

Review Article

Signal Transduction in Astrocytes during Chronic or Acute Treatment with Drugs (SSRIs, Antipolar Drugs, GABA-ergic Drugs, and Benzodiazepines) Ameliorating Mood Disorders

Leif Hertz, Dan Song, Baoman Li, Ting Du, Junnan Xu, Li Gu, Ye Chen, and Liang Peng

Department of Clinical Pharmacology, China Medical University, No. 92 Beier Road, Heping District, Shenyang, China

Correspondence should be addressed to Liang Peng; hkkid08@yahoo.com

Received 30 October 2013; Accepted 16 December 2013; Published 24 February 2014

Academic Editor: Joseph I. Shapiro

Copyright © 2014 Leif Hertz et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Chronic treatment with fluoxetine or other so-called serotonin-specific reuptake inhibitor antidepressants (SSRIs) or with a lithium salt “lithium”, carbamazepine, or valproic acid, the three classical antipolar drugs, exerts a multitude of effects on astrocytes, which in turn modulate astrocyte-neuronal interactions and brain function. In the case of the SSRIs, they are to a large extent due to 5-HT_{2B}-mediated upregulation and editing of genes. These alterations induce alteration in effects of cPLA₂, GluK2, and the 5-HT_{2B} receptor, probably including increases in both glucose metabolism and glycogen turnover, which in combination have therapeutic effect on major depression. The ability of increased levels of extracellular K⁺ to increase [Ca²⁺]_i, is increased as a sign of increased K⁺-induced excitability in astrocytes. Acute anxiolytic drug treatment with benzodiazepines or GABA_A receptor stimulation has similar glycogenolysis-enhancing effects. The antipolar drugs induce intracellular alkalization in astrocytes with lithium acting on one acid extruder and carbamazepine and valproic acid on a different acid extruder. They inhibit K⁺-induced and transmitter-induced increase of astrocytic [Ca²⁺]_i, and thereby probably excitability. In several cases, they exert different changes in gene expression than SSRIs, determined both in cultured astrocytes and in freshly isolated astrocytes from drug-treated animals.

1. Introduction

Signal transduction in astrocytes is of fundamental, but largely unrecognized, importance for brain function under normal and abnormal conditions. Due to the extreme difficulty in studying specifically astrocytic signaling in the brain *in vivo*, many studies have been carried out in isolated preparations of astrocytes, that is, freshly isolated cells obtained directly from the brain or cultured astrocytes. Claims by the Kimelberg that astrocyte cultures are misleading [1] are unfortunately often correct. However, these researchers have unjustifiably ignored that many types of astrocyte cultures exist and that they are not similar in their characteristics. The fact that the cultures used by us are well suited to study drug-induced signaling changes is demonstrated in Table 1 (slightly modified from [2]). It shows identical changes in gene expression and editing induced by chronic treatment with drugs used to treat mood disorders (fluoxetine; carbamazepine) in cultured astrocytes and in astrocytes freshly dissociated from the brains of animals chronically treated with the same

drugs [3]. Not a single gene was affected differently in the two situations. The freshly dissociated cells are probably not sufficiently intact to show the mechanisms involved in these gene changes or their functional consequences, which for this reason have been elucidated in the cultured cells. Such studies have indicated important correlations between different effects on gene expression or editing shown in Table 1 and they have enabled tentative functional interpretations.

2. Effects of SSRIs

2.1. Acute Effects

2.1.1. Pathway. The prototypes of antidepressant drugs (which also have anxiolytic effect) are the serotonin-specific reuptake inhibitors (SSRIs). When fluoxetine, the first of the presently used SSRIs, was approved for clinical use in 1987, two effects of the drug had been established: inhibition of serotonin reuptake by the serotonin transporter (SERT) [6] and partial displacement of serotonin (5-HT) binding

TABLE 1: Comparison between effects on gene expression and editing of chronic treatment with the SSRI fluoxetine or the antibipolar drug carbamazepine in cultured mouse astrocytes and in astrocytes freshly isolated from drug-treated mice.

Gene	Drug	FACS, astrocytes	Culture astrocytes
ADAR2	Fluoxetine	Up	Up
5-HT _{2B} receptor expression	Fluoxetine	Up	Up
5-HT _{2B} editing	Fluoxetine	Up	Up
5-HT _{2c} receptor expression	Fluoxetine	Unchanged	Unchanged
cPLA _{2a}	Fluoxetine	Up	Up
sPLA ₂	Fluoxetine	Unaltered	Unaltered
GluK2 expression	Fluoxetine	Up	Up
GluK2 editing	Fluoxetine	Up	Up
GluK4 expression	Fluoxetine	Unchanged	Unchanged
cfos expression	Fluoxetine	Up	Up
fosB expression	Fluoxetine	Up	Up
Ca _v 1.2	Fluoxetine	Up	Up
NBCe1	Carbamazepine	Up	Up
GluK2	Carbamazepine	Down	Down
cPLA ₂	Carbamazepine	Up	Up

Table 1 shows all experiments in which drug effects were compared in cultured astrocytes and in astrocytes freshly obtained by FACS as described by Lovatt et al. [3]. For FACS, astrocytes had been obtained from mice stained with GFP, based on expression of the astrocyte-specific GFAP, in transgenic animals and the stained cells were separated after cell dissociation by means of their fluorescent signal. The cultures were treated chronically with either fluoxetine or carbamazepine, and the animals had been treated chronically for a similar length of time (2 weeks). Complete agreement was found. From Peng et al. [2], with the exception of Ca_v1.2, which is from Du et al. [4], using similar techniques.

to cultured astrocytes [7], which have no SERT expression [8]. However, at that time, astrocytes were supposed to be unimportant for brain function, and inhibition of SERT has since then been regarded as the mechanism responsible for SSRIs effects. In 1987, the 5-HT_{2B} receptor was unknown, but it is now established to have the highest affinity between 5-HT receptors for SSRIs, with a K_i for displacement of serotonin binding to cultured astrocytes of 70 nM [5]. This is identical to its K_i for inhibition of serotonin uptake via the human placental SERT [9]. Moreover, all five SSRIs are equipotent in their effect on astrocytes [10]. This is a distinct difference from the large potency difference in their effect on SERT, although the therapeutic doses are of roughly comparable magnitudes, a difference, which can only partly be explained by differences in drug kinetics and protein binding.

The metabolic pathway activated in cultured astrocytes by fluoxetine was first established by Li et al. [11] and an expanded version is shown in Figure 1. With increasing realization of the importance of glycogenolysis for signaling in astrocytes [12–14], it may be important that fluoxetine acutely stimulates glycogenolysis, an effect that is secondary to an increase in [Ca²⁺]_i [15]. Fluoxetine might also affect glycogen synthesis, since it stimulates the AKT pathway (see below). The involvement of 5-HT_{2B} receptor-stimulated glycogenolysis has been established during learning, where acute administration of serotonin can rescue long-term learning in a one trial aversive learning paradigm in day-old chickens under conditions when the aversive stimulus was otherwise too weak to establish more than transient long-term memory retention [16]. Fluoxetine and paroxetine have a similar effect and are equipotent, showing that the rescue was not due to inhibition of SERT, and the rescuing effect was

inhibited by an inhibitor of glycogenolysis (Gibbs and Hertz, submitted, *Frontiers in Pharmacology* (Neuropharmacology) after invitation to Research Topic).

In accordance with Figure 1, fluoxetine acutely phosphorylates AKT in cultured astrocytes [5]. AKT-mediated phosphorylation of glycogen synthase kinase-3 β (GSK-3 β) inhibits its enzymatic activity. AKT phosphorylation may stimulate glycogen synthesis, since phosphorylation and activation of glycogen synthase by GSK-3 decreases the activity of glycogen synthase [17]. It is consistent with these observations that Li et al. [18], from the Jope group, found in whole brain that fluoxetine administration increases the levels of phosphorylated GSK3 β . Later, Beurel et al. [19] from the same group confirmed that fluoxetine rapidly and robustly increases serine phosphorylation of GSK3 but that these responses in young mice are blunted or absent. This is consistent with an effect on astrocytes, since these glial cells are generated postnatally [20]. An increased glycogen synthesis following 5-HT_{2B} receptor activation has been directly shown in hepatocytes, whereas agonists of 5-HT₁ and 5-HT_{2A} receptors had the opposite effect [21].

2.2. Chronic Effects

2.2.1. Affected Genes. The effects of SSRIs that are important in connection with major depression are the chronic effects, since the effect of clinical treatment takes several weeks to appear. To-date, all gene effects caused by chronic treatment with fluoxetine (the only SSRI studied on astrocytes in vivo) after mice had been treated for 14 days with i.p. injection of fluoxetine (10 mg/kg per day) have been identical to those in cultured astrocytes chronically treated with fluoxetine or

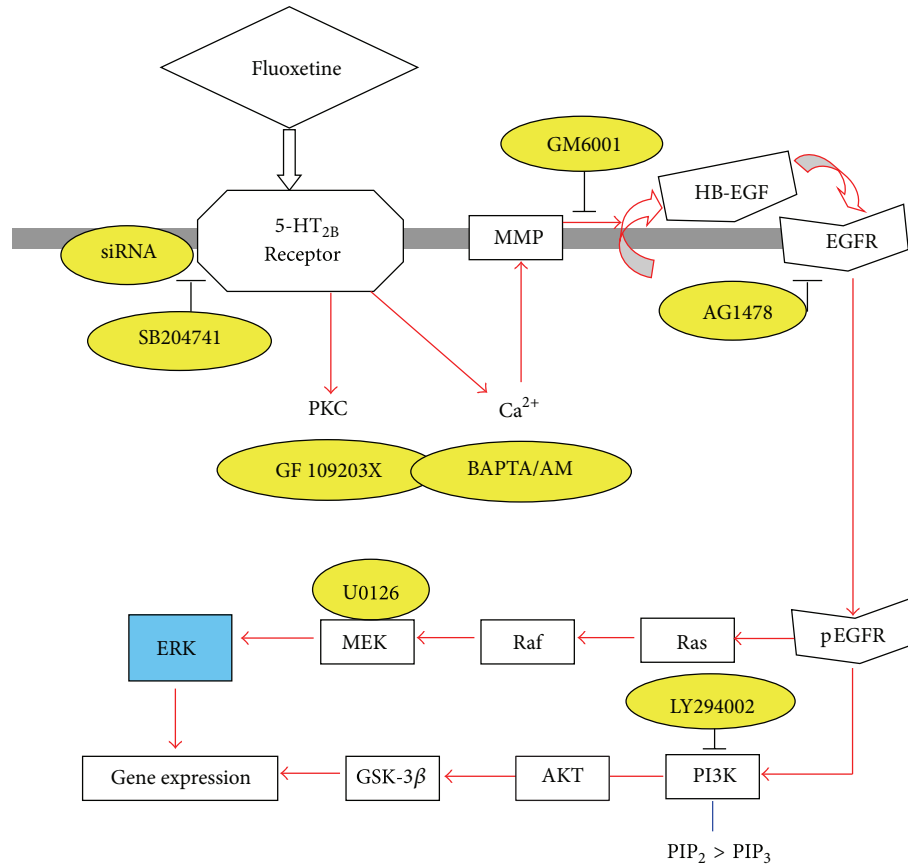


FIGURE 1: Schematic illustration of pathways leading to stimulation of ERK and AKT phosphorylation by fluoxetine in astrocytes. Fluoxetine binds to 5-HT_{2B} receptors. The activation of the receptors in turn induces an enhancement of protein kinase C (PKC) activity and of intracellular Ca²⁺ concentration by Ca²⁺ release from intracellular stores. The latter activates Zn-dependent metalloproteinases (MMPs) and leads to shedding of growth factor(s). The released epidermal growth factor receptor (EGFR) ligand stimulates phosphorylation of the EGFR. The downstream target of EGFR, extracellular-regulated kinase (ERK) (shown in blue) is phosphorylated via the Ras/Raf/MEK pathway, and AKT is phosphorylated via PI3K pathway. During chronic fluoxetine administration, inhibitors (shown in yellow) of the 5-HT_{2B} receptor (SB204741), or siRNA against this receptor, of PKC (GF 109293X), of intracellular Ca²⁺ homeostasis (BAPTA/AM, an intracellular Ca²⁺ chelator), of Zn-dependent metalloproteinases (GM6001), of the receptor-tyrosine kinase of the EGFR (AG1478), of ERK phosphorylation (U0126, a mitogen-activated kinase (MEK) inhibitor), or of the AKT pathway (LY294002, a PI3K inhibitor) prevent changes in gene expression and editing. PI3K catalyzes the formation of PIP₃ from PIP₂, from Hertz et al. [5].

other SSRIs, as was shown in Table 1. They have also been similar to those found by other authors or ourselves in total brain with the exception of gene expression of sPLA₂ and GluK4, genes that were upregulated in neurons of the treated animals and accordingly also in whole brain [22]. Anhedonia, one of the components of major depression, caused oppositely directed changes in expression of some of the same genes [22]. Other gene expression changes established in the cultured cells did not occur, but it should be remembered that anhedonia is only one component of major depression. Fluoxetine can reverse both the anhedonia and the gene expression changes (B. Li and L. Peng's unpublished results).

2.2.2. ADAR2. The upregulation of ADAR2 by chronic treatment with fluoxetine (Table 1) depends on 5-HT_{2B} receptor stimulation, since the 150–200% increase of mRNA and protein expression of ADAR2 was prevented in astrocytes treated

with 5-HT_{2B} receptor siRNA [23]. In contrast to the upregulation of cPLA₂ (see below), it is not known if additional steps of the fluoxetine-mediated pathways are also required. ADARs constitute a family of adenosine deaminases catalyzing deamination of adenosine to inosine in double-stranded regions of mRNAs, thus changing the translated protein sequence, since inosine is read by the cells as guanosine [24]. There are three members in the ADAR family: ADAR1, ADAR2, and ADAR3 [25]. All three types of ADAR are expressed in the brain [26]. ADAR2 upregulation in fluoxetine-treated mice was specific for this subtype, observed only in astrocytes and occurred within 3 days [23]. In the brain ADAR2 has been shown in hippocampal pyramidal neurons and cerebellar Purkinje cells and Bergmann glial cells, with less expression of ADAR1 and ADAR3 [27]. ADAR2 and probably also its upregulation are essential for the editing changes shown in Table 1; this has been directly demonstrated in cultured astrocytes by the use of cells treated with siRNA against ADAR2 [23].

2.2.3. *cPLA₂*. Astrocytes are among the cells that express calcium-dependent phospholipase 2 (*cPLA₂*) [27, 28], and in the brain *in vivo*, they may even be enriched in this phospholipase [29–31]. Its activation specifically releases arachidonic acid from the *sn*-2 position of membrane-bound phospholipid substrate in neural preparations [32–35], including glioma cells [36]. Arachidonic acid strongly stimulates glucose metabolism in cultured astrocytes [37]. So does treatment with 10 μ M fluoxetine for 24 h, which might have sufficed to induce an increase in *cPLA₂* [38], whereas acute exposure of astrocyte cultures to the same concentration of fluoxetine has no corresponding effect (L. Peng and L. Hertz's unpublished experiments). Arachidonic acid also stimulates glycogenolysis [39, 40].

Rapoport and coworkers [41–43] showed that chronic administration of fluoxetine leads to stimulation and enhanced mRNA and protein expression in rat brain of *cPLA₂*, but not of the two other phospholipases *A₂* (secretory *PLA₂* (*sPLA₂*) and intracellular *PLA₂* (*iPLA₂*)). Li et al. [28] confirmed a slow and selective upregulation of mRNA and protein expression of *cPLA_{2a}*, the major isoform of *cPLA₂*, in mouse astrocytes in primary cultures during chronic incubation with 1 or 10 μ M fluoxetine. The upregulation was abrogated by the 5-HT_{2B} receptor antagonist SB 204741, the metalloproteinase inhibitor GM6001, and the inhibitor of EGF receptor tyrosine phosphorylation AG1478 and by U0126, the inhibitor of ERK_{1/2} phosphorylation. These are all inhibitors of the signaling pathway that was shown in Figure 1 for fluoxetine, showing that upregulation of mRNA and protein of *cPLA₂* were inhibited by the same drugs that acutely inhibit ERK_{1/2} phosphorylation and by inhibition of the phosphorylation itself. As was shown in Table 1, upregulation, specifically of *cPLA_{2a}*, has also been found in freshly dissociated astrocytes isolated by FACS after 2-week treatment of rats with fluoxetine, whereas no corresponding effect was found in neurons [22]. This finding strongly suggests that the enhanced *cPLA₂* activity demonstrated in whole brain after chronic fluoxetine treatment [42] selectively occurs in astrocytes.

