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Magnetic Resonance Image Example Based Contrast Synthesis

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Abstract

The performance of image analysis algorithms applied to magnetic resonance images is strongly influenced by the pulse sequences used to acquire the images. Algorithms are typically optimized for a targeted tissue contrast obtained from a particular implementation of a pulse sequence on a specific scanner. There are many practical situations, including multi-institution trials, rapid emergency scans, and scientific use of historical data, where the images are not acquired according to an optimal protocol or the desired tissue contrast is entirely missing. This paper introduces an image restoration technique that recovers images with both the desired tissue contrast and a normalized intensity profile. This is done using patches in the acquired images and an atlas containing patches of the acquired and desired tissue contrasts. The method is an example-based approach relying on sparse reconstruction from image patches. Its performance is demonstrated using several examples, including image intensity normalization, missing tissue contrast recovery, automatic segmentation, and multimodal registration. These examples demonstrate potential practical uses and also illustrate limitations of our approach.

Index Terms

Neuroimaging; sparse reconstruction; image restoration; magnetic resonance imaging (MRI)

I. Introduction

Magnetic resonance (MR) imaging (MRI) is widely used to image the brain. Postprocessing of MR brain images, e.g., image segmentation [1]–[6] and image registration [7]–[11], has been used for many scientific purposes such as furthering our understanding of normal aging [12], [13], disease progression [14], [15], population analysis [16], and prognosis [17]. It is well known that image analysis algorithms are routinely optimized for a particular type of scan (or collection of scans), which could include specification of the tissue contrast (e.g., T1-weighted (T1-w) versus T2-weighted (T2-w)), image resolution, pulse sequence parameters, field strength of the scanner, and even the particular scanner manufacturer. Although many algorithms claim to be robust to variations in the input images, inevitably

there will be performance degradation as the input images deviate from a perfect match to the images that were used to carry out the algorithm optimization.

There are many practical situations, including multi-institution trials [18]–[20], rapid emergency scans [21], and scientific use of historical data [12], where the images are not acquired according to an optimal protocol for a given image analysis algorithm. As well, a particular tissue contrast that is necessary for a given algorithm might have been omitted for a given study or for a particular patient acquisition. In such cases, investigators and clinicians have three choices. First, they might apply the algorithm on available images and accept a sub-optimal result. Second, they might remove patients with inadequate image data from the study. Third, they might design a new image analysis algorithm that will work equally well on the image data that is available to them. Upon consideration of these options, it seems that each choice leads to concerns, and an alternative is highly desirable. This paper is concerned with a fourth alternative: to recover an image with the desired tissue contrast and intensity profile using the available images and prior information. There are three general approaches described in the literature to carry out this fourth alternative, which we briefly describe next. The method we describe in this paper offers an approach with significant advantages over existing approaches including improved performance and greater applicability.

The most common approach to recovery of images with a desired tissue contrast and intensity profile is histogram matching [22]–[29]. Some manner of histogram matching is used as a preprocessing step in nearly every neuroimage segmentation and registration pipeline because they do improve the accuracy and consistency of results [30], [31]. However, its use becomes questionable when applied to data acquired with different pulse sequences, especially when pathology is present, and its use in volumetric analysis may lead to incorrect results simply because the histograms are forced to follow that of a target image which itself contains certain ratios of tissue types. One way to address this last concern is to first segment brain structures and match their individual histograms to a registered atlas [32]. Although this method produces consistent subcortical segmentations over data sets acquired under different scanners, it requires a detailed segmentation of the images *before intensity normalization has been applied*, which leads to a “chicken and egg” situation. It is clearly more desirable to normalize the image intensities prior to any significant image analysis algorithms have been applied.

A second approach to recover images with a desired tissue contrast and intensity profile is to acquire multiple images of the subject with predefined, precisely calibrated acquisition parameters [33]. Given this set of images, tissue proton density P_D and relaxation parameters T_1 , T_2 , and T_2^* can be estimated by inverting the pulse sequence equations [34], and calibrated images with the desired tissue contrast can be computed by using these estimated parameters in the appropriate mathematical equation derived from the imaging physics [35]. This approach has three disadvantages. First, the imaging equations are typically only an approximation to what is used in practice, and therefore the acquired data may not be accurately related to the underlying tissue parameters by the assumed imaging equation. Second, the required voxel-wise mathematical inversion is generally poorly conditioned, which means that noise and artifacts can have a large impact on the accuracy of estimating

the underlying tissue parameters. Third, in large imaging studies with many clinical scenarios it is difficult to acquire the source image data with consistent, predefined image acquisition parameters. Because of these problems, most image processing algorithms are not designed to exploit the existence of underlying tissue parameter estimates nor are most studies and clinical scans designed to exploit this approach to tissue contrast normalization.

Another way to recover a target tissue contrast and intensity profile is to register the subject to a multiple-contrast atlas and to transfer the desired contrast intensities from the atlas space to the subject space [36]. This approach enjoys great popularity not only for transferring tissue contrasts but also for transferring segmented tissue labels [37], and is tacitly assumed by many researchers to be the best current approach for generation of images with alternate tissue contrast. However, this approach does not produce an acceptable result if the registration result is poor or if there are anomalies or unknown tissues in the subject that are not present or spatially distributed differently in the atlas. In the example shown in Fig. 1, a T1-w spoiled gradient recalled (SPGR) atlas (Fig. 1(a)) was registered to a similar contrast subject image (Fig. 1(c)) using the (inverse consistent and diffeomorphic) SyN registration algorithm [7]. The atlas magnetization prepared rapid gradient echo (MPRAGE) contrast intensities (Fig. 1(b)) were then mapped through the transformation to the subject space, yielding a synthetic subject MPRAGE image (Fig. 1(e)). By comparing this result to the subject's true MPRAGE image (Fig. 1(d)), it is apparent that this approach failed to adequately define the ventricles (Fig. 1(d), arrow adjacent to the ventricles) due to shortcomings in the registration, and it also failed to represent the WM lesions that are present in the original image (Fig. 1(d), arrow denoting the lesion posterior to the ventricles) because such lesions are not present in the atlas. The image produced using our approach (Fig. 1(f)) shows neither of these problems and compares quite favorably in overall contrast and appearance to the true MPRAGE.

The method proposed in this paper, **MR image example-based contrast synthesis** (MIMECS), combines the idea of example-based image hallucination [38], [39] with non-local means [40]–[42] and sparse priors [43], [44] to synthesize MR contrasts using patches—i.e., small regions of the image—from a multiple-contrast atlas (also called a textbook in [36]). The core idea of MIMECS is that each subject patch can be matched to a combination of a few relevant patches from a dictionary in a non-local fashion [43], [45], [46]. It is different from classical histogram matching in the sense that a patch can be thought of as a feature vector of its center voxel, thus including local neighborhood information for that voxel. We show the results of seven case studies which demonstrate potential uses and limitations of MIMECS in realistic scientific and clinical scenarios.

There are several advantages of MIMECS compared to previous synthesis methods. First, it is a completely automatic pre-processing step that can precede any image processing task. Second, there is no need to use multiple, calibrated pulse sequences in order to estimate underlying tissue parameters. Third, even though MIMECS uses an atlas, there is no need to carry out subject-to-atlas registration, thus avoiding potential errors due to misregistration or missing tissues. Finally, it does not require any segmentation of the subject. This last point, detailed in Sec. III-C, is a unique feature of this work improving on our previous patch based

results [47]–[49], and is important in the field of sparse reconstruction as it alleviates the need for atlas training.

The paper is organized as follows. First, we summarize the idea of sparse priors and establish some notation that will be used throughout the paper. Then the atlas based patch matching synthesis algorithm is explained in a sparse priors paradigm, Fig. 2 provides a high level overview of the algorithm. Finally, we demonstrate the applicability of MIMECS on a variety of case studies.

II. Background

Sparse signal reconstruction maintains that because most signals are sparse in some way it is not necessary to observe the full signal in order to accurately reconstruct it. The idea has been successfully applied to many image processing algorithms including denoising [46], image restoration [50], [51], and super-resolution [43], often improving on state-of-the-art algorithms. For example in [46], authors use a K-SVD algorithm [52] to generate an overcomplete set of bases from natural image patches and apply it to a denoising problem. Dictionary based patch matching methods [53], [54] are also popular for natural image restoration. In our case, we build an overcomplete dictionary using patches from human brain images to both normalize a given contrast and synthesize alternate MR tissue contrasts. This section presents the mathematical formulation and key results that are needed to explain our approach.

Suppose we want to reconstruct a vector $\mathbf{x} \in \mathbb{R}^N$ that is s -sparse, i.e., having at most s non-zero elements, and our observations are $\mathbf{b} = A\mathbf{x}$, where $\mathbf{b} \in \mathbb{R}^d$, $s < d < N$, and $A \in \mathbb{R}^{d \times N}$. Since this is an under-determined system, we cannot directly invert the system to find \mathbf{x} , and additional penalties or constraints must therefore be used. Since \mathbf{x} is known to be sparse, it makes sense to formulate an objective function that tries to find a sparse solution that is also consistent with the measurements, as follows,

$$\hat{\mathbf{x}} = \min \|\mathbf{x}\|_0 \quad \text{such that} \quad \|\mathbf{b} - A\mathbf{x}\|_2^2 < \varepsilon_1. \quad (1)$$

Here, ε_1 is the noise level in the measurements, $\|\cdot\|_p$ is the ℓ_p norm; in particular, $\|\mathbf{x}\|_0$ is the number of non-zero elements in \mathbf{x} . There are relatively simple conditions on A that guarantee a feasible solution to (1), however the solution of this problem is combinatorial in nature, and therefore NP-hard.

