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Effect of In Utero and Early-Life Conditions on Adult Health and Disease

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Along latency period between an environmental trigger and the onset of subsequent disease is widely recognized in the etiology of certain cancers, yet this phenomenon is not generally considered in the etiology of other conditions such as cardiovascular disease, metabolic disease, or osteoporosis. However, many lines of evidence, including epidemiologic data and data from extensive clinical and experimental studies, indicate that early life events play a powerful role in influencing later susceptibility to certain chronic diseases. An increased understanding of developmental plasticity (defined as the ability of an organism to develop in various ways, depending on the particular environment or setting) provides a conceptual basis for these observations.¹

Developmental plasticity requires stable modulation of gene expression, and this appears to be mediated, at least in part, by epigenetic processes such as DNA methylation and histone modification. Thus, both the genome and the epigenome interactively influence the mature phenotype and determine sensitivity to later environmental factors and the subsequent risk of disease. In this review, we synthesize evidence from several disciplines to support the contention that environmental factors acting during development should be accorded greater weight in models of disease causation.

EPIDEMIOLOGIC AND CLINICAL OBSERVATIONS

The epidemiologic observations that smaller size or relative thinness at birth and during infancy is associated with increased rates of coronary heart disease, stroke, type 2 diabetes mellitus, adiposity, the metabolic syndrome, and osteoporosis in adult life²⁻⁶ have been extensively replicated. Perinatal events appear to exert effects that are independent of environmental risk factors in adults^{7,8} or may be amplified by other risk factors.⁹ Slow growth in utero may be associated with increased allocation of nutrients to adipose tissue during development and may then result in accelerated weight gain during childhood,^{10,11} which may contribute to a relatively greater risk of coronary heart disease, hypertension, and type 2 diabetes mellitus. There is a continuous relation between birth weight and future risk — not just for extreme weights but also for normal weights.¹² Prematurity itself, independent of size for gestational age, has been associated with insulin resistance and

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glucose intolerance in prepubertal children¹³ that may track into young adulthood and may be accompanied by elevated blood pressure.¹⁴

In mammalian development, the mother transduces environmental information such as nutritional status to her embryo or fetus through the placenta or to her infant through lactation. Fetal growth is generally matched to the mother's body size (rather than to genetic potential) through what is termed maternal constraint.^{15,16} Maternal constraint may be mediated, in part, by the limiting effects of placental size in utero or perfusion on fetal nutrition, but imprinted genes, particularly those linked to the expression of growth factors, may also play a role in the allocation of nutritional resources.¹⁷ Maternal constraint is increased with short maternal stature, young or old maternal age, first pregnancy, or multiple pregnancy; in addition, the effects of unbalanced maternal diet or excessive maternal thinness or fatness influence fetal nutrition in the absence of other disease. Beyond these mechanisms, fetal development may be further impaired by poor placental function or maternal disease, each of which can influence several points along the pathway from the mother's intake of food to the delivery of nutrients to growing fetal tissues.¹⁸

The developmental-origins hypothesis proposes that an altered long-term risk of disease is initially induced through adaptive responses that the fetus or infant makes to cues from the mother about her health or physical state. Fetal or perinatal responses may include changes in metabolism, hormone production, and tissue sensitivity to hormones that may affect the relative development of various organs, leading to persistent alterations in physiologic and metabolic homeostatic set points. Thus, the association between reduced fetal growth rate, small body size at birth, and a later risk of disease may be interpreted as reflecting the long-term consequences of fetal adaptive responses. However, reduced overall fetal body growth is seen not as causing the long-term consequences but rather as constituting a marker of a coordinated fetal response to a limiting intrauterine environment, resulting in changes in tissue and organ development that are not necessarily evident at birth but that result in perturbed responses later in life.¹⁹

The effects of subsequent environmental exposures during infancy, childhood, and adult life may be influenced by these past exposures and may condition the later risk of disease. For example, there are hints from a cross-sectional study that insulin resistance develops at a lower body-mass index in British children of South Asian ancestry than in British children of European ancestry,²⁰ perhaps reflecting the lower birth weight of the South Asian children, which is the result of different statures and nutritional states of the mothers.

When undernutrition during early development is followed by improved nutrition later in development, whether during late gestation or the early postnatal period, many mammals retain some capacity to compensate, by increasing their growth rate. Life-history theory predicts that such compensatory changes will carry costs — for example, a reduced life span as a result of diversion of resources from repair capacity to growth.²¹ This may explain why rapid childhood growth, especially in people who were born small or were thin in infancy, appears to have deleterious effects on later health.^{10,11}

Although it has been proposed that the associations between fetal and infant growth and later adult disease represent the multiple (pleiotropic) effects of genes transmitted from mother to child,²² maternally mediated environmental modulation of gene expression in offspring may be more important than purely heritable genetic risk. Studies of osteoporosis provide one example. Currently identified genetic markers explain only a small proportion of the variation in individual bone mass and risk of fracture,²³ as exemplified by the relatively weak associations between, on the one hand, single-nucleotide polymorphisms in the genes for the vitamin D receptor, type 1 collagen, or growth hormone and, on the other

hand, adult bone density or bone loss. In a study of a small cohort of elderly subjects,²⁴ no significant association was found between either birth weight or vitamin D receptor genotype and bone mineral density; however, the relationship between lumbar spine bone mineral density and vitamin D receptor genotype varied according to birth weight (Fig. 1). These data hint that genetic influences on vitamin D response, and therefore on adult bone mineral density, might be modified by undernutrition in utero. The results of studies involving twins appear to support these observations: in a cohort study of female twins (4008 subjects), there was significant residual intrapair concordance between birth weight and bone mass, even between monozygous twins, suggesting that a larger proportion of variation in birth weight and bone mass in the population may result from the intrauterine environment than from genomic inheritance.²⁵

Much attention has been focused on fetal undernutrition as a facilitator of predisposition to later disease, but there is evidence that excessive energy supply to the fetus or infant also has adverse consequences. Maternal hyperglycemia, for example, may lead to fetal hyperinsulinemia and fat deposition, and substantial data suggest that the offspring of obese women or women with diabetes are at greater risk for developing metabolic disorders themselves, even during childhood.^{26,27} Thus, the relation between prenatal nutrition and later metabolic disease is likely to be U-shaped, with increased risk at both ends of the birthweight curve. Infants who are fed formula have a higher energy intake and, in general, greater early gain in body weight than breast-fed infants, and they appear to have a greater risk of obesity in later life,²⁸ findings that suggest further complexity of the long-term effects of prenatal and early-life nutrition. In addition, epidemiologic studies have drawn other associations between higher birth weights and greater risk in adults of other conditions, such as breast cancer.²⁹

PHYSIOLOGICAI, CELLULAR, AND MOLECULAR BASES OF DEVELOPMENTAL PLASTICITY

INTEGRATED RESPONSES

The biologic basis for invoking developmental plasticity as an influence on the risk of disease derives from numerous studies in animals in which dietary, endocrine, or physical challenges at various times from conception until weaning induce persistent changes in cardiovascular and metabolic function in the offspring. The most commonly used animal models involve a prenatal nutrient imbalance, which can be induced by a global reduction in overall maternal food intake³⁰ or by protein restriction in an isocaloric diet,³¹ or glucocorticoid exposure (without any change in diet).³²