The stimulation of glucose metabolism may be important in the pathophysiology of depressive illness and its pharmacological treatment. Glucose metabolism in brain is reduced in many regions, primarily in the frontotemporal parts, in patients suffering from unipolar depression [44–47], with a correlation between the degree of hypometabolism and severity of the illness [48], and normalization following treatment with an SSRI [49–51]. Figure 2 shows that arachidonic acid may play a role in determining rates of cerebral glucose metabolism as seen from a rectilinear correlation in depressed patients between plasma concentration of arachidonic acid and rate of cerebral glucose utilization in one of the regions affected metabolically by depression [52]. Moreover, genetic associations are found between *cPLA₂* and major depression [53, 54]. Depending on the site(s) of impairment of glucose metabolism, glycogen metabolism could also be affected. However, the effect of arachidonic acid is not necessarily exerted on glucose metabolism itself but might also be exerted on energy-requiring reactions. The importance of mitochondrial abnormalities in major

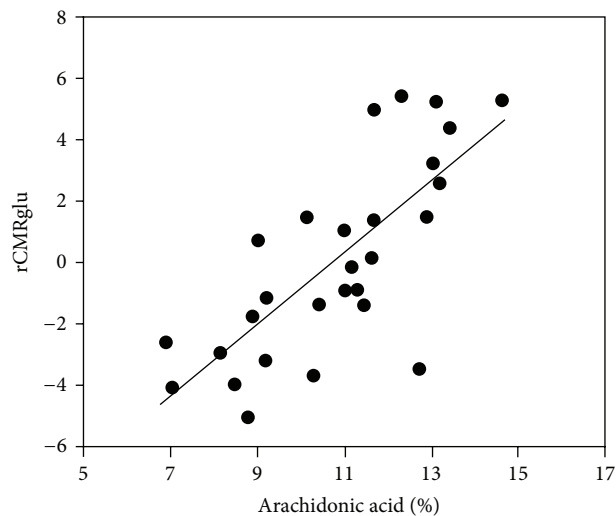


FIGURE 2: Correlation between cerebral metabolic rates of glucose metabolism and plasma arachidonic acid levels. Cerebral metabolic rates of glucose metabolism (rCMRglu) measured by NMR *in vivo* are shown to be correlated with plasma arachidonic acid, expressed as a percentage of total phospholipid polyunsaturated fatty acids. The correlation is statistically significant ($P < 0.0005$), from Elizabeth Sublette et al. [52].

depression is shown by a significantly reduced number of mitochondria, but larger mean mitochondrial volume, in hippocampus in rats sensitive to stress-induced depression than in control rats [55]. Following treatment with the antidepressant imipramine, a significant increase in the number of mitochondria occurred in the stress-sensitive group. Clinical studies also suggest that psychiatric features can be prominent features of mitochondrial disorders, but additional methodologically rigorous and adequately powered studies are needed before definitive conclusions can be drawn [56].

Arachidonic acid metabolites, including prostaglandins, may also exert other beneficial effects important for the amelioration of depression [57]. Prostaglandin synthesis inhibitors in doses used to treat pain [58, 59] may cause fear, agitation, and affective lability, and one euthymic bipolar patient repeatedly developed depression during such exposure [60]. Thus, the prostaglandins are likely to exert oppositely directed effects.

2.2.4. *Kainate Receptors*. The only glutamate receptors shown in Table 1 are the kainate receptors GluK, among which GluK2 was upregulated and edited in astrocytes, whereas GluK4 is upregulated, but not edited, in neurons following chronic fluoxetine treatment [22]. mRNA expression analysis has demonstrated that the human GluK2c splice variant in brain is mainly expressed in nonneuronal cells and barely expressed in neurons [61]. GluK2 can operate not only in an inotropic, but also in a metabotropic mode [62]. Three positions can be edited in the genome-encoded GluK2 mRNA at 3 sites, the I/V site, the Y/C site, and the Q/R site. The editing was increased at all three sites by chronic treatment with fluoxetine [22]. This observation is consistent with another previous observation by the Barbon group that editing of the Q/R

site in intact brain is slightly decreased by chronic fluoxetine treatment [63]. In most cases, inclusion of subunits containing the edited R form of the Q/R site lowers Ca^{2+} permeability [64], and in fluoxetine-treated cultures, a normally occurring increase in free cytosolic Ca^{2+} concentration of 300–400% of control value in response to 100 μM glutamate was abolished by the fluoxetine treatment [23]. This increase is evoked by the GluK2 receptor operating in its metabotropic manner [5]. Glutamate-mediated inhibition of a slow neuronal afterhyperpolarization current I (sAHP) is blocked by kainate receptor(s) in the metabotropic mode [65], and inhibition of sAHP might be secondary to transmitter effects on astrocytes [66]. Inhibition of a slow neuronal afterhyperpolarization by glutamate would contribute to the importance of acute effects of glutamate in learning [16] and could also have a bearing on the mechanism by which GluK2 upregulation and editing may be therapeutically beneficial in major depression. Nevertheless, it is often assumed that increase in $[\text{Ca}^{2+}]_i$ in astrocytes is mediated by the metabotropic glutamate receptor mGluR5, but this is only in immature brain [67]. Moreover, mGluR5 is at best minimally upregulated in either astrocytes or neurons after chronic *in vivo* treatment of adult mice with fluoxetine (Figure 3). Mice in which the GluK2 receptor is knocked-out exhibit less anxious or more risk-taking type behavior and less manifestation of despair [68]. Obsessive-compulsive disorder is also genetically linked to abnormalities in *Grik2*, the gene coding for GluK2 [69, 70].

The role of glutamate in major depression and its drug treatment has repeatedly been discussed [5, 22, 71, 72], and some concepts are continuously changing. Hertz et al. [5] reviewed synaptic potentiation by glutamatergic stimulation of astrocytes, a topic which has been further mathematically analyzed by Tewari and Majumdar [73]. Correlations between drug effects on major depression and on glutamatergic activities have attracted special interest in connection with the rapid but short-lasting therapeutic effects of ketamine and riluzole in depressed patients. Recently, the effects of riluzole and of ketamine have been reviewed by Murrrough and colleagues, Lapidus et al. [74] mainly discuss neuronal effects, although disregarding the GluK4 receptor (which according to Table 1 becomes upregulated in neurons after chronic fluoxetine treatment) but mentioning mGluRs (of which we found at least mGluR5 to be virtually unaffected by fluoxetine treatment). Murrrough et al. [75] note very interesting correlations between ketamine, major depression, and cognitive function.

2.2.5. The 5-HT_{2B} Receptor. Unpublished experiments in cultured astrocytes by Li et al. have shown that upregulation of the 5-HT_{2B} receptor itself was specific for this 5-HT₂ receptor, since neither 5-HT_{2A} nor 5-HT_{2c} receptors were upregulated. The lack of upregulation of the latter 5-HT₂ receptor was confirmed in freshly isolated astrocytes from fluoxetine-treated mice [22] as shown in Table 1. In contrast to the changes in gene expression of ADAR, cPLA₂ and GluK2 described above and those in Ca^{2+} homeostasis to be discussed below, that of the 5-HT_{2B} receptor occurred very slowly (Figures 4(a) and 4(b)), but as usual with the latency

depending upon the fluoxetine concentration. 5-HT_{2B} protein was not upregulated after 3 days, even at the highest fluoxetine concentration, whereas mRNA expression occurred somewhat faster (Figures 4(a) and 4(b)). In contrast editing of the 5-HT_{2B} receptor (Figure 4(c)) was obvious after 3 days of treatment, and after 7 days the edited receptor no longer responded to serotonin by an increase in activity measured as the ability of serotonin to evoke release of ³H-inositol phosphate (IP) from labeled IP₃ (Figure 4(d)) or phosphorylation of ERK_{1/2} (not shown). To ascertain that the inhibition of 5-HT_{2B} receptor activity was a direct result of receptor editing and not due to other effects by chronic fluoxetine administration, COS-7 cells were infected with receptor plasmids of either normal 5-HT_{2B} receptors or receptors with 8 RNA sites RNA edited, and a similar inhibition was shown (Figure 4(e)).

2.2.6. Ca²⁺ Homeostasis. In contrast to the ability of fluoxetine (and many transmitters) to acutely cause an increase in $[\text{Ca}^{2+}]_i$, chronic treatment with fluoxetine rapidly abolishes or reduces transmitter and fluoxetine-induced $[\text{Ca}^{2+}]_i$ increase [23]. However, a corresponding increase by elevation of extracellular concentrations of K⁺ above 15 mM [76] is *not* reduced, but rather increased, by chronic treatment with fluoxetine [4, 23]. The reason for this is a fluoxetine-mediated upregulation of the L-channel gene Ca_v1.2, shown both in cultured cells and in astrocytes freshly obtained from fluoxetine-treated animals (Table 1). This may compensate for a downregulation of capacitative Ca²⁺ uptake via store-operated channels, Socs [23], as will be described below. In contrast, the Ca_v1.3 gene, which shows higher expression in freshly isolated astrocytes than in corresponding neurons, is unaffected by treatment of mice with fluoxetine for 2 weeks [4].

Socs are very important for regulation of the amount of Ca²⁺ stored in the endoplasmic reticulum (ER) and thus the amount of Ca²⁺ released by activation of inositol trisphosphate (IP₃) receptors (IP₃R) or ryanodine receptors (RyR) (Figure 5). After IP₃R- or RyR-mediated unloading of ER-bound Ca²⁺, some Ca²⁺ may enter mitochondria [77], where it exerts a stimulatory effect on several tricarboxylic acid enzymes [78], and some Ca²⁺ exits via the Na⁺/Ca²⁺ exchanger. Accordingly, the cell suffers loss of intracellular Ca²⁺, which under normal conditions is compensated for by capacitative Ca²⁺ uptake via Socs. A major component of Socs is the transient receptor potential channel (TRPC) protein TRPC1 [79]. In cells in which TRPC1 had been knocked down by treatment with antisense oligonucleotides TRPC1 antibody the capacitative Ca²⁺ uptake is also greatly reduced. The same occurs after short-lasting chronic treatment with fluoxetine and many other drugs (see below) and reduces or abrogates the ability of transmitters to increase astrocytic $[\text{Ca}^{2+}]_i$ [80]. Thus, treatment with SSRIs inhibits the ability of transmitters, at least temporarily (see below), but on account of the upregulation of Ca_v1.2, not that of elevated K⁺ concentrations to increase astrocytic $[\text{Ca}^{2+}]_i$. This is consistent with the increased ability of elevated extracellular K⁺ concentration to increase $[\text{Ca}^{2+}]_i$ reported in [4, 23], and it is a prerequisite for transmitter effects, for example, on glycogenolysis.

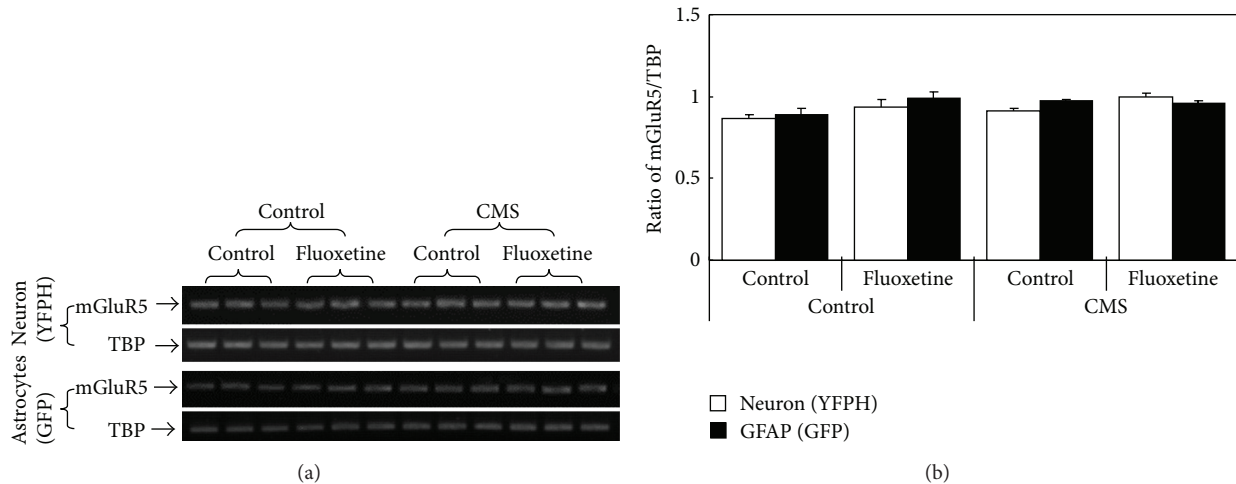


FIGURE 3: mRNA of mGluR5 is not upregulated in either astrocytes or neurons freshly isolated from animals treated with fluoxetine (10 mg fluoxetine hydrochloride/kg, i.p.) for 14 days. In both cases, astrocytes had been stained with GFP and neurons with YPHF in transgenic animals and they were separated after cell dissociation by means of the different fluorescent signals. (a) Blot showing mGluR5 RNA measured by reverse transcription polymerase chain reaction (RT-PCR) in all experiments together with that of TATA-binding protein (TBP) used as housekeeping gene (as a further check for application of similar amounts of total mRNA). mGluR5 PCR product is 513 bp. Primer sequence for mGluR5 is FWD: 5'GTCTCCTGATGTCAAGTGGTT3'; REV: 5'GGACCACACTTCATCATCATC3'. (b) mGluR5/TBP expression ratio is virtually unaffected by fluoxetine treatment regardless of whether normal mice (left part of (b)) or mice that showed some signs of depression after exposure to chronic mild stress (CMS) (right part of (b)) were studied. Unpublished experiments by B. Li and L. Peng, using methodology similar to that used by Li et al. [22].

2.2.7. Glycogenolysis and Glycogen Synthesis. An increase in $[Ca^{2+}]_i$ is a prerequisite for stimulation of glycogenolysis, which is consistent with the reduced or abrogated ability of fluoxetine to increase $[Ca^{2+}]_i$ after 1 week of treatment with 10 μ M fluoxetine. However, after treatment with this concentration for 2-3 weeks acute administration of fluoxetine causes a larger increase in glycogenolysis than in control cultures [8] as shown in Figure 6. This is a very long treatment with a very high concentration, and it cannot be excluded that this finding is an artifact. However, another possibility is that it is a consequence of the late upregulation of the 5-HT_{2B} receptor. If this is the case, the inability of transmitters to increase $[Ca^{2+}]_i$ might also be reversed after longer treatment. However, the effects of the gene changes of ADAR2, cPLA₂, and GluK2 are probably not further altered.

AKT phosphorylation by serotonin or 10 μ M fluoxetine is also impaired or abolished after 3 days of treatment with 10 μ M fluoxetine (B. Li and L. Peng's unpublished experiments) but may similarly recover with an increased response after 2 weeks [5]. Since GSK3 β is phosphorylated by AKT (Figure 1), this might imply that glycogen synthesis is also increased after treatment with fluoxetine for a sufficiently long period. The importance of GSK3 β is shown by observations that reduced immobility in the forced swim test (a sign a fear reduction) occur after administration of GSK3 inhibitors to normal animals and in GSK3 β haploinsufficient mice and that a selective GSK3 β inhibitor can alter serotonin-associated behavioral phenotypes in tests evaluating 5-HT-related antidepressant and anxiolytic drug effects (reviewed in [81]). More recently, Liu et al. [82] found that rats exposed to chronic mild stress showed depression-like behaviors and

decreased levels of phosphorylated GSK3 β in the hippocampus. Chronic citalopram treatment alleviated the depression-like behaviors and reversed the disruptions of the phosphorylated GSK3 β in these animals (Figure 7). In contrast, Karege et al. [83] reported decrease in phosphorylated GSK3 β protein and a reduced ratio between phosphorylated and total GSK3 β in postmortem brain tissue from patients having suffered from major depression.