It has been shown that if $\|\mathbf{x}\|_0$ is small [55], the optimal solution to (1) can also be found by solving,

$$\hat{\mathbf{x}} = \min \|\mathbf{x}\|_1 \quad \text{such that} \quad \|\mathbf{b} - A\mathbf{x}\|_2^2 < \varepsilon_2,$$

This is a convex programming problem and can be solved in polynomial time. If ε_2 is unknown, we can rewrite the above in the following form,

$$\hat{\mathbf{x}} = \arg \min_{\mathbf{x}} \{ \|\mathbf{b} - A\mathbf{x}\|_2^2 + \lambda \|\mathbf{x}\|_1 \}, \quad (2)$$

where λ is a weighting factor. The resulting sparsity in $\hat{\mathbf{x}}$ decreases as λ increases. The formulation in (2) is now the standard way to frame a ‘‘sparse prior’’ estimation problem and is the core idea behind MIMECS.

III. Method

The MIMECS method is based on analysis of image patches, which are $p \times q \times r$ 3D subimages associated with each voxel of the image. Patches are typically small and centered on the voxel of interest—e.g., $p = q = r = 3$ or $p = q = r = 5$. For convenience we write a patch as the 1D vector \mathbf{b} of size $d \times 1$, where $d = pqr$. The voxels within a patch are always ordered in the same way, using a consistent rasterization, in order to create this 1D representation. Fig. 2 provides an illustrative overview of the MIMECS algorithm.

A. Atlas description

We define an atlas as an $(n + 1)$ -tuple of images, $\{a_1, a_2, \dots, a_n, a_{n+1}\}$, where a_k has tissue contrast c_k , $k = 1, \dots, n+1$. All atlas images a_k are co-registered and sampled on the same voxel locations in space. At each voxel, 3D patches can be defined on each image and are denoted by the $d \times 1$ vectors $\mathbf{a}_k(i)$, where $i = 1, \dots, N$, is an index over the voxels of a_k . Since a_k 's are co-registered, each of them has the exact same number of voxels N . The primary aim of MIMECS is to synthesize the c_{n+1} contrast image using a set of n subject images having contrasts c_1 to c_n . We define the $c_{1:n}$ contrast dictionary A_1 , $A_1 \in \mathbb{R}^{nd \times N}$, where the i^{th} column of A_1 is $\mathbf{f}(i) = [\mathbf{a}_1(i)^T \dots \mathbf{a}_n(i)^T]^T$, i.e., an ordered (according to their contrasts c_1 through c_n) column vector composed of patches $\mathbf{a}_k(i)$, which is the i^{th} patch from the atlas a_k . The contrast dictionary A_2 , $A_2 \in \mathbb{R}^{d \times N}$, is constructed in a similar manner but only contains the patches found in the atlas a_{n+1} , which correspond to the c_{n+1} tissue contrast. By construction, the i^{th} columns of A_1 and A_2 correspond to the same spatial location and represent the c_1 through c_n and c_{n+1} contrasts, respectively.

B. Contrast synthesis algorithm

We assume that there are n subject (input) images $\{s_1, \dots, s_n\}$ available with contrasts c_1 to c_n . They are co-registered and sampled on the same voxel locations in space. We note that $\{s_1, \dots, s_n\}$ are not registered to $\{a_1, \dots, a_{n+1}\}$. Also, if there are fewer subject images than the number of images in the atlas, then the atlas contrasts that do not match the subject contrasts are removed, n is reduced (to a minimum of $n = 1$), and MIMECS is carried out on the available subject images. We first decompose the subject images s_k into $d \times 1$ patches $\mathbf{s}_k(j)$, where j is an index over the voxels of s_k . For every j , we stack all the patches $\mathbf{s}_k(j)$ in a $nd \times 1$ vector $\mathbf{b}_1(j)$, in the order of their contrasts c_1 through c_n , $\mathbf{b}_1(j) = [\mathbf{s}_1(j)^T \dots \mathbf{s}_n(j)^T]^T$,

$\mathbf{b}_1(j) \in \mathbb{R}^{nd}$. For simplicity of terminology, we also call any $nd \times 1$ vector a ‘‘patch’’. The aim is to obtain a synthetic c_{n+1} contrast subject image \hat{s}_{n+1} using $\{s_1, \dots, s_n\}$ and $\{a_1, \dots, a_{n+1}\}$.

We assume that for a subject image patch $\mathbf{b}_1(j)$, a small number of relevant and similar examples can always be found from a rich and overcomplete patch dictionary, A_1 , which has the same contrasts (and assembled into patch vectors in the same order) as $\mathbf{b}_1(j)$. It is unlikely that a single patch from A_1 will perfectly match $\mathbf{b}_1(j)$, but it is quite likely that an optimal linear combination of a small number of patches will yield a very close approximation. The problem of finding a few patches to form the linear combination can be solved either by comparing noise characteristics, as derived in non-local means [40]–[42], or by assuming the weight vector $\mathbf{x}(j)$ is sparse [44], which is the approach we take here. This leads to the following generic expression for the desired solution

$$\mathbf{b}_1(j) \approx A_1 \mathbf{x}(j), \text{ for some } \mathbf{x}(j) \in \mathbb{R}^N, \|\mathbf{x}(j)\|_0 \ll N, \forall j, \quad (3)$$

where A_1 is a subset of ‘‘relevant’’ patches taken from the atlas images $\{a_1, \dots, a_n\}$. The relevant patches are chosen according to their ℓ_2 distance from the subject patch, $\mathbf{b}_1(j)$, as suggested in [45]. The details are given in Sec. IV. Once the sparse weight $\mathbf{x}(j)$ is found, the c_{n+1} contrast subject patch estimate is found by applying the same linear combination to the corresponding c_{n+1} contrast dictionary, A_2 , i.e.,

$$\hat{\mathbf{b}}_2(j) = A_2 \mathbf{x}(j). \quad (4)$$

The idea of transferring estimated coefficients from one dictionary to another, as we do here, has been explored previously in the context of super-resolution [43] and label fusion [39], [41].

It is important to consider why we seek a sparse representation. The sparsest representation $\mathbf{x}(j)$ is just a single column of A_1 which is approximately equal to $\mathbf{b}_1(j)$. In this case, the corresponding column of A_2 gives the c_{n+1} contrast of $\mathbf{b}_1(j)$. However, our formulation will not yield unit sparsity in general, but instead a small number of patches from A_1 whose linear combination gives $\mathbf{b}_1(j)$. The motivation behind choosing a sparse $\mathbf{x}(j)$ to reconstruct $\mathbf{b}_1(j)$ is twofold. First, to reconstruct a subject patch, we want to pick those atlas patches that are close in intensity, i.e., those that are likely to be from the same tissue classes, so it would be undesirable to pick a large number of patches that might tend to mix the tissue classes. Second, if too many similar atlas patches are used to reconstruct $\mathbf{b}_1(j)$, then the corresponding c_{n+1} contrast patch will be overly smooth due to the cumulative effects of small mismatches in each patch. Empirically, we have found that non-sparse $\mathbf{x}(j)$ tends to produce smooth images acting much like a denoising process, while highly sparse $\mathbf{x}(j)$ produces noisy images. In Sec. IV, an empirical way of choosing the sparsity level is described.

Since the combinatorics of (3) makes it infeasible to solve this problem directly, we use an ℓ_1 minimization strategy and reformulate the problem as,

$$\hat{\mathbf{x}}(j) = \arg \min_{\mathbf{x} \geq 0} \left[\|\mathbf{b}_1(j) - A_1 \mathbf{x}\|_2^2 + \lambda \|\mathbf{x}\|_1 \right], \forall j, \quad \text{subject to} \quad \|\mathbf{f}(i)\|_2^2 = 1, i=1, \dots, N, \quad (5)$$

where $\mathbf{f}(i)$ denotes the i^{th} column of A_1 , also defined in Sec. III-A. As in (2), λ is a weighting factor that encourages a sparse solution when it is large. Given this optimal combination of the patches in A_1 , which best approximates the subject patch $\mathbf{b}_1(j)$, the c_{n+1} contrast patch is estimated using (4). The full contrast image \hat{s}_{n+1} is reconstructed by taking the union of all central voxels in the reconstructed patches.

There are two key differences in our solution (5) as compared to (2). The first, is the positivity constraint on the coefficients \mathbf{x} in (5). The reason we impose this condition is to encourage the selected patches—i.e., those used in the sparse reconstruction of the subject patch—to have the same “texture” as the subject patch. For example, we do not want a boundary that is GM on the left and WM on the right to be approximated with a negative coefficient multiplying a patch having exactly the opposite orientation. The image being approximated may “look” the same, but the underlying tissues would be wrong and a patch that is synthesized would have incorrect tissues and thus have the wrong appearance. This condition is therefore designed to encourage “gray matter patches” to be used to synthesize “gray matter patches” and so on. We note that computational aspects of this positivity constraint has been previously explored in Lasso [56], and that this constrained ℓ_1 minimization procedure has been shown to be equivalent to an ℓ_0 minimization procedure (like (1) with a similar positivity constraint) if $\mathbf{x}(j)$ is sufficiently sparse [57].

