

Preparation and evaluation of chitosan microspheres containing nicorandil

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Abstract

Objectives: The objective of present study was to develop chitosan-based sustained release nicorandil microspheres to reduce the dosing frequency. **Materials and Methods:** The nicorandil-loaded chitosan microspheres were formulated by emulsion crosslinking method. A 3² factorial design was employed to study the influence of drug: Polymer ratio and volume of glutaraldehyde (GA) on percentage entrapment efficiency, particle size, and % drug release at 8 h. **Results:** The entrapment efficiency was found to be 41.67 ± 1.43 - $77.33 \pm 1.97\%$ and particle size range 65.67 ± 2.08 - $146.67 \pm 2.18 \mu\text{m}$. The batch CH5 showed 79.11 ± 2.23 and $96.21 \pm 2.41\%$ drug release at 8 and 12 h, respectively. **Conclusions:** Drug: Polymer ratio and volume of GA had significant effect on % entrapment efficiency, particle size, and % drug release. From the scanning electron microscopy (SEM) study observed that microspheres were spherical and fairly smooth surface. Fickian diffusion was the mode of drug release from nicorandil-loaded chitosan microspheres formulations.

Key words: Chitosan, emulsion crosslinking method, glutaraldehyde, microspheres, nicorandil

INTRODUCTION

Nicorandil is a nitrate derivative of nicotinamide that is used in the treatment of angina pectoris. It is a potassium channel opener providing vasodilatation of arterioles and large coronary arteries. Its nitrate component produces venous vasodilatation.^[1] Nicorandil has an elimination half-life of 1 h, and the therapeutic dose is in the range of 5-40 mg taken twice daily.^[2,3] To reduce the frequency of administration and to improve patient compliance, sustained-release formulations of nicorandil are desirable. One of the methods of sustained drug delivery system is by microencapsulation, which is microsphere drug delivery system.

Microspheres have been explored extensively for their use in the field of drug delivery and various polymers have been utilized

for the formulation of the microspheres, which in turn have been assessed for different purposes. Microspheres are one of the multiple unit dosage forms. Eventually the total dose and few adverse reactions may be reduced since a steady plasma concentration is maintained. Microspheres are potential drug delivery carrier systems in the segment of novel drug delivery and are prepared using assorted polymers.^[4,5] Chitosan which is deacetylated derivative of (-4)2-acetamido-2-deoxy-b-d-glucose or chitin has been extensively explored for its various biomedical and pharmaceutical applications. Properties such as biodegradability, low toxicity, and good biocompatibility make it suitable for use in drug delivery and biomedical field. As a drug carrier, chitosan has been investigated for the sustained delivery of many oral formulations and parenteral formulations.^[6] Chitosan microspheres have been prepared by emulsion crosslinking, ion-induced coagulation, and spray-drying methods. Of these methods, the most common method used to prepare chitosan microspheres is the emulsion chemical crosslinking method.^[7] There are numerous reports on the use of glutaraldehyde (GA) as a crosslinking agent in the preparation of microspheres.^[8,9] The chemical crosslinking method for preparation of chitosan microspheres involves emulsification followed by crosslinking with a suitable crosslinking agent (e.g., GA). The aim of this study was to prepare chitosan microspheres containing nicorandil by emulsion chemical crosslinking method to achieve a controlled drug release profile and to study the effect of different formulation variables such as drug:polymer ratio and GA on particle size, encapsulation efficiency, and its *in vitro* release behavior.

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MATERIALS AND METHODS

Materials

Nicorandil was obtained as a gift sample from Torrent Research Center. Chitosan was procured from Yarrow Chemicals Limited. Liquid paraffin, acetic acid, and Span 80 were purchased from RFCL. GA was purchased from Estron Chemicals.

Drug and excipient compatibility study by differential scanning calorimetry (DSC)

The DSC study was carried out using DSC-60 (Shimadzu Corporation, Japan). The instrument comprises of calorimeter, flow controller, thermal analyzer, and operating software. The samples were heated in sealed aluminum pans under air flow (30 ml/min) at a scanning rate of 10°C/min from 35 to 250°C. Empty aluminum pan was used as a reference. The heat flow as a function of temperature was measured for the samples.