Embryos of pregnant rats fed a low-protein diet during the preimplantation period (0 to 4.25 days) show altered development in multiple organ systems, and if the gestation was permitted to reach term, the offspring had reduced birth weights, relatively increased postnatal growth, or adult-onset hypertension.³³ This outcome may reflect a direct effect on the environment of the fertilized ovum, since other rodent studies have shown that in vitro culture from the two-cell stage to the blastocyst followed by embryo transfer, or even transfer at the blastocyst stage without previous culture, may result in elevated blood pressure in adult offspring.³⁴ The periconceptional period is clearly one of particular sensitivity, since even specific nutrient deficiencies (of B₁₂, folate, or methionine) at this stage can have effects on later metabolism and blood pressure in sheep³⁵; imbalance in maternal B₁₂ and folate status during pregnancy has recently been reported to contribute to childhood insulin resistance in humans.³⁶

The administration of glucocorticoids to the pregnant rat at specific points during gestation has been reported to cause hypertension³⁷ and insulin resistance³⁸ in the offspring in later

life, as well as alterations in gene expression in the developing brain of the offspring and increased sensitivity to postnatal stress.³⁹ In the rat, maternal undernutrition during pregnancy may result in offspring that later show central obesity and reduced skeletal-muscle mass, altered insulin sensitivity, altered hepatic metabolism, reduced numbers of nephrons, hypertension, and altered endothelial function, together with altered appetite regulation, level of activity, and neuroendocrine control.^{30,31,40,41} Postnatal stress, in the form of reduced grooming and licking by the mother, has been shown to induce neurodevelopmental changes in rat pups that lead to excessive behavioral and hypothalamic–pituitary axis responses to stress later in life; such variations in maternal behavior appear to have effects on glucocorticoid-receptor gene expression in the hippocampus of the offspring.⁴² As in humans, however, the effects of early cues are complex. For example, in the offspring of rats, increases in blood pressure induced by a maternal low-protein diet are influenced by sex,^{33,43} estrogen level,⁴⁴ and the particular composition of the diet⁴⁵ and are subject to postnatal environmental factors.^{46,47}

There are several reported similarities, such as induction of hypertension and altered insulin sensitivity, between the effects of maternal nutritional challenges and glucocorticoid challenges on the offspring, findings that suggest common mechanisms. One hypothesis is that unbalanced maternal nutrition might lead to increased fetal cortisol levels or might alter the expression of the glucocorticoid receptor,^{48,49} influencing growth and maturation of fetal organs. Such alterations might cause preterm delivery and might also affect the long-term function of many organs.⁵⁰ However, an elevated fetal cortisol level is unlikely to account for all the effects produced in animal models by manipulation of the intrauterine milieu, especially those induced by imbalanced periconceptional nutrition.⁵¹

EXPERIMENTAL DATA RELEVANT TO HUMAN DISEASE

There are critical periods in the differentiation and maturation of the tissues and cells involved in organogenesis throughout gestation and early postnatal life. We illustrate this concept using the examples of the kidney, heart, and pancreas, since their functional units are formed prenatally in the human fetus. The subject of environmental perturbations, organogenesis, and perinatal effects is extensively reviewed elsewhere.^{19,52}

In the kidney, maternal dietary imbalance may lead to developmentally induced deviations from the optimal ratio of body mass to nephron number. A relative deficiency in the number of nephrons is thought to create an increased risk of inadequate renal function and hypertension in later life^{31,53} and, ultimately, a predisposition to renal failure and a potentially reduced life span.⁵⁴ The severity of the hypertension in rodent models appears to depend on sex, with males having higher risk.⁴³ The molecular mechanisms are incompletely understood. In the rat, the intrarenal renin–angiotensin system appears to be critical for normal nephrogenesis and may be altered by maternal dietary imbalance, both during the neonatal stage⁵⁵ and at later time points.⁵⁶ Other studies have implicated reduced activity of the antiapoptotic homeobox gene product paired box 2 (Pax-2) in reduced number of nephrons^{57,58} or have suggested that hypertension in later life caused by maternal dietary imbalance results from up-regulated sodium transport in the distal nephron, possibly triggered by increased oxidative stress.⁵⁹

Nutritional stress in pregnant rats reduces the growth of the endocrine pancreas during organogenesis and increases beta-cell apoptosis,⁶⁰ leading to hyperglycemia and impaired insulin secretion when the offspring become adults. Glucocorticoids may be involved in inducing phenotypic changes and have been shown to inhibit the transcription factor pancreatic and duodenal homeobox 1 (Pdx-1) in beta-cell precursors, which may affect the resultant number of beta cells.⁶¹ In the adult male rat offspring of mothers on a protein-restricted diet, low birth weight is associated with reduced expression of components of the

insulin signal-transduction pathway in skeletal muscle (including the protein kinase C zeta isoform, the p85 regulatory subunit of phosphoinositide-3 kinase, and the insulin-sensitive glucose transporter type 4 [GLUT4]).⁶² Similar abnormalities have been reported in infants of low birth weight,⁶² and together with the developmentally induced reduction in skeletal muscle mass,³ these abnormalities might contribute to later insulin resistance.

In the rat model of nutritional imbalance, the offspring of rats fed an imbalanced diet during pregnancy later had elevated blood pressure, reduced nephron number, and increased responses to salt loading⁵⁵ as well as reduced vasodilator function in the systemic arteries.⁴⁰ Rat pups subjected to hypoxic conditions during gestation appear to have fewer but larger cardiomyocytes than pups exposed to normal oxygen levels and are more susceptible to infarction during periods of ischemia and reperfusion as adults.⁶³ Increased blood pressure in fetal sheep stimulates cardiomyocytes to leave the cell cycle prematurely and hypertrophy,⁶⁴ which may affect cardiac function in adult life. Cardiac hypertrophy is also evident in lambs born to ewes undernourished during early gestation.⁶⁵ Chronic fetal anemia alters the developing coronary vascular tree in the near-term sheep fetus, and the remodeled coronary tree persists into adulthood.⁶⁶ In one study, carotid intima-media thickness at 9 years of age in 216 children of European ancestry whose mothers had energy intake in the lowest quartile during early or late pregnancy was higher than that of children whose mothers had intake in the highest quartile, a finding that implies that maternal nutrition within an unexceptional range during pregnancy can affect the subsequent risk of atherogenesis in the offspring.67

EPIGENETIC MECHANISMS

There is growing evidence that epigenetic mechanisms are responsible for tissue-specific gene expression during differentiation and that these mechanisms underlie the processes of developmental plasticity. Examples of epigenetic mechanisms include coordinated changes in the methylation of cytidine–guanosine (CpG) nucleotides in the promoter regions of specific genes, changes in chromatin structure through histone acetylation and methylation, and post-transcriptional control by microRNA (Fig. 2).⁶⁸ Epigenetic modifications are gene-specific and cell-type–specific, and since only a small set of enzymes is involved in making these modifications, it is likely that this specificity is directed by interactions between DNA and small RNA molecules. Widespread epigenetic reprogramming occurs after fertilization to ensure totipotency of the developing embryo, although methylation patterns associated with imprinting are maintained.⁶⁹ Developmentally induced epigenetic modifications of DNA are generally stable during the mitotic cell divisions that continue throughout a lifetime.

