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# Bayesian Model Aggregation for Ensemble-Based Estimates of Protein $pK_a$ Values

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# Abstract

This paper investigates an ensemble-based technique called Bayesian Model Averaging (BMA) to improve the performance of protein amino acid  $pK_a$  predictions. Structure-based  $pK_a$  calculations play an important role in the mechanistic interpretation of protein structure and are also used to determine a wide range of protein properties. A diverse set of methods currently exist for  $pK_a$ prediction, ranging from empirical statistical models to *ab initio* quantum mechanical approaches. However, each of these methods are based on a set of conceptual assumptions that can effect a model's accuracy and generalizability for  $pK_a$  prediction in complicated biomolecular systems. We use BMA to combine eleven diverse prediction methods that each estimate  $pK_a$  values of amino acids in staphylococcal nuclease. These methods are based on work conducted for the  $pK_a$ Cooperative and the p $K_a$  measurements are based on experimental work conducted by the García-Moreno lab. Our cross-validation study demonstrates that the aggregated estimate obtained from BMA outperforms all individual prediction methods with improvements ranging from 45-73% over other method classes. This study also compares BMA's predictive performance to other ensemble-based techniques and demonstrates that BMA can outperform these approaches with improvements ranging from 27-60%. This work illustrates a new possible mechanism for improving the accuracy of  $pK_a$  prediction and lays the foundation for future work on aggregate models that balance computational cost with prediction accuracy.

# Keywords

titration;  $pK_a$ ; prediction; statistics; model aggregation

# **1** Introduction

The calculation of  $pK_a$  values and titration behavior plays an important role in the analysis of biomolecular structure and function, including catalytic activity [19], ligand binding [59], and protein stability [6,9,46,71]. Accurate  $pK_a$  predictions, however, are challenging to calculate due to a variety of computational factors including appropriate treatment of electronic, solvation, and electrostatic effects [22,23,65] as well as adequate sampling of the biomolecular ensemble and response to titration state change [12,25,31,36,57,63,69,74]. A wide range of approaches have been developed for estimating the  $pK_a$  and titration behavior of proteins [1] and other biological molecules [60]. These approaches range from physicsbased methods and simulations [3,37,63,68], to data-driven methods that are primarily based on statistical models [8, 44, 56]. To differentiate between these approaches, we will use the term *method* throughout this paper to indicate what many computational chemists would call a *model* for predicting  $pK_a$ s, and we reserve the word *model* to indicate a *statistical model*. The topic of  $pK_a$  prediction has been thoroughly addressed in other articles [42].

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Common across all  $pK_a$  methods is the uncertainty associated with selecting, specifying, and evaluating a set of processes, parameters, and mathematical systems in order to accurately estimate  $pK_a$ . This type of uncertainty, referred to as *method selection uncertainty*, is arguably the greatest source of error and risk associated with model-based estimation and can affect a wide range of scientific and mathematical disciplines [2, 15, 51]. One of the most powerful ways to address selection uncertainty is through ensemble-based estimates [5, 28, 45, 52]. In ensemble approaches, estimates from a collection of methods are combined (e.g., through a weighted average) to form a single aggregated estimate. The motivation behind ensemble-based approaches is based on two principles: 1) all methods in the ensemble possess some unique, useful information; and, 2) no single method is sufficient to fully account for all uncertainties. Proponents of ensemble-based approaches assert that the best method to use for estimation is a combination of all of the methods. The underlying premise behind this tenet is that the information and strengths of individual methods can be combined, and their corresponding weaknesses and biases can be overcome by the strength of the group [28, 47, 49, 55]. Ensemble-based estimates are therefore expected to be more reliable and potentially more accurate than individual methods, an expectation that has been upheld in numerous examples [5,28,41,45,48,52,55,61,73].

Recently, an informal " $pK_a$  Cooperative" group has been established to explore the strengths and weaknesses of titration state prediction methods in the context of well-characterized experimental systems [42]. This paper uses the results of predictions from the Cooperative to investigate the utility of an ensemble-based approach called Bayesian Model Averaging (BMA) [28] to estimate  $pK_a$  values measured by the García-Moreno lab in staphylococcal nuclease [4, 6, 9, 10, 18, 26, 27, 31–35]. Although other statistical approaches have been used to train [40, 53] and analyze [8, 70]  $pK_a$  prediction algorithms, and BMA itself has been applied successfully for prediction tasks across many domains [41,48,61,72], this is the first application of the BMA approach to this problem domain.

## 2 Methods

#### 2.1 Bayesian Model Averaging

For  $pK_a$  prediction, a basic BMA approach is to consider a set of prediction methods as a linear system [28, 47, 49]. Let  $y_i$  for i = 1, ..., N be a series of  $pK_a$  observations, and let  $x_{ij}$  denote the  $i^{th}$  estimate obtained from the  $j^{th}$  prediction method for these observations.For example, given that  $y_i$  is the experimentally measured  $pK_a$  of Arg 313, each  $x_{ij}$  for j = 1, ..., P would be a specific method's estimate for this value. Given P prediction methods, the combination of all  $x_{ij}$  forms the numerical ensemble estimate matrix that, along with  $y_i$ , defines a linear regression model

$$y_i = \sum_{j=1}^{P} x_{ij} \beta_j + \epsilon_i \quad (1)$$

Here, the parameter vector  $\beta_j$  defines the unknown relationship between the ensemble's *P* constituents and  $\epsilon_i$  is the disturbance term that captures all factors (e.g., noise and measurement error) that influence the dependent variable  $y_i$  other than the regressors  $x_{ij}$ .

In evaluating Equation 1, the objective is to estimate the values  $\beta_j$  that will both fit the known p $K_a$  data in  $y_i$  and facilitate the ability to make inferences on unknown p $K_a$  values. Many different regression techniques can estimate  $\beta_j$  [7,30,38,50]; however, these techniques commonly generate estimates that vary in their ability to model and infer [13,21,28,47,49]. The risk and uncertainty associated with using one of these estimates over any other estimate (i.e., for statistical inference) is called *statistical model uncertainty*. Like

method selection uncertainty, statistical model uncertainly is also a common source of error in predictive modeling [28,47–49,62].

BMA addresses the challenge of statistical model uncertainty by first evaluating all possible models that can be formed from the *P* prediction methods, and then combining each model's estimates for  $\beta_i$  through a weighted average. This aggregation process generates an

aggregate-based parameter vector,  $\beta_j^{\text{BMA}}$  (Equation 2) that can provide more accurate and reliable estimates than any ensemble method, and can also outperform other ensemble-based strategies (e.g., stepwise regression) [13,21,28,64].