Preparation of microspheres

Weigh amount chitosan and 150 mg of drug was dissolved in 15 ml 5% acetic acid. The drug-polymer dispersion was added in a 150 ml liquid paraffin (100 ml light liquid paraffin + 50 ml heavy liquid paraffin) containing 1.5 ml span 80 and it was stirred with the help of mechanical stirrer at 1,500 revolutions per minute (rpm). After 10 and 40 min, GA (25% aqueous solution) was added and stirred continuously till 2 h. Suspension of chitosan microspheres in paraffin oil, thus obtained was allowed to stand for 15 min to let the microspheres settle down under gravity. Supernatant was decanted and filtered. Microspheres obtained as residue were washed four times with solvent n-hexane to remove traces of the oil. They were finally washed with water to remove excess GA. The microspheres were dried at 40°C for 24 h. A total of nine batches, each in triplicate, were prepared as per the factorial design (3²). The amount of crosslinking agent and drug:polymer ratio was varied in batch no CH1 to CH9.

Experimental design

In this design, two factors were evaluated each at three levels and experimental trials were performed using all possible nine combination. In this present study, drug:polymer ratio (X₁) and volume of GA (X₂) were selected as independent variables. The % entrapment efficiency, particle size, and % *in vitro* drug release at 8 h was selected as dependent variables. A statistical model, incorporating interactive and polynomial terms was used to evaluate the response.

$$Y = b_0 + b_1X_1 + b_2X_2 + b_{12}X_1X_2 + b_{11}X_1^2 + b_{22}X_2^2 \quad \dots(1)$$

Where Y is the dependent variable, b₀ is the arithmetic mean response and b₁ and b₂ are the estimated coefficient for the factor X₁ and X₂, respectively. The main effect (X₁ and X₂) represents the average result of changing one factor at a time from its low to high value. The interaction term (X₁X₂) shows how the responses changes when two factors are changed simultaneously. The polynomial terms (X₁², X₂²) are included to investigate nonlinearity [Table 1 and 2].

Evaluation of microspheres

Percentage entrapment efficiency

A total of 25 mg microspheres were crushed and dispersed in 100 ml phosphate buffer pH 6.8 and sonicated for 20 min. Dispersion was stirred on magnetic stirrer for 6 h. The dispersion was filtered and drug content was analyzed spectrophotometrically at 262 nm. The percentage drug entrapment efficiency was calculated using following equation:

$$\% \text{ Entrapment Efficiency} = \frac{\text{Practical drug content}}{\text{Theoretical drug content}} \times 100$$

Particle size analysis

Particle size analysis of drug-loaded chitosan microspheres was performed by optical microscopy using a compound microscope. A small amount of dry microspheres was suspended in purified

Table 1: Variables in 3² factorial design

Independent variables	Level		
	-1	0	+1
X1: Drug:polymer ratio	1:2 (150 mg:300 mg)	1:3 (150 mg:450 mg)	1:4 (150 mg:600 mg)
X2: Volume of glutaraldehyde (ml)	2	3	4

Dependent variables: Y₁: % entrapment efficiency, Y₂: particle size, Y₃: % drug release at 8 h

Table 2: Composition of batches nicorandil microspheres

Batch no.	Drug (mg)	Polymer (mg)	Amount of cross linking agent glutaraldehyde (ml)		
			After 10 min	After 40 min	Total glutaraldehyde
CH1	150	300	1	1	2
CH2	150	450	1	1	2
CH3	150	600	1	1	2
CH4	150	300	1.5	1.5	3
CH5	150	450	1.5	1.5	3
CH6	150	600	1.5	1.5	3
CH7	150	300	2	2	4
CH8	150	450	2	2	4
CH9	150	600	2	2	4

water (10 ml). The suspension was ultrasonicated for 5 s. A small drop of suspension thus obtained was placed on a clean glass slide. The slide containing chitosan microspheres was mounted on the stage of the microscope and diameter of at least 300 particles was measured using a calibrated ocular micrometer.

In vitro drug release

A total of 20 mg nicorandil equivalent microspheres were weighed and filled in the empty capsule shells. Dissolution tests were performed in a USP Dissolution Tester Apparatus I (Basket method) at $37 \pm 0.5^\circ\text{C}$. The baskets were rotated at a speed of 50 rpm. The dissolution medium consisted of 0.1N hydrochloric acid for the first 2 h and the phosphate buffer pH 6.8 for 3-12 h (900 ml). Aliquots of 5 ml were withdrawn at different time intervals, filtered through Whatman filter paper and the content of nicorandil was determined spectrophotometrically at a wavelength of 262 nm using ultraviolet (UV) spectrophotometer.^[1]

In vitro release kinetics

The drug release data of controlled-release microspheres was fitted to kinetics models, that is, zero order, first order, and Higuchi to find out drug release pattern and mechanism.