2.2.8. Conclusion. Chronic treatment with fluoxetine, a 5-HT_{2B} agonist (confirmed by Diaz et al. [84]), exerts a multitude of effects on astrocytes, which in turn modulate astrocyte-neuronal interactions and brain function. These are to a large extent due to an upregulation and editing of genes, with rapid editing but late upregulation of the 5-HT_{2B} receptor gene itself. These alterations induce alteration in effects of cPLA₂, GluK2, and the 5-HT_{2B} receptor, probably including increases in both glucose metabolism and glycogen turnover, which in combination have therapeutic effect on major depression. The fact that gene effects are involved probably contributes to the late manifestation of the therapeutic effect of SSRIs. Moreover, the ability of increased levels of extracellular K⁺ to increase $[Ca^{2+}]_i$ is increased as a sign of increased K⁺-induced excitability in astrocytes and therefore probably also of an enhanced ability of elevated K⁺ to stimulate glycogenolysis.

3. Effects of Antibipolar Drugs

3.1. Effects Shared by Different Antibipolar Drugs. The three classical drugs that have effect against bipolar disorder, and

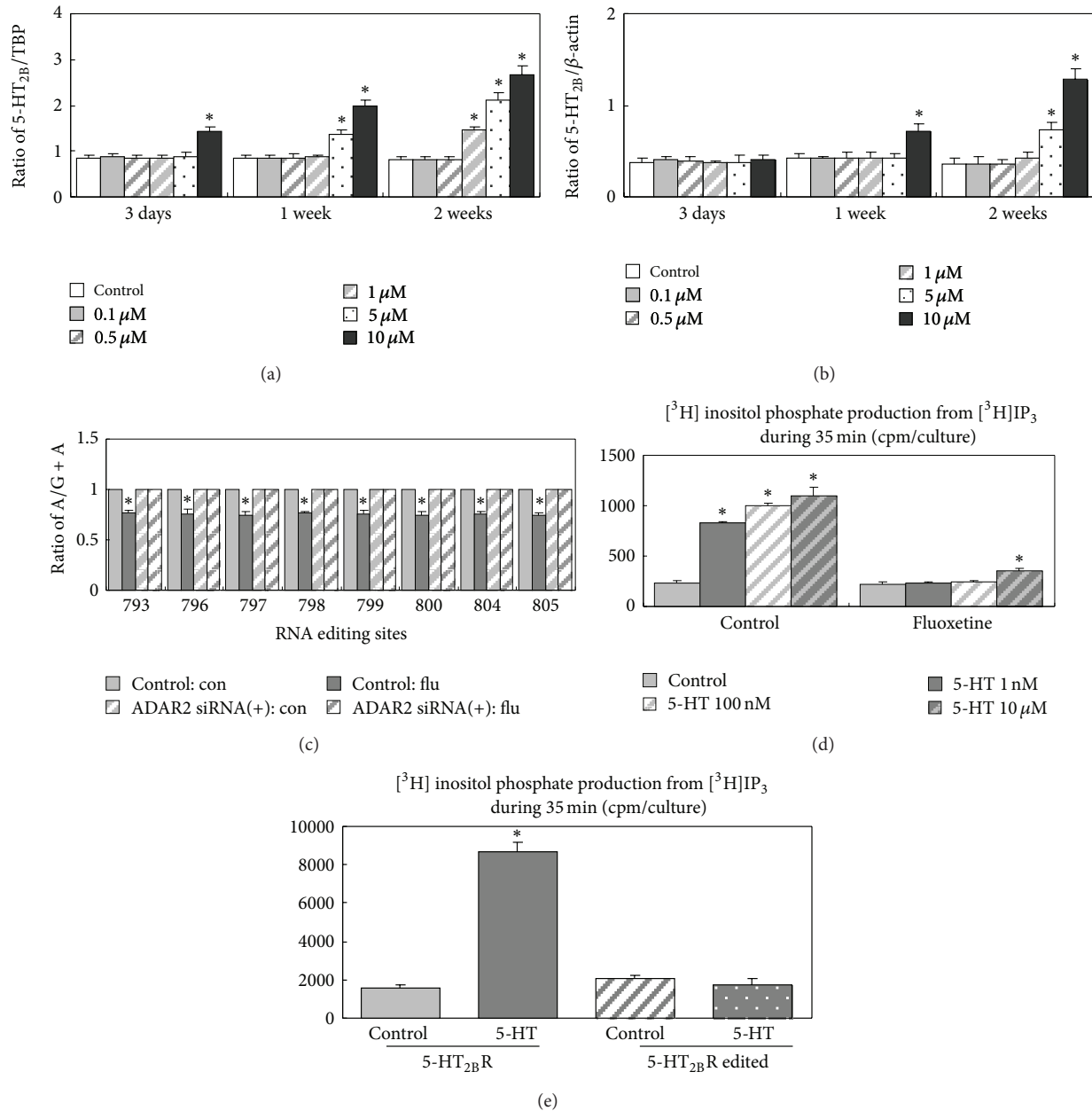


FIGURE 4: ((a), (b)) Time course for upregulation of 5-HT_{2B} receptor mRNA (a) and protein (b) during treatment of cultured mouse astrocytes with different concentrations of fluoxetine. (c) Editing of 5-HT_{2B} receptor after 3 days of treatment with 10 μM fluoxetine. ((d), (e)) Reduction of effect of 5-HT_{2B} receptor stimulation after downregulation of cultured astrocytes and transfected COS-7 cells with 10 μM fluoxetine for 7 days. Unpublished experiments by B. Li and L. Peng. Methodologies for (a)–(c) were as in Li et al. [22]. Response of the receptor to serotonin was measured as increase in the ability of serotonin to evoke release of ³H-inositol phosphate (IP) from labeled IP₃ in cultured astrocytes (d) and in cos-7 cells infected with receptor plasmids of either normal 5-HT_{2B} receptors or receptors with 8 RNA sites RNA (e). Unpublished experiments by B. Li and L. Peng.

especially mania, are the lithium ion “lithium”, carbamazepine, and valproic acid. A common feature of these three drugs is that they, like SSRIs, must be administered for a couple of weeks for the therapeutic action to become manifest. In studies of mechanism(s) of action for antibipolar drugs, it is therefore again important to determine effects that only appear after chronic administration. Moreover, the mechanisms of drug action may with advantage be elucidated

by studying shared effects of all three classical antibipolar drugs.

A common effect of chronic treatment with these otherwise very different drugs is a gradual development of intracellular alkalization in astrocytes, caused by stimulation of acid extruders. Lithium [85] increases intracellular pH (Figure 8(a1)) by directly stimulating the Na⁺/H⁺ exchanger NHE1, which leads to a compensatory decrease in its gene

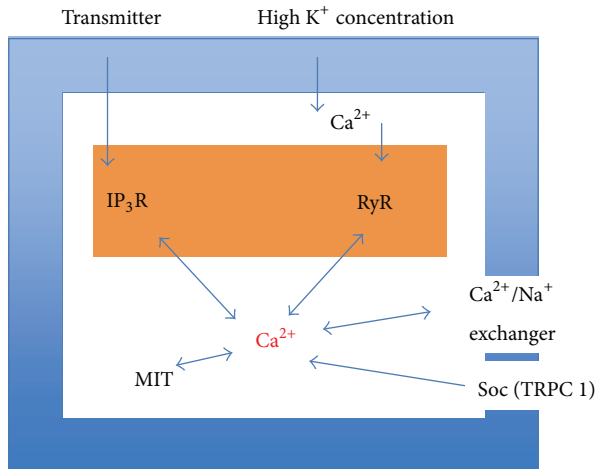


FIGURE 5: Aspects of Ca^{2+} homeostasis in astrocytes discussed in this paper. Many transmitters increase $[\text{Ca}^{2+}]$ in astrocytes by triggering release of intracellularly bound Ca^{2+} (brown) by stimulation of inositol trisphosphate (IP_3) receptors. Highly elevated extracellular K^+ concentrations (≥ 15 mM) cause L-channel-mediated Ca^{2+} entry, and additional Ca^{2+} is released by stimulation of Ca^{2+} -activated ryanodine receptors (RyR). Free intracellular Ca^{2+} (red) is accumulated by ER or mitochondria or leaves the cell via a $\text{Ca}^{2+}/\text{Na}^+$ exchanger. This creates a need for Ca^{2+} entry via store-operated channels (Socs), of which TRPC1 is an important component in astrocytes.

expression (Figure 8(a2)). Carbamazepine and valproic acid upregulate the expression and thus the function of the Na^+ /bicarbonate cotransporter NBCe1 [85, 86, 89] in astrocytes but not in neurons, as seen from Figure 8(b1). However, the carbamazepine treatment increases NHE1 expression in neurons, but not in astrocytes Figure 8(b2). It is likely that pH becomes increased in extracellular fluid as a result of the increased astrocytic NBCe1 function induced by carbamazepine. The neuronal increase in NHE1 expression may therefore be a defense against the development of a resulting neuronal acidosis. The intracellular astrocytic alkalinization may be the cause of an upregulation of cPLA₂ in astrocytes shown in Table 1, since the activity of this enzyme increases with rising pH [90]. This effect is similar to that seen after chronic fluoxetine administration and may have similar consequences, including stimulation of glucose metabolism. However, cPLA₂ is downregulated in neurons [86] and in whole brain [91] after carbamazepine treatment, which has contributed to the probably exaggerated concept of a role in neuroinflammation in bipolar disorder [57]. In contrast to the effect of fluoxetine, GluK2 expression is downregulated. Some potential effects of this downregulation are discussed by Song et al. [86], and it would be important to know the detailed differences between the consequences of the downregulated GluK2 function and the function of the upregulated, but edited GluK2 observed after chronic treatment with fluoxetine. The most important consequence of the intracellular alkalosis is probably the effect it has on *myo*-inositol uptake and thereby on phosphatidylinositide signaling. Moreover, antibipolar drugs have effects on ion homeostasis: Na^+ and K^+ by altering expression of Na^+ , K^+ -ATPase

subtype expression, and Ca^{2+} by downregulating TRPC1, apparently with no concomitant upregulation of $\text{Ca}_v1.2$, since K^+ effects are also reduced or abrogated as shown below.

3.2. Inositol Uptake and Metabolism: Transporters and pH Effects

3.2.1. Depletion of *myo*-Inositol. A classical proposition, the Berridge hypothesis for lithium's therapeutic effects in bipolar disorder is that *myo*-inositol is depleted by lithium [92]. Transmitter-mediated phospholipase C (PLC) stimulation hydrolyzes phosphatidyl-4,5-bisphosphate (PIP_2) in the cell membrane. This produces two second messengers, the cytosolic IP_3 and the membrane-associated 1,2-diacylglycerol (DAG). IP_3 triggers Ca^{2+} release from intracellular Ca^{2+} stores via stimulation of the IP_3 receptor, and DAG activates protein kinase C. Subsequently, IP_3 is stepwise dephosphorylated, eventually to *myo*-inositol by inositol phosphatases. *myo*-Inositol cannot resynthesize PIP_2 without contribution from a DAG metabolite. Initially, DAG is converted to phosphatidic acid (PA), an ester between a diglyceride and phosphoric acid, which condenses with cytidine triphosphate to cytidine monophosphoryl-phosphatidate (CMP-PA), which combines with *myo*-inositol to form PIP_2 via phosphatidyl-4-monophosphate (PIP) [93, 94]. According to the "Berridge hypothesis," inhibition of the regeneration of PIP_2 by lithium-induced reduction of inositol formation from IP decreases renewed responses to PLC-linked receptor agonists. It is consistent with this concept that a normally occurring noradrenaline-induced $[\text{Ca}^{2+}]_i$ increase in cultured astrocytes is inhibited by chronic exposure to lithium [87] (Figure 9). Moreover, in nonmedicated bipolar patients a significant increase in local *myo*-inositol concentration was recently shown by NMR [95]. However, the lithium effect on inositol phosphate hydrolysis to free *myo*-inositol is not shared by either carbamazepine or valproic acid. The original Berridge hypothesis is therefore insufficient to explain effects by anti-bipolar drugs, but a modified and expanded Berridge-like hypothesis will be presented as follows.

3.2.2. Cellular Contents of *myo*-Inositol. The concentration of inositol in plasma is 30–60 μM [96, 97], and the intracellular concentration is at a low millimolar level [98]. Accordingly, there is a steep gradient between extracellular and intracellular *myo*-inositol levels, necessitating transport mechanisms for continuous resupply of *myo*-inositol, since *myo*-inositol and many of the PIP_2 metabolites are further degraded. This is partly effectuated by dietary uptake following a slow transfer across the blood-brain barrier [99] and partly by synthesis in brain, which occurs only in the vasculature [100]. Therefore, inhibition of cellular uptake would also deplete *myo*-inositol in astrocytes and neurons.

3.2.3. Inositol Transporters. Wolfson et al. [101] showed that treatment of cultured mouse astrocytes with 1 mM LiCl for 8 days reduced the cellular content of *myo*-inositol after exposure to 50 μM *myo*-inositol for 24 hours. Acute administration of lithium had no similar effect. Lubrich and van Calker [102] expanded this observation by demonstrating

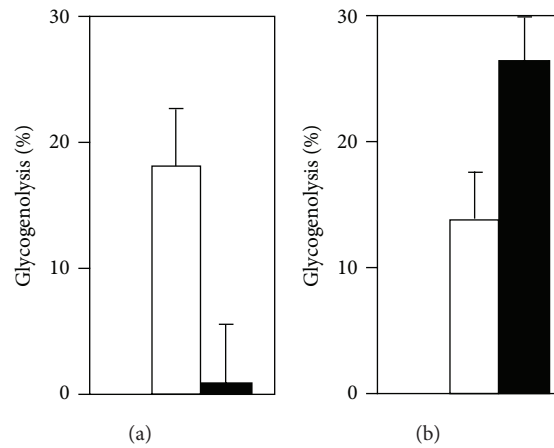


FIGURE 6: Effects on glycogenolysis (as percent of total glycogen) by 10 min acute exposure to $10 \mu\text{M}$ fluoxetine in cultured mouse astrocytes chronically treated (filled columns) with $10 \mu\text{M}$ fluoxetine for either 1 week (a) or for 2-3 weeks (b), compared to the effect of acute fluoxetine administration to similar untreated cultures from the same batches measured in the same experiments (open columns). In both (a) and (b) the treatment effect is significant ($P < 0.05$ or better). Glycogenolytic rates in (a) and (b) for the treated cultures were different at $P < 0.0005$, whereas there was no significant difference in the untreated cultures, from Kong et al. [8].

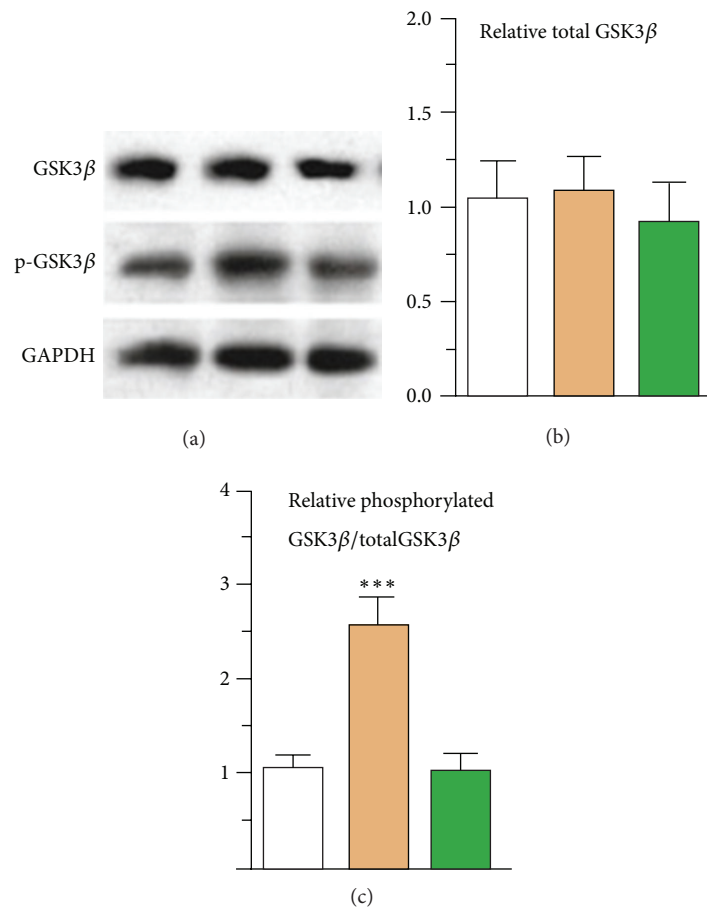


FIGURE 7: Treatment with citalopram (15 mg/kg) for 14 days (brown columns) has no effect on total GSK3 β expression in hippocampus (a,b), compared to that in untreated control animals (white columns), but dramatically increases its phosphorylation (c). This effect is abolished by sulindac, an inhibitor of the stimulated pathway (green columns), from Liu et al. [82], with permission.

