

## Original Article

# Expression and prognostic value of c-Myc and Fas (CD95/APO1) in patients with pancreatic cancer

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Received November 27, 2013; Accepted December 12, 2013; Epub January 15, 2014; Published February 1, 2014

**Abstract:** The identification of molecular prognostic markers for pancreatic cancer patients could provide insightful information for their management in the clinic. The aim of the study is to investigate whether or not the expressions of c-Myc and Fas (CD95/APO1) have prognostic relevance for overall survival (OS) in patients with pancreatic cancer. We used immunohistochemistry on tissue microarrays containing 162 pancreatic cancer specimens to assess the protein expression levels of c-Myc and Fas. Kaplan-Meier survival analysis demonstrated that high level of c-Myc cytoplasmic expression was significantly correlated with worse survival in patients with pancreatic cancer ( $P = 0.012$ ), while high level of Fas cytoplasmic expression was significantly associated with better outcome of pancreatic cancer ( $P = 0.046$ ). However, multivariate Cox model analysis showed that tumor differentiation, lymph node status and c-Myc cytoplasmic expression were significant independent prognostic factors for OS ( $P < 0.001$ ,  $P = 0.023$ ,  $P = 0.001$ , respectively). On the contrary, Fas cytoplasmic expression did not independently influence patient's prognosis ( $P = 0.249$ ). Our data suggested that high level of c-Myc cytoplasmic expression may be considered as a valuable marker for prognosis of pancreatic cancer.

**Keywords:** Pancreatic cancer, c-Myc, Fas (CD95/APO1), immunohistochemistry, survival analysis

## Introduction

Pancreatic cancer is a fourth or fifth leading cause of cancer-related death for both men and women in the western world [1, 2]. For all stages combined, the 5-year survival rates is < 5% if left untreated [3]. Complete surgical resection is the only chance for long-term survival, improving five-year relative survival rates to 25-35% after surgery [4, 5]. Unfortunately, because pancreatic cancer is often advanced at the time of diagnosis, only 15 to 20 out of every 100 diagnosed cases can be considered candidates for potentially curative resection [6]. Although there has been a remarkable improvement in the treatment of the disease, the prognosis in patients with pancreatic cancer remains extremely poor.

Identification of prognostic factors may provide useful information for clinical management.

The current prognosis prediction for pancreatic cancer very greatly depend on the TNM staging system (Tumor, Node, Metastasis). However, prognosis varies among patients with a similar tumor stage, therefore the TNM classification alone can not accurately predict the outcome for individual patients. Over recent years, researchers have extensively investigated the potential predictive molecular markers, and also have identified some molecular markers are significantly correlated with clinicopathological variables and survival rates of pancreatic cancer [7-9]. So far, however, no prognostic and predictive molecular marker in pancreatic cancer is recommended in clinical practice.

Previous studies have demonstrated that aberrant expressions of c-Myc and Fas (CD95/APO1) have been described in multiple malignancies and implicated in the pathogenesis of various malignancies, which include epithelial

## c-Myc and Fas (CD95/APO1) in patients with pancreatic cancer

**Table 1.** Characteristics features of the 162 patients with pancreatic cancer

Clinicopathologic features	Number	Percentage (%)
Age (years)		
< 60	89	54.9
≥ 60	73	45.1
Gender		
female	60	37
male	102	63
Size (cm)		
< 4	74	45.7
≥ 4	88	54.3
Location		
head	117	72.2
body and rear	45	27.8
Differentiation		
well	117	72.2
moderate	14	8.7
poor	31	19.1
Nodal status		
Negative	97	59.9
Positive	65	40.1
Perineural Invasion status		
Negative	79	48.8
Positive	83	51.2
Stage		
stage I	80	49.4
stage II	80	49.4
stage III	0	0
stage IV	2	1.2

malignancies as well as pancreatic cancer [10-12]. Many studies have shown a significant association between the over-expression of c-Myc and poor prognosis in numerous tumor types [13, 14], whereas some other reports do not find such a relationship [15, 16]. Moreover, a recent study has shown that higher levels of c-Myc mRNA in breast cancer are correlated with better survival [17]. Currently, the final prognostic value of the c-Myc expression with respect to overall survival in pancreatic cancer remains uncertain. In addition, only a few studies regarding the expression of Fas in human pancreatic cancer tissues have been reported, and the available data for prognostic value of Fas in pancreatic cancer is very limited. Recently, a previous study showed that the over-expression of c-Myc and low-expression of Fas were significantly correlated with the PNI of pancreatic cancer [18]. Consequently, we spec-

ulated that the expressions of c-Myc and Fas may have potential prognostic value in patients with pancreatic cancer. Therefore, in order to clarify our thought, we performed an immunohistochemical study on 162 pancreatic cancer using tissue microarrays technology. In addition, we also examined the association of c-Myc and Fas expression with each other and their association with patient clinicopathological characteristics.