Challenges during pregnancy or early neonatal life in experimental models of programming result in changes in promoter methylation and thus directly or indirectly affect gene expression in pathways associated with a range of physiologic processes. For example, in the rat, altered promoter methylation and gene expression have been shown for the hepatic glucocorticoid receptor and the peroxisome proliferator-activated receptor a (PPAR-a),^{49,70} influencing carbohydrate and lipid metabolism (Fig. 3).⁷¹ Similar epigenetic changes have been observed in p53 in the kidney⁷² and the angiotensin II type 1b receptor in the adrenal gland,⁷³ influencing renal apoptosis and pressor responses, respectively, and in the hypothalamic glucocorticoid receptor,⁴² influencing stress responses. The phenotypic effects of epigenetic modifications during development may not manifest until later in life, especially if they affect genes modulating responses to later environmental challenges, such as a high-fat diet. The extent of the developmental window for the induction of epigenetic change in key systems is not known, but it appears to extend from the periconceptional period³⁵ into postnatal life.⁴² There is also evidence from studies in twins for changes in the

human epigenome related to age and the environment.⁷⁴ Many of the genes affected by epigenetic change do not appear to be classically imprinted (expressed according to the parental origin of the allele), although some imprinted genes may show altered expression after perturbations during early development, such as if blastocyst culture in vitro is prolonged.⁷⁵

The effects of maternal nutrition and behavior clearly target the promoter regions of specific genes rather than being associated with global changes in DNA methylation. For example, in a rat model, protein undernutrition in mothers affects the expression and promoter methylation of PPAR- α in the liver but does not affect methylation of the related transcription factor PPAR- γ (a regulator of adipogenesis).⁴⁹ Additional studies of this model indicate that altered promoter methylation appears to result from reduced capacity of the specific DNA methyltransferase that maintains methylation patterns through cell division⁷⁰ and that the changes in gene expression and promoter methylation can be transmitted to the F₂ generation without further nutritional challenge to the F₁ generation.⁷⁶

REVERSIBILITY

Recent laboratory studies have explored the reversibility of induced phenotypic effects and whether aberrant phenotypes induced in utero or during early development can be rescued. Corrective effects on phenotypic changes, gene expression, and associated methylation changes in PPAR-*a* have been reported after exogenous leptin administration to the neonatal offspring of undernourished rats (Fig. 4).^{77,78} Other studies suggest that hyperleptinemia and hypertension may be reversed by dietary intervention with n–3 fatty acids⁷⁹ and that altered behavioral responses can be reversed by pharmacologic manipulation of epigenetic status.⁸⁰ Research exploring methods of restoring aberrant phenotypes to normal has led to promising speculation that, ultimately, susceptible people might be identified by means of screening for epigenetic markers during early life and that customized interventions might then be instituted.

DEVELOPMENTAL PLASTICITY AND LATER DISEASE

Responses to environmental cues during early human development appear to initiate a range of overlapping effects that are induced according to the nature, size, and timing of those cues.^{1,81} One example is pathologic disruption (teratogenesis) by toxins or by extreme conditions such as poorly controlled maternal diabetes, which ultimately leads to cardiac abnormalities.⁸² Another type of example is a nondisruptive yet substantial developmental challenge such as inadequate maternal nutrition, which can induce a range of phenotypes that have been called "thrifty,"⁵ which means that the response of the developing fetus is a defense against an immediate challenge. The defensive fetal response usually involves a reduction in somatic growth, which may be specific to an organ or tissue, such as diminished skeletal muscle mass and restricted numbers of nephrons and neurons. Once such a challenged fetus has been born, it has to cope with the consequences of altered body composition, often through tradeoffs affecting other functions such as ultimate adult size or the timing of reproductive function.

An environmental cue that does not require an immediate defensive response, such as mild nutritional stress not substantially affecting birth weight, may nevertheless cause the developing organism to make phenotypic modifications that have a later fitness advantage in terms of tuning its physiology to better match aspects of the predicted adult environment. Such modifications have been termed predictive adaptive responses,⁸³ and striking examples have been reported in humans (e.g., the early-life establishment of patterns of thermoregulation⁸⁴) and other species. The adaptive advantage of such responses is determined by the probability that the choices made in early development are appropriate for

the environment that the organism will experience during its maturation and reproductive life.⁸¹ If the prediction is accurate, then the organism is matched to its subsequent environment and will cope adequately, ensuring positive selection for the mechanisms mediating such predictive responses. One example is a poor intrauterine environment inducing the reduced development of skeletal muscle and increased visceral fat deposition, a pattern that favors survival in a poor postnatal environment. This pattern has been observed in some South Asian babies, such as those in India.⁸⁵ But if the developmental choices made are not appropriate for the subsequent environment, the person may be more susceptible to later disease. For instance, sarcopenia and visceral obesity in a nutritionally rich postnatal environment that favors overconsumption are likely to promote further obesity, insulin resistance, and development of the metabolic syndrome (Fig. 5). The same "match–mismatch" theory can be applied to other systems, such as those affecting fluid balance⁸⁶ and the timing of puberty.⁸⁷

HERITABLE ENVIRONMENTAL INFLUENCES

The developmental cue is not limited to the nutritional environment during the period of gestation; rather, the information passed to the fetus or neonate from conception to weaning is a summation of maternal nutritional experience, integrating a lifetime of signals from the mother and perhaps even the grandmother.⁸⁸⁻⁹⁰ Such intergenerational transfer of environmental information may confer an adaptive advantage, even if the environment changes between generations, as shown in modeling studies.⁹¹ For example, in rat models, exposure during pregnancy to glucocorticoids⁹² or a low-protein diet⁷⁶ results in altered expression of liver enzymes, elevated blood pressure, and endothelial dysfunction in the F_1 generation. These changes can be transmitted to the F₂ generation without further challenge to members of the F1 generation during their lives. Limited clinical data are concordant with these experimental observations: epidemiologic studies have linked grandpaternal nutrition in one generation to the risk of diabetes in the F_2 generation.⁹³ The mechanism of intergenerational transfer is not clear, although it is known that postfertilization erasure of epigenetic marks such as DNA methylation and histone modification is incomplete for imprinted genes and similar processes may operate for some nonimprinted genes.⁹⁴ In addition, inheritance mediated by microRNA in the gametes, as recently shown in the mouse,⁹⁵ may act by altering post-transcriptional processing of factors affecting early embryonic development. Epigenetic changes induced in developing oocytes in the F1 fetus would be lost after the F₂ generation, as shown experimentally.⁹² Environmental influences during the F₀ pregnancy could also be transmitted nonepigenetically through the pregnancies of F_1 female offspring. These effects might involve the size⁹⁶ or vascular responses⁹⁷ of the reproductive tract, maternal behavior,⁹⁸ or body composition.⁹⁹ These considerations raise the possibility that familial clusters of metabolic disease may have an environmental and epigenetic basis, rather than a purely multigenic basis. In humans, there is a considerable contribution from familial and learned behaviors, such as eating patterns.¹⁰⁰

MEDICAL AND PUBLIC HEALTH IMPLICATIONS

Observational and experimental evidence increasingly supports a relation between growth and development during fetal and infant life and health in later years. This relation has two major implications. First, it reinforces the growing awareness that investment in the health and education of young people in relation to their responsibilities during pregnancy and parenthood is of fundamental importance. Second, any rational approach to health care should embrace a life-course perspective. These considerations have been recognized by the World Health Organization in their consultations on diet, nutrition, and chronic disease¹⁰¹ and on promoting optimal fetal development.¹⁰² Thus, the outcome of a pregnancy must be

considered in terms of maternal and neonatal health, the growth and cognitive development of the infant, its health as an adult, and even the health of subsequent generations.