Formally, there are  $k = 1, ..., 2^P - 1$  distinct combinations of the *P* methods, each with a corresponding statistical model,  $M^{(k)}$ , and parameter vector,  $\beta_j^{(k)}$ . BMA combines each  $\beta_j^{(k)}$ , through a weighted average that weights each  $\beta_j^{(k)}$  by the probability that its statistical model,  $M^{(k)}$ , is the "true" model.

$$\beta_{j}^{\text{BMA}} = E\left[\beta_{j}|\mathbf{y}\right] = \sum_{k=1}^{2^{P}-1} E\left[\beta_{j}^{(k)}|\mathbf{y}, M^{(k)}\right] \Pr\left(M^{(k)}|\mathbf{y}\right) \quad (2)$$

In Equation 2,  $E\left[\beta_{j}^{(k)}|y, M^{(k)}\right]$  is the expected value of the posterior distribution of  $\beta_{j}^{(k)}$  that is weighted by the posterior probability  $Pr(M^{(k)}|y)$  (i.e., the probability that  $M^{(k)}$  is the *true* 

statistical model given  $y_i$ ). The expected posterior distribution of  $\beta_j^{(k)}$  is approximated through the linear least squares solution of the given model  $M^{(k)}$  and  $pK_a$  response variable,  $y = [y_1, \dots, y_N]$ . The posterior probability term is estimated from information criteria [47]

$$\Pr\left(M^{(k)}|\mathbf{y}\right) \propto \frac{e^{-\frac{1}{2}B^{(k)}}}{\sum\limits_{l=1}^{2^{P}-1} e^{-\frac{1}{2}B(l)}} \quad (3)$$

where  $B^{(k)}$  is the Bayesian Information Criteria for model  $M^{(k)}$ , and the information criteria itself is estimated [47]

$$B^{(k)} \approx N \log\left(1 - R^{2(k)}\right) + p^{(k)} \log N \quad (4)$$

Here  $R^{2(k)}$  is the  $R^2$  correlation value for model  $M^{(k)}$ ,  $p^{(k)}$  is the number of methods used by the model (not including the intercept), and N is the number of p $K_a$  values to be predicted.

BMA's aggregation thus weights each model's expected parameter vector  $\beta_j^{(k)}$  with the probability value that is based on that model's ability to balance trade-offs between model complexity (i.e., the number of methods used) and goodness of fit. Models that use a larger number of methods, or that do not fit the observations well, are penalized and can be eliminated from the final aggregation process (i.e., their posterior probabilities are effectively 0). In this context, BMA combines the *best* models to provide an accurate estimate for the *true* parameter terms,  $\beta_i$ .

The resulting parameter vector,  $\beta_j^{\text{BMA}}$ , obtained from Equation 2 helps to address model uncertainty by accounting for all systems of linear equations that can model the relationship between the measured p $K_a$  values  $y_i$  and values  $x_{ij}$  predicted by each method *j*. More importantly,  $\beta_j^{\text{BMA}}$  can be used to estimate new p $K_a$  values for unmeasured residues by combining new  $x_{ii}$  estimates.

# 2.2 pKa data and prediction methods

We apply the BMA approach to a set of prediction methods that estimate 83 p $K_a$  values for Lys, Asp, and Glu residues in staphylococcal nuclease mutants measured by the García-Moreno lab in a series of studies [4, 6, 9, 10, 26, 27, 31–35]. As this data is the largest set of systematic p $K_a$  values available for any protein system, it provides an extremely valuable resource for the development of new computational methods for p $K_a$  prediction.

The set of prediction methods for this data comes from a large-scale, collaborative exercise run by the  $pK_a$  Cooperative to assess and compare contemporary strategies for estimating  $pK_a$  values [42]. In this study, we have used only a subset of the methods demonstrated in the  $pK_a$  Cooperative tests and a subset of the original 83 "ground truth" experimental  $pK_a$ measurements. This restriction is due to the fact that most methods used in the  $pK_a$ Cooperative tests provided predictions for only a subset of the 83 residues and mutants characterized by the García-Moreno lab; most methods only provide estimates for half of the data. In order to include the maximum number of methods in the aggregation process, we only consider locations on the nuclease protein where *all* ensemble members provide a predicted estimate. As a result, our study is based on 36 measurements listed in Table I.

The subset of  $pK_a$  Cooperative methods used in the BMA approach were chosen based on two criteria. First, we select those methods that predict the greatest number of common residues/mutants. Second, in the event that multiple method variants exist, we chose only a single variant to eliminate the number of highly correlated methods in the aggregate. We perform this second step to ensure that multicollinearity does not inflate the significance that certain methods have during model selection and averaging; such bias can create unstable

estimates for  $\beta_j^{BMA}$  that can reduce BMA's predictive accuracy [11]. Table II summarizes the 11 methods that constitute our ensemble; each method predicts the 36 measurments in Table I. This table also lists each method's approach for conformational sampling and its solvation model.

Four types of sampling strategies were used by the methods considered in this study: molecular dynamics (MD) simulations [63,69], Monte Carlo (MC) sampling [57,69], Rosetta model refinement [57], and static structures [39, 40, 43, 53, 66, 70]. Three basic types of implicit solvation models were used: generalized Born [17,58], Poisson-Boltzmann and related methods [14,20,29], and empirical approaches [40,53]. Finally, a "null" model consisting of model compound  $pK_a$  values (ASP = 3.8, GLU = 4.3, LYS = 10.5) is also included in the analysis. The results of these methods are shown in Figure 1.

# 2.3 Training and estimating with BMA

To train the BMA model, we began by randomly sampling (without replacement) 18 of the original 36 experimental  $pK_a$  measurements. Collectively, these sampled values form the observation vector  $y_i$ . The estimates from each of the 11  $pK_a$  prediction methods in Table II for these measurements define the ensemble estimate matrix,  $x_{ij}$ . The observation vector and the ensemble estimate matrix form the linear system in Equation 1. Next, we estimated the  $\beta_j^{\text{BMA}}$  parameter from Equation 2 by assembling all  $k = 1, ..., 2^{11} - 1 = 2047$  possible statistical models  $M^{(k)}$  and estimating each model's posterior probability (Equation 3) based

statistical models  $M^{(k)}$  and estimating each model's posterior probability (Equation 3) based on its associated  $R^{2(k)}$  value and its number of independent parameters (Equation 4). The calculated parameters  $\beta_i^{\text{BMA}}$  are the weighted average of each statistical model's ordinary

least squares solution based on that model's posterior probability. Finally, we use  $\beta_j^{\text{BMA}}$  to estimate the remaining 18 pK<sub>a</sub> measurements that were *not* used to train the BMA model.

This task is accomplished by combining the estimates of all methods in Table II for the validation data with  $\beta_i^{\text{BMA}}$  to produce an aggregate prediction.

# 3 Results and discussion

We exercised the training and estimation process in Section 2.3 repeatedly in a 100-fold cross-validation study to assess the performance of several prediction approaches. In each of the study's 100 tests, we randomly selected (without replacement) 18 of the 36 experimental measurements as training data and used the remaining 18 measurements as validation data. Thus for a given test in our cross-validation study, we determined the root mean squared error (RMSE) of each prediction approach based on the validation data. The distribution of the RMSE for all 100 tests are reported for each predictive approach in our results (see Figures 1-3).

