abovementioned CysLT₁R antagonists have been shown to inhibit the permeability of peripheral capillaries, thereby preventing tumor metastasis in a Lewis lung carcinoma metastasis model^[102]. Proliferation and migration of endothelial cells are needed to form new vessels, a process required in cancer development. Montelukast has been shown to reduce LTD₄-CysLT₁R-mediated migration of the endothelial cell line EA.hy926

via the Erk1/2 pathway^[103]. In line with aforementioned data, we have demonstrated in a nude mouse xenograft model of colon cancer that reduction of tumor growth can be accomplished with CysLT₁R antagonist treatment. The molecular mechanisms underlying the observed inhibition of tumor growth was attributed to the reduction in proliferation, induction of apoptosis and impairment of angiogenesis^[104].

We have also shown that the CysLT₁R antagonist ZM198,615 reduces proliferation in the colon cancer cell lines Caco-2 and SW480^[105]. Cianchi *et al.*^[106] reported the additive effects of the COX-2 selective inhibitor celecoxib, when combined with either the 5-LOX inhibitor MK886 or CysLT₁R antagonist LY171883, in reducing the proliferative ability of Caco-2 and HT29 cells. The combined treatment was also shown to induce apoptosis, whereas none of these compounds had any effect alone.

The COX pathway is the most extensively studied among eicosanoid pathways in CRC prevention and/or therapy. However, the cardiovascular side effects associated with long-term usage of NSAIDs and selective COX-2 inhibitors have raised some concerns. Other approaches are being explored, such as inhibition of 5-LOX activity. Simultaneous dual inhibition of COX-2 and 5-LOX activity could possibly provide a more effective and tolerable therapy than COX-inhibition alone. Accordingly, the anti-tumor effects of celecoxib in colon cancer cells were augmented when combined with inhibition of 5-LOX activity using the FLAP inhibitor MK886^[106]. The combined inhibition of COX-2 and 5-LOX activity have also shown a more pronounced anti-tumor growth effect in a cigarette smoke-promoting mouse xenograft model of CRC^[107]. However, targeting either the COX or LOX pathway alone resulted in a shunt toward the other pathway, except in the latter study, in which a shunt was observed when COX-2 activity was targeted with celecoxib. The abovementioned studies investigated the effects on CRC growth targeting the eicosanoid production in epithelial cells. However, the activity of 5-LOX of mast cells was also shown to be important in intestinal polyp formation in APCA⁴⁶⁸ mice. The mast cells were found to utilize 5-LOX to promote proliferation of intestinal epithelial cells and recruit myeloid-derived suppressor cells to the polyp site^[108]. Another possible effective chemopreventive option against CRC could be the modification of AA metabolism. Apc^{Nim/+} mice with the fed diets containing highly purified ω -3 polyunsaturated fatty acids were shown to have their mucosal AA replaced, presumably with a reduction in the production of pro-inflammatory mediators. Reduced polyp formation could be observed in both the intestine and the colon of these mice. These effects were associated with significantly decreased proliferation, COX-2 expression, and nuclear β -catenin accumulation, as well as a concomitant increase in apoptosis in the intestinal epithelium^[109]. A reduction in size and number of rectal polyps has also been observed in patients with hereditary CRC (familial adenomatous polyposis, FAP) who have undergone colectomy and received highly purified ω -3-polyunsaturated fatty acids^[110].

CONCLUSION

Deregulated AA metabolism creates an imbalance in the tissue homeostatic events of proliferation, regeneration and repair, and host defense. Additionally, deregulated AA metabolism contributes to sustained inflammatory processes that could result in CRC development. Among the implicated inflammatory mediators are the eicosanoids, such as CysLTs. Thus, the modification of CysLT signaling could pave the way for the development of new personalized medicine for patients with CRC^[104].