### Materials and methods

#### *Patients and tissue samples*

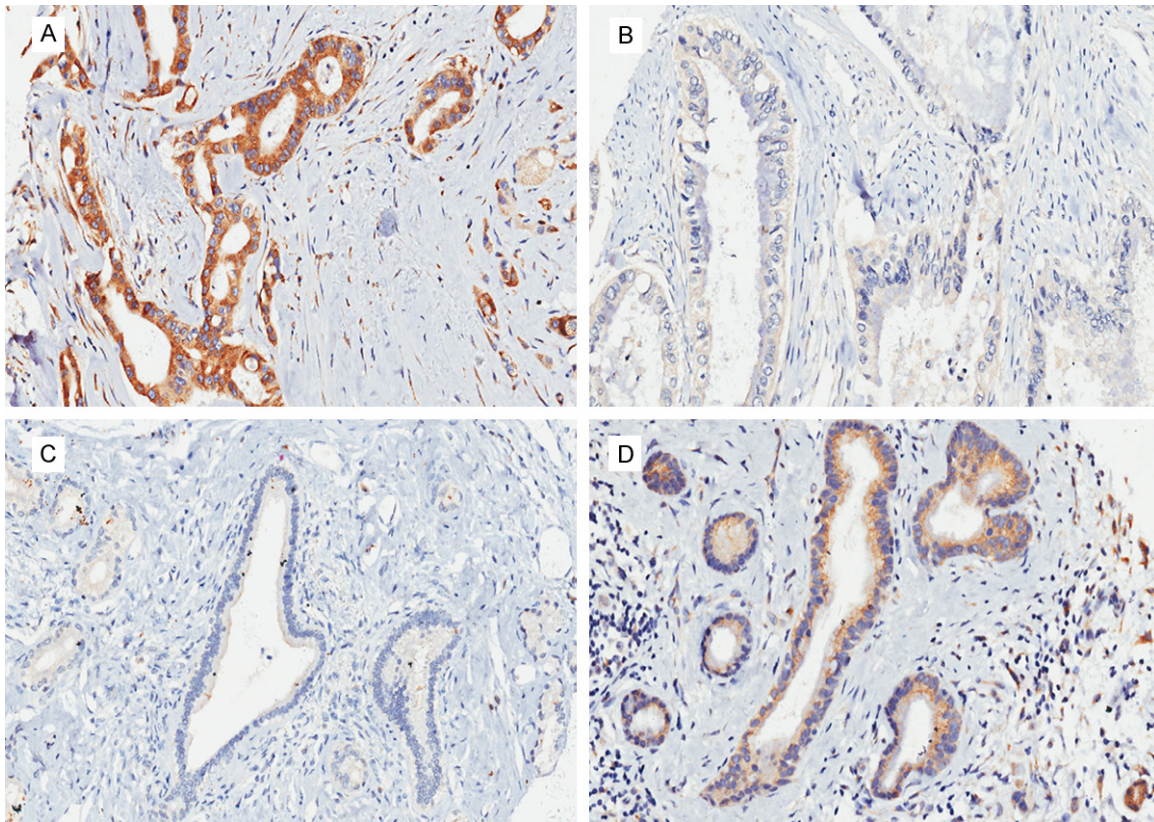
The paraffin-embedded samples from patients investigated in this study were collected retrospectively from archival material stored in the biobank center in National Engineering Center for Biochip at Shanghai. Samples included a total of 162 patients who had undergone a resection surgery between 1995 and 2009. Written informed consent for the tissue specimens was received from all participants, and the study was approved by the ethical committee of biobank center related hospitals.

The following clinicopathological data was obtained from original pathology reports, including age, gender, tumor size, location, and invasion, LN metastases, grade of differentiation, and tumor stage. Staging of pancreatic cancer were assessed according to the American Joint Committee on Cancer (AJCC) criteria. A detailed description of clinical and pathological data of these 162 patients is provided in **Table 1**.

All follow-up times was measured from the date of surgery to patient's death from pancreatic cancer. Unfortunately, at the time of last follow-up ended in December 2011, 92 patients were lost to follow-up and excluded from the 5-year survival analysis, only 70 patient samples were available for the survival analysis. Among remaining 70 patients, 46 patients died during the follow-up period.