Based on these RMSE distributions, we assessed the performance of BMA to a given prediction approach *X* through a Wilcoxon rank sum paired comparison test [67]. This non-parametric approach tests the hypothesis that the RMSE distributions of BMA and *X* are equal:  $H_0: \mu_{BMA} = \mu_X$ . To control the familywise error rate of our tests (there were 56 paired tests in total), we applied a Bonferroni correction to determine a p-value threshold of

 $\alpha = \frac{0.05}{56} = 9.0E - 4$ . Thus when comparing BMA to *X*, a Wilcoxon-generated p-value that is greater than 9.0E - 4 indicates we fail to reject  $H_0$ : the distributions are thus equal and we conclude that BMA and *X* are equivalent in their predictive accuracy. On the other hand, Wilcoxon-generated p-values that are less than 9.0E - 4 indicate we should reject  $H_0$ . In this latter case, we then compared the mean RMSE for BMA (Tables III, IV, and V) and the given technique to assess performance.

We report the results of our study in three stages. Stage 1 compares the predictive results of BMA to predictions of different methods in Table II. Stage 2 assesses the robustness of BMA by evaluating BMA predictions that are based on different ensembles of methods from Table II. Stage 3 compares BMA's performance to the performance of other aggregation approaches (e.g., Stepwise regression) to examine the benefits of addressing statistical model uncertainty.

# 3.1 Stage 1: Comparing BMA to the ensemble of pKa methods

We have chosen to analyze the results by overlapping classes of methods (see Table II), rather than by individual method, for multiple reasons. First, we intend to confound the analysis of individual model performance out of respect for the authors of the methods who freely contributed their results to the  $pK_a$  Cooperative. The goal of the Cooperative is to encourage open conversation and exchange of ideas on improving biomolecular solvation models – not to rank or select "winners" from the pool of prediction methods. Second, the goal of this paper is to illustrate the potential benefits of model aggregation via BMA rather than analyze the performance of any single prediction method.

Table III and Figure 1 both provide an overview of the cross-validation  $pK_a$  prediction errors for BMA, the null model, and the various sampling and solvation methods. There are specific models in each of the categories (empirical, physics-based, static, and sampled) that will typically perform with errors < 2  $pK_a$  units. For example, in Figure 1 we can see that for GLU the expected performance for all empirical models is < 2.

However, from these results, we see that the BMA-based approach substantially outperforms all other classes of methods for all amino acids: BMA-based estimates reduce error by approximately 65% in comparison to methods that use conformational sampling, and by

about 73% in comparison to the static-conformation and physics-based solvation methods. In comparison to BMA, empirical solvation methods provide the next-best estimates; these estimates are approximately 45% higher in error than BMA estimates. The statistical significance of BMA's performance is based on p-values shown in Table VI. Based on an a value of 9.0E - 4, this table indicates that we *reject H*<sub>0</sub> for all paired comparison tests. By comparing  $\mu_{BMA}$  to other mean RMSE scores (see Table III), we concluded that BMA provides better performance than any method class in Figure 1.

#### 3.2 Stage 2: Comparing different ensembles of methods for BMA

We applied BMA to five distinct ensembles, where each ensemble was based on a unique set of methods listed in Table II. We then compared the performance of each of these BMA *instances* to assess how changes in ensemble constituents affect BMA's predictive accuracy. The first instance, labeled "BMA", is based on an ensemble that includes all methods in Table II. This instance is the same BMA instance used in Section 3.1 and Section 3.3. The next four instances were based on a subset of the methods in Table II. Specifically, the instance labeled "BMA-Static" is based on an ensemble that only used methods that relied on static-conformations (see Table II, second column). Similarly, the instance labeled "BMA-Sample" utilized an ensemble that was defined by methods that only use conformational sampling. The last two instances, BMA-Physics and BMA-Empirical, were defined exclusively by methods that used physics-based or empirical solvation approaches respectively (see Table II, third column).

Figure 2 and Table IV summarize the cross-validation  $pK_a$  prediction errors for the various BMA instances. From the RMSE distributions in Figure 2, it is clear that all instances provide excellent predictive capability. The significance of the results in Figure 2 and Table IV are base on the p-values in Table VII. Based on an *a* value of 9.0E - 4, these p-values indicate that we *accept H*<sub>0</sub> for all comparison tests save two: the BMA-Physics ensemble when predicting LYS residues and the BMA-Static ensemble when predicting ASP residues. Thus for almost all cases, the performance of the BMA instances are equivalent. In the two exceptions where  $H_0$  was rejected, the BMA instance based on the entire ensemble of methods reduced error by approximately 8% compared to the other instances. The results of these comparisons indicate that the BMA approach is robust and that BMA can overcome deficiencies observed in certain classes of methods (see Figure 1) to provide consistent predictive performance.

#### 3.3 Stage 3: Comparing BMA to other ensemble techniques

There are other approaches besides BMA that can combine an ensemble of methods to make an aggregate prediction. In our cross-validation study, we evaluated five common approaches for aggregating an ensemble and evaluated their predictive benefits in comparison to BMA. These methods included: forward regression, backward regression, step-wise regression, ordinary least squares (based on all methods in Table II), and Mallow's Cp statistic. These techniques were chosen as they have all been used successfully for a variety of inference tasks [24, 54]. As all of these approaches constructed an estimate for  $\beta_i$ , training and predicting with these approaches was performed identically to how we trained and predicted with BMA (Section 2.3). As a result, we also followed the same procedure for comparing BMA's predictive capability to these alternate ensemble-based prediction techniques.

Figure 3 and Table V provide an overview of the cross-validation  $pK_a$  prediction errors for the various ensemble-based prediction approaches and BMA (BMA used an ensemble that included all methods in Table II). The statistical significance of BMA's performance in Figure 3 is based on p-values shown in Table VIII. Based on an *a* value of 9.0*E*-4, Table

VIII indicates that we *reject*  $H_0$  for all paired comparison tests. BMA's RMSE distribution is therefore not equivalent to the RMSE distribution of any other ensemble-based technique

As the distributions are not equal, we compared mean RMSE distributions of BMA to the other ensemble-based approaches in Figure 3 and Table V. From these mean RMSE, it is clear that the BMA-based approach outperforms all other ensemble-based prediction approaches: BMA-based estimates reduced error by approximately 27% in comparison to Backward and Stepwise regression techniques and by 35% in comparison to Forward regression. In comparison to an ordinary least squares approach that uses all the methods in Table II, BMA reduces error by approximately 46%. Finally, in comparison to Mallow's  $C_p$  technique, BMA reduces error by approximately 60%.