Surface morphology

Morphological characterization of the microspheres was carried out by using scanning electron microscopy (SEM) under higher and lower resolution.

RESULTS AND DISCUSSION

Drug-excipient compatibility study by DSC

DSC of the nicorandil and nicorandil:chitosan mixtures show endothermic peak at 93.52 and 92.68°C, respectively. There was no change in the melting endotherm of the drug and drug-chitosan mixture. So, it was concluded that drug and chitosan was compatible with the each other [Figures 1 and 2].

Result of batches of nicorandil microspheres

[Tables 3 and 4].

Percentage entrapment efficiency

[Table 5].

$$Y_1 = + 60.72 + 13.28X_1 + 3.72X_2 + 0.71X_1X_2 - 0.27X_1^2 - 1.60X_2^2 \quad \dots(2)$$

The entrapment efficiency in chitosan microspheres was found 41.67-77.67. Here drug:polymer ratio and volume of GA had significant effect on entrapment efficiency.

From the above equation (2) shows that drug:polymer ratio and volume of GA had positive effect on entrapment of drug.

Effect of drug:polymer ratio: Entrapment of drug was increased

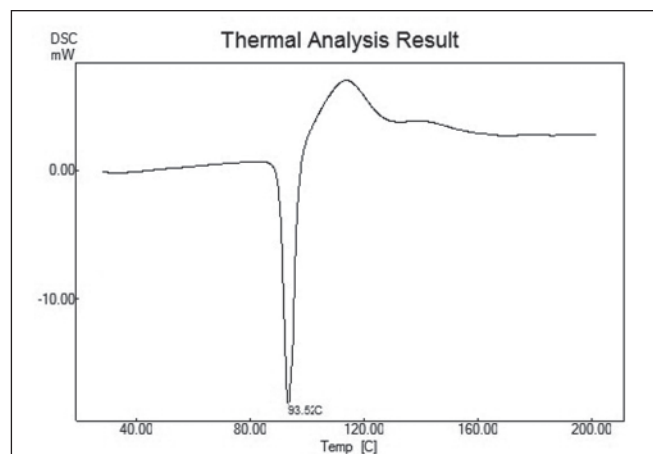


Figure 1: Differential scanning calorimetry (DSC) of nicorandil

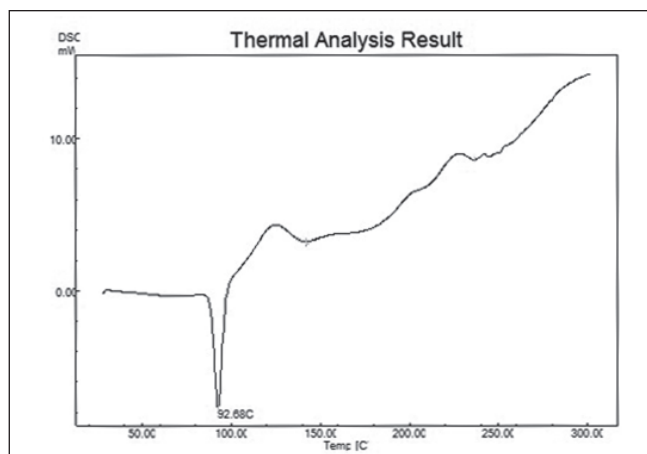


Figure 2: DSC of nicorandil:chitosan

Table 3: Observed response in 3² full factorial design for nicorandil microspheres

Batch no.	Independent variables		Dependent variables		
	X ₁ (mg)	X ₂ (ml)	Y ₁ : % entrapment efficiency	Y ₂ : Particle size (mm)	Y ₃ : % drug release at 8 h
CH1	-1	-1	41.67±1.43	73.67±1.52	101.91±1.58
CH2	0	-1	56.67±1.48	93.33±1.63	92.14±2.53
CH3	+1	-1	67.33±1.53	146.67± 2.18	71.08±2.67
CH4	-1	0	47.83±1.04	68.67±0.58	95.78±1.78
CH5	0	0	60.67±1.89	91.33±2.52	79.11±2.23
CH6	+1	0	73.33±1.87	137.33± 2.12	66.7±2.68
CH7	-1	+1	48.83±0.76	65.67±2.08	83.22±2.41
CH8	0	+1	61.83±1.04	83.00±2.00	67.64±2.97
CH9	+1	+1	77.33±1.97	129.67± 2.08	55.16±2.23