REFERENCES

- 1 **Tenesa A**, Dunlop MG. New insights into the aetiology of colorectal cancer from genome-wide association studies. *Nat Rev Genet* 2009; **10**: 353-358 [PMID: 19434079 DOI: 10.1038/nrg2574]
- 2 **Beller TC**, Friend DS, Maekawa A, Lam BK, Austen KF, Kanaoka Y. Cysteinyl leukotriene 1 receptor controls the severity of chronic pulmonary inflammation and fibrosis. *Proc Natl Acad Sci USA* 2004; **101**: 3047-3052 [PMID: 14970333 DOI: 10.1073/pnas.0400235101]
- 3 **Capra V**, Thompson MD, Sala A, Cole DE, Folco G, Rovati GE. Cysteinyl-leukotrienes and their receptors in asthma and other inflammatory diseases: critical update and emerging trends. *Med Res Rev* 2007; **27**: 469-527 [PMID: 16894531 DOI: 10.1002/med.20071]
- 4 **Riccioni G**, Bäck M, Capra V. Leukotrienes and atherosclerosis. *Curr Drug Targets* 2010; **11**: 882-887 [PMID: 20388065 DOI: 10.2174/138945010791320881]
- 5 **Schain F**, Schain D, Mahshid Y, Liu C, Porwit A, Xu D, Claesson HE, Sundström C, Björkholm M, Sjöberg J. Differential expression of cysteinyl leukotriene receptor 1 and 15-lipoxygenase-1 in non-Hodgkin lymphomas. *Clin Lymphoma Myeloma* 2008; **8**: 340-347 [PMID: 19064398 DOI: 10.3816/CLM.2008.n.049]
- 6 **Matsuyama M**, Funao K, Hayama T, Tanaka T, Kawahito Y, Sano H, Takemoto Y, Nakatani T, Yoshimura R. Relationship between cysteinyl-leukotriene-1 receptor and human transitional cell carcinoma in bladder. *Urology* 2009; **73**: 916-921 [PMID: 19167045 DOI: 10.1016/j.urology.2008.11.005]
- 7 **Sperling RI**. Eicosanoids in rheumatoid arthritis. *Rheum Dis Clin North Am* 1995; **21**: 741-758 [PMID: 8619097]
- 8 **Nicosia S**, Capra V, Rovati GE. Leukotrienes as mediators of asthma. *Pulm Pharmacol Ther* 2001; **14**: 3-19 [PMID: 11162414 DOI: 10.1006/pupt.2000.0262]
- 9 **Hendel J**, Nielsen OH. Expression of cyclooxygenase-2 mRNA in active inflammatory bowel disease. *Am J Gastroenterol* 1997; **92**: 1170-1173 [PMID: 9219792]
- 10 **Coussens LM**, Werb Z. Inflammation and cancer. *Nature* 2002; **420**: 860-867 [PMID: 12490959 DOI: 10.1038/nature0132]
- 11 **Rakoff-Nahoum S**. Why cancer and inflammation? *Yale J Biol Med* 2006; **79**: 123-130 [PMID: 17940622]
- 12 **Karin M**, Lawrence T, Nizet V. Innate immunity gone awry: linking microbial infections to chronic inflammation and cancer. *Cell* 2006; **124**: 823-835 [PMID: 16497591 DOI: 10.1016/j.cell.2006.02.016]
- 13 **Balkwill F**, Mantovani A. Inflammation and cancer: back to Virchow? *Lancet* 2001; **357**: 539-545 [PMID: 11229684 DOI: 10.1016/S0140-6736(00)04046-0]
- 14 **Wang D**, Dubois RN. Eicosanoids and cancer. *Nat Rev Cancer* 2010; **10**: 181-193 [PMID: 20168319 DOI: 10.1038/nrc2809]
- 15 **Algra AM**, Rothwell PM. Effects of regular aspirin on long-term cancer incidence and metastasis: a systematic comparison of evidence from observational studies versus randomised trials. *Lancet Oncol* 2012; **13**: 518-527 [PMID:

- 22440112 DOI: 10.1016/S1470-2045(12)70112-2]
- 16 **Burn J**, Gerdes AM, Macrae F, Mecklin JP, Moeslein G, Olschwang S, Eccles D, Evans DG, Maher ER, Bertario L, Bisgaard ML, Dunlop MG, Ho JW, Hodgson SV, Lindblom A, Lubinski J, Morrison PJ, Murday V, Ramesar R, Side L, Scott RJ, Thomas HJ, Vasen HF, Barker G, Crawford G, Elliott F, Movahedi M, Pylvanainen K, Wijnen JT, Fodde R, Lynch HT, Mathers JC, Bishop DT. Long-term effect of aspirin on cancer risk in carriers of hereditary colorectal cancer: an analysis from the CAPP2 randomised controlled trial. *Lancet* 2011; **378**: 2081-2087 [PMID: 22036019 DOI: 10.1016/S0140-6736(11)61049-0]
- 17 **Moysich KB**, Menezes RJ, Ronsani A, Swede H, Reid ME, Cummings KM, Falkner KL, Loewen GM, Bepler G. Regular aspirin use and lung cancer risk. *BMC Cancer* 2002; **2**: 31 [PMID: 12453317 DOI: 10.1186/1471-2407-2-31]