# 4 Conclusions

This study demonstrates a proof-of-principle application of BMA to  $pK_a$  prediction using a single protein (staphylococcus nuclease) as a test case. While the performance of BMA is expected to generalize to a much broader set of  $pK_a$  prediction problems, the specific BMA model trained in this study is likely to be dependent on the staph nuclease system. In particular, the staph nuclease system has been engineered for stability and tolerates many internal titratable residues by shifting their  $pK_a$  values towards neutral titration states when covered with solvent. We note that this work is complementary to the hybrid methods developed by Witham et al. [69] which suggest the use of multiple or hybrid  $pK_a$  prediction methods based on the nature of the residue under consideration. For example, Witham et al. use structural features and the molecular environment to help select the best sampling and prediction method for a specific titratable residue. While our BMA approach is purely statistical in nature, the BMA method described here could also be trained to modify the aggregation process based on structural and environmental features (e.g., only look at ensembles of empirical methods for certain structural features and consider all methods for other structures). In future work we will look at penalizing computationally expensive methods that provide minimal accuracy benefits. Finally, we will explore larger datasets with a diverse set of bimolecular systems to provide a trained BMA  $pK_a$  prediction model using a variety of calculation methods.

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# References

- [1]. Alexov E, Mehler E, Baker N, Baptista A, Huang Y, Milletti F, Nielsen JE, Farrell D, Carstensen T, Olsson M, Shen J, Warwicker J, Williams S, Word M. Progress in the prediction of pK<sub>a</sub> values in proteins. Proteins. 2011; 79(12):3260–3275. [PubMed: 22002859]
- [2]. Apostolakis G. The concept of probability in safety assessments of technological systems. Science. 1990; 250(4986):1359–1364. [PubMed: 2255906]
- [3]. Arthur E, Yesselman J, Brooks C. Predicting extreme  $pK_a$  shifts in staphylococcal nuclease mutants with constant pH molecular dynamics. Proteins. 2011; 79(12):3276–3286. [PubMed: 22002886]
- [4]. Baran K, Chimenti M, Schlessman J, Fitch C, Herbst K, Garcia-Moreno B. Electrostatic effects in a network of polar and ionizable groups in staphylococcal nuclease. Journal of Molecular Biology. 2008; 379(5):1045–1062. [PubMed: 18499123]

- [5]. Bates JM, Granger CWJ. The combination of forecasts. Operational Research Quarterly. 1969; 20(4):451–468.
- [6]. Bell-Upp P, Robinson A, Whitten S, Wheeler E, Lin J, Stites W, García-Moreno B. Thermodynamic principles for the engineering of pH-driven conformational switches and acid insensitive proteins. Biophysical Chemistry. 2011
- [7]. Candes E, Tao T. The Dantzig selector: Statistical estimation when *p* is much larger than *n*. Annals of Statistics. 2007; 35(6):2313–51.
- [8]. Carstensen T, Farrell D, Huang Y, Baker N, Nielsen J. On the development of protein pKa calculation algorithms. Proteins. 2011; 79(12):3287–3298. [PubMed: 21744393]
- [9]. Castañeda C, Fitch C, Majumdar A, Khangulov V, Schlessman J, García-Moreno B. Molecular determinants of the pK<sub>a</sub> values of asp and glu residues in staphylococcal nuclease. Proteins. 2009; 77(3):570–588. [PubMed: 19533744]
- [10]. Chimenti M, Castañeda C, Majumdar A, García-Moreno. Structural origins of high apparent dielectric constants experienced by ionizable groups in the hydrophobic core of a protein. Journal of Molecular Biology. 2011; 405(2):361–377. [PubMed: 21059359]
- [11]. Clyde M. Bayesian Model Averaging and Model Search Strategies. Bayesian Statistics. 1999; volume 6:157–185.
- [12]. Damjanovic A, Wu X, García-Moreno, Brooks B. Backbone relaxation coupled to the ionization of internal groups in proteins: A self-guided Langevin dynamics study. Biophys J. 2008; 95(9): 4091–4101. [PubMed: 18641078]
- [13]. Davidson, I.; Fan, W. When Efficient Model Averaging Out-Performs Boosting and Bagging. Vol. volume 4213 of Lecture Notes in Computer Science. Springer; Berlin Heidelberg: 2006. p. 478-486.
- [14]. Davis M, McCammon A. Electrostatics in biomolecular structure and dynamics. Chem. Rev. 1990; 90(3):509–521.
- [15]. Devooght J. Model uncertainty and model inaccuracy. Reliability Engineering and System Safety. 1998; 59(2):171–185.
- [16]. Dolinsky T, Czodrowski P, Li H, Nielsen J, Jensen J, Klebe G, Baker N. PDB2PQR: expanding and upgrading automated preparation of biomolecular structures for molecular simulations. Nucleic Acids Research. 2007; 35(Web Server issue)
- [17]. Dominy B, Brooks C. Development of a generalized Born model parametrization for proteins and nucleic acids. J. Phys. Chem. B. 1999; 103(18):3765–3773.
- [18]. Dwyer JJ, Gittis AG, Karp DA, Lattman EE, Spencer DS, Stites WE, García-Moreno E B. High apparent dielectric constants in the interior of a protein reflect water penetration. Biophysical Journal. 2000; 79(3):1610–1620. [PubMed: 10969021]
- [19]. Fersht, A. Enzyme Structure and Mechanism. W. H. Freeman and Co.; San Francisco, CA: 1985.
- [20]. Fixman M. The Poisson–Boltzmann equation and its application to polyelectrolytes. Journal of Chemical Physics. 1979; 70(11):4995–146.
- [21]. Genell A, Nemes S, Steineck G, Dickman PW. Model selection in medical research: A simulation study comparing Bayesian model averaging and stepwise regression. BMC Medical Research Methodology. 2010; 10:108. [PubMed: 21134252]
- [22]. Ghosh N, Cui Q. pK<sub>a</sub> of residue 66 in staphylococal nuclease. I. insights from qm/mm simulations with conventional sampling. The Journal of Physical Chemistry B. 2008; 112(28): 8387–8397. [PubMed: 18540669]
- [23]. Ghosh N, Prat-Resina X, Gunner M, Cui Q. Microscopic pK<sub>a</sub> analysis of glu286 in cytochrome c oxidase (rhodobacter sphaeroides): Toward a calibrated molecular model. Biochemistry. 2009; 48(11):2468–2485. [PubMed: 19243111]
- [24]. Gilmour SG. The Interpretation of Mallows's Cp Statistic. J. of the Royal Statistical Society. 1996; 45(1):49–56.
- [25]. Gunner M, Zhu X, Klein M. MCCE analysis of the pKas of introduced buried acids and bases in staphylococcal nuclease. Proteins. 2011; 79(12):3306–3319. [PubMed: 21910138]
- [26]. Harms M, Castañeda C, Schlessman J, Sue G, Isom D, Cannon B, García-Moreno E B. The  $pK_a$  values of acidic and basic residues buried at the same internal location in a protein are governed by different factors. Journal of Molecular Biology. 2009; 389(1):34–47. [PubMed: 19324049]