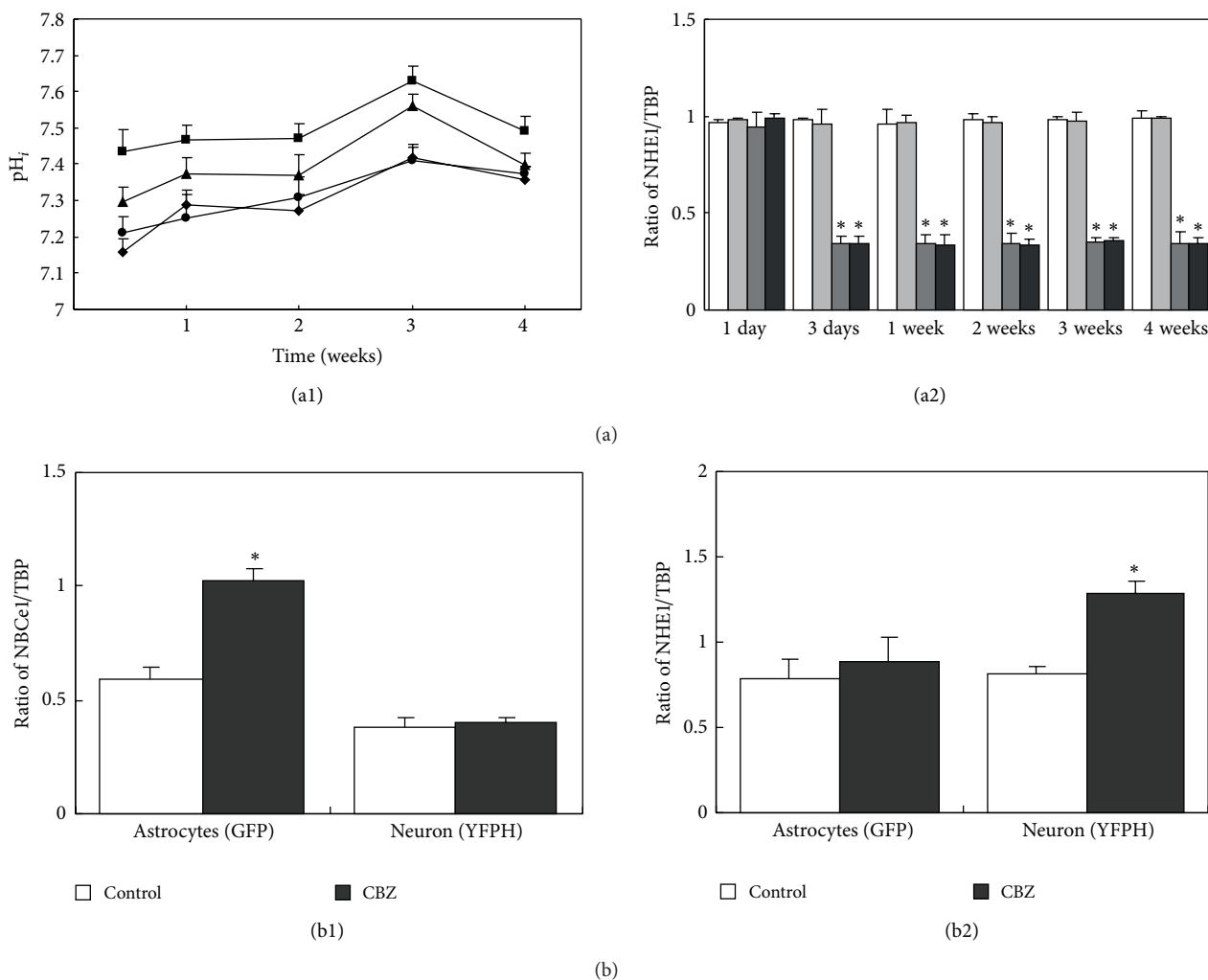


FIGURE 8: (a1) Effects of chronic treatment with different concentrations of lithium on intracellular pH measured as a function of the length of the treatment and the lithium concentration (diamonds: control; circles 0.5 mM Li^+ ; triangles 1 mM Li^+ ; or squares 2 mM Li^+). (a2) mRNA expression of NHE1 in control cultures (white); cultures treated with 0.5 mM Li^+ (light gray); 1 mM Li^+ (darker gray) or 2 mM Li^+ (black). * indicates statistically significant difference ($P < 0.05$ from control cultures and cultures treated with 0.5 mM Li^+ for the same length of time). (b) mRNA expression of the acid extruders NBCe1 and NHE1 in astrocytes and neurons from astrocytes, obtained from untreated animals or animals treated for 14 days with carbamazepine (CBZ). In both cases, astrocytes had been stained with GFP and neurons with YFPH in transgenic animals, and they were separated after cell dissociation by means of the different fluorescent signals. The size of the PCR products of NBCe1 is 298 bp, of NHE1 is 422 bp, and of TBP is 236 bp. The primers for NBCe1 were (FWD) 5'CTCACTTCTCCTGTGCTTGCCT3' and (REV) 5'GTGGTTGGAAAATAGCGGCTGG3', and those for NHE1 and TBP the same as used by Song et al. [85]. (a1) and (a2) from Song et al. [86], ((b1) and (b2)) Unpublished experiments by B. Li, D. Song, and L. Peng, using methodology similar to that used by Li et al. [22].

that chronic treatment of cultured rat astrocytes with lithium, carbamazepine, or valproic inhibited *myo*-inositol uptake. Wolfson et al. [88] confirmed this observation in cultured human astrocytoma cells at a *myo*-inositol concentration of 50 μ M, but at 25 μ M *myo*-inositol, the uptake was stimulated (Figure 10). Such an effect can be explained by the combined effects of (i) an induced alkalinization, (ii) the presence of two different *myo*-inositol transporters in astrocytes, the high-affinity Na^+ -dependent transporter (SMIT), and the lower-affinity H^+ -dependent (HMIT), and (iii) stimulation of SMIT but inhibition of HMIT at increased pH [103, 104]. We have recently confirmed that uptake of *myo*-inositol in cultured astrocytes at normal extracellular *myo*-inositol

levels mainly is catalyzed by a lower-affinity, higher-capacity HMIT-mediated uptake (K_m 143 μ M and V_{max} 358 pmol/mg protein), whereas a higher-affinity, lower-capacity SMIT-mediated uptake (K_m 16.7 μ M and V_{max} 60.2 pmol/mg protein) plays a minor role [105]. Moreover, the uptake of 100 μ M *myo*-inositol was inhibited at increased intracellular pH, whereas that at 10 μ M *myo*-inositol was enhanced. The observed increase in uptake at 25 μ M *myo*-inositol but reduction of uptake at 50 μ M reported by Wolfson et al. [88] can therefore be explained by changes in relative contribution of the two *myo*-inositol transporters to total uptake on account of the induced intracellular alkalinization [85, 86]. Since di Daniel et al. [106] reported that HMIT in neurons

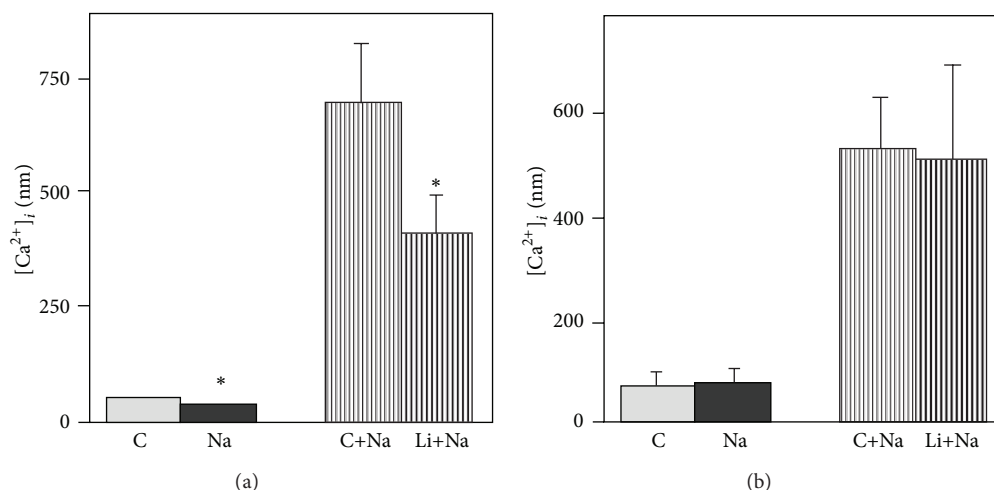


FIGURE 9: $[Ca^{2+}]_i$ in mouse astrocytes during basal conditions control (C) and during exposure to $1 \mu M$ noradrenaline (Na) in untreated control cultures and in sister cultures which had been treated for either 7–14 days (a) or 30–45 min (b) with 1 mM lithium chloride (Li). The chronic lithium treatment decreased $[Ca^{2+}]_i$ significantly ($*P < 0.05$) both under basal condition and during exposure to noradrenaline (a), whereas the short-lasting exposure to Li^+ had no effect, from Chen and Hertz [87].

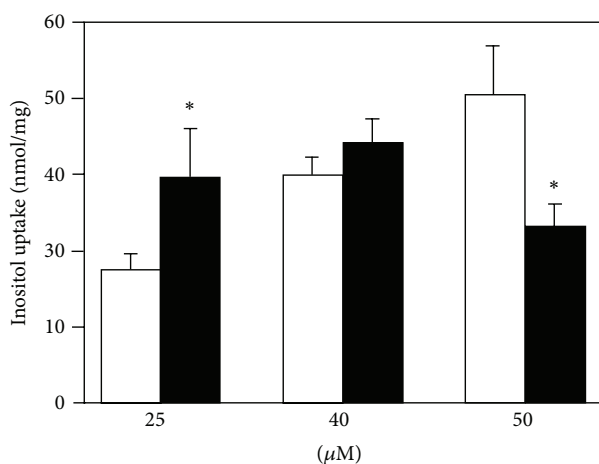


FIGURE 10: Uptake of $[^3H]$ myo-inositol at concentrations of 25, 40, and $50 \mu M$ during 60 min in U251 MG astrocytoma cells treated with 1 mM lithium chloride for 2 weeks before the uptake experiment as well as during the uptake (filled columns) and in untreated control cultures (open columns). Note lithium-induced decrease of uptake at $50 \mu M$ myo-inositol versus increase at $25 \mu M$. SEM values are shown by vertical bars. *Statistically significant difference between lithium-treated and control cultures ($P < 0.05$), from Wolfson et al. [88].

can transport IP_3 and Gossman and Zhao [107] and Wang et al. [108] found in the cochlea that IP_3 can be released from nonneuronal cells through hemichannels, it cannot be excluded that astrocytically generated IP_3 might be transferred to neurons. In that case, a decrease in astrocytic IP_3 formation in response to antibipolar drug treatment might also affect neuronal IP_3 . Effects of transmitters operating via the protein kinase C (PKC) and the phosphatidylinositide signaling system might therefore become reduced primarily in astrocytes but possibly also in neurons by antibipolar

drug treatment. Effects exerted via protein kinase A (PKA), like those of adrenaline acting on β -adrenergic receptors, may also be affected in astrocytes on account of a G_s/G_i shift in their pathway [109]. This might include β -adrenergic glycogenolysis, which is dependent on the increase in $[Ca^{2+}]_i$ occurring after this shift. Since β -adrenergic glycogenolysis is a prerequisite for neuronal glutamatergic signaling [12, 110], reduced glycogenolysis might have considerable dampening effect on neuronal excitability. Although $GSK3\beta$ is very often discussed in connection with this disease and its treatment, possible enhancement of glycogen synthesis seems not to have been considered or tested.

3.3. Ion Homeostasis. Like fluoxetine chronic treatment with any of the three antibipolar drugs downregulates TRPC1 and therefore interferes with Ca^{2+} homeostasis. However, as shown in Figure 11, the ability of elevated extracellular K^+ concentrations to increase $[Ca^{2+}]_i$ is also inhibited [76]. Thus, both transmitter-induced and depolarization-mediated changes in astrocytic $[Ca^{2+}]_i$ are reduced. This represents an important difference from astrocytes treated with an SSRI, where only the transmitter-mediated responses are inhibited [80], and the inhibition of the transmitter-induced effect might even be transient.

An effect on TRPC1 may affect not only Ca^{2+} homeostasis, but also Na^+ homeostasis in astrocytes, because the TRPC1 channel has equal permeability for Ca^{2+} and Na^+ [111]. This may be of major importance in astrocytes, since these nonexcitable cells may become deficient in intracellular Na^+ required for Na^+ , K^+ -ATPase activity [14]. Paradoxically, antibody-mediated partial inactivation of TRPC1's Ca^{2+} permeability is accompanied by an increase of the intracellular concentration of Na^+ , suggesting that the binding of the antibody to the channel decreases Ca^{2+} flux but increases Na^+ flux [112]. Findings of decreased Na^+ , K^+ -ATPase activity in

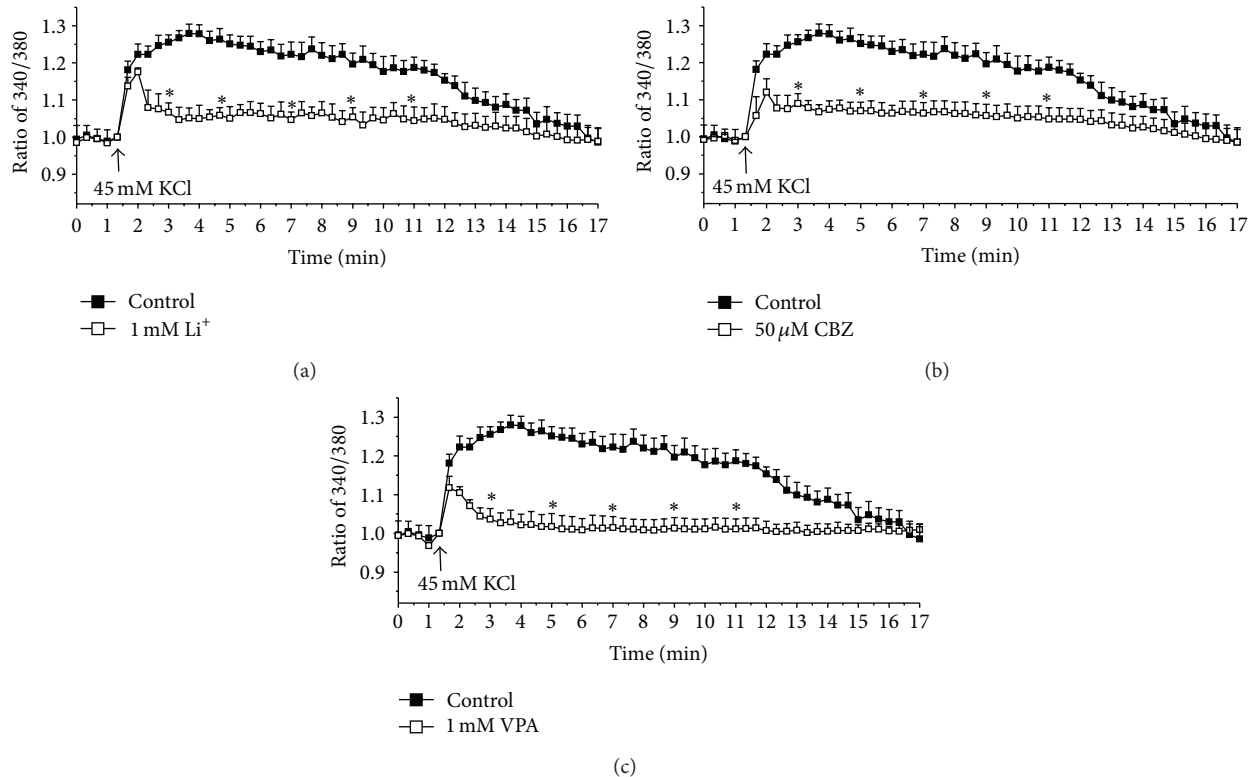


FIGURE 11: $[\text{Ca}^{2+}]_i$ in astrocytes in arbitrary units (fluorescence ratio) in response to addition of 45 mM KCl in control cultures and in cultures treated during 14 days with 1 mM lithium (a), 50 mM carbamazepine (CBZ) (b), or 1 mM valproic acid (VPA) (c), from Yan et al. [76].