- 18 **Cossack M**, Ghaffary C, Watson P, Snyder C, Lynch H. Aspirin Use is Associated with Lower Prostate Cancer Risk in Male Carriers of BRCA Mutations. *J Genet Couns* 2013; Epub ahead of print [PMID: 23881471 DOI: 10.1007/s10897-013-9629-8]
- 19 **Negus RP**, Stamp GW, Hadley J, Balkwill FR. Quantitative assessment of the leukocyte infiltrate in ovarian cancer and its relationship to the expression of C-C chemokines. *Am J Pathol* 1997; **150**: 1723-1734 [PMID: 9137096]
- 20 **Condeelis J**, Pollard JW. Macrophages: obligate partners for tumor cell migration, invasion, and metastasis. *Cell* 2006; **124**: 263-266 [PMID: 16439202 DOI: 10.1016/j.cell.2006.01.007]
- 21 **Egeblad M**, Littlepage LE, Werb Z. The fibroblastic co-conspirator in cancer progression. *Cold Spring Harb Symp Quant Biol* 2005; **70**: 383-388 [PMID: 16869775 DOI: 10.1101/sqb.2005.70.007]
- 22 **Egeblad M**, Werb Z. New functions for the matrix metalloproteinases in cancer progression. *Nat Rev Cancer* 2002; **2**: 161-174 [PMID: 11990853 DOI: 10.1038/nrc745]
- 23 **Wang D**, Dubois RN. Prostaglandins and cancer. *Gut* 2006; **55**: 115-122 [PMID: 16118353 DOI: 10.1136/gut.2004.047100]
- 24 **Zlotnik A**. Chemokines and cancer. *Int J Cancer* 2006; **119**: 2026-2029 [PMID: 16671092 DOI: 10.1002/ijc.22024]
- 25 **Karin M**, Greten FR. NF-kappaB: linking inflammation and immunity to cancer development and progression. *Nat Rev Immunol* 2005; **5**: 749-759 [PMID: 16175180 DOI: 10.1038/nri1703]
- 26 **Singh RK**, Gupta S, Dastidar S, Ray A. Cysteinyl leukotrienes and their receptors: molecular and functional characteristics. *Pharmacology* 2010; **85**: 336-349 [PMID: 20516735 DOI: 10.1159/000312669]
- 27 **Samuelsson B**. The discovery of the leukotrienes. *Am J Respir Crit Care Med* 2000; **161**: S2-S6 [PMID: 10673217 DOI: 10.1164/ajrccm.161.supplement_1.lta-1]
- 28 **Peters-Golden M**, Henderson WR. Leukotrienes. *N Engl J Med* 2007; **357**: 1841-1854 [PMID: 17978293 DOI: 10.1056/NEJMr071371]
- 29 **Cathcart MC**, Lysaght J, Pidgeon GP. Eicosanoid signalling pathways in the development and progression of colorectal cancer: novel approaches for prevention/intervention. *Cancer Metastasis Rev* 2011; **30**: 363-385 [PMID: 22134655 DOI: 10.1007/s10555-011-9324-x]
- 30 **Mezhybovska M**, Wikström K, Ohd JF, Sjölander A. The inflammatory mediator leukotriene D4 induces beta-catenin signaling and its association with antiapoptotic Bcl-2 in intestinal epithelial cells. *J Biol Chem* 2006; **281**: 6776-6784 [PMID: 16407243 DOI: 10.1074/jbc.M509999200]
- 31 **Yudina Y**, Parhamifar L, Bengtsson AM, Juhas M, Sjölander A. Regulation of the eicosanoid pathway by tumour necrosis factor alpha and leukotriene D4 in intestinal epithelial cells. *Prostaglandins Leukot Essent Fatty Acids* 2008; **79**: 223-231 [PMID: 19042113 DOI: 10.1016/j.plefa.2008.09.024]
- 32 **Mezhybovska M**, Yudina Y, Abhyankar A, Sjölander A. Beta-catenin is involved in alterations in mitochondrial activ-

- ity in non-transformed intestinal epithelial and colon cancer cells. *Br J Cancer* 2009; **101**: 1596-1605 [PMID: 19826421 DOI: 10.1038/sj.bjc.6605342]
- 33 **Lam BK**, Austen KF. Leukotriene C4 synthase: a pivotal enzyme in cellular biosynthesis of the cysteinyl leukotrienes. *Prostaglandins Other Lipid Mediat* 2002; **68-69**: 511-520 [PMID: 12432940]
- 34 **Fabre JE**, Goulet JL, Riche E, Nguyen M, Coggins K, Offenbacher S, Koller BH. Transcellular biosynthesis contributes to the production of leukotrienes during inflammatory responses in vivo. *J Clin Invest* 2002; **109**: 1373-1380 [PMID: 12021253 DOI: 10.1172/JCI14869]
- 35 **Lynch KR**, O'Neill GP, Liu Q, Im DS, Sawyer N, Metters KM, Coulombe N, Abramovitz M, Figueroa DJ, Zeng Z, Conolly BM, Bai C, Austin CP, Chateauneuf A, Stocco R, Greig GM, Kargman S, Hooks SB, Hosfield E, Williams DL, Ford-Hutchinson AW, Caskey CT, Evans JF. Characterization of the human cysteinyl leukotriene CysLT1 receptor. *Nature* 1999; **399**: 789-793 [PMID: 10391245 DOI: 10.1038/21658]