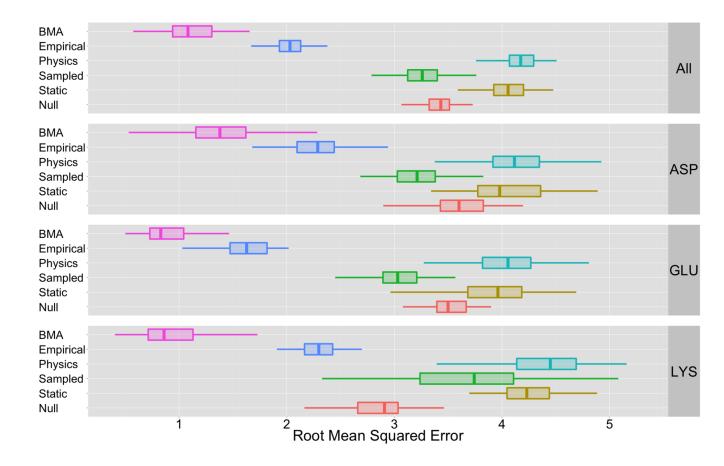
- [28]. Hoeting JA, Madigan D, Raftery AE, Volinsky CT. Bayesian model averaging: a tutorial. Statistical Science. 1999; 14(4):382–417.
- [29]. Honig B, Nicholls A. Classical electrostatics in biology and chemistry. Science. 1995; 268(5214): 1144–1149. [PubMed: 7761829]
- [30]. Hosmer, DW.; Lemeshow, S. Applied Logistic Regression. Wiley; New York: 1989.
- [31]. Isom D, Castañeda C, Cannon B, García-Moreno E. Bertrand. Large shifts in pK<sub>a</sub> values of lysine residues buried inside a protein. Proceedings of the National Academy of Sciences. 2011; 108(13):5260–5265.
- [32]. Isom D, Castañeda C, Cannon B, Velu P, García-Moreno E B. Charges in the hydrophobic interior of proteins. Proceedings of the National Academy of Sciences of the United States of America. 2010; 107(37):16096–16100. [PubMed: 20798341]
- [33]. Isom DG, Cannon BR, Castañeda CA, Robinson A, García-Moreno E B. High tolerance for ionizable residues in the hydrophobic interior of proteins. Proceedings of the National Academy of Sciences. 2008; 105(46):17784–17788.
- [34]. Karp DA, Gittis AG, Stahley MR, Fitch CA, Stites WE, E BGM. High apparent dielectric constant inside a protein reflects structural reorganization coupled to the ionization of an internal Asp. Biophysical Journal. 2007; 92(6):2041–2053. [PubMed: 17172297]
- [35]. Karp DA, Stahley MR, García-Moreno B. Conformational consequences of ionization of Lys, Asp, and Glu buried at position 66 in staphylococcal nuclease. Biochemistry. 2010; 49(19):4138– 4146. [PubMed: 20329780]
- [36]. Kato M, Pisliakov AV, Warshel A. The barrier for proton transport in aquaporins as a challenge for electrostatic models: The role of protein relaxation in mutational calculations. Proteins. 2006; 64(4):829–844. [PubMed: 16779836]
- [37]. Machuqueiro M, Baptista A. Is the prediction of pK<sub>a</sub> values by constant-pH molecular dynamics being hindered by inherited problems? Proteins. 2011; 79(12):3437–3447. [PubMed: 22072522]
- [38]. Mallows CL. Some comments on C<sub>p</sub>. Technometrics. 1973; 15(4):661–675.
- [39]. Meyer T, Kieseritzky G, Knapp E-W. Electrostatic pK<sub>a</sub> computations in proteins: role of internal cavities. Proteins. 2011; 79(12):3320–3332. [PubMed: 21744394]
- [40]. Milletti F, Storchi L, Cruciani G. Predicting protein pK<sub>a</sub> by environment similarity. Proteins. 2009; 76(2):484–495. [PubMed: 19241472]
- [41]. Morales-Casique E, Neuman SP, Vesselinov VV. Maximum likelihood Bayesian averaging of airflow models in unsaturated fractured tuff using Occam and variance windows. Stochastic Environmental Research and Risk Assessment. 2010; 24(6):863–880.
- [42]. Nielsen J, Gunner M, García-Moreno. The pK<sub>a</sub> cooperative: A collaborative e ort to advance structure-based calculations of pK<sub>a</sub> values and electrostatic effects in proteins. Proteins. 2011; 79(12):3249–3259. [PubMed: 22002877]
- [43]. Nielsen JE, Vriend G. Optimizing the hydrogen-bond network in poisson-boltzmann equationbased  $pK_a$  calculations. Proteins-Structure Function and Genetics. 2001; 43(4):403–412.
- [44]. Olsson M. Protein electrostatics and  $pK_a$  blind predictions; contribution from empirical predictions of internal ionizable residues. Proteins. 2011; 79(12):3333–3345. [PubMed: 22072518]
- [45]. Opitz D, Maclin R. Popular ensemble methods: An empirical study. Journal of Artificial Intelligence Research. 1999; 11:169–198.
- [46]. Privalov PL. Stability of proteins: small globular proteins. Adv Protein Chem. 1979; 33:167–241.[PubMed: 44431]
- [47]. Raftery A. Bayesian Model Selection in Social Research. Sociological Methodology. 1995; 25:111–163.
- [48]. Raftery AE, Gneiting T, Balabdaoui F, Polakowski M. Using Bayesian model averaging to calibrate forecast ensembles. Monthly Weather Review. 2005; 133(5):1155–1174.