bipolar disorder are robust: (i) in brain tissue from bipolar disorder a significantly lower Na^+ , K^+ -ATPase density was found in tissues from patients having suffered from major depression or schizophrenia [113]; (ii) expression of the $\alpha 2$ gene is decreased in temporal cortex from bipolar disorder patients [114]; (iii) Na^+ - K^+ -ATPase activity is significantly reduced in erythrocytes from patients with bipolar disorder [115], and (iv) lymphoblastoid cell lines originating from bipolar patients express less Na^+ , K^+ -ATPase in response to an increase in intracellular Na^+ concentration than similar cell lines from unaffected siblings. A possible genomic involvement of Na^+ , K^+ -ATPase dysfunction in bipolar disorder is also suggested by a genetic association between bipolar disorder and variants of the genes encoding the Na^+ , K^+ -ATPase subunits $\alpha 1$, $\alpha 2$, and $\alpha 3$ [116]. Brains in bipolar patients show increased brain acidity in vivo [117]. This might have influenced Na^+ , K^+ -ATPase activity and expression, since Deigweier et al. [118] demonstrated that in cuttlefish gill tissues increases in water CO_2 tension, which must decrease pH, led to a decline in Na^+ , K^+ -ATPase activity. Previously, Cummins and Hydén [119] had shown a pH maximum around 8.0 for ATP hydrolysis in microdissected astrocytes. Based on all this evidence, we investigated whether chronic treatment with carbamazepine could alter expression of Na^+ , K^+ -ATPase subtypes and of its auxiliary protein FXYD in neurons and astrocytes of normal mice [120]. Mice which coexpress one fluorescent marker with a neuron-specific gene and another marker with an astrocyte-specific gene [86] were either treated with carbamazepine

for 2 weeks or received no treatment but were used as controls. From Figure 12, it can be seen that $\alpha 3$ expression was unchanged and $\alpha 2$ increased in the treated animals both in astrocytes and in neurons, where it is normally not expressed, whereas $\alpha 1$ increased only in neurons. FXYD expression was reduced by the carbamazepine treatment, but only in neurons, and $\beta 1$ was upregulated in astrocytes, but not in neurons (results not shown). These changes should facilitate K^+ uptake in neurons (see discussion in [120]), without compromising preferential uptake in astrocytes at increased extracellular K^+ concentrations, a process which is important for K^+ homeostasis of the cellular level of the brain [14]. Although an overall increase in Na^+ , K^+ -ATPase expression and presumably activity could be related to pH changes, the induced isoform expressions are not a direct consequence of the intracellular pH changes, since $\alpha 2$ was upregulated in both neurons and astrocytes, and $\alpha 1$, which shows a slight increase in activity at increased pH [121], showed increased expression in neurons but not in astrocytes. However, Na^+ , K^+ -ATPase activity, and thus expression, is regulated by a multitude of different factors, many of which may have been altered in the carbamazepine-treated mice [122].

3.4. Conclusion. The intracellular alkalinization induced in astrocytes by the three otherwise very dissimilar drugs lithium, carbamazepine, and valproic acid suggests that this effect is therapeutically important. If anything, this concept is strengthened by the finding that lithium acts on one acid extruder and carbamazepine and valproic acid on a different

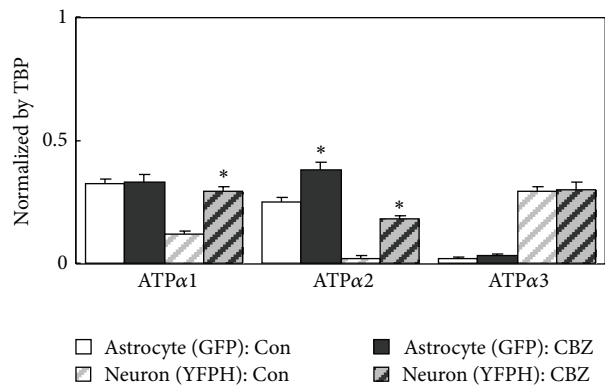


FIGURE 12: Expression of mRNA for the Na⁺, K⁺-ATPase subunits α 1, α 2, and α 3, expressed as ratios between mRNA of the subtype gene and TATA-binding protein (TBP) used as a housekeeping gene in astrocytes and neurons of mice treated for 14 days with carbamazepine (25 mg/kg per day). As in Figures 3 and 8(b), the astrocytes had been stained with GFP, and the neurons with YPHF in transgenic animals, and they were separated after cell dissociation by means of the different fluorescent signals, from Li et al. [120].

acid extruder, and by the finding of increased intracellular acidity in the brains of bipolar patients. In contrast to the ability of chronic treatment with antidepressant medication to enhance the increase of $[Ca^{2+}]_i$ and thus probably glycogenolysis by exposure to elevated extracellular K⁺ concentrations (and after sufficiently long treatment perhaps also to transmitters), chronic treatment with antibipolar drugs inhibit both K⁺-induced and transmitter-induced increase of astrocytic $[Ca^{2+}]_i$ and thereby probably excitability. The effect on astrocytic cPLA₂ expression is similar to that after antidepressant treatment and may have similar beneficial effects, but the effect on the neuronal enzyme is the opposite. The expression of GluK2, which is upregulated (and edited) by antidepressant therapy, is downregulated, but the therapeutic consequences have not been determined with certainty. Further studies of GluK2 function in brain and drug effects on this kainate receptor under normal and pathological conditions are urgently needed. Together with the opposite effect of SSRIs and antibipolar drugs on the ability of elevated extracellular K⁺ concentrations to affect astrocytic $[Ca^{2+}]_i$, they may constitute important differences in the directions of changes caused by the antidepressant SSRIs and the mainly antimanic antibipolar drugs.

4. Acute Anxiolytic Drug Effects

4.1. GABA_A Receptor Stimulation. Because of a high intracellular Cl⁻ concentration in astrocytes [123, 124], GABA_A receptor-mediated increase of Cl⁻ fluxes causes a depolarization, resulting in GABA_A-induced $[Ca^{2+}]_i$ increases [124, 125] and in glycogenolysis (Figure 13). These effects are strikingly similar to those seen after chronic treatment with fluoxetine and are consistent with the observation that GABA_A but not GABA_B receptor stimulation has anxiolytic-like effects in rats tested in the elevated-plus-maze [126].

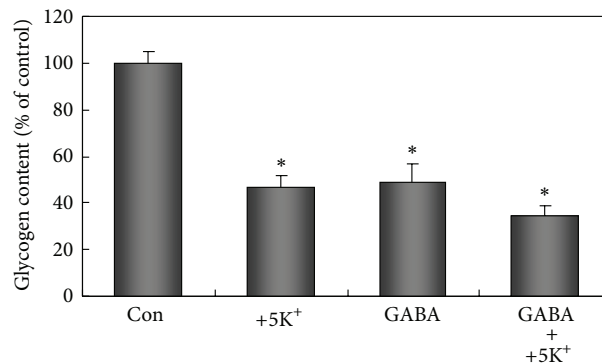


FIGURE 13: Glycogenolytic effect of GABA and/or addition of 5 mM K⁺, indicated as reduction of glycogen content, in astrocytes. Cultured astrocytes were incubated for 20 min in DMEM (containing 7.5 mM glucose) without any addition (control), addition of 5 mM K⁺ to a final extracellular concentration of 10 mM (+5 K⁺), of γ -aminobutyric acid (GABA) to a final concentration of 100 μ M (GABA), or of GABA and K⁺ (GABA plus +5 K⁺). After the incubation, the astrocytes were washed three times with ice-cold phosphate-buffered saline (PBS) and sonicated in 30 mM HCl. The suspension was used to measure nonhydrolyzed glycosyl units of glycogen. Three 50 μ L aliquots were sampled. In the first aliquot, 150 μ L of acetate buffer (0.1 M, pH 4.65) was added. In the second, 150 μ L of a solution containing 1% amyloglucosidase (10 mg/mL) in the acetate buffer was added in order to degrade remaining glycogen to glucose, and the mixture was incubated at room temperature for 30 min. Subsequently, the two aliquots were treated identically. Two mL of Tris-HCl buffer (0.1 M, pH 8.1) containing 3.3 mM MgCl₂, 0.2 mM ATP, 25 μ g/mL NADP, 4 μ g/mL hexokinase, and 2 μ g/mL glucose-6-phosphate dehydrogenase was added to each, and the mixture was incubated at room temperature for 30 min. The fluorescence of the NADPH formed in amounts equivalent to glucose metabolized by hexokinase was then read (excitation 340 nm; emission 450 nm). The first aliquot measures the sum of glucose and glucose-6-phosphate in the tissue, whereas the second aliquot in addition to those also measures the glycosyl units from glycogen remaining in the tissue. Determination of the difference between these two aliquots provides a measurement of the amount of the latter. The third aliquot was used to measure the protein content by the Lowry method to normalize the glycogen contents (nmol) per mg protein. Average glycogen contents were indicated as percentages of those under control conditions. All values were expressed as means \pm S.E.M indicated by vertical bars and were from three-five individual cultures. *Statistically significant ($P < 0.05$) difference from control. Results are unpublished experiments by J. Xu, D. Song, L. Hertz, and L. Peng.

Recent publications suggest that the role of GABA effects on astrocytes is currently greatly underestimated [124, 127]. This is in spite of astrocytic subunit composition of the GABA_A receptor indicative of an anxiolytic role.

GABA_A receptors are made up of α , β , and γ subunits, each of which can be further subdivided. In the present context, the γ 1 subunit, which is densely expressed in basal ganglia [130] but scarcely in cortex, may be of special interest. A comparison between mRNA expression levels of different GABA_A receptor subunits in freshly isolated astrocytes and neurons [131] showed that >90 percent of β 1 cortical mRNA

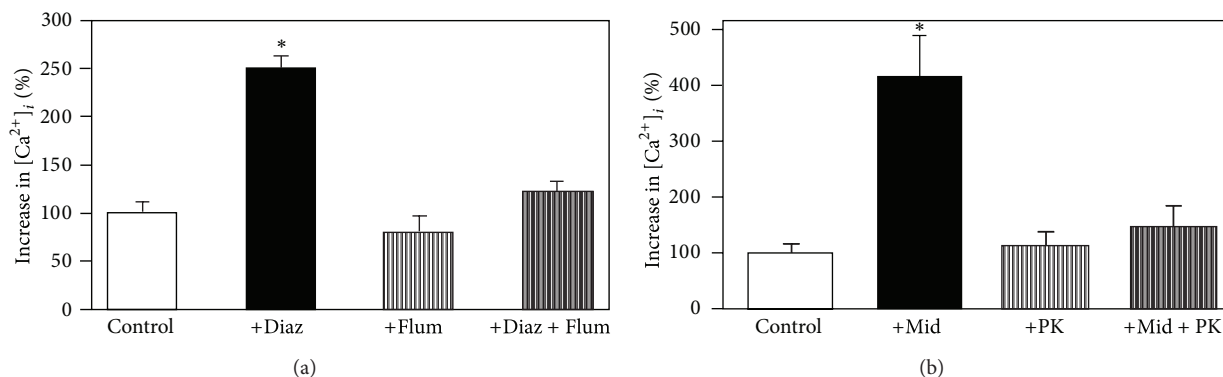


FIGURE 14: (a) Effects of 500 nM diazepam (+Diaz.), of 1 μ M of the “neuronal-type benzodiazepine antagonist” flumazenil (+Flum), and of diazepam plus flumazenil (+Diaz +Flum) on the increase in $[Ca^{2+}]_i$ evoked by an increase in the extracellular K^+ concentration to 20 mM. The control value (100%) represents the increase by the elevated K^+ concentration alone, which approximately doubled resting $[Ca^{2+}]_i$ (about 100 nM). Diazepam more than doubles the response to the increase in extracellular K^+ , and this effect is abrogated by flumazenil. The value in the presence of diazepam is statistically significantly different from all other values, none of which differs from any of the other. (b) Effects of 10 nM midazolam (+Mid.), of 1 μ M of the “mitochondrial benzodiazepine antagonist” PK 11195 (+PK), and of midazolam plus PK 11195 (+Mid. +PK) on the increase in $[Ca^{2+}]_i$ evoked by an increase in the extracellular K^+ concentration to 20 mM. Midazolam almost quadruples the response to the increase in extracellular K^+ , and this effect is abrogated by PK 11195. The value in the presence of midazolam is statistically significantly different from all other values, none of which differs from any of the other, from Hertz et al. [128].

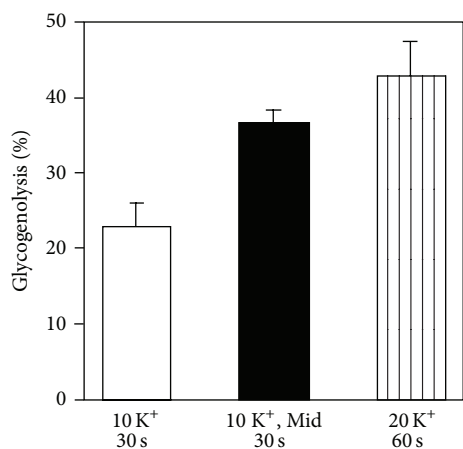


FIGURE 15: Release of previously incorporated label in glycogen during exposure to 10 mM extracellular K^+ for 30 sec in the presence and absence of 20 nM midazolam and to 20 mM K^+ without midazolam for 60 sec. In the presence of midazolam, glycogenolysis during exposure to 10 mM K^+ for 30 sec is significantly increased and its magnitude becomes similar to that seen after 1 min of exposure to 20 mM K^+ alone, from Subbarao et al. [129].

was astrocytic, and astrocytes also accounted for ~70 percent of $\alpha 2$ expression, ~60% of $\beta 1$, and ~40% of $\alpha 4$ (Table 2). Since limbic areas are enriched in the $\gamma 1$ subunit, GABA_A receptors containing this subtype might serve affective functions [132], and the $\alpha 2$ subunit is known to mediate anxiolysis [133]. Future attempts to develop anxiolytics acting on GABA_A receptors [133] should include potential effects on astrocytes.

4.2. Effects of Benzodiazepines: Emphasis on a Membrane Receptor.

It was initially believed that all anxiolytic effects

TABLE 2: Percentage astrocytic expression (astrocytic expression/(astrocytic + neuronal expression)) of GABA_A receptor subunits as found by Cahoy et al. [131] in astrocytic and neuronal cell fractions obtained from murine brain*.

GABA _A receptor subunit	Percentage astrocytic expression
$\alpha 1$	0
$\alpha 2$	69.6 \pm 3.65
$\alpha 4$	43.9 \pm 2.15
$\alpha 5$	0
$\beta 1$	61.4 \pm 8.73
$\beta 3$	0
$\gamma 1$	92.6 \pm 0.50
$\gamma 2$	0

*No information was provided about $\beta 2$.

of benzodiazepines were exerted on the neuronal “central” benzodiazepine receptor [134], which facilitated GABAergic transmission by binding to its receptor. Since this effect was abrogated by the benzodiazepine antagonist flumazenil, it was often concluded that all benzodiazepine effects that were inhibited by this antagonist were exerted on the “central” benzodiazepine receptor. However, the peripheral-type benzodiazepine receptor localized in mitochondria, the translocator protein (18 kDa) [135], is also important in anxiolysis [136]. It is stimulated not only by exogenously administered benzodiazepines but also by derivatives of the 86-amino acid polypeptide diazepam-binding inhibitor (DBI), which is found in brain [137]. It can be cleaved into several biologically active peptides, including the triakontatetrapeptide, TTN, and the octadecaneuropeptide ODN [138]. Gandolfo et al. [139] showed (i) that binding of the peripheral-type benzodiazepine receptor ligand [³H]Ro5-4864 to intact astrocytes

cultures was displaced by TTN, whereas ODN did not compete for [^3H]Ro5-4864 binding and (ii) that TTN provoked a concentration-dependent increase in $[\text{Ca}^{2+}]_i$ in the cultured astrocytes, which was blocked by chelation of extracellular Ca^{2+} by EGTA or blockage of Ca^{2+} channels with Ni^{2+} and significantly reduced by the L-type calcium channel blocker nifedipine. Patch-clamp studies showed that TTN induced a sustained depolarization, and the authors suggested that TTN acted through the peripheral benzodiazepine binding site and opening of Cl^- channels, that is, a mechanism similar to that of stimulation of GABA_A receptors.