- 36 **Heise CE**, O'Dowd BF, Figueroa DJ, Sawyer N, Nguyen T, Im DS, Stocco R, Bellefeuille JN, Abramovitz M, Cheng R, Williams DL, Zeng Z, Liu Q, Ma L, Clements MK, Coulombe N, Liu Y, Austin CP, George SR, O'Neill GP, Metters KM, Lynch KR, Evans JF. Characterization of the human cysteinyl leukotriene 2 receptor. *J Biol Chem* 2000; **275**: 30531-30536 [PMID: 10851239 DOI: 10.1074/jbc.M003490200]
- 37 **Ciana P**, Fumagalli M, Trincavelli ML, Verderio C, Rosa P, Lecca D, Ferrario S, Parravicini C, Capra V, Gelosa P, Guerrini U, Belcredito S, Cimino M, Sironi L, Tremoli E, Rovati GE, Martini C, Abbracchio MP. The orphan receptor GPR17 identified as a new dual uracil nucleotides/cysteinyl-leukotrienes receptor. *EMBO J* 2006; **25**: 4615-4627 [PMID: 16990797 DOI: 10.1038/sj.emboj.7601341]
- 38 **Paruchuri S**, Tashimo H, Feng C, Maekawa A, Xing W, Jiang Y, Kanaoka Y, Conley P, Boyce JA. Leukotriene E4-induced pulmonary inflammation is mediated by the P2Y12 receptor. *J Exp Med* 2009; **206**: 2543-2555 [PMID: 19822647 DOI: 10.1084/jem.20091240]
- 39 **Maekawa A**, Kanaoka Y, Xing W, Austen KF. Functional recognition of a distinct receptor preferential for leukotriene E4 in mice lacking the cysteinyl leukotriene 1 and 2 receptors. *Proc Natl Acad Sci USA* 2008; **105**: 16695-16700 [PMID: 18931305 DOI: 10.1073/pnas.0808993105]
- 40 **Jans DA**, Xiao CY, Lam MH. Nuclear targeting signal recognition: a key control point in nuclear transport? *Bioessays* 2000; **22**: 532-544 [PMID: 10842307 DOI: 10.1002/(SICI)1521-1878(200006)]
- 41 **Servant MJ**, Tenover B, Lin R. Overlapping and distinct mechanisms regulating IRF-3 and IRF-7 function. *J Interferon Cytokine Res* 2002; **22**: 49-58 [PMID: 11846975 DOI: 10.1089/107999002753452656]
- 42 **Magnusson C**, Ehrnström R, Olsen J, Sjölander A. An increased expression of cysteinyl leukotriene 2 receptor in colorectal adenocarcinomas correlates with high differentiation. *Cancer Res* 2007; **67**: 9190-9198 [PMID: 17909024 DOI: 10.1158/0008-5472.CAN-07-0771]
- 43 **Chan CC**, McKee K, Tagari P, Chee P, Ford-Hutchinson A. Eosinophil-eicosanoid interactions: inhibition of eosinophil chemotaxis in vivo by a LTD4-receptor antagonist. *Eur J Pharmacol* 1990; **191**: 273-280 [PMID: 1964905 DOI: 10.1016/0014-2999(90)94159-U]
- 44 **Barnes NC**, Piper PJ, Costello JF. Comparative effects of inhaled leukotriene C4, leukotriene D4, and histamine in normal human subjects. *Thorax* 1984; **39**: 500-504 [PMID: 6463929 DOI: 10.1136/thx.39.7.500]
- 45 **Drazen JM**, Austen KF, Lewis RA, Clark DA, Goto G, Marfat A, Corey EJ. Comparative airway and vascular activities of leukotrienes C-1 and D in vivo and in vitro. *Proc Natl Acad Sci USA* 1980; **77**: 4354-4358 [PMID: 6933488 DOI: 10.1073/pnas.77.7.4354]
- 46 **Marom Z**, Shelhamer JH, Bach MK, Morton DR, Kaliner M. Slow-reacting substances, leukotrienes C4 and D4, increase the release of mucus from human airways in vitro. *Am Rev Respir Dis* 1982; **126**: 449-451 [PMID: 7125334]
- 47 **Ford-Hutchinson AW**. Leukotriene B4 in inflammation. *Crit Rev Immunol* 1990; **10**: 1-12 [PMID: 2155000]
- 48 **Funk CD**. Prostaglandins and leukotrienes: advances in eicosanoid biology. *Science* 2001; **294**: 1871-1875 [PMID: 11729303 DOI: 10.1126/science.294.5548.1871]
- 49 **Yokomizo T**, Izumi T, Chang K, Takuwa Y, Shimizu T. A G-protein-coupled receptor for leukotriene B4 that mediates chemotaxis. *Nature* 1997; **387**: 620-624 [PMID: 9177352 DOI: 10.1038/42506]
- 50 **Yokomizo T**, Kato K, Terawaki K, Izumi T, Shimizu T. A second leukotriene B(4) receptor, BLT2. A new therapeutic target in inflammation and immunological disorders. *J Exp Med* 2000; **192**: 421-432 [PMID: 10934230 DOI: 10.1084/jem.192.3.421]