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- [49]. Raftery AE, Madigan D, Hoeting JA. Bayesian model averaging for linear regression models. Journal of the American Statistical Association. 1998; 92:179–191.
- [50]. Reiss PT, Huang L, Cavanaugh JE, Roy AK. Resampling-based information criteria for bestsubset regression. Annals of the Institute of Statistical Mathematics. 2012; 64(6):1161–1186.
- [51]. Rojas R, Batelaan O, Feyen L, Dassargues A. Assessment of conceptual model uncertainty for the regional aquifer Pampa del Tamarugal – North Chile. Hydrology and Earth System Sciences. 2010; 14(2):171–192.
- [52]. Rokach L. Ensemble-based classifiers. Artif. Intell. Rev. 2010; 33(1-2):1-39.
- [53]. Rostkowski M, Olsson M, Søndergaard C, Jensen J. Graphical analysis of pH-dependent properties of proteins predicted using PROPKA. BMC structural biology. 2011; 11(1):6. [PubMed: 21269479]
- [54]. Seber, G.; Lee, A. Linear Regression Analysis. Vol. volume 1 of Wiley Series in Probability and Statistics. John Wiley & Sons; 2012.
- [55]. Seni G, Elder JF. Ensemble methods in data mining: Improving accuracy through combining predictions. Synthesis Lectures on Data Mining and Knowledge Discovery. 2010; 2(1):1–126.
- [56]. Shan J, Mehler E. Calculation of  $pK_a$  in proteins with the microenvironment modulated-screened coulomb potential. Proteins. 2011; 79(12):3346–3355. [PubMed: 21748803]
- [57]. Song Y. Exploring conformational changes coupled to ionization states using a hybrid Rosetta-MCCE protocol. Proteins. 2011; 79(12):3356–3363. [PubMed: 22072519]
- [58]. Still C, Tempczyk A, Hawley R, Hendrickson T. Semianalytical treatment of solvation for molecular mechanics and dynamics. J. Am. Chem. Soc. 1990; 112(16):6127–6129.
- [59]. Szabo A, Karplus M. A mathematical model for structure-function relations in hemoglobin. Journal of Molecular Biology. 1972; 72(1):163–197. [PubMed: 4648112]
- [60]. Tang C, Alexov E, Pyle AM, Honig B. Calculation of pK<sub>a</sub>s in rna: on the structural origins and functional roles of protonated nucleotides. Journal of Molecular Biology. 2007; 366(5):1475– 1496. [PubMed: 17223134]
- [61]. Vlachopoulou M, Gosink L, Pulsipher T, Ferryman T, Zhou N, Tong J. An ensemble approach for forecasting net interchange schedule. IEEE PES General Meeting. 2013
- [62]. Volinsky CT, Madigan D, Raftery AE, Kronmal RA. Bayesian model averaging in proportional hazard models: Assessing the risk of a stroke. Journal of the Royal Statistical Society: Series C (Applied Statistics). 1997; 46(4):433–448.
- [63]. Wallace J, Wang Y, Shi C, Pastoor K, Nguyen B-L, Xia K, Shen J. Toward accurate prediction of pK<sub>a</sub> values for internal protein residues: The importance of conformational relaxation and desolvation energy. Proteins. 2011; 79(12):3364–3373. [PubMed: 21748801]
- [64]. Wang D, Zhang W, Bakhai A. Comparison of Bayesian model averaging and stepwise methods for model selection in logistic regression. Statistics in Medicine. 2004; 23(22):3451–3467. [PubMed: 15505893]
- [65]. Warshel A, Sharma PK, Kato M, Parson WW. Modeling electrostatic effects in proteins. Biochimica et Biophysica Acta (BBA) - Proteins & Proteomics. 2006; 1764(11):1647–1676.
- [66]. Warwicker J. pK<sub>a</sub> predictions with a coupled finite difference Poisson-Boltzmann and Debye-Hückel method. Proteins. 2011; 79(12):3374–3380. [PubMed: 21661058]
- [67]. Wilcoxon F. Individual Comparisons by Ranking Methods. Biometrics Bulletin. 1945; 1(6):80– 83.
- [68]. Williams S, Blachly P, McCammon A. Measuring the successes and deficiencies of constant pH molecular dynamics: A blind prediction study. Proteins. 2011; 79(12):3381–3388. [PubMed: 22072520]
- [69]. Witham S, Talley K, Wang L, Zhang Z, Sarkar S, Gao D, Yang W, Alexov E. Developing hybrid approaches to predict pK<sub>a</sub> values of ionizable groups. Proteins. 2011; 79(12):3389–3399. [PubMed: 21744395]
- [70]. Word M, Nicholls A. Application of the Gaussian dielectric boundary in Zap to the prediction of protein pK<sub>a</sub> values. Proteins. 2011; 79(12):3400–3409. [PubMed: 21661059]
- [71]. Yang AS, Honig B. On the pH dependence of protein stability. Journal of Molecular Biology. 1993; 231(2):459–474. [PubMed: 8510157]

- [72]. Ye M, Neuman SP, Meyer PD. Maximum likelihood Bayesian averaging of spatial variability models in unsaturated fractured tuff. Water Resources Research. 2004; 40(5)
- [73]. Zhang GP. Time series forecasting using a hybrid ARIMA and neural network model. Neurocomputing. 2003; 50:159–175.
- [74]. Zheng L, Chen M, Yang W. Random walk in orthogonal space to achieve efficient free-energy simulation of complex systems. Proceedings of the National Academy of Sciences. 2008; 105(51):20227–20232.

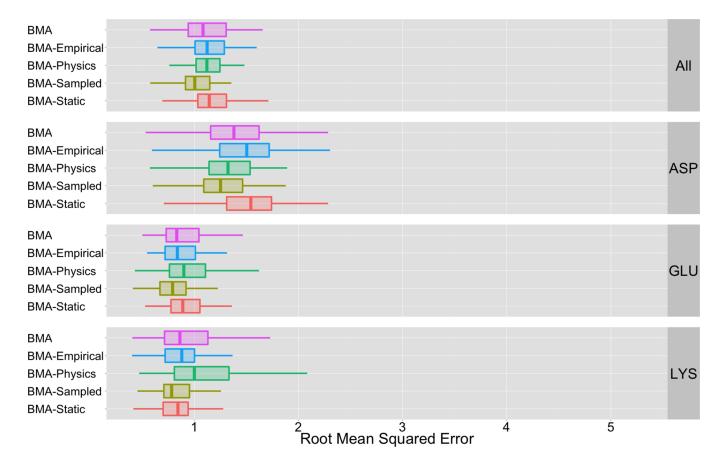
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#### Figure 1.

This figure depicts a summary of root-mean-squared error for various conformational sampling and solvation methods (see Table II). The label "BMA" reflects performance for a BMA instance that uses all methods in Table II to construct  $\beta_{BMA}$ ; we examine the performance effects of constraining the ensemble to specific methods (e.g., BMA based only on Static methods) in Figure 2 and Table VII. The "Sampled" set contains results from methods that use MD, MC, and Rosetta sampling methods; the "Static" set contains results from methods that did not use conformational sampling. The "Physics" set contains results from methods that use GB and PB solvation models; the "Empirical" set includes methods with empirical solvation models. From these results, we see that the BMA-based approach substantially outperforms all other classes of methods: BMA-based estimates reduce error by approximately 65% in comparison to methods that use conformational sampling, and by about 73% in comparison to the static-conformation and physics-based solvation methods. In comparison to BMA, empirical solvation methods provide the next-best estimates; these estimates are approximately 45% higher in error than BMA estimates. The mean RMSE values shown in this figure are listed in Table III. The statistical significance of BMA's performance is based on p-values shown in Table VI.

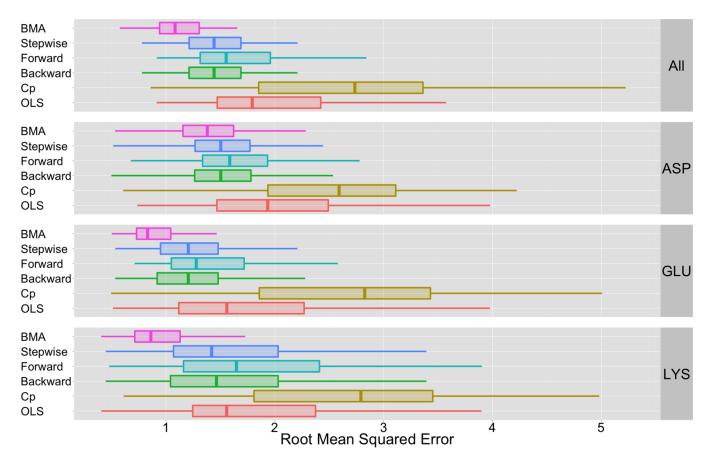
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#### Figure 2.