#### *Tissue microarray construction*

Tissue microarrays (TMAs) were constructed using appropriate tissue cores from formalin-fixed and paraffin-embedded (FFPE) samples as previously described [19]. Briefly, appropriate tumor areas and its corresponding non-



**Figure 1.** Immunohistochemical staining of c-Myc and Fas in human pancreatic cancer tissues and paracancerous tissues. A: Showing the positive expression of c-Myc in cancer tissues. B: Showing the negative expression of Fas in cancer tissues. C: Showing the negative expression of c-Myc in paracancerous tissues. D: Showing the positive expression of Fas in paracancerous tissues. All images were taken at 100× magnification.

tumor pancreatic samples were selected by the pathologists, and then a single core with a diameter of 0.6 mm was taken from each case. TMA blocks were constructed using an automated tissue arrayer (Beecher Instruments, Sun Prairie, WI). The array blocks were cut into five-micron sections, and then Sections were stained with hematoxylin-eosin to verify the presence of tumor cells. In all cases, tissue cores obtained from normal adjacent pancreas were severed as internal controls.

#### *Immunohistochemistry and scoring*

The TMA sections were deparaffinized in xylene, rehydrated with graded ethanol, washed in Tris-buffered saline. Antigen retrieval was conducted at high temperature under high pressure in sodium citrate buffer (pH 6) for 10 minutes. After quenching of endogenous peroxidase activity, c-Myc polyclone antibody (Santa company, expression in cytoplasm and cytomembrane) and Fas monoclonal antibody (Abcam

company, expression in cytoplasm) was used at 1:300 dilution, respectively, and then specimens were incubated with the antibodies overnight at 4°C. Slides were then incubated with an appropriate dilution of the corresponding secondary biotinylated rabbit antibody for 30 minutes at room temperature. And then slides were washed 3 times in Tris-buffered saline and incubated in streptavidin-horseradish peroxidase (1:100, Dako) at room temperature for 30 minutes. Chromogenic immunolocalization was conducted using 3,3-diaminobenzidine (Dako, inc, CA, USA). Slides were then counterstained with hematoxylin. After washing, slides were dehydrated, and then mounted with coverslips. Other tissue cores containing pancreatic cancer tissues used as positive controls. The negative control consisted of normal serum substituted for primary antibody.

Each slide was scored semi-quantitatively on the basis of percentage and intensity of stained normal or neoplastic epithelial cells as



## c-Myc and Fas (CD95/APO1) in patients with pancreatic cancer

**Table 2.** Associations between the various clinicopathological factors and the expression of c-Myc and Fas

Features		NO. of cases	c-Myc cytoplasmic expression			Fas cytoplasmic expression		
			High	Low	p-value	High	Low	p-value
Age (years)	< 60	89	50	39	0.296	54	35	0.686
	≥ 60	73	35	38		42	31	
Gender	female	60	30	30	0.629	31	29	0.131
	male	102	55	47		65	37	
Location	head	117	59	58	0.401	68	49	0.634
	body/rear	45	26	19		28	17	
Size (cm)	< 4	74	39	35	0.956	46	28	0.490
	≥ 4	88	46	42		50	38	
Differentiation	poor	31	13	18	0.192	22	9	0.140
	moderate/well	131	72	59		74	57	
LNM	positive	65	32	33	0.499	37	28	0.620
	negative	97	53	44		59	38	
PNI	positive	83	51	32	0.019*	42	41	0.022*
	negative	79	34	45		54	25	
Stage	stage I	80	35	45	0.048*	48	32	0.952
	stage II	80	48	32		47	33	
	stage IV	2	2	0		1	1	

Note: LNM: Lymph node metastasis; PNI: perineural invasion. \*c-Myc had significantly higher cytoplasmic expression level in patients with PNI ( $P = 0.019$ ) and high stage ( $P = 0.048$ ) than those in patients with low stage and without PNI. Fas had significantly lower cytoplasmic expression level in patients with PNI ( $P = 0.022$ ) than those in patients without PNI.

described previously [20]. The percentages of stained cells were scored as following: 0 points for no staining; 1 point for < 20%; 2 points for 20-75%; 3 points for > 75% of cells stained. The intensity of staining was graded on the following scale: 0, negative; 1, low; 2, moderate; and 3, strong intensity. The total score was the product of the scores for the intensity and positive rate of staining. In this study, a final total score of 0-4 and 5-9 in c-Myc expression was considered to be low or high expression, while a total score > 2 in Fas expression was defined as high-expression. The stained TMA slides were scored independently by two experienced pathologists in a blinded manner.

### Statistical analysis

Associations between clinicopathological parameters and expressions of c-Myc and Fas were compared using the  $\chi^2$ -test. Relationship between c-Myc and Fas protein expressions were compared using the Spearman correlation coefficient analysis. Overall survival was calculated and survival curves were plotted using Kaplan-Meier method, and the differences between groups were compared using log-

rank test. The significant variables in univariate models were further analyzed by the multivariate Cox proportional hazards regression models for independent prognostic value. All analyses were performed using the SPSS software package (SPSS Inc, Chicago, IL, USA, version 17.0). All tests were two-sided and  $P$  values < 0.05 were considered statistically significant.