It has long been known—and virtually ignored—that peripheral benzodiazepine receptors are present not only in mitochondria, but also at the plasma membrane of astrocytes [140–142]. In addition, the peripheral benzodiazepine binding site shows multiplicity and has also been demonstrated on a plasma membrane-enriched preparation from rat astrocytes [143]. Backus et al. [140] suggested the same mechanism for Ca^{2+} entry as Gandolfo et al. [139], that is, a depolarizing effect. In contrast, Bender and Hertz [141] and Zhao et al. [142], using cultures which had been differentiated by treatment with dibutyryl cyclic AMP and therefore express L-channels, suggested a direct interaction between the benzodiazepine and this channel. It seems consistent with this interpretation that midazolam had no significant effect at nonelevated extracellular K^+ concentrations in the experiments by Zhao et al. [142]. More conclusive evidence has been reached in cardiac cells, where direct activation of the channel by the Ca^{2+} channel activator BAY K 8644 was blocked by the benzodiazepine antagonist PK11195 [144, 145]. Nevertheless, the most important finding, an increase in astrocytic $[\text{Ca}^{2+}]_i$, is identical in all these studies.

As shown in Figures 14(a) and 14(b), the benzodiazepine-mediated augmentation of the response to elevated K^+ concentrations is selectively prevented in the presence of $1\ \mu\text{M}$ of the “peripheral-type” benzodiazepine antagonist PK 11195 and notably also by the supposedly “central-type” antagonist flumazenil [128, 142, 146]. The inhibition by flumazenil confirms previous observation by Backus et al. [140] and emphasizes again that inhibition of a benzodiazepine response by flumazenil does *not* necessarily indicate that the response is exerted on neuronal receptors. The dihydropyridine L-channel blocker nimodipine abolishes both the effect of elevated K^+ and that of the benzodiazepine [142], although it should have no effect on the latter if only an L-channel-independent depolarization was involved. Figure 15 shows that midazolam also increases the glycogenolytic effect of elevated extracellular K^+ concentrations [129]. The high potency of benzodiazepines in increasing L-channel opening in astrocytes (500 nM diazepam and 10 nM midazolam in Figure 14) is an indication that the response may be relevant for clinically observed benzodiazepine effects, because the effective concentrations are similar to those obtained clinically [147, 148]. In contrast, stimulation of GABA -mediated Cl^- influx in cortical neurons by 100–1000 nM diazepam causes only a 5–40% increase [149]. Peripheral-type receptors are also present on several additional, nonneural cell types as discussed by Gandolfo et al. [139], and mature erythrocytes, which have

no mitochondria, express peripheral-type benzodiazepine receptors [150]. These sites should *not* be identified as translocator proteins and have been referred to as the “Joker” receptor by Hertz et al. [128] and Hertz and Chen [146].

Yet another indication that drugs increasing $[\text{Ca}^{2+}]_i$ and causing glycogenolysis can provide acute anxiolytic effects is that acute treatment with the 5-HT_{2B} receptor agonist, BW 723C86, has been found to exert anxiolytic effects in animal experiments [151–153]. This is the same receptor as that activated by fluoxetine and other SSRIs, and acute administration of fluoxetine increases $[\text{Ca}^{2+}]_i$ and causes glycogenolysis [154]. Why some degree of anxiolysis can be obtained by acute 5-HT_{2B} activation, but the antidepressant effects require weeks of treatment and gene editing and up-regulation, remains to be shown.

4.3. Conclusion. Acute anxiolytic drug therapy induces Ca^{2+} -dependent anxiolysis and enhances glycogenolysis evoked by exposure to elevated extracellular K^+ concentrations, an effect which is strikingly similar to that of chronic treatment with an SSRI, which also has anxiolytic effect.

The response to GABA_A receptor stimulation is secondary to depolarization and that to benzodiazepines to a direct effect on the L-channel. One might wonder about the importance of L-channel opening, since it, in experiments where $[\text{K}^+]_e$ is the only factor changed, requires $[\text{K}^+]_e$ values above those usually seen in response to normal stimulation. However, it should be remembered that the $[\text{K}^+]_e$ is probably normally not changed alone but that the change might be accompanied by changes in transmitters, like GABA , that also contribute to the depolarization, as seen by the tendency to a more pronounced glycogenolysis in Figure 13. Accordingly, activation of Ca^{2+} uptake by elevated $[\text{K}^+]_e$ may occur much more frequently than should be anticipated based on the experiments in which $[\text{K}^+]_e$ alone is increased. This may also be of importance for the oppositely directed changes in effects of increase in $[\text{K}^+]_e$ after treatment with SSRIs and with antibipolar drugs.

5. Concluding Remarks

Astrocytic increase in $[\text{Ca}^{2+}]_i$ and in glycogenolysis after chronic treatment with SSRIs (which also have anxiolytic effects) and during acute GABA_A stimulation or administration of benzodiazepines is probably important for antidepressant-anxiolytic drug effects, and astrocytic glycogenolysis is essential to support astrocytic signaling. The glycogenolytic effect is unlikely to be shared by antibipolar drugs, which all cause intracellular alkalization in astrocytes, although by stimulation of different acid extruders. These drugs inhibit effects of both elevated K^+ concentrations and transmitters on astrocytic $[\text{Ca}^{2+}]_i$. Carbamazepine was also shown to have important effects on Na^+ , K^+ -ATPase in both astrocytes and neurons. Both antidepressant drugs and antibipolar drugs affect GluK2 , but the effects are different, and further studies of this kainate receptor are warranted in both major depression and bipolar disease. So is that of the astrocytic upregulation of cPLA_2 .

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgment

Mrs. Elna Hertz is cordially thanked for her very careful reading of the paper and correction of a multitude of typos and other errors.

References

- [1] H. K. Kimelberg, "Functions of mature mammalian astrocytes: a current view," *Neuroscientist*, vol. 16, no. 1, pp. 79–106, 2010.
- [2] L. Peng, C. Guo, T. Wang, B. Li, L. Gu, and Z. Wang, "Methodological limitations in determining astrocytic gene expression," *Frontiers in Endocrinology*, vol. 4, article 176, 2013.
- [3] D. Lovatt, U. Sonnewald, H. S. Waagepetersen et al., "The transcriptome and metabolic gene signature of protoplasmic astrocytes in the adult murine cortex," *The Journal of Neuroscience*, vol. 27, no. 45, pp. 12255–12266, 2007.
- [4] T. Du, C. Liang, B. Li, L. Hertz, and L. Peng, "Chronic fluoxetine administration increases expression of the L-channel gene $Ca_v1.2$ in astrocytes from the brain of treated mice and in culture and augments K^+ -induced increase in $[Ca^{2+}]_i$," *Cell Calcium*. In press.
- [5] L. Hertz, B. Li, D. Song et al., "Astrocytes as a 5-HT_{2B}-mediated, SERT-independent SSRI target, slowly altering depression-associated genes and functions," *Current Signal Transduction Therapy*, vol. 7, no. 1, pp. 65–80, 2012.
- [6] D. T. Wong, J. S. Horng, F. P. Bymaster, K. L. Hauser, and B. B. Molloy, "A selective inhibitor of serotonin uptake, Lilly 110140, 3 (p trifluoromethylphenoxy) n methyl 3 phenylpropylamine," *Life Sciences*, vol. 15, no. 3, pp. 471–479, 1974.
- [7] L. Hertz, F. Baldwin, and A. Schousboe, "Serotonin receptors on astrocytes in primary cultures: effects of methysergide and fluoxetine," *Canadian Journal of Physiology and Pharmacology*, vol. 57, no. 2, pp. 223–226, 1979.
- [8] E. K. C. Kong, L. Peng, Y. Chen, A. C. H. Yu, and L. Hertz, "Up-regulation of 5-HT_{2B} receptor density and receptor-mediated glycogenolysis in mouse astrocytes by long-term fluoxetine administration," *Neurochemical Research*, vol. 27, no. 1-2, pp. 113–120, 2002.
- [9] D. R. Cool, F. H. Lieblich, and V. Ganapathy, "Interaction of fluoxetine with the human placental serotonin transporter," *Biochemical Pharmacology*, vol. 40, no. 9, pp. 2161–2167, 1990.
- [10] S. Zhang, B. Li, D. Lovatt et al., "5-HT_{2B} receptors are expressed on astrocytes from brain and in culture and are a chronic target for all five conventional serotonin-specific reuptake inhibitors," *Neuron Glia Biology*, vol. 6, no. 2, pp. 113–125, 2010.
- [11] B. Li, S. Zhang, H. Zhang et al., "Fluoxetine-mediated 5-HT_{2B} receptor stimulation in astrocytes causes EGF receptor transactivation and ERK phosphorylation," *Psychopharmacology*, vol. 201, no. 3, pp. 443–458, 2008.
- [12] M. E. Gibbs, D. G. Anderson, and L. Hertz, "Inhibition of glycogenolysis in astrocytes interrupts memory consolidation in young chickens," *Glia*, vol. 54, no. 3, pp. 214–222, 2006.
- [13] L. F. Obel, M. S. Müller, A. B. Walls et al., "Brain glycogen—new perspectives on its metabolic function and regulation at the subcellular level," *Front Neuroenergetics*, vol. 4, no. 3, 2012.
- [14] J. Xu, D. Song, Z. Xue, L. Gu, L. Hertz, and L. Peng, "Requirement of glycogenolysis for uptake of increased extracellular K^+ in astrocytes: potential implications for K^+ homeostasis and glycogen usage in brain," *Neurochemical Research*, vol. 38, no. 3, pp. 472–485, 2013.
- [15] Y. Chen, L. Peng, X. Zhang, J.-U. Stolzenburg, and L. Hertz, "Further evidence that fluoxetine interacts with a 5-HT_{2C} receptor in glial cells," *Brain Research Bulletin*, vol. 38, no. 2, pp. 153–159, 1995.
- [16] M. E. Gibbs, D. Hutchinson, and L. Hertz, "Astrocytic involvement in learning and memory consolidation," *Neuroscience & Biobehavioral Reviews*, vol. 32, no. 5, pp. 927–944, 2008.
- [17] N. Embi, D. B. Rylatt, and P. Cohen, "Glycogen synthase kinase-3 from rabbit skeletal muscle. Separation from cyclic-AMP-dependent protein kinase and phosphorylase kinase," *European Journal of Biochemistry*, vol. 107, no. 2, pp. 519–527, 1980.
- [18] X. Li, W. Zhu, M.-S. Roh, A. B. Friedman, K. Rosborough, and R. S. Jope, "In vivo regulation of glycogen synthase kinase-3 β (GSK3 β) by serotonergic activity in mouse brain," *Neuropsychopharmacology*, vol. 29, no. 8, pp. 1426–1431, 2004.
- [19] E. Beurel, M. A. Mines, L. Song, and R. S. Jope, "Glycogen synthase kinase-3 levels and phosphorylation undergo large fluctuations in mouse brain during development," *Bipolar Disorders*, vol. 14, no. 8, pp. 822–830, 2012.
- [20] A. Schousboe, "Development of potassium effects on ion concentrations and indicator spaces in rat brain-cortex slices during postnatal ontogenesis," *Experimental Brain Research*, vol. 15, no. 5, pp. 521–531, 1972.
- [21] S. J. Tudhope, C.-C. Wang, J. L. Petrie et al., "A novel mechanism for regulating hepatic glycogen synthesis involving serotonin and cyclin-dependent kinase-5," *Diabetes*, vol. 61, no. 1, pp. 49–60, 2012.
- [22] B. Li, L. Dong, B. Wang, L. Cai, N. Jiang, and L. Peng, "Cell type-specific gene expression and editing responses to chronic fluoxetine treatment in the in vivo mouse brain and their relevance for stress-induced anhedonia," *Neurochemical Research*, vol. 37, no. 11, pp. 2480–2495, 2012.
- [23] B. Li, S. Zhang, H. Zhang, L. Hertz, and L. Peng, "Fluoxetine affects GluK2 editing, glutamate-evoked Ca^{2+} influx and extracellular signal-regulated kinase phosphorylation in mouse astrocytes," *Journal of Psychiatry and Neuroscience*, vol. 36, no. 5, pp. 322–338, 2011.
- [24] B. L. Bass, "RNA editing by adenosine deaminases that act on RNA," *Annual Review of Biochemistry*, vol. 71, pp. 817–846, 2002.
- [25] L. Valente and K. Nishikura, "ADAR gene family and A-to-I RNA editing: diverse roles in posttranscriptional gene regulation," *Progress in Nucleic Acid Research and Molecular Biology*, vol. 79, pp. 299–338, 2005.
- [26] Y. Kawahara, K. Ito, H. Sun, M. Ito, I. Kanazawa, and S. Kwak, "Regulation of glutamate receptor RNA editing and ADAR mRNA expression in developing human normal and Down's syndrome brains," *Developmental Brain Research*, vol. 148, no. 1, pp. 151–155, 2004.
- [27] G. Y. Sun, J. Xu, M. D. Jensen, and A. Simonyi, "Phospholipase A2 in the central nervous system: implications for neurodegenerative diseases," *Journal of Lipid Research*, vol. 45, no. 2, pp. 205–213, 2004.
- [28] B. Li, S. Zhang, M. Li, L. Hertz, and L. Peng, "Chronic treatment of astrocytes with therapeutically relevant fluoxetine concentrations enhances cPLA₂ expression secondary to 5-HT_{2B}-induced, transactivation-mediated ERK_{1/2} phosphorylation," *Psychopharmacology*, vol. 207, no. 1, pp. 1–12, 2009.