Even in a developed nation, an imprudent diet before or during pregnancy may be common.¹⁰³ Interventions could involve correction of micronutrient and macronutrient imbalances in the mother before conception or at critical periods of early development⁸⁹ or, more broadly, could involve aspects of social structure, education, health information, nutrition, and behavior modification both before and after birth. Such complex interventions require novel thinking about trial design in a socially and culturally appropriate context.

CONCLUSIONS

The high incidence of metabolic disease in modern populations has been explained by selection for thrifty metabolism during evolution in an uncertain nutritional environment,¹⁰⁴ yet anthropologic evidence suggests that nutrition was not a primary challenge for preagricultural humans.¹⁰⁵ Molecular epidemiology has, to date, failed to define strong genetic determinants of the risk of developing metabolic disease.¹⁰⁶ Perhaps epigenetics will provide some explanations of how subtle early-life influences can produce longterm functional and structural changes. Furthermore, the concept of developmental plasticity could contribute an adaptive model that includes the effects of environmental factors during early development.^{5,81}

Human demographics are changing, with smaller families and older mothers as well as more teenage pregnancies; these demographic changes are concurrent with dramatic shifts in nutritional and workload environments of many populations. Against this background, it is essential to learn how influences on early development will interact with the physiologic processes of developmental plasticity to determine patterns of noncommunicable chronic disease.

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References

- 1. Bateson P, Barker D, Clutton-Brock T, et al. Developmental plasticity and human health. Nature. 2004; 430:419–21. [PubMed: 15269759]
- Barker DJ, Osmond C. Infant mortality, childhood nutrition, and ischaemic heart disease in England and Wales. Lancet. 1986; 1:1077–81. [PubMed: 2871345]
- Kensara OA, Wootton SA, Phillips DI, et al. Fetal programming of body composition: relation between birth weight and body composition measured with dual-energy X-ray absorptiometry and anthropometric methods in older Englishmen. Am J Clin Nutr. 2005; 82:980–7. [PubMed: 16280428]
- 4. Osmond C, Barker DJP, Winter PD, Fall CHD, Simmonds SJ. Early growth and death from cardiovascular disease in women. BMJ. 1993; 307:1519–24. [PubMed: 8274920]
- 5. Hales CN, Barker DJ. Type 2 (non-insulin-dependent) diabetes mellitus: the thrifty phenotype hypothesis. Diabetologia. 1992; 35:595–601. [PubMed: 1644236]
- Cooper C, Fall C, Egger P, Hobbs R, Eastell R, Barker D. Growth in infancy and bone mass in later life. Ann Rheum Dis. 1997; 56:17–21. [PubMed: 9059135]

- Rich-Edwards JW, Stampfer MJ, Manson JE, et al. Birth weight and risk of cardiovascular disease in a cohort of women followed up since 1976. BMJ. 1997; 315:396–400. [PubMed: 9277603]
- 8. Frankel S, Elwood P, Sweetnam P, Yarnell J, Smith GD. Birthweight, body-mass index in middle age, and incident coronary heart disease. Lancet. 1996; 348:1478–80. [PubMed: 8942776]
- Barker DJP, Forsén T, Uutela A, Osmond C, Eriksson JG. Size at birth and resilience to effects of poor living conditions in adult life: longitudinal study. BMJ. 2001; 323:1273–6. [PubMed: 11731388]
- Bhargava SK, Sachdev HS, Fall CHD, et al. Relation of serial changes in childhood body-mass index to impaired glucose tolerance in young adulthood. N Engl J Med. 2004; 350:865–75. [PubMed: 14985484]
- Barker DJP, Osmond C, Forsén TJ, Kajantie E, Eriksson JG. Trajectories of growth among children who have coronary events as adults. N Engl J Med. 2005; 353:1802–9. [PubMed: 16251536]
- Barker DJP. In utero programming of chronic disease. Clin Sci (Lond). 1998; 95:115–28. [PubMed: 9680492]
- Hofman PL, Regan F, Jackson WE, et al. Premature birth and later insulin resistance. N Engl J Med. 2004; 351:2179–86. [Erratum, N Engl J Med 2004;351:2888.]. [PubMed: 15548778]
- 14. Hovi P, Andersson S, Eriksson JG, et al. Glucose regulation in young adults with very low birth weight. N Engl J Med. 2007; 356:2053–63. [PubMed: 17507704]
- Gluckman PD, Hanson MA. Maternal constraint of fetal growth and its consequences. Semin Fetal Neonatal Med. 2004; 9:419–25. [PubMed: 15691778]
- Trevathan W. Fetal emergence patterns in evolutionary perspective. Am Anthropol. 1988; 90:674– 81.
- Charalambous M, da Rocha ST, Ferguson-Smith AC. Genomic imprinting, growth control and the allocation of nutritional resources: consequences for postnatal life. Curr Opin Endocrinol Diabetes Obes. 2007; 14:3–12. [PubMed: 17940412]
- Godfrey K, Robinson S, Barker DJ, Osmond C, Cox V. Maternal nutrition in early and late pregnancy in relation to placental and fetal growth. BMJ. 1996; 312:410–4. [PubMed: 8601112]
- Gluckman PD, Hanson MA. Developmental origins of disease paradigm: a mechanistic and evolutionary perspective. Pediatr Res. 2004; 56:311–7. [PubMed: 15240866]
- Whincup PH, Gilg JA, Papacosta O, et al. Early evidence of ethnic differences in cardiovascular risk: cross sectional comparison of British South Asian and white children. BMJ. 2002; 324:635. [PubMed: 11895820]
- Kirkwood TBL, Shanley DP. Food restriction, evolution and ageing. Mech Ageing Dev. 2005; 126:1011–6. [PubMed: 15893805]
- Hattersley AT, Tooke JE. The fetal insulin hypothesis: an alternative explanation of the association of low birthweight with diabetes and vascular disease. Lancet. 1999; 353:1789–92. [PubMed: 10348008]
- Javaid MK, Cooper C. Prenatal and childhood influences on osteoporosis. Best Pract Res Clin Endocrinol Metab. 2002; 16:349–67. [PubMed: 12064897]
- 24. Dennison EM, Arden NK, Keen RW, et al. Birthweight, vitamin D receptor genotype and the programming of osteoporosis. Paediatr Perinat Epidemiol. 2001; 15:211–9. [PubMed: 11489147]
- 25. Antoniades L, MacGregor AJ, Andrew T, Spector TD. Association of birth weight with osteoporosis and osteoarthritis in adult twins. Rheumatology (Oxford). 2003; 42:791–6. [PubMed: 12730541]
- Boney CM, Verma A, Tucker R, Vohr BR. Metabolic syndrome in childhood: association with birth weight, maternal obesity, and gestational diabetes mellitus. Pediatrics. 2005; 115(3):e290– e296. [PubMed: 15741354]
- Hillier TA, Pedula KL, Schmidt MM, Mullen JA, Charles MA, Pettitt DJ. Childhood obesity and metabolic imprinting: the ongoing effects of maternal hyperglycemia. Diabetes Care. 2007; 30:2287–92. [PubMed: 17519427]
- Harder T, Bergmann R, Kallischnigg G, Plagemann A. Duration of breastfeeding and risk of overweight: a meta-analysis. Am J Epidemiol. 2005; 162:397–403. [PubMed: 16076830]