### Results

#### Expression of c-Myc and Fas in pancreatic cancer tissues and paracancerous tissues

Immunohistochemistry staining showed that c-Myc protein was mainly located in the cytoplasm of the cell, but also a small fraction (11.7%) was detected in the cell membrane. While, staining of Fas was detected only in the cytoplasm of the cell. The representative photographs were shown in **Figure 1**. In the present study, we found a significant increase of c-Myc expression in pancreatic cancer tissues compared with paracancerous tissues ( $P < 0.001$ ). On the contrary, the expression level of Fas was significantly decreased in cancer tissues compared with paracancerous tissues ( $P < 0.001$ ).

## c-Myc and Fas (CD95/APO1) in patients with pancreatic cancer

**Table 3.** Relationship between c-Myc and Fas protein expression

		c-Myc cytoplasmic expression			c-Myc cell membrane expression		
		High	Low	<i>p</i> -value	High	Low	<i>p</i> -value
Fas cytoplasmic expression	High	54	42	0.248	3	93	0.641
	Low	31	35		3	63	

**Table 4.** Relationship between cytoplasmic and cell membrane expression of c-Myc

		c-Myc cytoplasmic expression		
		High	Low	<i>p</i> -value
c-Myc cell membrane expression	High	5	1	0.174
	Low	80	76	

### *Relationships between expression of c-Myc and Fas and clinicopathological features in pancreatic cancer*

The cytoplasmic expression of c-Myc was significantly increased in pancreatic cancers with high tumor stage and with perineural invasion (PNI) ( $P = 0.019$ ,  $P = 0.048$ , respectively), while no significant difference was observed between the cell membrane expression of c-Myc and all the clinicopathological features (all  $P > 0.05$ ). In addition, the cytoplasmic expression level of Fas tended to be lower in pancreatic cancers with PNI than in those without ( $P = 0.022$ ). However, the results showed that there was no significant correlation between the cytoplasmic expressions of c-Myc and Fas and the other variables, including age, gender, tumor size, location, grade of differentiation and lymph node status (all  $P > 0.05$ ; **Table 2**).

### *Relationship between c-Myc and Fas expression in pancreatic cancer*

The results showed that both the cytoplasmic expression and cell membrane expression of c-Myc were not significantly correlated with the cytoplasmic expression of Fas in pancreatic cancer tissues ( $r = 0.091$ ,  $r = 0.037$ , respectively;  $P = 0.248$ ,  $P = 0.641$ , respectively; **Table 3**). In addition, no significant correlation was found between the c-Myc expression in cytoplasm and in cell membrane ( $r = 0.107$ ,  $P = 0.174$ ; **Table 4**).

### *Survival analysis*

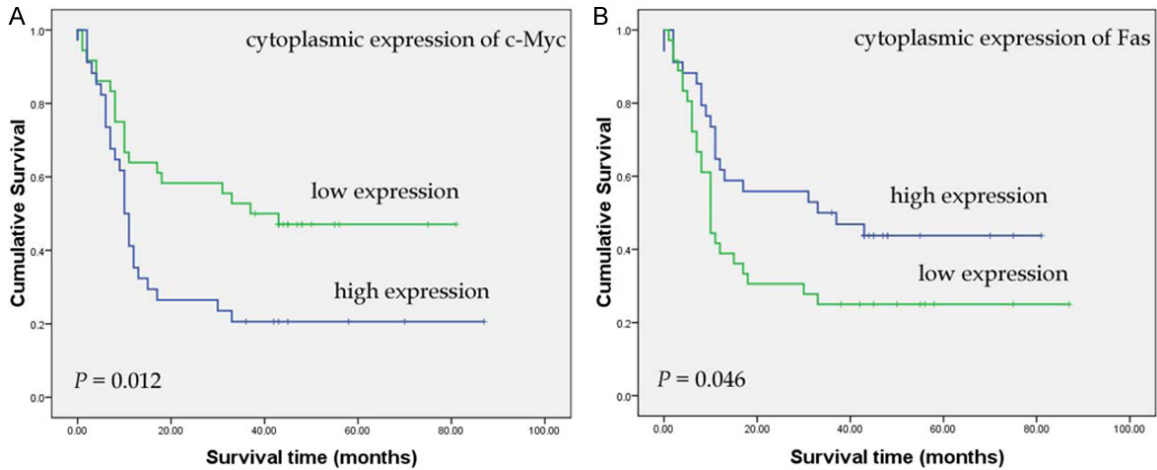
The median overall survival (OS) in the study cohort was 14 months. Kaplan-Meier analysis

demonstrated that LNM, grade of differentiation and AJCC stage were negatively significant prognostic predictors for overall survival of pancreatic cancer ( $P = 0.01$ ,  $P = 0.003$ ,  $P = 0.036$ , respectively), whereas, the other clinicopathological characteristics including age, gender, tumor size and location were not significantly associated with prognosis (data not shown).