The second difference between (5) and (2) is the requirement that the columns of A_1 (denoted by $\mathbf{f}(i)$, $i = 1, \dots, N$) be normalized to unity. This constraint is necessary in order to guarantee the uniqueness of the solution (by removing the scaling ambiguity), and is a common feature of patch-based techniques in computer vision and image processing [43], [45]. However, unit normalization of patches taken from the atlas MR images, removes the relationship between overall patch intensity and the patch texture, which is essential to distinguish between a “pure” GM patch and a “pure” WM patch, which can only be differentiated by their intensities and not the texture. Therefore, in Sec. III-C we propose a novel approach that normalizes patches in the $(nd + 1)$ -dimensional space rather than the nd -dimensional space in order to preserve the desired intensity information.

C. Normalization of A_1

To guarantee a unique solution to (5), the columns of A_1 are normalized such that $\|\mathbf{f}(i)\|_2^2 = 1$, $\forall i = 1, \dots, N$. However, if scale is directly removed from an MR image patch, then a key feature in distinguishing tissue types is lost and patches used in the sparse reconstruction are less likely to be selected from the same tissue type. It is common in the sparse reconstruction literature to use prior training to learn how to select a subset of patches from which to synthesize a given subject patch. In our prior work [47]–[49] we used an atlas selection

method based on image classification in order to restrict the dictionary to patches that are likely to come from appropriate tissue classes [47], [48]. We note that the present method supports this strategy since we use a subset A_1 of patches from which to select a sparse collection (see Sec. IV). However, an important goal in the present work is to eliminate both the dependency on prior training and the requirement for a classification step prior to synthesis. We have accomplished this using the “trick” of normalizing the patches (e.g., $\mathbf{b}_1(j)$'s or $\mathbf{f}(i)$'s) in a $(nd + 1)$ -dimensional space rather than the nd -dimensional space, which we now describe.

All the atlas and subject patches are first globally normalized such that their maximum of their norms is unity. Define,

$$m = \max_{i,j} \{ \|\mathbf{f}(i)\|_2, \|\mathbf{b}_1(j)\|_2 \}.$$

Then the patches are scaled as follows,

$$\begin{aligned} \tilde{\mathbf{f}}(i) &= \frac{\mathbf{f}(i)}{m}, & \forall i, \\ \tilde{\mathbf{b}}_1(j) &= \frac{\mathbf{b}_1(j)}{m}, & \forall j. \end{aligned}$$

This global scaling guarantees that relative intensities are preserved and that all intensities of both subject and atlas fall in the range $[0, 1]$. Now, we project both the subject patch and all atlas patches to the unit sphere in \mathbb{R}^{nd+1} as follows,

$$\begin{aligned} \mathbf{f}'(i) &= \begin{bmatrix} \tilde{\mathbf{f}}(i) \\ \sqrt{1 - \|\tilde{\mathbf{f}}(i)\|^2} \end{bmatrix}, \forall i, \\ \mathbf{b}'_1(j) &= \begin{bmatrix} \tilde{\mathbf{b}}_1(j) \\ \sqrt{1 - \|\tilde{\mathbf{b}}_1(j)\|^2} \end{bmatrix}, \forall j. \end{aligned}$$

At this point, both the subject patch and the atlas patches are normalized to unity, satisfying $\|\mathbf{f}'(i)\|_2^2 = 1, \forall i$, and $\|\mathbf{b}'_1(j)\|_2^2 = 1, \forall j$. The normalization criterion in (5) is therefore automatically satisfied if the $c_{1,n}$ contrast dictionary A_1 is changed to $A'_1 \in \mathbb{R}^{(nd+1) \times N}$ such that the columns of A'_1 are $\mathbf{f}'(i), i = 1, \dots, N$.

Given this new normalization, we rewrite (5) as

$$\hat{\mathbf{x}}(j) = \arg \min_{\mathbf{x} \geq \mathbf{0}} \left[\|\mathbf{b}'_1(j) - A'_1 \mathbf{x}\|_2^2 + \lambda \|\mathbf{x}\|_1 \right], \forall j. \quad (6)$$

The solution of (6) yields a nonnegative combination of the columns of A'_1 that sparsely approximates $\mathbf{b}'_1(j)$. The resulting reconstructed patch matches both the pattern and intensities within the target patch. We use this solution $\mathbf{x}(\hat{j}) \in \mathbb{R}^N$ to synthesize the c_{n+1} contrast subject patch in \mathbb{R}^d . This works correctly because we maintain the association of columns in A_1 , A_2 and A'_1 . Once $\hat{\mathbf{b}}_2(j)$ is found from (4), we use only its central value in the reconstructed image \hat{s}_{n+1} .

IV. Implementation

A. Patch size and ℓ_1 Solvers

For all the experiments reported in this and subsequent sections, we used patches of dimension $3 \times 3 \times 3$ voxels, i.e. $d=27$. We used two freely available ℓ_1 solver packages, the large-scale ℓ_1 -regularized least-squares (ℓ_1 _ls) [58] and CVX [59], to optimize (6). From our experience, ℓ_1 _ls is the faster of the two, typically having ~ 1 ms run-time per patch, while CVX on average takes ~ 10 ms. However, CVX is more robust, managing to produce reasonable results in cases where ℓ_1 _ls does not converge. Thus, we use ℓ_1 _ls as our ℓ_1 solver except in the following three cases:

1. the algorithm does not converge and gives null output,
2. the output $\mathbf{x}(\hat{j})$ contains a complex number,
3. experimentally we have found that for typical SPGR images, $\|\mathbf{x}(\hat{j})\|_1$ has a Laplacian distribution [60], [61] with mean 1 and small variance (~ 0.005). Thus if $\|\mathbf{x}(\hat{j})\|_1 \gg 1$ for some j , the algorithm is assumed to converge to an undesirable solution.

For these cases, ℓ_1 _ls having failed, we use CVX.

B. Dictionary Selection

A typical $256 \times 256 \times 199$ isotropic MR image, with voxels of size 1 mm^3 , contains about one million non-background patches. It is computationally intractable to solve (6) with all such patches included in the dictionary A'_1 . Recall that for $3 \times 3 \times 3$ patches, our dictionary $A'_1 \in \mathbb{R}^{28 \times N}$, could potentially be a $28 \times 1,000,000$ matrix, if N is assumed to be indexing over all patches in the atlas image a_1 . To avoid the computational bottleneck of large N we construct a dictionary for each patch $\mathbf{b}'_1(j)$, that is of size $N=100$. This is done by selecting 100 patches from the global collection of one million patches, thus drastically reducing run time. In keeping with the literature [45], we construct our patch dictionary, A'_1 , to consist of only those atlas patches that are close to $\mathbf{b}'_1(j)$ in an ℓ_2 sense. It is also in accordance with the assumption that the atlas patches should be close to the subject patch. To achieve this in our setting, we sort the atlas patches, $\mathbf{f}'_1(i)$, $i=1, 2, \dots, N$, by their ℓ_2 distance to the current patch, $\mathbf{b}'_1(j)$. The nearest 100 patches, $\mathbf{f}'_1(i)$, $i=1, 2, \dots, 100$, are then used to generate the dictionary, A'_1 , for the current patch, then $\mathbf{x}(\hat{j})$ is found using (6). To accelerate the search for the 100 nearest patches, we use a k-d tree [62]. We construct A'_2 as the corresponding c_2

contrast patches for the collection $\mathbf{a}_2(i)$, $i = 1, 2, \dots, 100$. Our selection of $N = 100$ is based on our experience, having observed that smaller N does not always yield the best candidate patches and larger N generally shows little improvement in the reconstruction result.

To demonstrate the validity of the choice of atlas patches using such an ℓ_2 proximity criteria between them, Table I shows the fractions of major atlas tissues used to reconstruct a subject patch, averaged over all subject patches from a $256 \times 256 \times 199$ real SPGR image, shown in Fig. 3. The classification of the center voxel of a patch is used to classify the patch. Ideally, the table should contain 1 at the diagonal entries with 0 otherwise. From the table, 78% of the atlas patches, that are used to reconstruct the subject ventricle patches are from atlas ventricle patches, while the remainder are from atlas CSF (16%) and GM (6%). However, we have observed that the non-zero fraction of atlas GM patches used to synthesize subject ventricles can be attributed to the partial volume effect at the boundary of WM and ventricles, which is reconstructed by GM patches. Similarly, the non-zero contributions of atlas CSF and WM patches (10%) to reconstruct subject GM patches can be attributed to the partial volume near CSF-GM and GM-WM boundary. On the other hand, the zero contribution of atlas WM patches to reconstruct subject CSF patches or atlas CSF patches for subject WM patches indicate that the tissues are not “interchanged” while reconstructing, which provides evidence that the $(d + 1)$ -dimensional normalization and the coefficient positivity aspects of the algorithm, as described in Sections III-B and III-C are working as desired.