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- 29. dos Santos Silva I, De Stavola BL, Hardy RJ, Kuh DJ, McCormack VA, Wadsworth ME. Is the association of birth weight with premenopausal breast cancer risk mediated through childhood growth? Br J Cancer. 2004; 91:519–24. [PubMed: 15266328]
- Vickers MH, Breier BH, Cutfield WS, Hofman PL, Gluckman PD. Fetal origins of hyperphagia, obesity, and hypertension and postnatal amplification by hypercaloric nutrition. Am J Physiol Endocrinol Metab. 2000; 279:E83–E87. [PubMed: 10893326]
- Langley-Evans SC, Welham SJ, Jackson AA. Fetal exposure to a maternal low protein diet impairs nephrogenesis and promotes hypertension in the rat. Life Sci. 1999; 64:965–74. [PubMed: 10201645]
- Seckl JR, Meaney MJ. Glucocorticoid programming. Ann N Y Acad Sci. 2004; 1032:63–84. [PubMed: 15677396]
- Kwong WY, Wild AE, Roberts P, Willis AC, Fleming TP. Maternal undernutrition during the preimplantation period of rat development causes blastocyst abnormalities and programming of postnatal hypertension. Development. 2000; 127:4195–202. [PubMed: 10976051]
- Watkins AJ, Platt D, Papenbrock T, et al. Mouse embryo culture induces changes in postnatal phenotype including raised systolic blood pressure. Proc Natl Acad Sci U S A. 2007; 104:5449– 54. [PubMed: 17372207]
- Sinclair KD, Allegrucci C, Singh R, et al. DNA methylation, insulin resistance, and blood pressure in offspring determined by maternal periconceptional B vitamin and methionine status. Proc Natl Acad Sci U S A. 2007; 104:19351–6. [PubMed: 18042717]
- Yajnik CS, Deshpande SS, Jackson AA, et al. Vitamin B₁₂ and folate concentrations during pregnancy and insulin resistance in the offspring: the Pune Maternal Nutrition Study. Diabetologia. 2008; 51:29–38. [PubMed: 17851649]
- Levitt NS, Lindsay RS, Holmes MC, Seckl JR. Dexamethasone in the last week of pregnancy attenuates hippocampal glucocorticoid receptor gene expression and elevates blood pressure in the adult offspring in the rat. Neuroendocrinology. 1996; 64:412–8. [PubMed: 8990073]
- Nyirenda MJ, Lindsay RS, Kenyon CJ, Burchell A, Seckl JR. Glucocorticoid exposure in late gestation permanently programs rat hepatic phosphoenolpyruvate carboxykinase and glucocorticoid receptor expression and causes glucose intolerance in adult offspring. J Clin Invest. 1998; 101:2174–81. [PubMed: 9593773]
- Welberg LAM, Seckl JR, Holmes MC. Prenatal glucocorticoid programming of brain corticosteroid receptors and corticotrophin-releasing hormone: possible implications for behaviour. Neuroscience. 2001; 104:71–9. [PubMed: 11311532]
- 40. Brawley L, Itoh S, Torrens C, et al. Dietary protein restriction in pregnancy induces hypertension and vascular defects in rat male offspring. Pediatr Res. 2003; 54:83–90. [PubMed: 12646717]
- 41. Vickers MH, Breier BH, McCarthy D, Gluckman P. Sedentary behavior during postnatal life is determined by the prenatal environment and exacerbated by postnatal hypercaloric nutrition. Am J Physiol Regul Integr Comp Physiol. 2003; 285:R271–R273. [PubMed: 12794001]
- 42. Weaver ICG, Cervoni N, Champagne FA, et al. Epigenetic programming by maternal behavior. Nat Neurosci. 2004; 7:847–54. [PubMed: 15220929]
- Woods LL, Ingelfinger JR, Rasch R. Modest protein restriction fails to program adult hypertension in female rats. Am J Physiol Regul Integr Comp Physiol. 2005; 289:R1131–R1136. [PubMed: 15961538]
- Musha Y, Itoh S, Hanson MA, Kinoshita K. Does estrogen affect the development of abnormal vascular function in offspring of rats fed a low-protein diet in pregnancy? Pediatr Res. 2006; 59:784–9. [PubMed: 16641213]
- 45. Langley-Evans SC. Critical differences between two low protein diet protocols in the programming of hypertension in the rat. Int J Food Sci Nutr. 2000; 51:11–7. [PubMed: 10746100]
- 46. Fernandez-Twinn DS, Ekizoglou S, Wayman A, Petry CJ, Ozanne SE. Maternal low-protein diet programs cardiac beta-adrenergic response and signaling in 3-mo-old male offspring. Am J Physiol Regul Integr Comp Physiol. 2006; 291:R429–R436. [PubMed: 16914429]
- Hoppe CC, Evans RG, Moritz KM, et al. Combined prenatal and postnatal protein restriction influences adult kidney structure, function, and arterial pressure. Am J Physiol Regul Integr Comp Physiol. 2007; 292:R462–R469. [PubMed: 16973940]