In addition, the prognosis of pancreatic cancer patients with high c-Myc cytoplasmic expression was significantly worse than that of pancreatic cancer patients with low c-Myc cytoplasmic expression ( $P = 0.012$ ; **Figure 2A**). Median survival times were 10.0 months for high cytoplasmic c-Myc expression and 37.0 months for low cytoplasmic c-Myc expression. After stratification of patients according to AJCC stage, high c-Myc cytoplasmic expression remained a significant predictor of poor survival in stage I ( $P = 0.005$ ). In contrast, patients with high Fas cytoplasmic expression had significantly better outcomes than those with low Fas cytoplasmic expression ones (high expression: median 33.0 months, low expression: median 10.0 months,  $P = 0.046$ ; **Figure 2B**). However, there was no significant correlation in prognosis of pancreatic cancer between the patients with high c-Myc cell membrane expression than those with low c-Myc cell membrane expression ( $P = 0.183$ ).

Furthermore, variables that were significantly associated with OS in univariate analysis were included in Cox proportional hazards multivariate regression analysis. The analysis demonstrated that poor differentiation [*hazard ratio* (*HR*), 3.98; 95% *CI*, 1.92-8.23;  $P < 0.001$ , LNM (*HR*, 3.30; 95% *CI*, 1.17-9.26;  $P = 0.023$ ) and high c-Myc cytoplasmic expression (*HR*, 3.36; 95% *CI*, 1.69-6.66;  $P = 0.001$ ) were independently correlated with increased risk of death, whereas Fas cytoplasmic expression (*HR*, 1.43; 95% *CI*, 0.77-2.63;  $P = 0.249$ ) and tumor stage

## c-Myc and Fas (CD95/APO1) in patients with pancreatic cancer



**Figure 2.** Kaplan-Meier curves show that high cytoplasmic expression level of c-Myc (A) was significantly correlated with worse survival of patients with pancreatic cancer, while high cytoplasmic expression level of Fas (B) was significantly associated with better survival ( $n = 70$ ;  $P = 0.012$ ,  $P = 0.046$ , respectively; log rank test).

**Table 5.** Multivariate Cox regression analysis of potential prognostic factors for survival of pancreatic cancer

variables	HR	95% CI	P-value
Differentiation, poor v moderate/well	3.98	1.92-8.23	< 0.001*
Stage, II v I	1.05	0.39-2.82	0.922
LNM, yes v no	3.3	1.17-9.26	0.023*
c-Myc cytoplasmic expression, high v low	3.36	1.69-6.66	0.001*
Fas cytoplasmic expression, low v high	1.43	0.77-2.63	0.249

Note: HR: hazard ratio; CI: confidence interval; LNM: Lymph node metastasis. \* $P < 0.05$ , statistically significant.

(HR, 1.05; 95% CI, 0.39-2.82;  $P = 0.123$ ) were not statistically significant (Table 5).

### Discussion

Finding molecular biomarkers useful for predicting outcomes of pancreatic cancer patients could provide important information for management in the clinic. The present study demonstrated that both the aberrant cytoplasmic expressions of c-Myc and Fas were significantly correlated with OS in patients with resected pancreatic cancer. Multivariate survival analysis showed that high cytoplasmic expression level of c-Myc protein was the independent factor predicting decreased overall survival. However, Fas protein expression level lost its independent prognostic power on multivariate analysis for OS.

The c-Myc is a member of the MYC family of transcription factors, which also includes

N-Myc and L-Myc. Mounting evidence suggests that c-Myc participates in most aspects of cellular function, including metabolism, growth, differentiation, apoptosis, adhesion, and migration [21]. Previous studies have demonstrated that the amplification and/or over-expression of c-Myc are frequently detected in various cancer cells including pancreatic cancer [22-24]. In

the present study, we found that the expression level of c-Myc was higher in pancreatic cancer tissues than that in paracancerous tissues. Our results were in agreement with previous reports that c-Myc was a very strong proto-oncogene and involved in the genesis of pancreatic cancer [10]. Fas belongs to a member of the death receptor subfamily of the tumor necrosis factor (TNFR) superfamily, and it is involved in apoptotic cell death. Previous studies have showed that down-regulation or loss of Fas expression has been also observed in many types of human cancer. Our data demonstrated that the expression level of Fas was lower in pancreatic cancer tissues than that in paracancerous tissues. The data presented here confirmed previous reports that absence or low-expression of Fas contributed to the pathogenesis of pancreatic cancer, and supported the idea that cancer cells might be resistant to apoptosis induced by the loss or low-expression of Fas protein. Moreover, our present results showed that high

level of c-Myc cytoplasmic expression and low level of Fas cytoplasmic expression were significantly correlated with PNI. These findings suggested that the aberrant expression of c-Myc and Fas played important roles not only in the pancreatic tumorigenesis, but also in the progression of pancreatic cancer, and thereby contributed to the poor clinical outcome.