This figure depicts a summary of root-mean-squared error for different BMA ensemble instances. The label "BMA" indicates that the BMA aggregate uses all methods from Table II to construct  $\beta_{BMA}$ . The "BMA-Empirical" and "BMA-Physics" labels indicate performance for BMA instances that are based on ensembles that only consider a subset of the total methods; e.g., the "BMA-Empirical" aggregation process only uses empirical methods to construct  $\beta_{BMA}$ . "BMA-Static" and "BMA-Sampling" labels indicate performance for ensembles that are restricted by sampling strategies. The mean RMSE values shown in this figure are listed in Table IV. The statistical significance of BMA's performance is based on p-values shown in Table VII. Based on Table IV and Table VII, we conclude that the performance of all instances are equivalent with the exception of two cases (BMA-Physics predicting LYS and BMA-Static predicting ASP). In these two cases, BMA based on a full ensemble outperforms these instances by approximately 8%.

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#### Figure 3.

This figure depicts a summary of root-mean-squared error for different ensemble approaches. The label "BMA" indicates that the BMA aggregate uses all methods from Table II to construct  $\beta_{BMA}$ . The other approaches are alternative ensemble techniques that are based on different strategies for model specification (see Section 3.3). The mean RMSE values shown in this figure are listed in Table V and the statistical significance of BMA's performance is based on p-values shown in Table VIII. Based on data in Table VIII and Table V, BMA-based approach outperform all other ensemble techniques: BMA-based estimates reduce error by approximately 27% in comparison to Backward and Stepwise regression techniques and by 35% in comparison to Forward regression. In comparison to an ordinary least squares approach that uses all methods in II, BMA reduces error by approximately 46%. Finally, in comparison to  $C_p$  methods, BMA reduces error by approximately 60%.

# Table I

This table lists the locations and experimental values for  $pK_a$  measurements on a series of mutant staphylococcus nuclease proteins used in our cross-validation study. References for each measurement are provided in the table:

Residue	$D20^d$	E23 <sup>c</sup>	E25 <sup>c</sup>	К25 <sup>b</sup>	К34 <sup>b</sup>	К36 <sup>b</sup>	E38 <sup>c</sup>	E39 <sup>c</sup>	E41 <sup>c</sup>
pK <sub>a</sub>	4.0	7.1	7.5	6.3	7.1	7.2	6.8	8.2	6.5
Residue	D41 <sup><i>d</i></sup>			D62 <sup><i>d</i></sup>	E62 <sup><i>c</i></sup>	D66 <sup>d</sup>		E72 <sup>c</sup>	К72 <sup>b</sup>
p <i>K<sub>a</sub></i>	4.0	6.8	8.1	8.7	7.7	8.1	8.5	7.3	8.6
Residue	D74 <sup><i>d</i></sup>	E74 <sup>C</sup>	$D90^d$	E91 <sup>c</sup>	К91 <sup><i>b</i></sup>	E92 <sup>c</sup>	E99 <sup>c</sup>	$D99^d$	$D100^d$
р <i>К</i> <sub>а</sub>	8.3	7.8	7.5	7.1	9.0	9.0	8.4	8.5	6.9
Residue	E100 <sup>c</sup>	K103 <sup>b</sup>	K104 <sup>b</sup>	D109 <sup>d</sup>	D118 <sup>d</sup>	E125 <sup>c</sup>	D125 <sup>d</sup>	D132 <sup>d</sup>	E132 <sup>c</sup>
1									

<sup>a</sup>Dwyer et al., 2000 [18]

<sup>b</sup>Isom et al., 2011 [31]

<sup>c</sup>Isom et al., 2010 [32]

<sup>d</sup>Bertrand García-Moreno, personal communication, 2009.

# Table II

This table lists the  $pK_a$  prediction methods used in our BMA approach. Sampling strategies include molecular dynamics (MD), Monte Carlo (MC), Rosetta structure refine ment (Rosetta), and static structure (Static). Solvation models include generalized Born (GB), Poisson-Boltzmann (PB), and empirical methods (Empirical).

Reference	Sampling strategy	Solvation model	Number of predictions made (out of 89 total)
"Null" model		_	89
Wallace J, et al, 2011 [63]	MD	GB	68
Warwicker J, 2011 [66]	Static	PB	83
Rostkowski M, et al, 2011 [53]	Static	Empirical	89
Milletti F, et al, 2009 [40]	Static	Empirical	74
Nielsen JE, et al, 2001 [43]	Static	PB	87
Witham S, et al, 2011 [69]	MD and MC	GB and PB	65
Word JM, et al, 2011 [70]	Static	PB	61
PDB2PKA [16, 43]	Static	PB	87
Song Y, 2011 [57]	MC and Rosetta	PB	69
Meyer T, et al, 2011 [39]	Static	PB	87

#### Table III

This table lists the average root-mean-squared-error performance results from our 100-fold cross-validation study, ordered by the overall error obtained by the model or method. The table shows performance for BMA, the Null model, as well as performance of categories of  $pK_a$  prediction methods. The "Sampled" set contains results from methods which used MD, MC, and Rosetta sampling methods; the "Static" set contains results from methods which did not use conformational sampling. The "Physics" set contains results from methods which used GB and PB solvation models; the "Empirical" set includes methods with empirical solvation models. The ensemble of prediction methods is broken into two comparative groups based on conformational sampling strategy (e.g., MD, MC, and Rosetta sampling vs. static) and solvation model (GB and PB solvation methods vs. empirical methods). Additionally, cross-validation results are shown for all amino acids as well as for the individual amino acid types ASP, GLU, and LYS. The statistical significance of BMA's performance is based on *p*-values shown in Table VI. Table VI indicates that BMA's mean RMSE is statistically significant to all other methods. Based on this table's mean RMSE scores, we see that the BMA-based approach outperforms all other ensemble techniques: BMA-based estimates reduce error by approximately 65% in comparison to methods that use conformational sampling, and by about 73% in comparison to the staticconformation and physics-based solvation methods. In comparison to BMA, empirical solvation methods provide the next-best estimates; these estimates are approximately 45% higher in error than BMA estimates.