- Kapoor A, Dunn E, Kostaki A, Andrews MH, Matthews SG. Fetal programming of the hypothalamo-pituitary-adrenal function: prenatal stress and glucocorticoids. J Physiol. 2006; 572:31–44. [PubMed: 16469780]
- 49. Lillycrop KA, Phillips ES, Jackson AA, Hanson MA, Burdge GC. Dietary protein restriction of pregnant rats induces and folic acid supplementation prevents epigenetic modification of hepatic gene expression in the offspring. J Nutr. 2005; 135:1382–6. [PubMed: 15930441]
- Bloomfield FH, Oliver MH, Giannoulias CD, Gluckman PD, Harding JE, Challis JRG. Brief undernutrition in late-gestation sheep programs the hypothalamic-pituitary-adrenal axis in adult offspring. Endocrinology. 2003; 144:2933–40. [PubMed: 12810548]
- Jaquiery AL, Oliver MH, Bloomfield FH, Connor KL, Challis JR, Harding JE. Fetal exposure to excess glucocorticoid is unlikely to explain the effects of periconceptional undernutrition in sheep. J Physiol. 2006; 572:109–18. [PubMed: 16469778]
- McMillen IC, Robinson JS. Developmental origins of the metabolic syndrome: prediction, plasticity, and programming. Physiol Rev. 2005; 85:571–633. [PubMed: 15788706]
- Zandi-Nejad K, Luyckx V, Brenner BM. Adult hypertension and kidney disease: the role of fetal programming. Hypertension. 2006; 47:502–8. [PubMed: 16415374]
- 54. Aihie-Sayer A, Dunn R, Langley-Evans S, Cooper C. Prenatal exposure to a maternal low protein diet shortens life span in rats. Gerontology. 2001; 47:9–14. [PubMed: 11244286]
- Woods LL, Ingelfinger JR, Nyengaard JR, Rasch R. Maternal protein restriction suppresses the newborn renin-angiotensin system and programs adult hypertension in rats. Pediatr Res. 2001; 49:460–7. [PubMed: 11264427]
- Vehaskari VM, Aviles DH, Manning J. Prenatal programming of adult hypertension in the rat. Kidney Int. 2001; 59:238–45. [PubMed: 11135076]
- 57. Zhang SL, Chen YW, Tran S, et al. Pax-2 and N-myc regulate epithelial cell proliferation and apoptosis in a positive autocrine feedback loop. Pediatr Nephrol. 2007; 22:813–24. [PubMed: 17357786]
- Quinlan J, Lemire M, Hudson T, et al. A common variant of the PAX2 gene is associated with reduced newborn kidney size. J Am Soc Nephrol. 2007; 18:1915–21. [PubMed: 17513325]
- Vehaskari VM. Developmental origins of adult hypertension: new insights into the role of the kidney. Pediatr Nephrol. 2007; 22:490–5. [PubMed: 17115193]
- 60. Reusens B, Remacle C. Programming of the endocrine pancreas by the early nutritional environment. Int J Biochem Cell Biol. 2006; 38:913–22. [PubMed: 16337425]
- Breant B, Gesina E, Blondeau B. Nutrition, glucocorticoids and pancreas development. Horm Res. 2006; 65(Suppl 3):98–104. [Erratum, Horm Res 2006;65:294.]. [PubMed: 16612121]
- Ozanne SE, Jensen CB, Tingey KJ, Storgaard H, Madsbad S, Vaag AA. Low birthweight is associated with specific changes in muscle insulin-signalling protein expression. Diabetologia. 2005; 48:547–52. [PubMed: 15729577]
- 63. Li G, Xiao Y, Estrella JL, Ducsay CA, Gilbert RD, Zhang L. Effect of fetal hypoxia on heart susceptibility to ischemia and reperfusion injury in the adult rat. J Soc Gynecol Investig. 2003; 10:265–74.
- Barbera A, Giraud GD, Reller MD, Maylie J, Morton MJ, Thornburg KL. Right ventricular systolic pressure load alters myocyte maturation in fetal sheep. Am J Physiol Regul Integr Comp Physiol. 2000; 279:R1157–R1164. [PubMed: 11003978]
- Cleal JK, Poore KR, Boullin JP, et al. Mismatched pre- and postnatal nutrition leads to cardiovascular dysfunction and altered renal function in adulthood. Proc Natl Acad Sci U S A. 2007; 104:9529–33. [PubMed: 17483483]
- 66. Davis L, Thornburg KL, Giraud GD. The effects of anaemia as a programming agent in the fetal heart. J Physiol. 2005; 565:35–41. [PubMed: 15760943]
- Gale CR, Jiang B, Robinson SM, Godfrey KM, Law CM, Martyn CN. Maternal diet during pregnancy and carotid intimamedia thickness in children. Arterioscler Thromb Vasc Biol. 2006; 26:1877–82. [PubMed: 16728657]
- Goldberg AD, Allis CD, Bernstein E. Epigenetics: a landscape takes shape. Cell. 2007; 128:635–8. [PubMed: 17320500]

- 69. Morgan HD, Santos F, Green K, Dean W, Reik W. Epigenetic reprogramming in mammals. Hum Mol Genet. 2005; 14:R47–R58. [PubMed: 15809273]
- 70. Lillycrop KA, Slater-Jefferies JL, Hanson MA, Godfrey KM, Jackson AA, Burdge GC. Induction of altered epigenetic regulation of the hepatic glucocorticoid receptor in the offspring of rats fed a protein-restricted diet during pregnancy suggests that reduced DNA methyltransferase-1 expression is involved in impaired DNA methylation and changes in histone modifications. Br J Nutr. 2007; 97:1064–73. [PubMed: 17433129]
- 71. Burdge GC, Lillycrop KA, Jackson AA, Gluckman PD, Hanson MA. The nature of the growth pattern and of the metabolic response to fasting in the rat are dependent upon the dietary protein and folic acid intakes of their pregnant dams and post-weaning fat consumption. Br J Nutr. in press.
- Pham TD, MacLennan NK, Chiu CT, Laksana GS, Hsu JL, Lane RH. Uteroplacental insufficiency increases apoptosis and alters p53 gene methylation in the full-term IUGR rat kidney. Am J Physiol Regul Integr Comp Physiol. 2003; 285:R962–R970. [PubMed: 12869365]
- Bogdarina I, Welham S, King PJ, Burns SP, Clark AJ. Epigenetic modification of the reninangiotensin system in the fetal programming of hypertension. Circ Res. 2007; 100:520–6. [PubMed: 17255528]
- 74. Fraga MF, Ballestar E, Paz MF, et al. Epigenetic differences arise during the lifetime of monozygotic twins. Proc Natl Acad Sci U S A. 2005; 102:10604–9. [PubMed: 16009939]
- Doherty AS, Mann MR, Tremblay KD, Bartolomei MS, Schultz RM. Differential effects of culture on imprinted H19 expression in the preimplantation mouse embryo. Biol Reprod. 2000; 62:1526– 35. [PubMed: 10819752]
- 76. Burdge GC, Slater-Jefferies JL, Torrens C, Phillips ES, Hanson MA, Lillycrop KA. Dietary protein restriction of pregnant rats in the F0 generation induces altered methylation of hepatic gene promoters in the adult male offspring in the F1 and F2 generations. Br J Nutr. 2007; 97:435–9. [PubMed: 17313703]
- 77. Vickers MH, Gluckman PD, Coveny AH, et al. Neonatal leptin treatment reverses developmental programming. Endocrinology. 2005; 146:4211–6. [PubMed: 16020474]
- Gluckman PD, Lillycrop KA, Vickers MH, et al. Metabolic plasticity during mammalian development is directionally dependent on early nutritional status. Proc Natl Acad Sci U S A. 2007; 104:12796–800. [PubMed: 17646663]
- 79. Wyrwoll CS, Mark PJ, Mori TA, Puddey IB, Waddell BJ. Prevention of programmed hyperleptinemia and hypertension by postnatal dietary ω-3 fatty acids. Endocrinology. 2006; 147:599–606. [PubMed: 16210371]
- Weaver IC, Meaney MJ, Szyf M. Maternal care effects on the hippocampal transcriptome and anxiety-mediated behaviors in the offspring that are reversible in adulthood. Proc Natl Acad Sci U S A. 2006; 103:3480–5. [PubMed: 16484373]
- Gluckman PD, Hanson MA, Spencer HG, Bateson P. Environmental influences during development and their later consequences for health and disease: implications for the interpretation of empirical studies. Proc Biol Sci. 2005; 272:671–7. [PubMed: 15870029]
- Aberg A, Westbom L, Källén B. Congenital malformations among infants whose mothers had gestational diabetes or preexisting diabetes. Early Hum Dev. 2001; 61:85–95. [PubMed: 11223271]
- Gluckman PD, Hanson MA, Spencer HG. Predictive adaptive responses and human evolution. Trends Ecol Evol. 2005; 20:527–33. [PubMed: 16701430]
- 84. Diamond J. Pearl Harbor and the Emperor's physiologists. Nat Hist. 1991; 12:2–7. [PubMed: 11612598]
- Yajnik CS, Fall CHD, Coyaji KJ, et al. Neonatal anthropometry: the thin-fat Indian baby: the Pune Maternal Nutrition Study. Int J Obes Relat Metab Disord. 2003; 27:173–80. [PubMed: 12586996]
- 86. Desai M, Ladella S, Ross MG. Reversal of pregnancy-mediated plasma hypotonicity in the nearterm rat. J Matern Fetal Neonatal Med. 2003; 13:197–202. [PubMed: 12820842]
- Gluckman PD, Hanson MA. Evolution, development and timing of puberty. Trends Endocrinol Metab. 2006; 17:7–12. [PubMed: 16311040]