With respect to prognosis, our results showed that high level of c-Myc expression was a highly significant independent predictor of reduced overall survival (*HR*, 3.36, 95% *CI*, 1.69 to 6.66, *P* = 0.001). Our data suggested that high cytoplasmic expression of c-Myc might be considered as a valuable marker for prognosis of pancreatic cancer. This result was consistent with the data of some previous studies. For example, a report by Naidu et al [25] suggested that elevated c-Myc expression played an important role in breast cancer progression and might act as a potential prognostic marker for predicting the prognosis in patients with breast cancer. Schrader et al [26] also found a significant correlation between c-Myc over-expression and poor prognosis in diffuse large B cell lymphoma (DLBCL). The data revealed that c-Myc was an independent negative prognostic factor in a subgroup of patients with DLBCL. However, some studies do not find any association between c-Myc expression and prognosis [27]. Moreover, a recent study [17] showed that over-expression of c-Myc was significantly correlated with a low incidence of LNM (*P* = 0.006) and increased DFS (*P* = 0.04). The results of this study suggested that tumors with higher expression level of c-myc were correlated with better survival. It is difficult to explain these inconsistent results. However, some researchers have suggested that c-Myc protein may induce cells to differentiation and apoptosis, and may suppress expression of vascular endothelial growth factor in tumor cells. This may in part explain why over-expression of c-myc is correlated with a better outcome.

Similarly, some researchers have also found that Fas may be an independent prognostic factor in cancers. For instance, Macher et al. [28] constructed a tissue microarray containing 617 patients with renal cell carcinoma (RCC) to investigate the relationship between the expression of Fas and prognosis of RCC. Authors found high expression of Fas was cor-

related with LNM and negatively associated with disease-specific survival. In multivariate analysis, the result identified high expression of Fas was a negative independent prognostic factor in RCC. However, some studies do not find any association between Fas expression and prognosis [29, 30]. Moreover, some other reports have demonstrated that high level of Fas expression was correlated with a worse prognosis. For instance, Sejima et al [31] conducted a study using mRNA quantification and immunohistochemistry to investigate the expressions of Fas, Fas ligand (FasL) and Bcl-2 in surgically resected tumors from 82 patients with renal cell carcinoma (RCC). Multivariate analysis showed that high level of Fas mRNA was significantly associated with poorer outcome in patients with RCC (*P* = 0.0002). In this study, our data showed low cytoplasmic expression of Fas had significant impact on overall survival of pancreatic cancer, but no statistically significant association was found between Fas expression and OS in multivariate analysis. Moreover, there was no association between Fas cytoplasmic expression and tumor stage. Therefore, we agree with Markovic et al. [32] who demonstrated that Fas negativity was an unfavorable factor for therapy response and worse survival in univariate analysis in patients with diffuse large B-cell lymphoma (DLBCL), but Fas does not affect overall survival independently.

In conclusion, the results of this study suggested that the abnormal expression of c-Myc and Fas were two of the most crucial factors in the process of pancreatic tumorigenesis and progression, and both aberrant cytoplasmic expression of c-Myc and Fas were significantly correlated with OS of pancreatic cancer in univariate analysis. Furthermore, multivariate analysis demonstrated that only cytoplasmic c-Myc expression was an independent prognostic biomarker in patients with pancreatic cancer. These data contribute to further improving prognostic stratification of patients with pancreatic cancer and provide us with a potential molecular target for anticancer therapy. Currently, only few data are available regarding the prognostic impact of c-Myc and Fas expression in patients with pancreatic cancer after surgery. Therefore, more large population-based studies with long-term follow-up are required to support our findings.

### Acknowledgements

This work was supported partly by Key Discipline Construction Project of Pudong Health Bureau of Shanghai, China (Grant No: PWZxkq2010-05) to HJ, and partly by China National 863 Project Foundation for Cancer Genomics (Pancreas Genomics) (Grant No: 1006AA02A302) to HG.

### Disclosure of conflict of interest

The authors declare that there is no conflict of interest.