Table 4: Summary of ANOVA result for the response of design formulation

Parameter	Sum of Squares	df	Mean square	F-value	P-value	Remark
Y ₁ (quadratic)						
Model	1152.21	5	230.44	334.30	<0.001	Significant
Residual	4.83	7	0.69			
Y ₂ (quadratic)						
Model	7843.43	5	1568.69	839.09	<0.001	Significant
Residual	13.09	7	1.87			
Y ₃ (linear)						
Model	1878.68	5	375.74	89.66	<0.001	Significant
Residual	29.34	7	4.19			

ANOVA: Analysis of variance; df: Degrees of freedom

Table 5: Regression analysis for Y₁ (% entrapment efficiency)

R ²	0.9958	
Adjusted R ²	0.9929	
Standard deviation	0.83	
Source	Sum of squares	P-value
X ₁	1057.62	<0.001
X ₂	83.03	<0.001
X ₁ ²	2.02	0.1310
X ₂ ²	0.2	0.6075
X ₁ X ₂	7.06	0.0151

Table 6: Regression analysis for Y₂ (particle size analysis)

R ²	0.9983	
Adjusted R ²	0.9971	
Standard deviation	1.37	
Source	Sum of squares	P-value
X ₁	7049.34	<0.001
X ₂	208.03	<0.001
X ₁ ²	20.25	0.0133
X ₂ ²	514.03	<0.001
X ₁ X ₂	3.93	0.1904

with increasing in drug:polymer ratio. It was occurred due to the increased in viscosity of aqueous phase with increasing the polymer concentration that stabilize droplets and which prevent out flow of drug during the hardening phase.^[10]

Effect of GA: Here % entrapment was increased with increasing the volume of GA. It can be explained by the higher degree of crosslinking occurred by higher concentration of GA. Increase in amount of GA produces much denser matrix due to increased crosslinking with chitosan that reduces the outflow of drug during stirring and increases the encapsulation efficiency.^[11]

Particle size analysis

$$Y_2 = +90.77 + 34.28X_1 - 5.89X_2 - 2.25X_1X_2 + 13.64X_1^2 - 1.19X_2^2 \quad \dots(3)$$

From the above equation (3) shows that drug:polymer ratio had positive effect and volume of GA had negative effect on particle size. Effect of drug:polymer ratio, the particle size of the

microspheres was found to be dependent on the concentration of chitosan [Table 6]. At lower concentration of chitosan (300 mg or 2%), the mean particle size of microspheres observed was $73.67 \pm 1.53 \mu\text{m}$. At medium concentration (450 mg or 3%), the mean particle size was observed to be $93.33 \pm 1.53 \mu\text{m}$. At higher concentration (600 mg or 4%), the chitosan solution was so viscous that it was difficult to pass through syringe and having mean particle size of $146.67 \pm 2.08 \mu\text{m}$. Under the same preparation conditions, the droplets formed from the higher viscosity chitosan solution will be larger in size and hence result in formation of larger microspheres. The mean particle size of microspheres was significantly increased when high concentration of chitosan (4%) was used.^[11]

Effect of GA: As the volume of GA was increased, very slight decrease in the particle size of microspheres was observed. The mean particle size of microspheres prepared with 2, 3, and 4 ml of GA was 73.67 ± 1.53 , 68.67 ± 0.58 , and $65.67 \pm 2.08 \mu\text{m}$, respectively. An increase in the volume GA can increase the efficiency of the stirrer due to decreased viscosity of the oil phase that may result in decreasing the particle size. Chitosan microspheres formulated with a higher GA concentration develop a greater number of covalent bonds, thus the polymeric matrix becomes stiffer, which is responsible for low microsphere size.^[12]

In vitro drug release

$$Y_3 = +79.17 - 14.66X_1 - 9.85X_2 \quad \dots(4)$$

From the above equation (4) shows that drug:polymer ratio and volume of GA had negative effect on % drug release [Table 7].