- Kuzawa CW. Fetal origins of developmental plasticity: are fetal cues reliable predictors of future nutritional environments? Am J Hum Biol. 2005; 17:5–21. [PubMed: 15611967]
- Morton, SMB. Maternal nutrition and fetal growth and development.. In: Gluckman, PD.; Hanson, MA., editors. Developmental origins of health and disease. Cambridge University Press; Cambridge, England: 2006. p. 98-129.
- Kuzawa, CW.; Gluckman, PD.; Hanson, MA.; Beedle, AS. Evolution, developmental plasticity, and metabolic disease.. In: Stearns, SC.; Koella, JC., editors. Evolution in health and disease. 2nd ed.. Oxford University Press; Oxford, England: 2008. p. 253-64.
- Jablonka E, Oborny B, Molnár I, Kisdi E, Hofbauer J, Czárán T. The adaptive advantage of phenotypic memory in changing environments. Philos Trans R Soc Lond B Biol Sci. 1995; 350:133–41. [PubMed: 8577857]
- 92. Drake AJ, Walker BR, Seckl JR. Intergenerational consequences of fetal programming by in utero exposure to glucocorticoids in rats. Am J Physiol Regul Integr Comp Physiol. 2005; 288:R34– R38. [PubMed: 15178540]
- Kaati G, Bygren LO, Edvinsson S. Cardiovascular and diabetes mortality determined by nutrition during parents' and grandparents' slow growth period. Eur J Hum Genet. 2002; 10:682–8. [PubMed: 12404098]
- 94. Chong S, Whitelaw E. Epigenetic germline inheritance. Curr Opin Genet Dev. 2004; 14:692–6. [PubMed: 15531166]
- Rassoulzadegan M, Grandjean V, Gounon P, Vincent S, Gillot I, Cuzin F. RNA-mediated nonmendelian inheritance of an epigenetic change in the mouse. Nature. 2006; 441:469–74. [PubMed: 16724059]
- 96. Ibáñez L, Potau N, Enriquez G, Marcos MV, de Zegher F. Hypergonadotrophinaemia with reduced uterine and ovarian size in women born small-for-gestational age. Hum Reprod. 2003; 18:1565–9. [PubMed: 12871863]
- Torrens C, Brawley L, Barker AC, Itoh S, Poston L, Hanson MA. Maternal protein restriction in the rat impairs resistance artery but not conduit artery function in pregnant offspring. J Physiol. 2003; 547:77–84. [PubMed: 12562942]
- Francis D, Diorio J, Liu D, Meaney MJ. Nongenomic transmission across generations of maternal behavior and stress responses in the rat. Science. 1999; 286:1155–8. [PubMed: 10550053]
- Aerts L, Van Assche FA. Animal evidence for the transgenerational development of diabetes mellitus. Int J Biochem Cell Biol. 2006; 38:894–903. [PubMed: 16118061]
- Brook JS, Whiteman M, Brook DW. Transmission of risk factors across three generations. Psychol Rep. 1999; 85:227–41. [PubMed: 10575989]
- 101. Diet, nutrition and the prevention of chronic diseases: report of a joint WHO/FAO expert consultation. World Health Organ Tech Rep Ser. 2003; 916:1–149.
- 102. Promoting optimal fetal development: report of a technical consultation. World Health Organization; Geneva: 2006.
- 103. Robinson SM, Crozier SR, Borland SE, Hammond J, Barker DJ, Inskip HM. Impact of educational attainment on the quality of young women's diets. Eur J Clin Nutr. 2004; 58:1174– 80. [PubMed: 15054431]
- 104. Neel JV. Diabetes mellitus: a "thrifty" genotype rendered detrimental by "progress"? Am J Hum Genet. 1962; 14:353–62. [PubMed: 13937884]
- 105. Benyshek DC, Watson JT. Exploring the thrifty genotype's food-shortage assumptions: a crosscultural comparison of ethnographic accounts of food security among foraging and agricultural societies. Am J Phys Anthropol. 2006; 131:120–6. [PubMed: 16485298]
- 106. Wellcome Trust Case Control Consortium. Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. Nature. 2007; 447:661–78. [PubMed: 17554300]

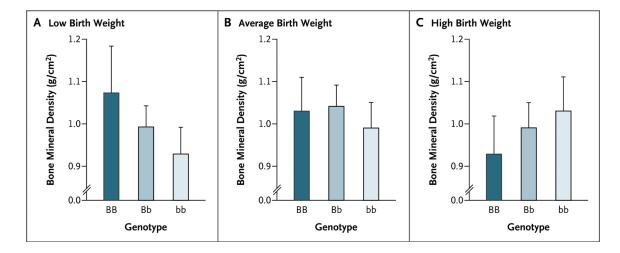


Figure 1. Birth Weight and the Relationship between Lumbar-Spine Bone Mineral Density and Vitamin D Receptor Genotype in Elderly Men and Women

Among persons in the lowest third of the birth-weight distribution, spine bone mineral density was significantly higher among persons of genotype BB than among persons of the Bb or bb genotype (P = 0.01) after adjustment for age, sex, and current weight. In contrast, in the highest third of the birth-weight distribution, spine bone mineral density was reduced among persons of genotype BB as compared with persons of the Bb or bb genotype (P = 0.04). A significant interaction was found between vitamin D receptor genotype and birth weight as determinants of bone mineral density (P = 0.02). Adapted from Dennison et al.²⁴

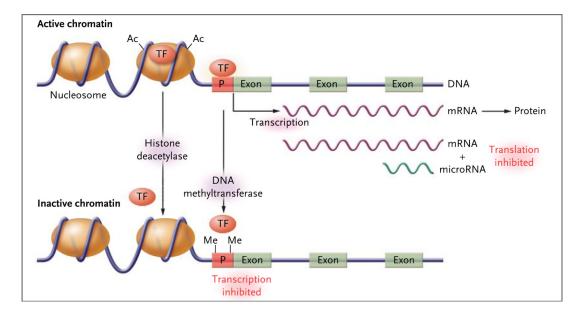


Figure 2. Regulation of Gene Expression through Epigenetic Processes

Epigenetic modification of histones or of DNA itself controls access of transcription factors (TFs) to the DNA sequence, thereby modulating the rate of transcription to messenger RNA (mRNA). Transcriptionally active chromatin (top) characterized by the presence of acetyl groups (Ac) on specific lysine residues of core histones in the nucleosome, which decreases their binding to DNA and results in a more open chromatin structure that permits access of transcription factors. In addition, cytidine–guanosine (CpG) sequences in the promoter regions (P) of actively transcribed genes are generally unmethylated, allowing for the binding of transcription factors. Transcriptionally inactive chromatin (bottom) is characterized by histone deacetylation, promoter CpG methylation (as indicated by methyl groups [Me]), and decreased binding of transcription factors. (For simplicity, other histone modifications [such as methylation] and additional regulatory factors [such as methyl-CpG binding proteins] are not shown.) A further level of epigenetic control is provided by microRNA molecules (19 to 22 nucleotides in length), which bind to complementary sequences in the 3' end of mRNA and reduce the rate of protein synthesis.