C. Sparsity regularization coefficient

The sparsity regularization term, λ , is an important parameter in our algorithm as it is used for tuning the sparsity of the reconstruction. Through empirical experiments involving hundreds of thousands of patch reconstructions, we have found the algorithm itself to be very stable for $\lambda \in [0, 0.85]$; but the quality of reconstruction varies within this range. To get a quantitative sense of the synthesis, we experimentally assessed how the error in synthesis is affected by the choice of λ . Accordingly, we created an atlas pair $\{a_1, a_2\}$ using patches from one part (several contiguous slices) of a subject’s SPGR (c_1) and MPRAGE (c_2) and then synthesized other slices of the MPRAGE image from the SPGR. Shown in Fig. 3 are three examples of reconstructed MPRAGEs using different values of λ and a graph of the root mean square error between the synthesized MIMECS MPRAGEs and the true MPRAGE. We conclude from this experiment that values of λ in the range $[0.05, 0.85]$ are visually acceptable and produce similar error levels. However, since our original goal was to recover a solution that satisfies an ℓ_0 criterion, we choose to use a larger λ from this range because sparser solutions produced by ℓ_1 minimization are more likely to agree with the desired ℓ_0 solution [55]. We have therefore used $\lambda = 0.80$ in all of our experimental results reported below.

V. Results

In order to verify the correctness of our MIMECS software implementation, to demonstrate its most basic behavior, and to compare it to histogram matching methods, we first made extensive use of the Brainweb phantom data [63], the Kirby-21 reproducibility data [64], and

the BIRN traveling subject data [18]. These results are omitted here for paper length considerations, but they can be found in [65]. The most important results from this set of experiments can be summarized as follows. First, we found that features and noise that are present in the subject s_1 image but not in the atlas a_1 image are suppressed to some degree in the reconstruction. This was true both in recovering the same tissue contrast (a kind of “sanity check”) and in recovering different tissue contrasts. This observation suggests that MIMECS can be used for noise suppression (though other methods such as non-local means might be better) and also serves as a caution that the MIMECS atlas should contain a patch dictionary that is rich enough to encompass the patches that are expected to be found in the source image. Second, the experiments confirmed that sparsity is important when reconstructing patches with significant detail because otherwise the detail will be smoothed out. Third, MIMECS is fairly robust to the specifics of the subject pulse sequence. In particular, we found that an atlas comprising both an SPGR and MPRAGE can be used to synthesize either an MPRAGE or SPGR image even if the subject image did not precisely match the SPGR or MPRAGE pulse sequence parameters of the atlas. Fourth, we found that MIMECS normalization is comparable or better than histogram matching when normalizing the same tissue contrast (e.g., T1-w from T1-w) and, of course, is far more general in that it can recover missing contrasts. Finally, we found that errors in MIMECS synthesis do not take on specific anatomical features, but instead resemble a typical noise pattern of MR images. This was true in both normalization of the same contrasts and synthesis of alternate contrasts. We note that most of the above observations will be evident in the results below, which is another reason why we did not include these results here.

In this section, we focus on potential uses of MIMECS in clinical and scientific studies rather than focusing solely on performance-oriented criteria (since we have previously reported such results in [47]–[49], [65]). Accordingly, each result should be thought of as a “case study” designed to demonstrate a potential use. Some studies provide numerical results that demonstrate improvement over the state of the art, while others only show compelling visual results. One case study is cautionary, demonstrating a limitation of MIMECS in a potential application.

The results presented here all involve real, not simulated, MR images. Using a standard preprocessing pipeline for neuroimages, the images were skull-stripped [66], [67], corrected for intensity inhomogeneities using N3 [68], and then normalized using a global linear scaling such that the mode of WM intensities were at unity [28]. The MIMECS algorithm, applied to a $256 \times 256 \times 199$ image, takes 1–2 hours on a 12-core 2.7GHz Linux machine using Matlab (2012a, The MathWorks, Natick, MA, USA). The bulk of the computational time is spent in the ℓ_1 solver.

A. Longitudinal analysis of multi-contrast data

In large longitudinal studies (such as the BLSA [19] or ADNI [20]), the scanner and/or acquisition protocol can change over time. An example of such phenomenon is observed in the BLSA data set where 14 scans of a normal subject were acquired in consecutive years. The first 11 scans were acquired using an SPGR protocol on a GE 1.5T scanner, while the last 3 scans were acquired using an MPRAGE protocol on a Philips 3T scanner. It is noted in

[69] that it is difficult to analyze these data consistently. There has been several recent publications trying to address this problem [70]–[72].

To account for the change in acquisition protocol, we used MIMECS to synthesize images having SPGR contrasts from the three MPRAGE scans of one BLSA subject. A subject from the BIRN traveling subject database [18] having both SPGR and MPRAGE scans was used as the atlas. Fig. 4 shows real and synthetic images from this subject. The MIMECS images clearly display the SPGR tissue contrast, which is observed to be very similar to the earlier time frames. The synthetic images are visibly less noisy and slightly blurrier than the original SPGR and MPRAGE images.

Since analysis of longitudinal changes in brain volume is the primary objective in the BLSA study (cf. [12]), we used a state-of-the-art longitudinal segmentation algorithm called *longitudinal FreeSurfer* [71] (longFS) to segment and label the resulting data. LongFS can only be applied to data having the same pulse sequence (either SPGR or MPRAGE), so we were able to apply it to 14 volumes comprising the original 11 SPGR volumes and the 3 MIMECS SPGR volumes. Plots showing the computed GM and ventricle volumes as a function of subject age are shown in Fig. 5 (red with asterisks). For comparison, we applied longFS to each of the longitudinal segments acquired using the same pulse sequence. Accordingly, Fig. 5 also shows the GM and ventricle volumes for the 11 time frames having SPGR (blue with triangles) data and the 3 time frames from both the original MPRAGE (using the *-mprage* flag in longFS) and the MIMECS SPGRs (pink with crosses and green with squares, respectively).

The first key observation to make from Fig. 5 is that longFS yields very different segmentation results and corresponding volume measurements given source data with different pulse sequences. Every neuroimage segmentation algorithm that we have ever tested shares this discrepancy, so this observation should not be interpreted as an indictment of longFS. The second key observation is that MIMECS synthesis permits longFS to make very consistent longitudinal segmentations (at least as far as the volume measured from these segmentations) across the entire collection. Even the segmentation results from the short sequence of synthetic SPGRs are more consistent with the older data. Therefore, we conclude that MIMECS may be a useful preprocessing step prior to analysis of volume changes in longitudinal neuroimaging studies.

B. Average atlas construction

While developing voxel based measures from a population, it is common practice to register all the images to an average atlas space. However, pooling data from two populations can be difficult when individual images from two populations are acquired with different pulse sequences or on heterogeneous scanners. In this case study, we show that by synthesizing contrasts using MIMECS, a better average atlas can be created while pooling subjects from two different data sets. This MIMECS case study represents a new capability; the use of multiple data sources in atlas construction enabling richer and more complete analyses.

As a straight forward demonstration of this, we selected five normal subjects from the BLSA database (1.5T GE, SPGR) and five normal subjects from the OASIS [73] databases (1.5T

Siemens, MPRAGE), all male and right-handed with age range 60–70 years. A state-of-the-art registration tool SyN [7] was used to create average atlases from these ten subjects. A cross-section and zoomed region of this result are shown in Figs. 6(a) and 6(b). We then synthesized volumes with MPRAGE contrast from the five BLSA subjects and created an average atlas from the ten MPRAGE contrast images. This result and a zoomed region are shown in Figs. 6(c) and 6(d). Visually, the atlas created from all MPRAGE contrasts has sharper features than the one with the mixture of SPGR and MPRAGE contrasts (compare Figs. 6(b) and (d)).

To show a quantitative improvement in atlas quality, Figs. 6(e) and 6(f) show the voxel-wise standard deviations of intensities from the ten subjects used to create each of the atlases (after deforming them into the atlas space). It can be seen that the standard deviations are much smaller when synthetic MPRAGEs are combined with OASIS MPRAGEs, especially around the ventricles and in the GM, indicating better feature matching in registration. We then segmented the two atlases using TOADS [2]. A t-test shows that the average standard deviations of CSF, ventricles and GM are significantly lower (p -value < 0.01) for the atlas obtained with all MPRAGE contrasts compared to the one obtained with the mixture of SPGR and MPRAGE contrasts. This average atlas construction case study represents an enhancement to existing technologies, which would provide for richer atlas construction and perhaps more accurate deformation fields carrying each subject into the average atlas space (and vice versa).

C. Segmentation bias in cross-sectional analysis

When pooling data from different studies, the choice of scanning protocols introduces algorithm bias in the quantification even the simplest biomarkers, such as GM and WM volume. To study this bias we used 42 age-matched normal subjects (male, right-handed, and 60–70 years of age), 21 from the BLSA database (all SPGR scans) and 21 from the OASIS [73] database (all MPRAGE scans). The images were then skull-stripped [67] and segmented with FreeSurfer [74]. Plots of average ventricle and brain (GM+WM) volume are shown in Fig. 7. It is known that FreeSurfer generally overestimates brain volume when using SPGR data [75], and we observe this same tendency here (Fig. 7(a)). To test the null hypothesis that volumes for each structure are the same, we use a Wilcoxon rank-sum test with a significance level of 0.01. The average brain volumes (GM+WM) computed from the SPGR scans are significantly higher (p -value < 0.01) than the MPRAGE brain volumes. There was no statistically significant difference for the ventricle volumes for the two populations.