Effect of drug:polymer ratio: The *in vitro* drug release testing [Figure 3] exhibited a biphasic mode of drug release from the microspheres. There was an initial rapid release of drug, known as burst effect, due to fast dissolution of drug molecules attached to the surface of the microspheres, and a subsequent slow release phase, during which drug molecules present in the core of the microspheres diffused out [Figure 3]. The initial burst effect decreased with an increase in polymer concentration. Theoretically, the rate of drug release from microspheres decreases with an increase in polymer concentration due to the prolongation of the diffusion route of drug. It was due to as increased in polymer concentration the matrix wall of microspheres became thicker with less number of pores. The

results of the present study are in accordance with figure (i.e., at 8 h for CH1, CH2, and CH3 are 101.94, 92.14, and 71.08%, respectively in case of 2 ml of GA) indicating a decrease in drug release with an increase in chitosan concentration.^[12]

Effect of volume of GA: The nicorandil microspheres showed the drug release after 8 h in CH2, CH5, and CH8 batches were 92.14, 79.11, and 67.14%, respectively. Microspheres prepared using 4 ml of GA releases the drug slowly compared to the microspheres in which 2 ml of GA is used. From the data it was found that the amount of GA had a strong effect on *in vitro* drug release. As the volume of GA increased the drug release decreased, which was well supported by the drug release profiles. Higher levels of GA favors the more cross-linking reaction and thus slower drug release. It can be explained by the increased cross linking density of chitosan with increased amount of GA. During the microspheres formation reaction takes place between $-NH_2$ group of chitosan and $-COO$ group of GA that forms new bonds and matrix structure form. Drug release from chitosan matrix occur only after swelling of the matrix, but increase in crosslinking density increases the hydrophobicity of chitosan matrix that increase the time for hydration and drug release decreases.^[13,14]

The below figure [Figure 4] shows that CH5 batch was matched with the drug release profile with marketed formulation.

Selection of optimized batch

Based on the proposed target, range constraints set on the dependent variables were 60.00-77.33% for Y_1 , 85.0-95.12 μm for Y_2 , and 75.72-83.72% for Y_3 . The contour plots are evolved for each response which divides the plot surface into desirable and not desirable zone. A contour for each response is then

superimposed to locate the area where the targets for the all response are achieved. Here in figure [Figure 5] shows the yellow area was the optimized area and batch CH5 was fall in the yellow region.

Kinetic models

The release kinetic of the formulation was checked by fitting the release data to various kinetic models. The release was best fitted to Higuchi model. Here, n value was 0.4851, so release mechanism was Fickian diffusion based [Table 8].

Surface morphology

From Figure 6a and b, it was observed that shape of microspheres seems to be spherical with fairly smooth surface.

Table 7: Regression analysis for Y_3 (*in vitro* drug release at 8 h)

R ²	0.9812
Adjusted R ²	0.9774
Standard deviation	1.89
Source	
Source	Sum of squares
X ₁	1289.79
X ₂	582.33
P-value	
X ₁	<0.001
X ₂	<0.001

Table 8: Kinetic values obtained from different plots of batch CH5

Batch code	Regression values (R ²)			
	Zero order	First order	Higuchi square root	Korse peppas
CH5	0.9904	0.9623	0.9974	0.9946