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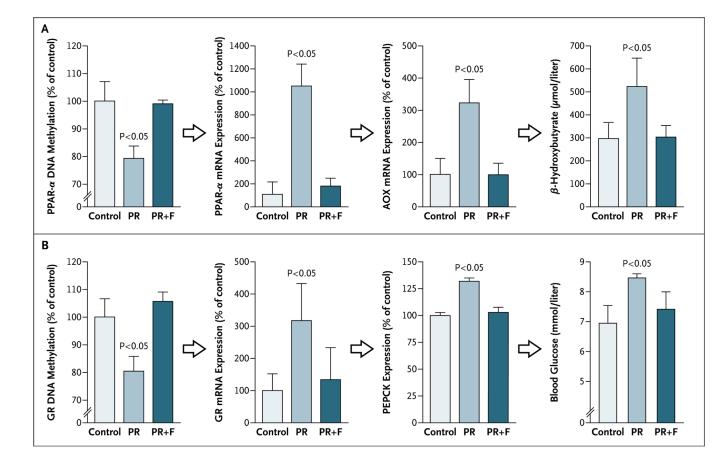


Figure 3. Metabolic Phenotype of Prenatally Undernourished Rats and Changes in Promoter Methylation and Gene Expression

The offspring of rats fed a protein-restricted diet (PR) show reduced promoter methylation and increased messenger RNA (mRNA) expression of the transcription factor peroxisome proliferator-activated receptor *a* (PPAR-*a*) in the liver, as compared with control rats fed a normal diet (Panel A). Increased PPAR-*a* expression is associated with increased expression of the downstream enzyme acyl-CoA oxidase (AOX), a key enzyme in fatty acid β oxidation, and increased circulating concentrations of the ketone β -hydroxybutyrate. These effects were prevented by supplementation of the maternal diet with folate (PR+F). The offspring of rats in the PR group show reduced promoter methylation and increased mRNA expression of the glucocorticoid receptor (GR) in the liver, associated with increased expression of the gluconeogenic enzyme phosphoenolpyruvate carboxykinase (PEPCK) and increased blood glucose levels (Panel B). Again, the effects are not seen in offspring of rats in the PR+F group. P values are shown for the comparison of the PR group with the control group; the P values for the comparison of the PR+F group with the control group indicated no significant difference. Data are means; T bars indicate SEs. Adapted from Lillycrop et al.^{49,70} and Burdge et al.⁷¹ Gluckman et al.

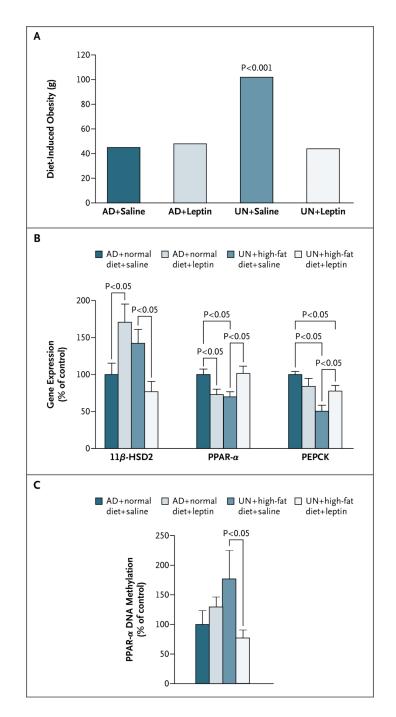


Figure 4. Effect of Neonatal Leptin Treatment on Metabolic Programming Caused by Maternal Undernutrition in the Rat

Female rats were subjected in utero to maternal undernutrition (UN) or ad libitum feeding (AD), treated with saline or leptin between days 3 and 13 of life, and fed a normal diet or a high-fat diet after having been weaned. Panel A shows the diet-induced obesity (defined as the difference in total body weight between rats fed a high-fat diet and those fed a normal diet) at 170 days of age. Neonatal leptin treatment prevented the increased susceptibility to diet-induced obesity associated with a high-fat diet after maternal undernutrition. The P value is for the comparison of the UN group with the other three groups. The expression of hepatic genes (for 11β -hydroxysteroid dehydrogenase type 2 [11β -HSD2], peroxisome

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proliferator-activated receptor *a* [PPAR-*a*], and phosphoenolpyruvate carboxykinase [PEPCK]) (Panel B) and promoter methylation of the PPAR-*a* gene (Panel C) are shown for female rats at 170 days of age. The data in Panels B and C are means, with T bars indicating SEs, for eight rats per group. The control groups in Panels B and C consisted of female offspring in the AD group, treated with saline and fed a normal diet after weaning. Adapted from Vickers et al.⁷⁷ and Gluckman et al.⁷⁸

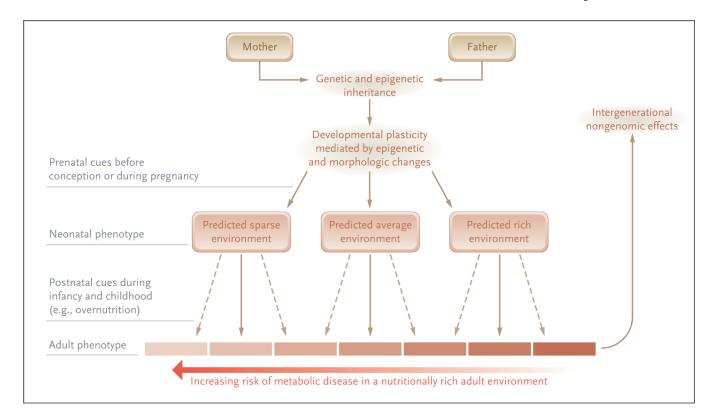


Figure 5. Environmental Cues during Development, Developmental Plasticity, and Determination of the Adult Phenotype

Prenatal cues predicting a nutritionally sparse environment will cause a shift in the trajectory of structural and functional development toward a phenotype matched to that environment. Such a phenotype will have a reduced capacity to cope with a nutritionally rich environment later in life, increasing the risk of metabolic disease. Postnatal cues, such as childhood overnutrition leading to compensatory growth, could further shift the positioning of the adult phenotype, exacerbating the mismatch (dashed lines) between phenotype and environment. Although there is a continuous range of possible developmental trajectories and multiple sequential cues that act during development, for simplicity only two developmental cues (before and after birth) and three trajectories are shown.