We then used MIMECS (with a BIRN SPGR/MPRAGE atlas, as above) to synthesize MPRAGE contrasts from the SPGR scans and then used FreeSurfer to segment the synthetic scans. As shown in Fig. 7(a), the brain volumes computed this way are comparable to those computed using OASIS/MPRAGE data; the same Wilcoxon rank-sum test shows that the MPRAGE and synthetic MPRAGE come from the same distribution. We also observe that the ventricle volumes do not change in a statistically significant way after synthesis, again, with respect to a Wilcoxon rank-sum test. This case study shows that MIMECS can remove

the systemic bias in volumetric analysis arising from scanner and pulse sequence differences in studies that pool multi-site and multi-center data.

D. Distortion correction in b_0 scans

Diffusion MR images are acquired using echo-planar techniques and are geometrically distorted by susceptibility and eddy currents unless corrected [76]. Although eddy current distortion correction can be carried out by consideration of the MR physics and pulse sequence alone, susceptibility causes subject-dependent distortion and is generally carried out retrospectively by deformably registering the b_0 image to a structural image such as a T1-w or T2-w image [77]. The b_0 image—a minimally-weighted diffusion image—has a tissue contrast similar to that of a conventional T2-w spin-echo image, thus making T2-w images the preferred target for distortion correction. But what happens if no T2-w image was acquired as part of the protocol? In this case, the correction might be attempted by registering the b_0 image to the T1-w image, leading generally to an inferior result. Here, we consider an alternative approach using MIMECS.

Figs. 8(c) and 8(e) show an MPRAGE image and a b_0 image, respectively, acquired in the same imaging session of a subject from the Kirby-21 database [64]. The b_0 image shows significant distortion in the anterior portion of the brain. Fig. 8(g) shows the b_0 image after it has been registered to the MPRAGE image using SyN [7] (with mutual information as the similarity metric). By comparing the GM/WM boundary generated by FreeSurfer segmentation of the MPRAGE (see Fig. 8(f)) to the underlying intensities of the registered b_0 image (see Fig. 8(h)), it is clear that the correction is inadequate.

Given the atlas shown in Figs. 8(a) and 8(b), we used MIMECS to synthesize T2-w image from the MPRAGE image; the result is shown in Fig. 8(d). We then registered the b_0 image to the synthetic T2-w using SyN (with the cross-correlation similarity metric), yielding the distortion-corrected b_0 image shown in Fig. 8(i). The red arrow in Fig. 8(i) indicates an area of improvement, which can be visually confirmed on the zoomed region containing the GM/WM contour shown in Fig. 8(j).

This case study demonstrates a potential use of MIMECS in cases where data is missing. It also reveals a new potential approach to registration of multimodal data, wherein synthesis of the alternate tissue contrast is used with a cross-correlation metric rather than the conventional mutual information metric applied to different tissue contrasts. This concept could have far more general applicability than the simple case example demonstrated here.

E. High resolution T2-w synthesis

Because of time constraints, T2-w scans are often acquired at lower resolution than T1-w scans. If high-resolution (hi-res) T2-w scans were available, however, they could be used for many purposes including lesion segmentation [78], [79] and b_0 dewarping (as described in the previous section). Super-resolution techniques using non-local means have previously been proposed to synthesize hi-res T2-w images [38], [80]. These methods use patches from both the hi-res T1-w image and a lo-res T2-w image and index into an atlas comprising these two images *and* a hi-res T2-w image.

MIMECS provides an alternate way to carry out super-resolution, where the hi-res T2-w image can be synthesized directly from a subject's high-res T1-w image. This means that acquisition of the lo-res T2-w image is unnecessary (for image processing purposes). As an example, consider the SPGR and T2-w images shown in Figs. 9(a) and 9(b), respectively. Both data sets were acquired axially with native resolutions of $0.94 \times 0.94 \times 1.5$ mm and $0.94 \times 0.94 \times 5$ mm, so the low through-plane resolutions are readily apparent as vertical blurring in both of these coronal images—and, clearly, it is much worse in the T2-w image. We use Brainweb [63] T1-w and T2-w images as the atlas images, a_1 and a_2 , as shown in Figs. 9(c) and 9(d), because both are available at high resolution. MIMECS is then used to synthesize a hi-res T2-w image from the SPGR image alone, and the result is shown in Fig. 9(f). For comparison, an upsampled (super-res) result using the non-local means method of [80] is shown in Fig. 9(e). It is readily apparent by visual inspection that MIMECS produces a superior hi-res T2-w image over both the original T2-w acquisition and the non-local means result.

F. MIMECS image segmentation

In this section, we show that MIMECS can also be used as a tissue classification and image segmentation method. In this experiment, we use five atlases, each containing a T1-w SPGR images and its segmentation into CSF, GM, and WM tissue classes. The segmentations, illustrated in the top center of Fig. 10, were carried out by FreeSurfer [74] followed by expert manual corrections.

In order to make use of MIMECS when label images rather than image intensities are involved, we must rethink its steps. Consider a single atlas (an image and its segmentation) and a single voxel (with its corresponding subject patch) in the MIMECS procedure (as described in Section III-B). Determination of the sparse coefficient vector $\mathbf{x}(\hat{j})$, which defines an optimal combination of atlas patches that best match the subject patch (see (5)), can be carried out as usual. But to use this vector to form a linear combination of label patches as in (4) does not make sense since the labels are discrete quantities. Two simple modifications are needed to solve this problem.

First, we form the contrast dictionary A_2 as usual; it comprises a single patch in each column and each patch is a collection of labels from a_2 . Let us represent the labels as follows: CSF ($k=1$), GM ($k=2$), and WM ($k=3$). We now form a vector contrast dictionary $\mathbf{A}_2=(A_2^1, A_2^2, A_2^3)$ with its elements defined by

$$A_2^k = \begin{cases} 1 & \text{if } A_2=k \\ 0 & \text{otherwise} \end{cases} \quad k=1, 2, 3. \quad (7)$$

This process is equivalent to viewing the a_2 dictionary image as a vector membership function rather than a label image. The elements of this dictionary are now interpreted as vector membership functions and we are free to form convex combinations of these vectors in order to produce new (fuzzy) membership functions.

This leads us to the second modification of the usual MIMECS process. It is desirable for the linear combination of vector membership functions to be convex combinations; in this way the elements of the resultant membership function will add up to unity and can also be viewed as either probabilities or partial fractions of the tissue classes. Since the elements of $\hat{\mathbf{x}}(j)$ are non-negative, we can guarantee its use in making a convex combination by simply dividing the inner products by the sum of the elements of $\hat{\mathbf{x}}(j)$. Accordingly, we form a MIMECS-based membership function as follows

$$\mu_k(j) = \frac{A_2^k \hat{\mathbf{x}}(j)}{\|\hat{\mathbf{x}}(j)\|_1}, \quad k=1, 2, 3. \quad (8)$$

The result of this whole process is a fuzzy tissue classification at each voxel and therefore a fuzzy (or soft) segmentation of the subject image. We have found that the result produced with just one atlas is not highly accurate. To produce a more robust result, we apply this exact approach to five atlases and average the resulting membership functions at each voxel. To produce a hard classification—as in the segmentations that are used in the atlas—we use the conventional maximum membership criterion.

An example of the MIMECS tissue segmentation process is shown in Fig. 10. Note that the membership functions (upper right) look very much like the soft segmentations produced by fuzzy C-means or mixture model methods; they can be used exactly as one might use such functions in subsequent segmentation or analysis stages. A hard segmentation is shown on the bottom right. Hard segmentations from SPM, fully-automatic FreeSurfer (with default parameters), and manually-corrected FreeSurfer are shown for comparison (see bottom row). The MIMECS segmentation has many desirable visual features and compares quite favorably to the manually-assisted FreeSurfer result.

For quantitative comparison, Dice coefficients for the SPM, fully-automatic FreeSurfer, and MIMECS results against the manually-corrected FreeSurfer result were computed, yielding 0.8630, 0.8811, 0.8837, respectively. MIMECS shows a small improvement in this example. The most important point to be made is that this MIMECS segmentation approach does not rely on any statistical (spatial or intensity) prior. Instead it obtains its result through the use of dictionary examples alone. Similar methods in the literature (e.g., [39], [81]–[83]) are gaining wide support because of their superior performance and generalizability in comparison to model-based methods.

G. FLAIR synthesis

FLAIR (fluid attenuated inversion recovery) images have become important in detecting lesions in the brain, especially in WM. Many older studies did not use the FLAIR pulse sequence, so image processing algorithms designed to exploit the FLAIR tissue contrast cannot be directly applied. Also, since the FLAIR pulse sequence is very sensitive to patient motion some FLAIR images are “spoiled” by motion artifacts and are also not amenable to image processing algorithms. We therefore carried out a study on the use MIMECS to synthesize FLAIR images.