Method class	All	ASP	GLU	LYS
ВМА	1.155	1.376	0.924	1.023
Empirical solvation	2.031	2.264	1.609	2.280
Physics-based solvation	4.167	4.129	4.051	4.414
Sampled conformations	3.257	3.209	3.034	3.628
Static conformations	4.057	4.061	3.936	4.246
Null	3.420	3.598	3.516	2.861

#### Table IV

This table lists the average root-mean-squared-error (RMSE) performance results from our 100-fold crossvalidation study based on the distributions in Figure 2. The label "BMA" indicates that the BMA aggregate uses all methods from Table II to construct BMA. The "BMA-Empirical" and "BMA-Physics" labels indicate performance for BMA instances that are based on ensembles that only consider a subset of the total methods; e.g., the "BMA-Empirical" aggregation process only uses methods from the Empirical class to construct BMA. Likewise, "BMA-Static" and "BMA-Sampling" labels indicate performance for ensembles that are restricted by sampling strategies. The statistical significance of these mean RMSE values is based on *p*-values shown in Table VII. Table VII indicates the performance for all BMA instances is equivalent with the exception of two cases: the BMA-Physics ensemble when predicting LYS residues; and, BMA-Static when predicting ASP residues.

Aggregate	All	ASP	GLU	LYS
BMA	1.155	1.376	0.924	1.023
BMA-Empirical	1.127	1.443	0.867	0.845
BMA-Physics	1.164	1.318	0.959	1.111
BMA-Sampled	1.017	1.260	0.809	0.828
BMA-Static	1.175	1.496	0.944	0.830

#### Table V

This table lists the average root-mean-squared-error performance results from our 100 X cross-validation study based on the distributions in Figure 3. The label "BMA" indicates that the BMA aggregate uses all methods from Table II to construct  $\beta_{BMA}$ . The other approaches are alternative ensemble techniques that are based on different strategies for model specification (see Section 3.3). The statistical significance of these mean RMSE is based on *p*-values shown in Table VIII. Table VIII indicates that BMA's mean RMSE is statistically significant to all other methods. Based on this table's mean RMSE scores, we see that the BMA-based approach outperforms all other ensemble techniques: BMA-based estimates reduce error by approximately 27% in comparison to Backward and Stepwise regression techniques and by 35% in comparison to Forward regression. In comparison to an ordinary least squares approach that uses all methods in II, BMA reduces error by approximately 46%. Finally, in comparison to  $C_p$  methods, BMA reduces error by approximately 60%.

Ensemble technique	All	ASP	GLU	LYS
BMA	1.155	1.376	0.924	1.023
Stepwise regression	1.564	1.541	1.298	1.766
Forward regression	1.740	1.680	1.436	2.044
Backward regression	1.565	1.543	1.295	1.768
C <sub>p</sub>	2.839	2.756	2.851	2.858
Ordinary Least Squares (OLS)	2.089	2.219	1.865	1.983

#### Table VI

This table contains p-values that indicate the statistical significance of BMA's RMSE distribution compared to other methods in Figure 1. *P*-values are based on a Wilcoxon rank sum paired comparison test that detects if data populations for BMA and method *X* (e.g., Sampled or Static) are identical: Based on the 56 paired tests (Figures 1- 3) we determine a *p*-value threshold of  $a = 0.05/56 = 9.0 \times 10^{-4}$ ; *p*-values less than  $9.0 \times 10^{-4}$  thus indicate that we reject the null hypothesis that the two distributions are identical. This table indicates that we reject this null hypothesis for all paired comparison tests and by comparing the mean BMA RMSE to other mean RMSE scores listed in Table III, As such, we conclude that BMA provides better performance than any method class in Figure 1. Repeated values in this table reflect floating point precision limits in the *p*-value calculation.

Method	All	ASP	GLU	LYS
Empirical	$3.95 \times 10^{-18}$	$4.73 \times 10^{-18}$	9.38×10 <sup>-18</sup>	$6.28 \times 10^{-17}$
Physics	$1.98 \times 10^{-18}$	$1.98 \times 10^{-18}$	$1.98 \times 10^{-18}$	$1.98 \times 10^{-18}$
Sampled	$1.98 \times 10^{-18}$	$1.98 \times 10^{-18}$	$1.98 \times 10^{-18}$	$2.17 \times 10^{-18}$
Static	$1.98 \times 10^{-18}$	$1.98 \times 10^{-18}$	$1.98 \times 10^{-18}$	$2.04 \times 10^{-18}$
Null	$1.98 \times 10^{-18}$	$2.04 \times 10^{-18}$	$1.98 \times 10^{-18}$	$8.09 \times 10^{-18}$

#### Table VII

This table contains *p*-values that indicate the statistical significance of performance for BMA based on the different ensembles shown in Figure 2. *P*-values are based on a Wilcoxon rank sum paired comparison test as described in Table VI. This table indicates that we accept the null hypothesis that the methods using different ensemble subsets behave identically for all comparison tests save two (italicized for contrast): the BMA-Physics ensemble when predicting LYS residues; and, BMA-Static ensemble when predicting ASP residues. The predictive performance of BMA based on different ensembles is therefore equivalent in almost all cases. For these two cases where we reject the null hypothesis, we compare the mean RMSE of BMA to the mean RMSE of BMA-Physics and BMA-Static (Table IV) to identify that BMA reduces error by approximately 8% compared to these instances.

Aggregate	All	ASP	GLU	LYS
BMA-Empirical	1.19×10 <sup>-1</sup>	$1.00 \times 10^{-3}$	4.92×10 <sup>-1</sup>	$9.58 \times 10^{-1}$
BMA-Physics	5.83×10 <sup>-2</sup>	9.90×10 <sup>-1</sup>	9.38×10 <sup>-3</sup>	$1.19 \times 10^{-4}$
BMA-Sampled	1.00	1.00	9.99×10 <sup>-1</sup>	9.98×10 <sup>-1</sup>
BMA-Static	2.20×10 <sup>-3</sup>	7.32×10 <sup>-6</sup>	8.08×10 <sup>-3</sup>	9.90×10 <sup>-1</sup>

#### Table VIII

This table contains *p*-values that indicate the statistical significance of BMA's RMSE distribution compared to the other ensemble-based methods in Figure 3. *P*-values are based on Wilcoxon rank sum paired comparison test as described in Table VI. This table indicates that we reject the null hypothesis for all paired comparison tests and by comparing and conclude that BMA provides better performance than any ensemble approach in Figure 3.

Ensemble technique	All	ASP	GLU	LYS
Stepwise	$3.85 \times 10^{-15}$	1.29×10 <sup>-4</sup>	$7.25 \times 10^{-17}$	5.78×10 <sup>-14</sup>
Forward	9.94×10 <sup>-17</sup>	4.10×10 <sup>-9</sup>	$6.57 \times 10^{-18}$	2.41×10 <sup>-16</sup>
Backward	$3.85 \times 10^{-15}$	$1.12 \times 10^{-4}$	$8.37 \times 10^{-17}$	6.76×10 <sup>-14</sup>
$C_p$	1.96×10 <sup>-17</sup>	$2.41 \times 10^{-16}$	$1.50 \times 10^{-17}$	1.86×10 <sup>-16</sup>
Ordinary Least Squares (OLS)	1.19×10 <sup>-17</sup>	1.19×10 <sup>-13</sup>	3.32×10 <sup>-17</sup>	4.12×10 <sup>-14</sup>