- 51 **Yokomizo T**, Izumi T, Shimizu T. Leukotriene B4: metabolism and signal transduction. *Arch Biochem Biophys* 2001; **385**: 231-241 [PMID: 11368003 DOI: 10.1006/abbi.2000.2168]
- 52 **Hussain SP**, Hofseth LJ, Harris CC. Radical causes of cancer. *Nat Rev Cancer* 2003; **3**: 276-285 [PMID: 12671666 DOI: 10.1038/nrc1046]
- 53 **Bernstein CN**, Blanchard JF, Kliewer E, Wajda A. Cancer risk in patients with inflammatory bowel disease: a population-based study. *Cancer* 2001; **91**: 854-862 [PMID: 11241255 DOI: 10.1002/1097-0142(20010215)]
- 54 **Hammerbeck DM**, Brown DR. Presence of immunocytes and sulfidopeptide leukotrienes in the inflamed guinea pig distal colon. *Inflammation* 1996; **20**: 413-425 [PMID: 8872504 DOI: 10.1007/BF01486743]
- 55 **Rubin DC**, Shaker A, Levin MS. Chronic intestinal inflammation: inflammatory bowel disease and colitis-associated colon cancer. *Front Immunol* 2012; **3**: 107 [PMID: 22586430 DOI: 10.3389/fimmu.2012.00107]
- 56 **Eaden J**. Review article: colorectal carcinoma and inflammatory bowel disease. *Aliment Pharmacol Ther* 2004; **20** Suppl 4: 24-30 [PMID: 15352890 DOI: 10.1111/j.1365-2036.2004.02046.x]
- 57 **Ekbohm A**, Helmick C, Zack M, Adami HO. Ulcerative colitis and colorectal cancer. A population-based study. *N Engl J Med* 1990; **323**: 1228-1233 [PMID: 2215606 DOI: 10.1056/NEJM199011013231802]
- 58 **Sharon P**, Stenson WF. Enhanced synthesis of leukotriene B4 by colonic mucosa in inflammatory bowel disease. *Gastroenterology* 1984; **86**: 453-460 [PMID: 6319219]
- 59 **Kim JH**, Tagari P, Griffiths AM, Ford-Hutchinson A, Smith C, Sherman PM. Levels of peptidoleukotriene E4 are elevated in active Crohn's disease. *J Pediatr Gastroenterol Nutr* 1995; **20**: 403-407 [PMID: 7636682 DOI: 10.1097/00005176-199505000-00005]
- 60 **Nishikawa M**, Hikasa Y, Hori K, Tanida N, Shimoyama T. Effect of leukotriene C4D4 antagonist on colonic damage induced by intracolonic administration of trinitrobenzene sulfonic acid in rats. *J Gastroenterol* 1995; **30**: 34-40 [PMID: 7719412 DOI: 10.1007/BF01211372]
- 61 **Shanahan F**. Inflammatory bowel disease: immunodiagnosics, immunotherapeutics, and ecotherapeutics. *Gastroenterology* 2001; **120**: 622-635 [PMID: 11179240 DOI: 10.1053/gast.2001.22122]
- 62 **Fuss IJ**, Neurath M, Boirivant M, Klein JS, de la Motte C, Strong SA, Fiocchi C, Strober W. Disparate CD4+ lamina propria (LP) lymphokine secretion profiles in inflammatory bowel disease. Crohn's disease LP cells manifest increased secretion of IFN-gamma, whereas ulcerative colitis LP cells manifest increased secretion of IL-5. *J Immunol* 1996; **157**: 1261-1270 [PMID: 8757634]
- 63 **Atreya I**, Neurath MF. Immune cells in colorectal cancer: prognostic relevance and therapeutic strategies. *Expert Rev*

- Anticancer Ther* 2008; **8**: 561-572 [PMID: 18402523 DOI: 10.1586/14737140.8.4.561]
- 64 **Gabrilovich DI**, Nagaraj S. Myeloid-derived suppressor cells as regulators of the immune system. *Nat Rev Immunol* 2009; **9**: 162-174 [PMID: 19197294 DOI: 10.1038/nri2506]
- 65 **Solinas G**, Germano G, Mantovani A, Allavena P. Tumor-associated macrophages (TAM) as major players of the cancer-related inflammation. *J Leukoc Biol* 2009; **86**: 1065-1073 [PMID: 19741157 DOI: 10.1189/jlb.0609385]
- 66 **Sica A**, Schioppa T, Mantovani A, Allavena P. Tumour-associated macrophages are a distinct M2 polarised population promoting tumour progression: potential targets of anti-cancer therapy. *Eur J Cancer* 2006; **42**: 717-727 [PMID: 16520032 DOI: 10.1016/j.ejca.2006.01.003]
- 67 **Zhang Y**, Sime W, Juhas M, Sjölander A. Crosstalk between colon cancer cells and macrophages via inflammatory mediators and CD47 promotes tumour cell migration. *Eur J Cancer* 2013; **49**: 3320-3334 [PMID: 23810249 DOI: 10.1016/j.ejca.2013.06.005]