The atlas we used for this experiment has three images: a_1 has an MPRAGE contrast, a_2 has a T2-w contrast, and a_3 has a FLAIR contrast. Cross-section of the atlas images are shown in the left-hand column of Fig. 11. The goal is to apply MIMECS using two subject images—an MPRAGE image and a T2-w image—to synthesize a FLAIR image. Both the atlas and the subject has WM lesions, so the presence of a unique lesion patch signature in the subject should also be present in the atlas.

Cross-sections of subject images and the synthetic MIMECS FLAIR image are shown in the right-hand column of Fig. 11. The true FLAIR image for this subject, which is not used in the synthesis algorithm, is shown for comparison. It is immediately evident that the lesions in the synthetic FLAIR image have been rendered as if they are GM—i.e., they are not bright like lesions in true FLAIR images. It is relatively easy to understand why this happens. If one visually identifies a lesion in the (atlas or subject) FLAIR image, it is readily apparent that these same regions on both the MPRAGE and T2-w have intensities that are similar to GM within their respective tissue contrasts. Thus, patches involving lesions in *both* MPRAGE and T2-w are indistinguishable from patches involving GM.

This experiment reveals a limitation of MIMECS synthesis. Stated simply, the set of pulse sequences used to synthesize a new image must contain intensity signatures that can uniquely identify the tissue classes that are present in the tissue contrast to be synthesized. This is certainly a cautionary note about MIMECS synthesis and it deserves further study to determine its specific limitations. On the other hand, the ability to synthesize images devoid of lesions presents an opportunity to enhance lesions by subtraction. The bottom right image in Fig. 11 is a subtraction image (true subject FLAIR minus synthetic FLAIR) that shows potential for visual enhancement of lesions or for computational assistance in finding lesions.

VI. Summary and Discussion

MIMECS is a new approach to image intensity normalization and tissue contrast synthesis. It is based on sparse dictionary reconstruction methods with a novel patch normalization approach that makes dictionary selection straightforward. For each voxel, a dictionary is found by casting all patches into one higher dimension and searching for “like” patches using a rapid kd-tree search with the ℓ_2 metric. A sparse and non-negative combination of dictionary patches is found to match the subject patch is found using an ℓ_1 solver. These same coefficients are then used to combine patches in another dictionary that is physically matched to the first dictionary. This process is shown in several provided examples to be capable of providing intensity normalization, tissue contrast synthesis, and tissue segmentation.

MIMECS shares strong similarities to non-local means methods [45], some of which have been applied in the medical imaging community for noise reduction [42], [84], [85], super-resolution [80], and segmentation [39], [41], [81]. Including our earlier conference publications on this topic [47]–[49], [86], MIMECS is the first approach of this class to be proposed and evaluated for contrast synthesis. The use of nonnegative coefficients in combining patches is an important aspect, heretofore not considered, in using the patch-based

framework for contrast synthesis. As well, the proposed dictionary selection technique, which uses a higher-dimensional space to normalize patches, is novel and important in practical scenarios since it avoids prior segmentation (as we used in earlier reports) or atlas selection training. Finally, the importance of sparsity was demonstrated in Section IV-C and Fig. 3.

Comparisons with histogram matching methods (cf. [25], [87]) were omitted from this paper for length considerations, but results are included in [65] and previous conference reports [47]–[49]. Histogram matching methods are used in nearly every neuroimage processing pipeline and remain important. In fact, MIMECS depends on a crude histogram matching method (linear normalization to the WM peak) at its outset. But histogram matching has serious limitations. For example, histogram matching in its most basic form changes the intensities of the subject image so that its histogram is exactly equal to that of the target. If a simple tissue classifier (such as the EM algorithm for a Gaussian mixture model) were applied to these two images, then the computed volumes of the corresponding tissue classes would be identical. This process would therefore disguise potential differences in a population of subject images. More sophisticated histogram matching methods have been designed, of course, and their results are more sensible. But the whole approach becomes more suspect as the pulse sequences vary significantly and it is entirely wrong to perform histogram matching between images that have fundamentally different tissue contrasts. MIMECS therefore provides an alternative to histogram matching which does not suffer from these fundamental problems.

Our results demonstrate a collection of potential applications of MIMECS. We particularly emphasize the potential for MIMECS to solve the “missing” or “mis-matched” tissue contrast problem in routine neuroimage processing tasks. For example, the BLSA longitudinal study has been active for so long that the SPGR images that were acquired at 1.5 T on older scanners has been replaced by MPRAGE images that are acquired at 3.0 T on modern scanners. MIMECS offers a straightforward way to carry out consistent volumetric analysis across this large time span. The use of diverse data from multiple institutions and multiple studies creates similar difficulties, as illustrated by our average atlas and segmentation bias examples. These examples showed how MIMECS could be used to synthesize “like” data for potential improvement in the statistical analysis of anatomical shape differences through average atlas or for merging volumetric analysis of normal subjects for greater power in cross-sectional studies.

The average atlas example demonstrated how registration could be improved by synthesizing images having the same tissue contrast. This was taken a step further with the example involving geometric distortion correction in diffusion MRI. Taken together, these two examples reveal a new approach to multimodal registration, one in which a “proxy” image is synthesized from either the subject or target image and registration is carried out using a simple image similarity criterion such as cross-correlation. This approach deserves further scrutiny since it provides an alternative to the mutual information similarity metric, which is problematic to optimize and often fails to provide accurate results, especially in deformable registration applications.

Super-resolution algorithms, and especially patch-based methods have gained a lot of attention in recent years [81], [83]. The MIMECS framework has strong similarity to other patch-based methods, but offers a twist—direct synthesis of an alternate tissue contrast—that could be advantageous. In fact, it is not difficult to envision putting some of the fundamental notions together, for example, having a multi-resolution atlas within the MIMECS framework. Again, certain elements of the MIMECS framework—non-negative patch combination and patch normalization in a higher-dimensional space—offer immediate advantages.

Use of patches for direct segmentation has been previously reported [81]. The advantages of MIMECS for this purpose are not clearly established in this paper as we did not include a comparison with these particular methods. However, our comparison to established state-of-the-art segmentation algorithms demonstrates a strong potential for its use in this way in the future. Like other patch-based methods, the use of examples rather than generative models may be advantageous when models are difficult to ascertain with precision as is often the case in medical imaging. A key limitation of this approach at present is that it provides only a tissue classification, not a structural labeling. FreeSurfer, to which it is compared subdivides the GM into anatomical structures with labels, a critically important set in many neuroimaging analyses.

Our final experiment involved FLAIR image synthesis, and it provides a cautionary lesson about the limitations of MIMECS. The FLAIR pulse sequence is unique in its ability to reveal WM lesions by nullifying the CSF signal using a 180° inversion recovery pulse. This accentuates the lesion signal relative to all other brain intensities. It is a nonlinear relationship to the signal coming from other pulse sequences because of the inversion recovery step, and this is (partly) what makes it impossible to synthesize using MIMECS directly. Lesions are not invisible in MPRAGE and T2-w images, but they are not distinguishable from GM by intensity and local pattern alone. We note that PD images, typically collected along with the T2-w images, do not help us since the lesions in a PD image also look like GM.

While we did illustrate how synthesizing images containing the *absence* of lesions could potentially be useful in lesion detection, the question still remains as to whether alternate methods or different source images might make FLAIR synthesis possible. It is also natural to ask what other anatomical features or pathologies might not be synthesized using standard MIMECS techniques and how these omissions might affect subsequent image processing steps such as segmentation and registration.

In summary, our case studies demonstrate the immediate utility of MIMECS for a wide variety of neuroimage processing tasks. There remains a great deal of flexibility in its use, particularly in the choice of atlas and source image(s) and new applications can be expected to be found. MIMECS represents a new class of neuroimage processing methods with a potentially rich future.