- [29] L. L. Lautens, X. G. Chiou, J. D. Sharp et al., "Cytosolic phospholipase A2 (cPLA₂) distribution in murine brain and functional studies indicate that cPLA₂ does not participate in muscarinic receptor-mediated signaling in neurons," *Brain Research*, vol. 809, no. 1, pp. 18–30, 1998.
- [30] M. A. Balboa and J. Balsinde, "Involvement of calcium-independent phospholipase A2 in hydrogen peroxide-induced accumulation of free fatty acids in human U937 cells," *The Journal of Biological Chemistry*, vol. 277, no. 43, pp. 40384–40389, 2002.
- [31] D. T. Stephenson, J. V. Manetta, D. L. White et al., "Phospholipase A2 in the central nervous system: implication for neurodegeneration diseases," *The Journal of Lipid Research*, vol. 46, pp. 205–213, 2004.
- [32] C. C. Felder, R. Y. Kanterman, A. L. Ma, and J. Axelrod, "Serotonin stimulates phospholipase A2 and the release of arachidonic acid in hippocampal neurons by a type 2 serotonin receptor that is independent of inositolphospholipid hydrolysis," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 87, no. 6, pp. 2187–2191, 1990.
- [33] B. D. Stout, W. P. Clarke, and K. A. Berg, "Rapid desensitization of the serotonin_{2c} receptor system: effector pathway and agonist dependence," *Journal of Pharmacology and Experimental Therapeutics*, vol. 302, no. 3, pp. 957–962, 2002.
- [34] Y. Qu, L. Chang, J. Klaff, R. Seemann, and S. I. Rapoport, "Imaging brain phospholipase A2-mediated signal transduction in response to acute fluoxetine administration in unanesthetized rats," *Neuropsychopharmacology*, vol. 28, no. 7, pp. 1219–1226, 2003.
- [35] S. I. Rapoport, "Brain arachidonic and docosahexaenoic acid cascades are selectively altered by drugs, diet and disease," *Prostaglandins Leukotrienes and Essential Fatty Acids*, vol. 79, no. 3–5, pp. 153–156, 2008.
- [36] M. C. Garcia and H.-Y. Kim, "Mobilization of arachidonate and docosahexaenoate by stimulation of the 5-HT_{2A} receptor in rat C6 glioma cells," *Brain Research*, vol. 768, no. 1–2, pp. 43–48, 1997.
- [37] N. Yu, J.-L. Martin, N. Stella, and P. J. Magistretti, "Arachidonic acid stimulates glucose uptake in cerebral cortical astrocytes," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 90, no. 9, pp. 4042–4046, 1993.
- [38] I. Allaman, H. Fiumelli, P. J. Magistretti, and J.-L. Martin, "Fluoxetine regulates the expression of neurotrophic/growth factors and glucose metabolism in astrocytes," *Psychopharmacology*, vol. 216, no. 1, pp. 75–84, 2011.
- [39] O. Sorg, L. Pellerin, M. Stolz, S. Beggah, and P. J. Magistretti, "Adenosine triphosphate and arachidonic acid stimulate glycogenolysis in primary cultures of mouse cerebral cortical astrocytes," *Neuroscience Letters*, vol. 188, no. 2, pp. 109–112, 1995.
- [40] L. Hertz, J. Xu, and L. Peng, "Glycogenolysis and purinergic 19 signaling," in *Glutamate and ATP at Interface of Metabolism and Signaling in the Brain*, V. Parpura, A. Schousboe, and A. Verkhratsky, Eds., Advances of Neurobiology, Springer. In press.
- [41] Y. Qu, L. Chang, J. Klaff, R. Seemann, D. Greenstein, and S. I. Rapoport, "Chronic fluoxetine upregulates arachidonic acid incorporation into the brain of unanesthetized rats," *European Neuropsychopharmacology*, vol. 16, no. 8, pp. 561–571, 2006.
- [42] J. S. Rao, R. N. Ertley, H. J. Lee, S. I. Rapoport, and R. P. Bazinet, "Chronic fluoxetine upregulates activity, protein and mRNA levels of cytosolic phospholipase A2 in rat frontal cortex," *The Pharmacogenomics Journal*, vol. 6, no. 6, pp. 413–420, 2006.
- [43] H.-J. Lee, J. S. Rao, L. Chang, S. I. Rapoport, and R. P. Bazinet, "Chronic N-methyl-D-aspartate administration increases the turnover of arachidonic acid within brain phospholipids of the unanesthetized rat," *The Journal of Lipid Research*, vol. 49, no. 1, pp. 162–168, 2008.
- [44] J. T. Little, T. A. Ketter, T. A. Kimbrell et al., "Venlafaxine or bupropion responders but not nonresponders show baseline prefrontal and paralimbic hypometabolism compared with controls," *Psychopharmacology Bulletin*, vol. 32, no. 4, pp. 629–635, 1996.
- [45] J. T. Little, T. A. Ketter, T. A. Kimbrell et al., "Bupropion and venlafaxine responders differ in pretreatment regional cerebral metabolism in unipolar depression," *Biological Psychiatry*, vol. 57, no. 3, pp. 220–228, 2005.
- [46] P. Videbeck, "PET measurements of brain glucose metabolism and blood flow in major depressive disorder: a critical review," *Acta Psychiatrica Scandinavica*, vol. 101, no. 1, pp. 11–20, 2000.
- [47] N. L. Rasgon, H. A. Kenna, C. Geist, G. Small, and D. Silverman, "Cerebral metabolic patterns in untreated postmenopausal women with major depressive disorder," *Psychiatry Research*, vol. 164, no. 1, pp. 77–80, 2008.
- [48] T. A. Kimbrell, T. A. Ketter, M. S. George et al., "Regional cerebral glucose utilization in patients with a range of severities of unipolar depression," *Biological Psychiatry*, vol. 51, no. 3, pp. 237–252, 2002.
- [49] M. S. Buchsbaum, W. Joseph, B. V. Siegel et al., "Effect of sertraline on regional metabolic rate in patients with affective disorder," *Biological Psychiatry*, vol. 41, no. 1, pp. 15–22, 1997.
- [50] H. S. Mayberg, S. K. Brannan, J. L. Tekell et al., "Regional metabolic effects of fluoxetine in major depression: serial changes and relationship to clinical response," *Biological Psychiatry*, vol. 48, no. 8, pp. 830–843, 2000.
- [51] S. H. Kennedy, K. R. Evans, S. Krüger et al., "Changes in regional brain glucose metabolism measured with positron emission tomography after paroxetine treatment of major depression," *The American Journal of Psychiatry*, vol. 158, no. 6, pp. 899–905, 2001.
- [52] M. Elizabeth Sublette, M. S. Milak, J. R. Hibbeln et al., "Plasma polyunsaturated fatty acids and regional cerebral glucose metabolism in major depression," *Prostaglandins Leukotrienes and Essential Fatty Acids*, vol. 80, no. 1, pp. 57–64, 2009.
- [53] C.-U. Pae, H.-S. Yu, J.-J. Kim et al., "BanI polymorphism of the cytosolic phospholipase A2 gene and mood disorders in the Korean population," *Neuropsychobiology*, vol. 49, no. 4, pp. 185–188, 2004.
- [54] K.-P. Su, S.-Y. Huang, C.-Y. Peng et al., "Phospholipase A2 and cyclooxygenase 2 genes influence the risk of interferon- α -induced depression by regulating polyunsaturated fatty acids levels," *Biological Psychiatry*, vol. 67, no. 6, pp. 550–557, 2010.
- [55] F. Chen, G. Wegener, T. M. Madsen, and J. R. Nyengaard, "Mitochondrial plasticity of the hippocampus in a genetic rat model of depression after antidepressant treatment," *Synapse*, vol. 67, no. 3, pp. 127–134, 2013.
- [56] R. E. Anglin, M. F. Mazurek, M. A. Tarnopolsky, and P. I. Rosebush, "The mitochondrial genome and psychiatric illness," *The American Journal of Medical Genetics B*, vol. 159, no. 7, pp. 749–759, 2012.
- [57] L. Hertz, D. Song, B. Li, E. Yan, and L. Peng, "Importance of "inflammatory molecules", but not necessarily of inflammation, in the pathophysiology of bipolar disorder and in the mechanisms of action of anti-bipolar drugs," *Neurology, Psychiatry and Brain Research*, vol. 19, no. 4, pp. 174–179, 2013.

- [58] D. Tharumaratnam, S. Bashford, and S. A. Khan, "Indomethacin induced psychosis," *Postgraduate Medical Journal*, vol. 76, no. 901, pp. 736–737, 2000.
- [59] M. Clunie, L.-A. Crone, L. Klassen, and R. Yip, "Psychiatric side effects of indomethacin in parturients," *Canadian Journal of Anesthesia*, vol. 50, no. 6, pp. 586–588, 2003.
- [60] H.-K. Jiang and D.-M. Chang, "Non-steroidal anti-inflammatory drugs with adverse psychiatric reactions: five case reports," *Clinical Rheumatology*, vol. 18, no. 4, pp. 339–345, 1999.
- [61] A. Barbon, A. Gervasoni, L. LaVia et al., "Human GluR6c, a functional splicing variants of GluR6, is mainly expressed in non-nervous cells," *Neuroscience Letters*, vol. 434, no. 1, pp. 77–82, 2008.
- [62] A. Rodríguez-Moreno and T. S. Sihra, "Metabotropic actions of kainate receptors in the CNS," *Journal of Neurochemistry*, vol. 103, no. 6, pp. 2121–2135, 2007.
- [63] A. Barbon, M. Popoli, L. la Via et al., "Regulation of editing and expression of glutamate α -amino-propionic-acid (AMPA)/kainate receptors by antidepressant drugs," *Biological Psychiatry*, vol. 59, no. 8, pp. 713–720, 2006.
- [64] J. Egebjerg and S. F. Heinemann, "Ca²⁺ permeability of unedited and edited versions of the kainate selective glutamate receptor GluR6," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 90, no. 2, pp. 755–759, 1993.
- [65] Z. Melyan, B. Lancaster, and H. V. Wheal, "Metabotropic regulation of intrinsic excitability by synaptic activation of kainate receptors," *The Journal of Neuroscience*, vol. 24, no. 19, pp. 4530–4534, 2004.
- [66] L. Hertz, J. Xu, D. Song, E. Yan, L. Gu, and L. Peng, "Astrocytic and neuronal accumulation of elevated extracellular K⁺ with a 2/3 K⁺/Na⁺ flux ratio-consequences for energy metabolism, osmolarity and higher brain function," *Frontiers in Computational Neuroscience*, vol. 7, article 114, 2013.
- [67] W. Sun, E. McConnell, J. F. Pare et al., "Glutamate-dependent neuroglial calcium signaling differs between young and adult brain," *Science*, vol. 339, no. 6116, pp. 197–200, 2013.
- [68] G. Shaltiel, S. Maeng, O. Malkesman et al., "Evidence for the involvement of the kainate receptor subunit GluR6 (GRIK2) in mediating behavioral displays related to behavioral symptoms of mania," *Molecular Psychiatry*, vol. 13, no. 9, pp. 858–872, 2008.
- [69] R. Delorme, M.-O. Krebs, N. Chabane et al., "Frequency and transmission of glutamate receptors GRIK2 and GRIK3 polymorphisms in patients with obsessive compulsive disorder," *NeuroReport*, vol. 15, no. 4, pp. 699–702, 2004.
- [70] A. S. Sampaio, J. Fagerness, J. Crane et al., "Association between polymorphisms in GRIK2 gene and obsessive-compulsive disorder: a family-based study," *CNS Neuroscience & Therapeutics*, vol. 17, no. 3, pp. 141–147, 2011.
- [71] G. Sanacora, G. Treccani, and M. Popoli, "Towards a glutamate hypothesis of depression: an emerging frontier of neuropsychopharmacology for mood disorders," *Neuropharmacology*, vol. 62, no. 1, pp. 63–77, 2012.
- [72] L. Peng, B. Li, T. Du, F. Wang, and L. Hertz, "Does conventional anti-bipolar and antidepressant drug therapy reduce NMDA-mediated neuronal excitation by downregulating astrocytic GluK2 function?" *Pharmacology Biochemistry and Behavior*, vol. 100, no. 4, pp. 712–725, 2012.
- [73] S. G. Tewari and K. K. Majumdar, "A mathematical model of the tripartite synapse: astrocyte-induced synaptic plasticity," *Journal of Biological Physics*, vol. 38, no. 3, pp. 465–496, 2012.
- [74] K. A. Lapidus, L. Soleimani, and J. W. Murrough, "Novel glutamatergic drugs for the treatment of mood disorders," *Journal of Neuropsychiatric Disease and Treatment*, vol. 9, pp. 1101–1112, 2013.
- [75] J. W. Murrough, A. M. Perez, S. Pillemer et al., "Rapid and longer-term antidepressant effects of repeated ketamine infusions in treatment-resistant major depression," *Biological Psychiatry*, vol. 74, no. 4, pp. 250–256, 2013.
- [76] E. Yan, B. Li, L. Gu, L. Hertz, and L. Peng, "Mechanisms for L-channel-mediated increase in [Ca²⁺]_i and its reduction by anti-bipolar drugs in cultured astrocytes combined with its mRNA expression in freshly isolated cells support the importance of astrocytic L-channels," *Cell Calcium*, vol. 54, no. 5, pp. 335–342, 2013.
- [77] G. Szabadkai and M. R. Duchen, "Mitochondria: the hub of cellular Ca²⁺ signaling," *Physiology*, vol. 23, no. 2, pp. 84–94, 2008.
- [78] R. M. Denton, "Regulation of mitochondrial dehydrogenases by calcium ions," *Biochimica et Biophysica Acta*, vol. 1787, no. 11, pp. 1309–1316, 2009.
- [79] S.-Z. Xu and D. J. Beech, "TrpC1 is a membrane-spanning subunit of store-operated Ca²⁺ channels in native vascular smooth muscle cells," *Circulation Research*, vol. 88, no. 1, pp. 84–87, 2001.
- [80] B. Li, L. Dong, H. Fu, B. Wang, L. Hertz, and L. Peng, "Effects of chronic treatment with fluoxetine on receptor-stimulated increase of [Ca²⁺]_i in astrocytes mimic those of acute inhibition of TRPC1 channel activity," *Cell Calcium*, vol. 50, no. 1, pp. 42–53, 2011.
- [81] J.-M. Beaulieu, R. R. Gainetdinov, and M. G. Caron, "Akt/GSK3 signaling in the action of psychotropic drugs," *Annual Review of Pharmacology and Toxicology*, vol. 49, pp. 327–347, 2009.
- [82] R. Liu, W. Dang, M. Jianting et al., "Citalopram alleviates chronic stress induced depression-like behaviors in rats by activating GSK3 β signaling in dorsal hippocampus," *Brain Research*, vol. 1467, pp. 10–17, 2012.
- [83] F. Karege, N. Perroud, S. Burkhardt et al., "Protein levels of β -catenin and activation state of glycogen synthase kinase-3 β in major depression. A study with postmortem prefrontal cortex," *Journal of Affective Disorders*, vol. 136, no. 1-2, pp. 185–188, 2012.
- [84] S. L. Diaz, S. Doly, N. Narboux-Nme et al., "5-HT_{2B} receptors are required for serotonin-selective antidepressant actions," *Molecular Psychiatry*, vol. 17, no. 2, pp. 154–163, 2012.
- [85] D. Song, T. Du, B. Li et al., "Astrocytic alkalization by therapeutically relevant lithium concentrations: implications for myo-inositol depletion," *Psychopharmacology*, vol. 200, no. 2, pp. 187–195, 2008.
- [86] D. Song, B. Li, E. Yan et al., "Chronic treatment with anti-bipolar drugs causes intracellular alkalization in astrocytes, altering their functions," *Neurochemical Research*, vol. 37, no. 11, pp. 2524–2540, 2012.
- [87] Y. Chen and L. Hertz, "Inhibition of noradrenaline stimulated increase in [Ca²⁺]_i in cultured astrocytes by chronic treatment with a therapeutically relevant lithium concentration," *Brain Research*, vol. 711, no. 1-2, pp. 245–248, 1996.
- [88] M. Wolfson, Y. Bersudsky, E. Zinger, M. Simkin, R. H. Belmaker, and L. Hertz, "Chronic treatment of human astrocytoma cells with lithium, carbamazepine or valproic acid decreases inositol uptake at high inositol concentrations but increases it at low inositol concentrations," *Brain Research*, vol. 855, no. 1, pp. 158–161, 2000.