- 68 **Sampson AP**. The role of eosinophils and neutrophils in inflammation. *Clin Exp Allergy* 2000; **30** Suppl 1: 22-27 [PMID: 10849470 DOI: 10.1046/j.1365-2222.2000.00092]
- 69 **Earashi M**, Noguchi M, Tanaka M. In vitro effects of eicosanoid synthesis inhibitors in the presence of linoleic acid on MDA-MB-231 human breast cancer cells. *Breast Cancer Res Treat* 1996; **37**: 29-37 [PMID: 8750525 DOI: 10.1007/BF01806629]
- 70 **Okano-Mitani H**, Ikai K, Imamura S. Human melanoma cells generate leukotrienes B4 and C4 from leukotriene A4. *Arch Dermatol Res* 1997; **289**: 347-351 [PMID: 9209681 DOI: 10.1007/s004030050203]
- 71 **Bittner S**, Wielckens K. Glucocorticoid-induced lymphoma cell growth inhibition: the role of leukotriene B4. *Endocrinology* 1988; **123**: 991-1000 [PMID: 2840273 DOI: 10.1210/endo-123-2-991]
- 72 **el-Hakim IE**, Langdon JD, Zakrzewski JT, Costello JF. Leukotriene B4 and oral cancer. *Br J Oral Maxillofac Surg* 1990; **28**: 155-159 [PMID: 1966928 DOI: 10.1016/0266-4356(90)90078-Y]
- 73 **Ihara A**, Wada K, Yoneda M, Fujisawa N, Takahashi H, Nakajima A. Blockade of leukotriene B4 signaling pathway induces apoptosis and suppresses cell proliferation in colon cancer. *J Pharmacol Sci* 2007; **103**: 24-32 [PMID: 17220595 DOI: 10.1254/jphs.FP0060651]
- 74 **Hennig R**, Ding XZ, Tong WG, Witt RC, Jovanovic BD, Adrian TE. Effect of LY293111 in combination with gemcitabine in colonic cancer. *Cancer Lett* 2004; **210**: 41-46 [PMID: 15172119 DOI: 10.1016/j.canlet.2004.02.023]
- 75 **Wang D**, Wang H, Shi Q, Katkuri S, Walhi W, Desvergne B, Das SK, Dey SK, DuBois RN. Prostaglandin E(2) promotes colorectal adenoma growth via transactivation of the nuclear peroxisome proliferator-activated receptor delta. *Cancer Cell* 2004; **6**: 285-295 [PMID: 15380519 DOI: 10.1016/j.ccr.2004.08.011]
- 76 **Kawamori T**, Uchiya N, Sugimura T, Wakabayashi K. Enhancement of colon carcinogenesis by prostaglandin E2 administration. *Carcinogenesis* 2003; **24**: 985-990 [PMID: 12771044 DOI: 10.1093/carcin/bgg033]
- 77 **Nakanishi M**, Montrose DC, Clark P, Nambiar PR, Belinsky GS, Claffey KP, Xu D, Rosenberg DW. Genetic deletion of mPGES-1 suppresses intestinal tumorigenesis. *Cancer Res* 2008; **68**: 3251-3259 [PMID: 18451151 DOI: 10.1158/0008-5472.CAN-07-6100]
- 78 **Nakanishi M**, Menoret A, Tanaka T, Miyamoto S, Montrose DC, Vella AT, Rosenberg DW. Selective PGE(2) suppression inhibits colon carcinogenesis and modifies local mucosal immunity. *Cancer Prev Res (Phila)* 2011; **4**: 1198-1208 [PMID: 21576350 DOI: 10.1158/1940-6207]
- 79 **Dreyling KW**, Hoppe U, Peskar BA, Morgenroth K, Kozushek W, Peskar BM. Leukotriene synthesis by human gastrointestinal tissues. *Biochim Biophys Acta* 1986; **878**: 184-193 [PMID: 3019409 DOI: 10.1016/0005-2760(86)90145-1]
- 80 **Ohd JF**, Nielsen CK, Campbell J, Landberg G, Löfberg H, Sjölander A. Expression of the leukotriene D4 receptor CysLT1, COX-2, and other cell survival factors in colorectal adenocarcinomas. *Gastroenterology* 2003; **124**: 57-70 [PMID: 12512030 DOI: 10.1053/gast.2003.50011]
- 81 **Soumaoro LT**, Iida S, Uetake H, Ishiguro M, Takagi Y, Higuchi T, Yasuno M, Enomoto M, Sugihara K. Expression of 5-lipoxygenase in human colorectal cancer. *World J Gastroenterol* 2006; **12**: 6355-6360 [PMID: 17072961]
- 82 **Wang D**, DuBois RN. The role of anti-inflammatory drugs in colorectal cancer. *Annu Rev Med* 2013; **64**: 131-144 [PMID: 23020877 DOI: 10.1146/annurev-med-112211-154330]
- 83 **Matsuyama M**, Hayama T, Funao K, Kawahito Y, Sano H, Takemoto Y, Nakatani T, Yoshimura R. Overexpression of cysteinyl LT1 receptor in prostate cancer and CysLT1R antagonist inhibits prostate cancer cell growth through apoptosis. *Oncol Rep* 2007; **18**: 99-104 [PMID: 17549353]