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References

1. Bezdek JC, Hall LO, Clarke LP. Review of MR image segmentation techniques using pattern recognition. *Med Physics*. 1993; 20(4):1033–1048.
2. Bazin PL, Pham DL. Topology-preserving tissue classification of magnetic resonance brain images. *IEEE Trans Med Imag*. Apr; 2007 26(4):487–496.
3. Roy S, Agarwal H, Carass A, Bai Y, Pham DL, Prince JL. Fuzzy c-means with variable compactness. *Intl Sym on Biomed Imag (ISBI)*. May.2008 :452–455.
4. Clark KA, Woods RP, Rottenberg DA, Toga AW, Mazziotta JC. Impact of acquisition protocols and processing streams on tissue segmentation of T1 weighted MR images. *NeuroImage*. 2006; 29(1): 185–202. [PubMed: 16139526]
5. Boesen K, Rehm K, Schaper K, Stoltzner S, Woods R, Lüders E, Rottenberg D. Quantitative comparison of four brain extraction algorithms. *NeuroImage*. Jul; 2004 22(3):1255–1261. [PubMed: 15219597]
6. Roy S, Carass A, Bazin PL, Resnick SM, Prince JL. Consistent Segmentation using a Rician Classifier. *Medical Image Analysis*. 2012; 16(2):524–535. [PubMed: 22204754]
7. Avants BB, Epstein CL, Grossman M, Gee JC. Symmetric diffeomorphic image registration with cross-correlation: evaluating automated labeling of elderly and neurodegenerative brain. *Medical Image Analysis*. 2008; 12(1):26–41. [PubMed: 17659998]
8. Rohde GK, Aldroubi A, Dawant BM. The adaptive bases algorithm for intensity-based nonrigid image registration. *IEEE Trans Med Imag*. 2003; 22:1470–1479.
9. Rorden C, Bonilha L, Fridriksson J, Bender B, Karnath H. Age-specific CT and MRI templates for spatial normalization. *NeuroImage*. 2012; 61(4):957–966. [PubMed: 22440645]
10. Chen, M.; Carass, A.; Bogovic, J.; Prince, P-LBJL. Distance transforms in multi channel MR image registration. *Proceedings of SPIE Medical Imaging (SPIE-MI 2011)*; Lake Buena Vista, FL. February 12–17, 2011; 2011.
11. Ellingsen LM, Prince JL. Mjolnir: Extending HAMMER using a diffusion transformation model and histogram equalization for deformable image registration. *International Journal of Biomedical Imaging*. 2009; 2009:281615. [PubMed: 19680457]
12. Resnick SM, Pham DL, Kraut MA, Zonderman AB, Davatzikos C. Longitudinal Magnetic Resonance Imaging Studies of Older Adults: A Shrinking Brain. *Journal of Neuroscience*. 2003; 23(8):3295–3301. [PubMed: 12716936]
13. Thambisetty M, Wan J, Carass A, An Y, Prince JL, Resnick SM. Longitudinal changes in cortical thickness associated with normal aging. *NeuroImage*. 2010; 52(4):1215–1223. [PubMed: 20441796]
14. Querbes O, Aubry F, Pariente J, Lotterie J, Démonet J, Duret V, Puel M, Berry I, Fort J-C, Celsis P, and The Alzheimer's Disease Neuroimaging Initiative. Early diagnosis of Alzheimer's disease using cortical thickness: impact of cognitive reserve. *Brain*. 2009; 132(8):2036–2047. [PubMed: 19439419]
15. Raz N, Rodrigue KM, Acker JD. Hypertension and the brain: Vulnerability of the prefrontal regions and executive functions. *Behavioral Neuroscience*. 2003; 117(6):1169–1180. [PubMed: 14674838]
16. Heckemann RA, Keihaninejad S, Aljabar P, Rueckert D, Hajnal JV, Hammers A, and The Alzheimer's Disease Neuroimaging Initiative. Improving intersubject image registration using

tissue-class information benefits robustness and accuracy of multi-atlas based anatomical segmentation. *NeuroImage*. 2010; 51(1):221–227. [PubMed: 20114079]

17. Unschuld PG, Edden RAE, Carass A, Liu X, Shanahan M, Wang X, Oishi K, Brandt J, Bassett SS, Redgrave GW, Margolis RL, van Zijl PCM, Barker PB, Ross CA. Brain metabolite alterations and cognitive dysfunction in early Huntington's disease. *Movement Disorders*. Jun; 2012 27(7):895–902. [PubMed: 22649062]
18. Friedman L, Stern H, Brown GG, Mathalon DH, Turner J, Glover GH, Gollub RL, Lauriello J, Lim KO, Cannon T, Greve DN, Bockholt HJ, Belger A, Mueller B, Doty MJ, He J, Wells W, Smyth P, Pieper S, Kim S, Kubicki M, Vangel M, Potkin SG. Test-Retest and Between-Site Reliability in a Multicenter fMRI Study. *Human Brain Mapping*. Aug; 2008 29(8):958–972. [PubMed: 17636563]
19. Kawas C, Gary S, Brookmeyer R, Fozard J, Zonderman A. Age-specific incidence rates of Alzheimer's disease: the Baltimore Longitudinal Study of Aging. *Neurology*. Jun; 2000 54(11):2072–2077. [PubMed: 10851365]
20. Mueller SG, Weiner MW, Thal LJ, Petersen RC, Jack C, Jagust W, Trojanowski JQ, Toga AW, Beckett L. The Alzheimer's disease neuroimaging initiative. *Neuroimaging Clin N Am*. Nov; 2005 15(4):869–877. [PubMed: 16443497]
21. Walter C, Kruessell M, Gindele A, Brochhagen HG, Gossmann A, Landwehr P. Imaging of renal lesions: evaluation of fast mri and helical ct. *Br J Radiol*. Oct; 2003 76(910):696–703. [PubMed: 14512329]
22. Bergeest, JP.; Jäger, F. *Bildverarbeitung für die Medizin 2008*, ser. Informatik Aktuell. Vol. ch. 8. Springer; Berlin Heidelberg: 2008. A Comparison of Five Methods for Signal Intensity Standardization in MRI; p. 36-40.
23. Jäger, F.; Nyúl, L.; Frericks, B.; Wacker, F.; Hornegger, J. *Bildverarbeitung für die Medizin 2008*, ser. Informatik Aktuell. Vol. ch. 20. Springer; Berlin Heidelberg: 2007. Whole Body MRI Intensity Standardization; p. 459-463.
24. Nyúl LG, Udupa JK. On Standardizing the MR Image Intensity Scale. *Mag Reson Med*. 1999; 42(6):1072–1081.
25. Nyúl LG, Udupa JK, Zhang X. New Variants of a Method of MRI Scale Standardization. *IEEE Trans Med Imag*. Feb; 2000 19(2):143–150.
26. Madabhushi A, Udupa JK. New methods of MR image intensity standardization via generalized scale. *Med Physics*. Sep; 2006 33(9):3426–3434.
27. He R, Datta S, Tao G, Narayana PA. Information measures-based intensity standardization of MRI. *Intl Conf Engg in Med and Biology Soc*. Aug.2008 :2233–2236.
28. Christensen JD. Normalization of brain magnetic resonance images using histogram even-order derivative analysis. *Mag Reson Im*. Sep; 2003 21(7):817–820.
29. Weisenfeld NL, Warfield SK. Normalization of Joint Image-Intensity Statistics in MRI Using the Kullback-Leibler Divergence. *Intl Symp on Biomed Imag (ISBI)*. Apr.2004 1:101–104.
30. Ellingsen LM, Prince JL. Mjólnir: Extending HAMMER Using a Diffusion Transformation Model and Histogram Equalization for Deformable Image Registration. *Int J Biomed Imaging*. 2009; 2009(281615)
31. Dai Y, Wang Y, Wang L, Wu G, Shi F, Shen D. and Alzheimer's Disease Neuroimaging Initiative, . aBEAT: A Toolbox for Consistent Analysis of Longitudinal Adult Brain MRI. *PLoS One*. 2012; 8(4):60344.
32. Han X, Fischl B. Atlas Renormalization for Improved Brain MR Image Segmentation Across Scanner Platforms. *IEEE Trans Med Imag*. 2007; 26(4):479–486.
33. Fischl B, Salat DH, van der Kouwe AJW, Makris N, Ségonne F, Quinn BT, Dale AM. Sequence-independent segmentation of magnetic resonance images. *NeuroImage*. 2004; 23(Suppl 1):S69–84. [PubMed: 15501102]
34. Prince JL, Tan Q, Pham DL. Optimization of MR Pulse Sequences for Bayesian Image Segmentation. *Med Physics*. Oct; 1995 22(10):1651–1656.
35. Deichmann R, Good CD, Josephs O, Ashburner J, Turner R. Optimization of 3-D MP-RAGE Sequences for Structural Brain Imaging. *NeuroImage*. Jul; 2000 12(1):112–127. [PubMed: 10875908]