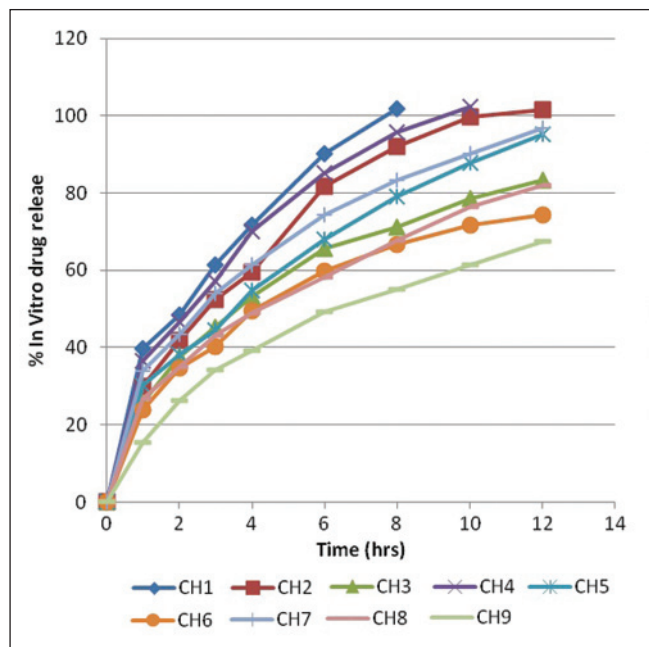


Figure 3: *In vitro* drug release profile of nicorandil microspheres batches

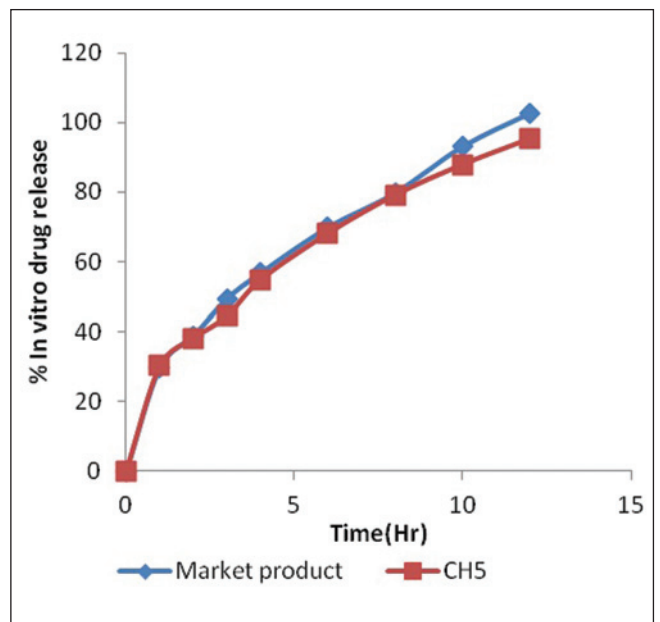


Figure 4: *In vitro* drug release profile of marketed formulation and batch CH5

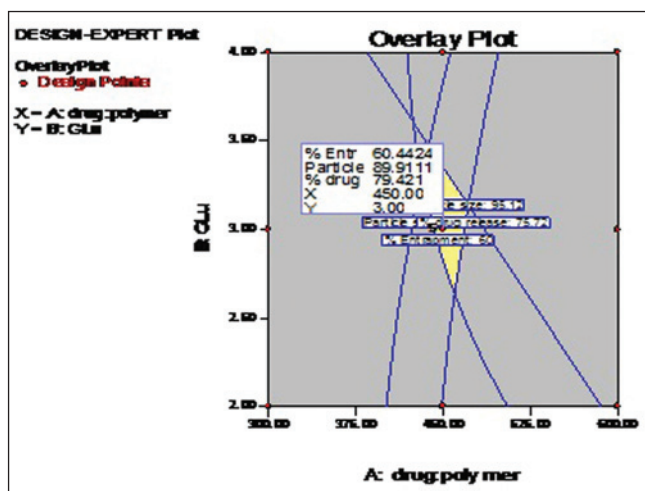


Figure 5: Overlay plot for optimization of nicorandil microspheres

CONCLUSION

Nicorandil microspheres were prepared successfully by emulsion crosslinking method. Drug:polymer ratio and volume of GA had significant effect on various parameters like percentage entrapment efficiency, particle size, and % *in vitro* drug release. It was found that increase in the drug:polymer ratio resulted an increase in particle size and % entrapment efficiency. Here, % drug release rate was decreased with increasing the concentration of polymer and volume of GA. The batch CH5 showed the drug release 96% after 12 h which was matched with market formulation release profile. From the SEM study, it was observed that microspheres were spherical and fairly smooth surface.

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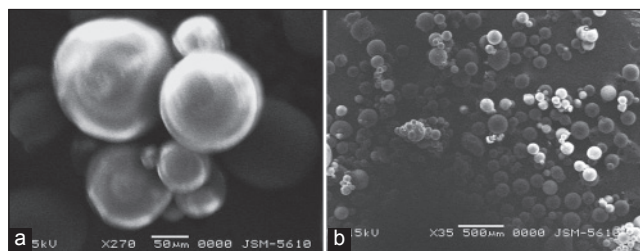


Figure 6: (a) Scanning electron microscopy (SEM) of Nicorandil microspheres. (b) SEM of nicorandil microspheres

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