- [89] D. Song, Y. Man, B. Li, J. Xu, L. Hertz, and L. Peng, "Comparison between drug-induced and K^+ -induced changes in molar acid extrusion fluxes J_H^+ and in energy consumption rates in astrocytes," *Neurochemical Research*, vol. 38, no. 11, pp. 2364–2374, 2013.
- [90] S. Kol, K. Ruutinen-Altman, I. Ben-Shlomo, D. W. Payne, M. Ando, and E. Y. Adashi, "The rat ovarian phospholipase A2 system: gene expression, cellular localization, activity characterization, and interleukin-1 dependence," *Endocrinology*, vol. 138, no. 1, pp. 322–331, 1997.
- [91] S. Ghelardoni, Y. A. Tomita, J. M. Bell, S. I. Rapoport, and F. Bosetti, "Chronic carbamazepine selectively downregulates cytosolic phospholipase A2 expression and cyclooxygenase activity in rat brain," *Biological Psychiatry*, vol. 56, no. 4, pp. 248–254, 2004.
- [92] M. J. Berridge, "Inositol trisphosphate and diacylglycerol as second messengers," *Biochemical Journal*, vol. 220, no. 2, pp. 345–360, 1984.
- [93] L. Hertz, Y. Chen, Y. Bersudsky, and M. Wolfson, "Shared effects of all three conventional anti-bipolar drugs on the phosphoinositide system in astrocytes," in *Non-Neuronal Cells of the Nervous System: Function and Dysfunction*, L. Hertz, Ed., pp. 1033–1048, Elsevier, 2004.
- [94] S. K. Fisher, J. E. Novak, and B. W. Agranoff, "Inositol and higher inositol phosphates in neural tissues: homeostasis, metabolism and functional significance," *Journal of Neurochemistry*, vol. 82, no. 4, pp. 736–754, 2002.
- [95] J. D. Port, S. S. Unal, D. A. Mrazek, and S. M. Marcus, "Metabolic alterations in medication-free patients with bipolar disorder: a 3T CSF-corrected magnetic resonance spectroscopic imaging study," *Psychiatry Research*, vol. 162, no. 2, pp. 113–121, 2008.
- [96] H. M. Aukema and B. J. Holub, "Inositol and pyrroloquinoline quinone. A : inositol," in *Modern Nutrition in Health and Disease*, M. E. Shils, J. A. Olson, and M. Shike, Eds., pp. 466–473, Lea and Febiger, 1994.
- [97] G. Agam, Y. Shapiro, Y. Bersudsky, O. Kofman, and R. H. Belmaker, "High-dose peripheral inositol raises brain inositol levels and reverses behavioral effects of inositol depletion by lithium," *Pharmacology Biochemistry and Behavior*, vol. 49, no. 2, pp. 341–343, 1994.
- [98] Y. Patishi, B. Lubrich, M. Berger, O. Kofman, D. van Calker, and R. H. Belmaker, "Differential uptake of *myo*-inositol in vivo into rat brain areas," *European Neuropsychopharmacology*, vol. 6, no. 1, pp. 73–75, 1996.
- [99] R. Spector and A. V. Lorenzo, "Myo-inositol transport in the central nervous system," *The American Journal of Physiology*, vol. 228, no. 5, pp. 1510–1518, 1975.
- [100] Y.-H. H. Wong, S. J. Kalmbach, B. K. Hartman, and W. R. Sherman, "Immunohistochemical staining and enzyme activity measurements show *myo*-inositol-1-phosphate synthase to be localized in the vasculature of brain," *Journal of Neurochemistry*, vol. 48, no. 5, pp. 1434–1442, 1987.
- [101] M. Wolfson, E. Hertz, R. H. Belmaker, and L. Hertz, "Chronic treatment with lithium and pretreatment with excess inositol reduce inositol pool size in astrocytes by different mechanisms," *Brain Research*, vol. 787, no. 1, pp. 34–40, 1998.
- [102] B. Lubrich and D. van Calker, "Inhibition of the high affinity *myo*-inositol transport system: a common mechanism of action of antibipolar drugs?" *Neuropsychopharmacology*, vol. 21, no. 4, pp. 519–529, 1999.
- [103] J. Matskevitch, C. A. Wagner, T. Rislis et al., "Effect of extracellular pH on the *myo*-inositol transporter SMIT expressed in *Xenopus oocytes*," *Pflügers Archiv*, vol. 436, no. 6, pp. 854–857, 1998.
- [104] M. Uldry, M. Ibberson, J.-D. Horisberger, J.-Y. Chatton, B. M. Riederer, and B. Thorens, "Identification of a mammalian H^+ -*myo*-inositol symporter expressed predominantly in the brain," *The EMBO Journal*, vol. 20, no. 16, pp. 4467–4477, 2001.
- [105] H. Fu, B. Li, L. Hertz, and L. Peng, "Contributions in astrocytes of SMIT1/2 and HMIT to *myo*-inositol uptake at different concentrations and pH," *Neurochemistry International*, vol. 61, no. 2, pp. 187–194, 2012.
- [106] E. di Daniel, M. H. S. Mok, E. Mead et al., "Evaluation of expression and function of the H^+ /*myo*-inositol transporter HMIT," *BMC Cell Biology*, vol. 10, article 54, pp. 1–12, 2009.
- [107] D. G. Gossman and H.-B. Zhao, "Hemichannel-mediated inositol 1,4,5-trisphosphate (IP_3) release in the cochlea: a novel mechanism of IP_3 intercellular signaling," *Cell Communication & Adhesion*, vol. 15, no. 4, pp. 305–315, 2008.
- [108] X.-H. Wang, M. Streeter, Y.-P. Liu, and H.-B. Zhao, "Identification and characterization of pannexin expression in the mammalian cochlea," *The Journal of Comparative Neurology*, vol. 512, no. 3, pp. 336–346, 2009.
- [109] T. Du, B. Li, H. Li, M. Li, L. Hertz, and L. Peng, "Signaling pathways of isoproterenol-induced $ERK_{1/2}$ phosphorylation in primary cultures of astrocytes are concentration-dependent," *Journal of Neurochemistry*, vol. 115, no. 4, pp. 1007–1023, 2010.
- [110] L. Hertz, J. Xu, D. Song, T. Du, E. Yan, and L. Peng, "Brain glycolysis, adrenoceptors, pyruvate carboxylase, Na^+ , K^+ -ATPase and Marie E. Gibbs' pioneering learning studies," *Frontiers in Integrative Neuroscience*, vol. 7, article 20, 2013.
- [111] G. Owsianik, K. Talavera, T. Voets, and B. Nilius, "Permeation and selectivity of TRP channels," *Annual Review of Physiology*, vol. 68, pp. 685–717, 2006.
- [112] R. C. Reyes, A. Verkhratsky, and V. Parpura, "TRPC1-mediated Ca^{2+} and Na^+ signalling in astroglia: differential filtering of extracellular cations," *Cell Calcium*, vol. 54, no. 2, pp. 120–125, 2013.
- [113] I. Goldstein, T. Levy, D. Galili et al., "Involvement of Na^+ , K^+ -ATPase and endogenous digitalis-like compounds in depressive disorders," *Biological Psychiatry*, vol. 60, no. 5, pp. 491–499, 2006.
- [114] A. M. Rose, B. J. Mellett, R. Valdes Jr. et al., "Alpha 2 isoform of the Na, K-adenosine triphosphatase is reduced in temporal cortex of bipolar individuals," *Biological Psychiatry*, vol. 44, no. 9, pp. 892–897, 1998.
- [115] U. Banerjee, A. Dasgupta, J. K. Rout, and O. P. Singh, "Effects of lithium therapy on Na^+ - K^+ -ATPase activity and lipid peroxidation in bipolar disorder," *Progress in Neuro-Psychopharmacology & Biological Psychiatry*, vol. 37, no. 1, pp. 56–61, 2012.
- [116] L. Mynett-Johnson, V. Murphy, J. McCormack et al., "Evidence for an allelic association between bipolar disorder and a Na^+ , K^+ adenosine triphosphatase alpha subunit gene (ATPIA3)," *Biological Psychiatry*, vol. 44, no. 1, pp. 47–51, 1998.
- [117] H. Hamakawa, J. Murashita, N. Yamada, T. Inubushi, N. Kato, and T. Kato, "Reduced intracellular pH in the basal ganglia and whole brain measured by ^{31}P -MRS in bipolar disorder," *Psychiatry and Clinical Neurosciences*, vol. 58, no. 1, pp. 82–88, 2004.
- [118] K. Deigweier, N. Koschnick, H.-O. Pörtner, and M. Lucassen, "Acclimation of ion regulatory capacities in gills of marine fish under environmental hypercapnia," *The American Journal of Physiology—Regulatory Integrative and Comparative Physiology*, vol. 295, no. 5, pp. R1660–R1670, 2008.

- [119] J. Cummins and H. Hydén, "Adenosine triphosphate levels and adenosine triphosphatases in neurons, glia and neuronal membranes of the vestibular nucleus," *Biochimica et Biophysica Acta*, vol. 60, no. 2, pp. 271–283, 1962.
- [120] B. Li, L. Hertz, and L. Peng, "Cell-specific mRNA alterations in Na⁺, K⁺-ATPase α and β isoforms and FXRD in mice treated chronically with carbamazepine, an anti-bipolar drug," *Neurochemical Research*, vol. 38, no. 4, pp. 834–841, 2013.
- [121] L. Segall, S. E. Daly, and R. Blostein, "Mechanistic basis for kinetic differences between the rat α 1, α 2, and α 3 isoforms of the Na,K-ATPase," *The Journal of Biological Chemistry*, vol. 276, no. 34, pp. 31535–31541, 2001.
- [122] A. G. Therien and R. Blostein, "Mechanisms of sodium pump regulation," *The American Journal of Physiology—Cell Physiology*, vol. 279, no. 3, pp. C541–C566, 2000.
- [123] L. K. Bekar and W. Walz, "Intracellular chloride modulates a-type potassium currents in astrocytes," *Glia*, vol. 39, no. 3, pp. 207–216, 2002.
- [124] K. Egawa, J. Yamada, T. Furukawa, Y. Yanagawa, and A. Fukada, "Cl⁻ homeodynamics in gap junction-coupled astrocytic networks on activation of GABAergic synapses," *The Journal of Physiology*, vol. 591, pp. 3901–3917, 2013.
- [125] S. D. Meier, K. W. Kafitz, and C. R. Rose, "Developmental profile and mechanisms of GABA-induced calcium signaling in hippocampal astrocytes," *Glia*, vol. 56, no. 10, pp. 1127–1137, 2008.
- [126] M.-R. Zarrindast, P. Rostami, and M. Sadeghi-Hariri, "GABA_A but not GABA_B receptor stimulation induces antianxiety profile in rats," *Pharmacology Biochemistry and Behavior*, vol. 69, no. 1–2, pp. 9–15, 2001.
- [127] B. E. Yoon, J. Woo, and C. J. Lee, "Astrocytes as GABA-ergic and GABA-ceptive cells," *Neurochemical Research*, vol. 37, no. 11, pp. 22474–22479, 2012.
- [128] L. Hertz, Z. Zhao, and Y. Chen, "The astrocytic GABA_A/benzodiazepine-like receptor: the Joker receptor for benzodiazepine-mimetic drugs?" *Recent Patents on CNS Drug Discovery*, vol. 1, no. 1, pp. 93–103, 2006.
- [129] K. V. Subbarao, J. U. Stolzenburg, and L. Hertz, "Pharmacological characteristics of potassium-induced, glycogenolysis in astrocytes," *Neuroscience Letters*, vol. 196, no. 1–2, pp. 45–48, 1995.
- [130] T. Goetz, A. Arslan, W. Wisden, and P. Wulff, "GABA_A receptors: structure and function in the basal ganglia," *Progress in Brain Research*, vol. 160, pp. 21–41, 2007.
- [131] J. D. Cahoy, B. Emery, A. Kaushal et al., "A transcriptome database for astrocytes, neurons, and oligodendrocytes: a new resource for understanding brain development and function," *The Journal of Neuroscience*, vol. 28, no. 1, pp. 264–278, 2008.
- [132] W. Wisden, D. J. Laurie, H. Monyer, and P. H. Seeburg, "The distribution of 13 GABA_A receptor subunit mRNAs in the rat brain. I. Telencephalon, diencephalon, mesencephalon," *The Journal of Neuroscience*, vol. 12, no. 3, pp. 1040–1062, 1992.
- [133] U. Rudolph and H. M. Möhler, "GABA_A receptor subtypes: therapeutic potential in down syndrome, affective disorders, schizophrenia, and autism," *Annual Review of Pharmacology and Toxicology*, vol. 54, pp. 483–507, 2014.
- [134] S. A. Bergman, "The benzodiazepine receptor," *Anesthesia Progress*, vol. 33, no. 5, pp. 213–219, 1986.
- [135] V. Papadopoulos and L. Lecanu, "Translocator protein (18 kDa) TSPO: an emerging therapeutic target in neurotrauma," *Experimental Neurology*, vol. 219, no. 1, pp. 53–57, 2009.
- [136] B. Costa, E. da Pozzo, and C. Martini, "Translocator protein as a promising target for novel anxiolytics," *Current Topics in Medicinal Chemistry*, vol. 12, no. 4, pp. 270–285, 2012.
- [137] P. Ferrero, E. Costa, B. Conti-Tronconi, and A. Guidotti, "A diazepam binding inhibitor (DBI)-like neuropeptide is detected in human brain," *Brain Research*, vol. 399, no. 1, pp. 136–142, 1986.
- [138] E. Slobodyansky, A. Guidotti, C. Wambebe, A. Berkovich, and E. Costa, "Isolation and characterization of a rat brain triakontatetrapeptide, a posttranslational product of diazepam binding inhibitor: specific action at the Ro 5-4864 recognition site," *Journal of Neurochemistry*, vol. 53, no. 4, pp. 1276–1284, 1989.
- [139] P. Gandolfo, E. Louiset, C. Patte et al., "The triakontatetrapeptide TTN increases [Ca²⁺]_i in rat astrocytes through activation of peripheral-type benzodiazepine receptors," *Glia*, vol. 35, no. 2, pp. 90–100, 2001.
- [140] K. H. Backus, H. Kettenmann, and M. Schachner, "Effect of benzodiazepines and pentobarbital on the GABA-induced depolarization in cultured astrocytes," *Glia*, vol. 1, no. 2, pp. 132–140, 1988.
- [141] A. S. Bender and L. Hertz, "Pharmacological evidence that the non-neuronal diazepam binding site in primary cultures of glial cells is associated with a calcium channel," *European Journal of Pharmacology*, vol. 110, no. 2, pp. 287–288, 1985.
- [142] Z. Zhao, L. Hertz, and W. E. Code, "Effects of benzodiazepines on potassium-induced increase in free cytosolic calcium concentration in astrocytes: interactions with nifedipine and the peripheral-type benzodiazepine antagonist PK 11195," *Canadian Journal of Physiology and Pharmacology*, vol. 74, no. 3, pp. 273–277, 1996.
- [143] Y. Itzhak, L. Baker, and M. D. Norenberg, "Characterization of the peripheral-type benzodiazepine receptors in cultured astrocytes: evidence for multiplicity," *Glia*, vol. 9, no. 3, pp. 211–218, 1993.
- [144] G. T. Bolger, A. H. Newman, K. C. Rice, H. W. M. Lueddens, A. S. Basile, and P. Skolnick, "Characterization of the effects of AHN 086, an irreversible ligand of 'peripheral' benzodiazepine receptors, on contraction in guinea-pig atria and ileal longitudinal smooth muscle," *Canadian Journal of Physiology and Pharmacology*, vol. 67, no. 2, pp. 126–134, 1989.
- [145] G. T. Bolger, S. Abraham, N. Oz, and B. A. Weissman, "Interactions between peripheral-type benzodiazepine receptor ligands and an activator of voltage-operated calcium channels," *Canadian Journal of Physiology and Pharmacology*, vol. 68, no. 1, pp. 40–45, 1990.
- [146] L. Hertz and Y. Chen, "The astrocytic GABA_A/benzodiazepine-like receptor: identifying the Joker receptor for benzodiazepine-mimetic drugs?" in *Frontiers in CNS Drug Discovery*, Atta-ur-Rahman and M. Iqbal Choudhary, Eds., pp. 342–361, Bentham, 2010.
- [147] J. Kanto, L. Kangas, and T. Siirtola, "Cerebrospinal fluid concentrations of diazepam and its metabolites in man," *Acta Pharmacologica et Toxicologica*, vol. 36, no. 4, pp. 328–334, 1975.
- [148] P. Polc, "Electrophysiology of benzodiazepine receptor ligands: multiple mechanisms and sites of action," *Progress in Neurobiology*, vol. 31, no. 5, pp. 349–423, 1988.
- [149] R. Yu and M. K. Ticku, "Chronic neurosteroid treatment decreases the efficacy of benzodiazepine ligands and neurosteroids at the γ -aminobutyric acid(A) receptor complex in mammalian cortical neurons," *The Journal of Pharmacology and Experimental Therapeutics*, vol. 275, no. 2, pp. 784–789, 1995.

- [150] J. M. M. Olson, B. J. Ciliax, W. R. Mancini, and A. B. Young, "Presence of peripheral-type benzodiazepine binding sites on human erythrocyte membranes," *European Journal of Pharmacology*, vol. 152, no. 1-2, pp. 47-53, 1988.
- [151] G. A. Kennett, F. Bright, B. Trail, G. S. Baxter, and T. P. Blackburn, "Effects of the 5-HT_{2B} receptor agonist, BW 723C86, on three rat models of anxiety," *British Journal of Pharmacology*, vol. 117, no. 7, pp. 1443-1448, 1996.
- [152] M. S. Duxon, G. A. Kennett, S. Lightowler, T. P. Blackburn, and K. C. F. Fone, "Activation of 5-HT_{2B} receptors in the medial amygdala causes anxiolysis in the social interaction test in the rat," *Neuropharmacology*, vol. 36, no. 4-5, pp. 601-608, 1997.
- [153] G. A. Kennett, B. Trail, and F. Bright, "Anxiolytic-like actions of BW 723C86 in the rat Vogel conflict test are 5-HT_{2B} receptor mediated," *Neuropharmacology*, vol. 37, no. 12, pp. 1603-1610, 1998.
- [154] X. Zhang, L. Peng, Y. Chen, and L. Hertz, "Stimulation of glycogenolysis in astrocytes by fluoxetine, an antidepressant acting like 5-HT," *NeuroReport*, vol. 4, no. 11, pp. 1235-1238, 1993.