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### References

[1] Jemal A, Siegel R, Xu JQ and Ward E. Cancer Statistics, 2010. *CA Cancer J Clin* 2010; 60: 277-300.

[2] Bond-Smith G, Banga N, Hammond TM and Imber CJ. Pancreatic adenocarcinoma. *BMJ* 2012; 344: e2476.

[3] Gong ZH, Holly EA and Bracci PM. Survival in Population-based Pancreatic Cancer Patients: San Francisco Bay Area, 1995-1999. *Am J Epidemiol* 2011; 174: 1373-1381.

[4] Mitchem JB, Hamilton N, Gao F, Hawkins WG, Linehan DC and Strasberg SM. Long-Term Results of Resection of Adenocarcinoma of the Body and Tail of the Pancreas Using Radical Antegrade Modular Pancreatoduodenectomy Procedure. *J Am Coll Surg* 2012; 214: 46-52.

[5] Evans DB, Varadhachary GR, Crane CH, Sun CC, Lee JE, Pisters PWT, Vauthey JN, Wang H, Cleary KR, Staerckel GA, Charnsangavej C, Lano EA, Ho L, Lenzi R, Abbruzzese JL and Wolff RA. Preoperative gemcitabine-based chemoradiation for patients with resectable adenocarcinoma of the pancreatic head. *J Clin Oncol* 2008; 26: 3496-3502.

[6] Vincent A, Herman J, Schulick R, Hruban RH and Goggins M. Pancreatic cancer. *Lancet* 2011; 378: 607-620.

[7] Hammad N, Heilbrun LK, Philip PA, Shields AF, Zalupski MM, Venkatramanamoorthy R and El-

Rayes BF. CA19-9 as a predictor of tumor response and survival in patients with advanced pancreatic cancer treated with gemcitabine based chemotherapy. *Asia Pac J Clin Oncol* 2010; 6: 98-105.

[8] Dabritz J, Preston R, Hanfler J and Oettle H. Follow-Up Study of K-ras Mutations in the Plasma of Patients With Pancreatic Cancer Correlation With Clinical Features and Carbohydrate Antigen 19-9. *Pancreas* 2009; 38: 534-541.

[9] Blackford A, Serrano OK, Wolfgang CL, Parmigiani G, Jones S, Zhang XS, Parsons DW, Lin JCH, Leary RJ, Eshleman JR, Goggins M, Jaffee EM, Iacobuzio-Donahue CA, Maitra A, Cameron JL, Olinio K, Schulick R, Winter J, Herman JM, Laheru D, Klein AP, Vogelstein B, Kinzler KW, Velculescu VE and Hruban RH. SMAD4 Gene Mutations Are Associated with Poor Prognosis in Pancreatic Cancer. *Clin Cancer Res* 2009; 15: 4674-4679.

[10] Skoudy A, Hernandez-Munoz I and Navarro P. Pancreatic ductal adenocarcinoma and transcription factors: role of c-Myc. *J Gastrointest Cancer* 2011; 42: 76-84.

[11] Grippo PJ and Sandgren EP. Acinar-to-ductal metaplasia accompanies c-myc-induced exocrine pancreatic cancer progression in transgenic rodents. *Int J Cancer* 2012; 131: 1243-1248.

[12] Bernstorff WV, Glickman JN, Odze RD, Farraye FA, Joo HG, Goedegebuure PS and Eberlein TJ. Fas (CD95/APO-1) and Fas ligand expression in normal pancreas and pancreatic tumors - Implications for immune privilege and immune escape. *Cancer* 2002; 94: 2552-2560.

[13] O'Toole SA, McNeil CM, Morey AL, Millar EK, Elena-Lopez-Knowles, Musgrove EA and Sutherland RL. 29. C-Myc Gene Amplification Is Associated With A Poor Prognosis in Invasive Ductal Carcinoma. *Pathology - Journal of the RCPA* 2010; 42: S89-S90.

[14] Wu X, Cai ZD, Lou LM and Zhu YB. Expressions of p53, c-MYC, BCL-2 and apoptotic index in human osteosarcoma and their correlations with prognosis of patients. *Cancer Epidemiol* 2012; 36: 212-216.

[15] Li YJ, Wei ZM, Meng YX and Ji XR. beta-catenin up-regulates the expression of cyclinD1, c-myc and MMP-7 in human pancreatic cancer: Relationships with carcinogenesis and metastasis. *World J Gastroenterol* 2005; 11: 2117-2123.

[16] Hawksworth D, Ravindranath L, Chen Y, Furusato B, Sesterhenn IA, McLeod DG, Srivastava S and Petrovics G. Overexpression of C-MYC oncogene in prostate cancer predicts biochemical recurrence. *Prostate Cancer Prostatic Dis* 2010; 13: 311-315.