- 84 **Sveinbjörnsson B**, Rasmuson A, Baryawno N, Wan M, Petersen I, Ponthan F, Orrego A, Haeggström JZ, Johnsen JL, Kogner P. Expression of enzymes and receptors of the leukotriene pathway in human neuroblastoma promotes tumor survival and provides a target for therapy. *FASEB J* 2008; **22**: 3525-3536 [PMID: 18591367 DOI: 10.1096/fj.07-103457]
- 85 **Zhang WP**, Hu H, Zhang L, Ding W, Yao HT, Chen KD, Sheng WW, Chen Z, Wei EQ. Expression of cysteinyl leukotriene receptor 1 in human traumatic brain injury and brain tumors. *Neurosci Lett* 2004; **363**: 247-251 [PMID: 15182953 DOI: 10.1016/j.neulet.2004.03.088]
- 86 **Magnusson C**, Liu J, Ehrnström R, Manjer J, Jirstrom K, Andersson T, Sjölander A. Cysteinyl leukotriene receptor expression pattern affects migration of breast cancer cells and survival of breast cancer patients. *Int J Cancer* 2011; **129**: 9-22 [PMID: 20824707 DOI: 10.1002/ijc.25648]
- 87 **Magnusson C**, Mezhybovska M, Löhrinc E, Fernebro E, Nilbert M, Sjölander A. Low expression of CysLT1R and high expression of CysLT2R mediate good prognosis in colorectal cancer. *Eur J Cancer* 2010; **46**: 826-835 [PMID: 20064706 DOI: 10.1016/j.ejca.2009]
- 88 **Parhamifar L**, Sime W, Yudina Y, Vilhardt F, Mörgelin M, Sjölander A. Ligand-induced tyrosine phosphorylation of cysteinyl leukotriene receptor 1 triggers internalization and signaling in intestinal epithelial cells. *PLoS One* 2010; **5**: e14439 [PMID: 21203429 DOI: 10.1371/journal.pone.0014439]
- 89 **Ohd JF**, Wikström K, Sjölander A. Leukotrienes induce cell-survival signaling in intestinal epithelial cells. *Gastroenterology* 2000; **119**: 1007-1018 [PMID: 11040187 DOI: 10.1053/gast.2000.18141]
- 90 **Paruchuri S**, Sjölander A. Leukotriene D4 mediates survival and proliferation via separate but parallel pathways in the human intestinal epithelial cell line Int 407. *J Biol Chem* 2003; **278**: 45577-45585 [PMID: 12912998 DOI: 10.1074/jbc.M302881200]
- 91 **Paruchuri S**, Broom O, Dib K, Sjölander A. The pro-inflammatory mediator leukotriene D4 induces phosphatidylinositol 3-kinase and Rac-dependent migration of intestinal epithelial cells. *J Biol Chem* 2005; **280**: 13538-13544 [PMID: 15657050 DOI: 10.1074/jbc.M409811200]
- 92 **Parker J**, Kaplon MK, Alvarez CJ, Krishnaswamy G. Prostaglandin H synthase expression is variable in human colorectal adenocarcinoma cell lines. *Exp Cell Res* 1997; **236**: 321-329 [PMID: 9344613 DOI: 10.1006/excr.1997.3741]
- 93 **Wikström K**, Ohd JF, Sjölander A. Regulation of leukotriene-dependent induction of cyclooxygenase-2 and Bcl-2. *Biochem Biophys Res Commun* 2003; **302**: 330-335 [PMID: 12604350 DOI: 10.1016/S0006-]
- 94 **Salim T**, Sand-Dejmek J, Sjölander A. The inflammatory mediator leukotriene D4 induces subcellular β -catenin translocation and migration of colon cancer cells. *Exp Cell Res* 2013; Epub ahead of print [PMID: 24211746 DOI: 10.1016/

- j.yexcr.2013.10.021]
- 95 **Magnusson C**, Bengtsson AM, Liu M, Liu J, Ceder Y, Ehrnström R, Sjölander A. Regulation of cysteinyl leukotriene receptor 2 expression—a potential anti-tumor mechanism. *PLoS One* 2011; **6**: e29060 [PMID: 22194989 DOI: 10.1371/journal.pone.0029060]
 - 96 **Shiota N**, Shimoura K, Okunishi H. Pathophysiological role of mast cells in collagen-induced arthritis: study with a cysteinyl leukotriene receptor antagonist, montelukast. *Eur J Pharmacol* 2006; **548**: 158-166 [PMID: 16949072 DOI: 10.1016/j.ejphar.2006.07.046]
 - 97 **Mueller CF**, Wassmann K, Widder JD, Wassmann S, Chen CH, Keuler B, Kudin A, Kunz WS, Nickenig G. Multidrug resistance protein-1 affects oxidative stress, endothelial dysfunction, and atherogenesis via leukotriene C4 export. *Circulation* 2008; **117**: 2912-2918 [PMID: 18506003 DOI: 10.1161/CIRCULATIONAHA.107.747667]