[17] Kanthan R, Fried I, Rueckl T, Senger JL and Kanthan SC. Expression of cell cycle proteins



## c-Myc and Fas (CD95/APO1) in patients with pancreatic cancer

- in male breast carcinoma. *World J Surg Oncol* 2010; 8: 10.
- [18] He C, Jiang H, Geng S, Sheng H, Shen X, Zhang X, Zhu S, Chen X, Yang C and Gao H. Expression of c-Myc and Fas correlates with perineural invasion of pancreatic cancer. *Int J Clin Exp Pathol* 2012; 5: 339-346.
- [19] Lin MS, Chen WC, Huang JX, Gao HJ, Zhang BF, Fang J, Zhou Q and Hu Y. Tissue Microarrays in Chinese Human Rectal Cancer: Study of Expressions of the Tumor-Associated Genes. *Hepatogastroenterology* 2011; 58: 1937-1942.
- [20] Ohuchida K, Mizumoto K, Ishikawa N, Fujii K, Konomi H, Nagai E, Yamaguchi K, Tsuneyoshi M and Tanaka M. The role of S100A6 in pancreatic cancer development and its clinical implication as a diagnostic marker and therapeutic target. *Clin Cancer Res* 2005; 11: 7785-7793.
- [21] Meyer N and Penn LZ. Reflecting on 25 years with MYC. *Nature reviews. Cancer* 2008; 8: 976-990.
- [22] Joensuu K, Hagström J, Leidenius M, Haglund C, Andersson L, Sariola H and Heikkilä P. Bmi-1, c-myc, and Snail expression in primary breast cancers and their metastases—elevated Bmi-1 expression in late breast cancer relapses. *Virchows Archiv* 2011; 459: 31-39.
- [23] Schleger C, Verbeke C, Hildenbrand R, Zentgraf H and Bleyl U. c-MYC activation in primary and metastatic ductal adenocarcinoma of the pancreas: incidence, mechanisms, and clinical significance. *Mod Pathol* 2002; 15: 462-469.
- [24] Li YJ, Wei ZM, Meng YX and Ji XR. Beta-catenin up-regulates the expression of cyclinD1, c-myc and MMP-7 in human pancreatic cancer: relationships with carcinogenesis and metastasis. *World J Gastroenterol* 2005; 11: 2117-2123.
- [25] Naidu R, Wahab NA, Yadav M and Kutty MK. Protein expression and molecular analysis of c-myc gene in primary breast carcinomas using immunohistochemistry and differential polymerase chain reaction. *Int J Mol Med* 2002; 9: 189-196.
- [26] Schrader A, Bentink S, Spang R, Lenze D, Hummel M, Kuo M, Arrand JR, Murray PG, Trumper L, Kube D and Vockerodt M. High myc activity is an independent negative prognostic factor for diffuse large B cell lymphomas. *Int J Cancer* 2012; 131: E348-E361.
- [27] Hayry V, Makinen LK, Atula T, Sariola H, Maki-tie A, Leivo I, Keski-Santti H, Lundin J, Haglund C and Hagstrom J. Bmi-1 expression predicts prognosis in squamous cell carcinoma of the tongue. *Br J Cancer* 2010; 102: 892-897.
- [28] Macher-Goeppinger S, Bermejo JL, Wagener N, Hohenfellner M, Haferkamp A, Schirmacher P and Roth W. Expression and prognostic relevance of the death receptor CD95 (Fas/APO1) in renal cell carcinomas. *Cancer Lett* 2011; 301: 203-211.
- [29] Gryko M, Guzinska-Ustymowicz K, Pryczynicz A, Cepowicz D, Kuklinski A, Czyzewska J, Kemona A and Kedra B. Correlation between Fas and FasL proteins expression in normal gastric mucosa and gastric cancer. *Folia Histochem Cytobiol* 2011; 49: 142-7.
- [30] Takikita M, Hu N, Shou JZ, Wang QH, Giffen C, Taylor PR and Hewitt SM. Biomarkers of apoptosis and survival in esophageal squamous cell carcinoma. *BMC Cancer* 2009; 9: 310.
- [31] Sejima T, Morizane S, Hinata N, Yao A, Isoyama T, Saito M and Takenaka A. Fas Expression in Renal Cell Carcinoma Accurately Predicts Patient Survival after Radical Nephrectomy. *Urol Int* 2012; 88: 263-270.
- [32] Markovic O, Marisavljevic D, Cemerikic V, Perunicic M, Savic S, Filipovic B and Mihaljevic B. Clinical and prognostic significance of apoptotic profile in patients with newly diagnosed nodal diffuse large B-cell lymphoma (DLBCL). *Eur J Haematol* 2011; 86: 246-255.