 - 98 **Bateman ED**, Hurd SS, Barnes PJ, Bousquet J, Drazen JM, FitzGerald M, Gibson P, Ohta K, O'Byrne P, Pedersen SE, Pizzichini E, Sullivan SD, Wenzel SE, Zar HJ. Global strategy for asthma management and prevention: GINA executive summary. *Eur Respir J* 2008; **31**: 143-178 [PMID: 18166595 DOI: 10.1183/09031936.00138707]
 - 99 **Matsuyama M**, Yoshimura R. Cysteinyl-leukotriene1 receptor is a potent target for the prevention and treatment of human urological cancer. *Mol Med Rep* 2010; **3**: 245-251 [PMID: 21472229 DOI: 10.3892/mmr_00000247]
 - 100 **Maekawa A**, Austen KF, Kanaoka Y. Targeted gene disruption reveals the role of cysteinyl leukotriene 1 receptor in the enhanced vascular permeability of mice undergoing acute inflammatory responses. *J Biol Chem* 2002; **277**: 20820-20824 [PMID: 11932261 DOI: 10.1074/jbc.M203163200]
 - 101 **Lee KS**, Kim SR, Park HS, Jin GY, Lee YC. Cysteinyl leukotriene receptor antagonist regulates vascular permeability by reducing vascular endothelial growth factor expression. *J Allergy Clin Immunol* 2004; **114**: 1093-1099 [PMID: 15536415 DOI: 10.1016/j.jaci.2004.07.039]
 - 102 **Nozaki M**, Yoshikawa M, Ishitani K, Kobayashi H, Houkin K, Imai K, Ito Y, Muraki T. Cysteinyl leukotriene receptor antagonists inhibit tumor metastasis by inhibiting capillary permeability. *Keio J Med* 2010; **59**: 10-18 [PMID: 20375653 DOI: 10.2302/kjm.59.10]
 - 103 **Yuan YM**, Fang SH, Qian XD, Liu LY, Xu LH, Shi WZ, Zhang LH, Lu YB, Zhang WP, Wei EQ. Leukotriene D4 stimulates the migration but not proliferation of endothelial cells mediated by the cysteinyl leukotriene cyslt(1) receptor via the extracellular signal-regulated kinase pathway. *J Pharmacol Sci* 2009; **109**: 285-292 [PMID: 19234368 DOI: 10.1254/jphs.08321FP]
 - 104 **Savari S**, Liu M, Zhang Y, Sime W, Sjölander A. CysLT(1)R antagonists inhibit tumor growth in a xenograft model of colon cancer. *PLoS One* 2013; **8**: e73466 [PMID: 24039952 DOI: 10.1371/journal.pone.0073466]
 - 105 **Paruchuri S**, Mezhybovska M, Juhas M, Sjölander A. Endogenous production of leukotriene D4 mediates autocrine survival and proliferation via CysLT1 receptor signalling in intestinal epithelial cells. *Oncogene* 2006; **25**: 6660-6665 [PMID: 16715140 DOI: 10.1038/sj.onc.1209666]
 - 106 **Cianchi F**, Cortesini C, Magnelli L, Fanti E, Papucci L, Schiavone N, Messerini L, Vannacci A, Capaccioli S, Perna F, Lulli M, Fabbri V, Perigli G, Bechi P, Masini E. Inhibition of 5-lipoxygenase by MK886 augments the antitumor activity of celecoxib in human colon cancer cells. *Mol Cancer Ther* 2006; **5**: 2716-2726 [PMID: 17121918 DOI: 10.1158/1535-7163.MCT-06-0318]
 - 107 **Ye YN**, Wu WK, Shin VY, Bruce IC, Wong BC, Cho CH. Dual inhibition of 5-LOX and COX-2 suppresses colon cancer formation promoted by cigarette smoke. *Carcinogenesis* 2005; **26**: 827-834 [PMID: 15637091 DOI: 10.1093/carcin/bgi012]
 - 108 **Cheon EC**, Khazaie K, Khan MW, Strouch MJ, Krantz SB, Phillips J, Blatner NR, Hix LM, Zhang M, Dennis KL, Salabat MR, Heiferman M, Grippo PJ, Munshi HG, Gounaris E, Bentrem DJ. Mast cell 5-lipoxygenase activity promotes intestinal polyposis in APCDelta468 mice. *Cancer Res* 2011; **71**: 1627-1636 [PMID: 21216893 DOI: 10.1158/0008-5472.CAN-10-1923]
 - 109 **Fini L**, Piazzi G, Ceccarelli C, Daoud Y, Belluzzi A, Munarini A, Graziani G, Fogliano V, Selgrad M, Garcia M, Gasbarrini A, Genta RM, Boland CR, Ricciardiello L. Highly purified eicosapentaenoic acid as free fatty acids strongly suppresses polyyps in Apc(Min/+) mice. *Clin Cancer Res* 2010; **16**: 5703-5711 [PMID: 21030497 DOI: 10.1158/1078-0432.CCR-10-1990]
 - 110 **West NJ**, Clark SK, Phillips RK, Hutchinson JM, Leicester RJ, Belluzzi A, Hull MA. Eicosapentaenoic acid reduces rectal polyp number and size in familial adenomatous polyposis. *Gut* 2010; **59**: 918-925 [PMID: 20348368 DOI: 10.1136/gut.2009.200642]

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