36. Miller MI, Christensen GE, Amit Y, Grenander U. Mathematical textbook of deformable neuroanatomies. *Proc Natl Acad Sci*. Dec; 1993 90(24):11 944–11 948. [PubMed: 8093556]
37. Sabuncu MR, Yeo BTT, Van Leemput K, Fischl B, Golland P. A generative model for image segmentation based on label fusion. *IEEE Trans Med Imag*. 2010; 29(10):1714–1729.
38. Rousseau, F. Brain Hallucination. *Proc. of the 10th European Conf. on Comp. Vision*; 2008. p. 497-508.
39. Coupé P, Eskildsen SF, Manjn JV, Fonov VS, Collins DL. and the Alzheimer's disease Neuroimaging Initiative. . Simultaneous segmentation and grading of anatomical structures for patient's classification: application to Alzheimer's disease. *NeuroImage*. 2012; 59(4):3736–3747. [PubMed: 22094645]
40. Buades A, Coll B, Morel JM. A Non-Local Algorithm for Image Denoising. *IEEE Comp Vision and Patt Recog*. 2005; 2:60–65.
41. Hu S, Coupé P, Pruessner JC, Collins DL. Nonlocal regularization for active appearance model: Application to medial temporal lobe segmentation. *Human Brain Mapping*. 2012; 00(00):00.
42. Kervrann, C.; Boulanger, J.; Coupé, P. Bayesian Non-local Means Filter, Image Redundancy and Adaptive Dictionaries for Noise Removal. *Proc. of 1st international conference on Scale Space and Variational Methods in Comp. Vision*; 2007. p. 520-532.
43. Yang J, Wright J, Huang TS, Ma Y. Image Super-Resolution Via Sparse Representation. *IEEE Trans Imag Proc*. Nov; 2010 19(11):2861–2873.
44. Candes EJ, Romberg JK, Tao T. Stable signal recovery from incomplete and inaccurate measurements. *Comm on Pure and Appl Math*. Aug; 2006 59(8):1207–1223.
45. Mairal J, Bach F, Ponce J, Sapiro G, Zisserman A. Non-local sparse models for image restoration. *IEEE Intl Conf on Comp Vision*. 2009:2272–2279.
46. Elad M, Aharon M. Image Denoising Via Sparse and Redundant Representations Over Learned Dictionaries. *IEEE Trans Imag Proc*. 2006; 15(12):3736–3745.
47. Roy S, Carass A, Prince JL. A Compressed Sensing Approach For MR Tissue Contrast Synthesis. *Inf Proc in Med Imaging (IPMI)*. 2011:371–383.
48. Roy, S.; Carass, A.; Prince, JL. Synthesizing MR contrast and resolution through a patch matching technique. *Proceedings of SPIE Medical Imaging (SPIE)*; 2010. p. 7623
49. Roy S, Carass A, Prince JL. Compressed sensing based intensity non-uniformity correction. *Intl Sym on Biomed Imag (ISBI)*. Apr.2011 :101–104.
50. Mairal J, Sapiro G, Elad M. Learning Multiscale Sparse Representations for Image and Video Restoration. *Multiscale Model Simul*. 2008; 7(1):214–241.
51. Elad M, Feuer A. Restoration of a Single Superresolution Image from Several Blurred, Noisy, and Undersampled Measured Images. *IEEE Trans Imag Proc*. 1997; 6(12):1646–1658.
52. Aharon M, Elad M, Bruckstein A. K-SVD: An Algorithm for Designing Overcomplete Dictionaries for Sparse Representation. *IEEE Trans Sig Proc*. 2006; 54(11):4311–4322.
53. Kwatra V, Schödl A, Essa I, Turk G, Bobick A. Graphcut Textures: Image and Video Synthesis Using Graph Cuts. *ACM Trans on Graphics*. 2003; 22(3):277–286.
54. Hertzmann, A.; Jacobs, CE.; Oliver, N.; Curless, B.; Salesin, DH. Image analogies. *Proc. of 28th Ann. Conf. on Comp. Graphics and Interactive Techniques, ser. SIGGRAPH '01*; 2001. p. 327-340.
55. Donoho DL. For most large underdetermined systems of linear equations, the minimal ℓ_1 -norm near-solution approximates the sparsest near-solution. *Communications on Pure and Applied Mathematics*. 2006; 59(7):907–934.
56. Tibshirani R. Regression Shrinkage and Selection via the Lasso. *Journal of Royal Stat Soc Series B (Methodological)*. 1996; 58(1):267–288.
57. Donoho DL, Tanner J. Sparse Nonnegative Solution of Underde-termined Linear Equations by Linear Programming. *Proc Natl Acad Sci*. 2005; 102(27):9446–9451. [PubMed: 15976026]
58. Koh K, Kim S, Boyd S. An interior-point method for large-scale ℓ_1 -regularized logistic regression. *Journal of Machine Learning Research*. 2007; 8:1519–1555.

59. Grant, M.; Boyd, S. Recent Advances in Learning and Control, ser. Lecture Notes in Control and Information Sciences. Springer-Verlag Limited; 2008. Graph implementations for nonsmooth convex programs; p. 95-110.
60. Elad M, Milanfar P, Rubinstein R. Analysis versus synthesis in signal priors. *Inverse Problems*. 2007; 23(3):947968.
61. Seeger MW. Bayesian Inference and Optimal Design for the Sparse Linear Model. *Journal of Machine Learning Research*. 2008; 9:759–813.
62. Friedman JH, Bentley JL, Finkel RA. An Algorithm for Finding Best Matches in Logarithmic Expected Time. *ACM Trans on Mathematical Software*. 1977; 3(3):209–226.
63. Cocosco CA, Kollokian V, Kwan RKS, Evans AC. BrainWeb: Online Interface to a 3D MRI Simulated Brain Database. *NeuroImage*. 1997; 5(4):S425.
64. Landman BA, Huang AJ, Gifford A, Vikram DS, Lim IL, Farrell JAD, Bogovic JA, Hua J, Chen M, Jarso S, Smith SA, Joel S, Mori S, Pekar JJ, Barker PB, Prince JL, van Zijl PCM. Multi-parametric neuroimaging reproducibility: A 3-T resource study. *NeuroImage*. 2011; 54(4):2854–2866. [PubMed: 21094686]
65. Roy, S. PhD dissertation. Johns Hopkins University; 2013. MR Image Contrast Synthesis for Consistent Segmentation. [Online]. Available: <http://iacl.ece.jhu.edu/~roy/roythesisjhu2013.pdf>
66. Carass A, Wheeler MB, Cuzzocreo J, Bazin P-L, Bassett SS, Prince JL. A Joint Registration and Segmentation Approach to Skull Stripping. *Intl Sym on Biomed Imag (ISBI)*. Apr.2007 :656–659.
67. Carass A, Cuzzocreo J, Wheeler MB, Bazin PL, Resnick SM, Prince JL. Simple paradigm for extra-cerebral tissue removal: Algorithm and analysis. *NeuroImage*. 2011; 56(4):1982–1992. [PubMed: 21458576]
68. Sled JG, Zijdenbos AP, Evans AC. A non-parametric method for automatic correction of intensity non-uniformity in MRI data. *IEEE Trans Med Imag*. 1998; 17(1):87–97.
69. Moghekar A, Kraut M, Elkins W, Troncoso J, Zonderman AB, Resnick SM, O'Brien RJ. Cerebral white matter disease is associated with Alzheimer pathology in a prospective cohort. *Alzheimer's & Dementia*. 2012; 8(5):S71–S77.
70. Xue Z, Shen D, Davatzikos C. CLASSIC: Consistent Longitudinal Alignment and Segmentation for Serial Image Computing. *NeuroImage*. 2006; 30(2):388–399. [PubMed: 16275137]
71. Reuter M, Schmansky NJ, Rosas HD, Fischl B. Within-Subject Template Estimation for Unbiased Longitudinal Image Analysis. *NeuroImage*. 2012; 61(4):1402–1418. [PubMed: 22430496]
72. Roy, S.; Carass, A.; Prince, JL. Longitudinal Intensity Normalization of Magnetic Resonance Images using Patches. *Proceedings of SPIE Medical Imaging (SPIE-MI 2013)*; Lake Buena Vista, FL. February 9–14, 2013; 2013. p. 86691J
73. Marcus DS, Wang TH, Parker J, Csernansky JG, Morris JC, Buckner RL. Open Access Series of Imaging Studies (OASIS): cross-sectional MRI data in young, middle aged, nondemented, and demented older adults. *Journal of Cognitive Neuroscience*. Sep; 2007 19(9):1498–1507. [PubMed: 17714011]
74. Dale AM, Fischl B, Sereno MI. Cortical Surface-Based Analysis I: Segmentation and Surface Reconstruction. *NeuroImage*. 1999; 9(2):179–194. [PubMed: 9931268]
75. Klauschen F, Goldman A, Barra V, Meyer-Lindenberg A, Lundervold A. Evaluation of Automated Brain MR Image Segmentation and Volumetry Methods. *Human Brain Mapping*. 2009; 30(4):1310–1327. [PubMed: 18537111]
76. Jezzard P, Balaban RS. Correction for geometric distortion in echo planar images from B_0 field variations. *Mag Reson Med*. 1995; 34(1):65–73.
77. Ardekani S, Sinha U. Geometric distortion correction of high-resolution 3T diffusion tensor brain images. *Mag Reson Med*. 2005; 54(5):1163–1171.
78. Souplet, J.; Lebrun, C.; Ayache, N.; Malandain, G. An Automatic Segmentation of T2-FLAIR Multiple Sclerosis Lesions. *Multiple Sclerosis Lesion Segmentation Challenge Workshop (MICCAI 2008 Workshop)*; 2008.
79. Wu Y, Warfield SK, Tan IL, Wells WM, Meier DS, van Schijndel RA, Barkhof F, Guttman CRG. Automated segmentation of multiple sclerosis lesion subtypes with multichannel MRI. *NeuroImage*. 2006; 32(3):1205–1215. [PubMed: 16797188]

80. Manjon JV, Coupé P, Buades A, Fonovan V, Collins DL, Robles M. Non-Local MRI Upsampling. *Medical Image Analysis*. 2010; 14(2):784–792. [PubMed: 20566298]
81. Coupé P, Manjon JV, Pruessner J, Robles M, Fonov V, Collins DL. Patch-based segmentation using expert priors: Application to hippocampus and ventricle segmentation. *NeuroImage*. 2011; 54(2): 940–954. [PubMed: 20851199]
82. Eskildsen SF, Coupé P, Fonov V, Manjon JV, Leung KK, Guizard N, Wassef SN, Ostergaard LR, Collins DL. and The Alzheimer’s Disease Neuroimaging Initiative, . BEaST: Brain extraction based on nonlocal segmentation technique. *NeuroImage*. 2012; 59(3):2362–2373. [PubMed: 21945694]
83. Rousseau F, Habas PA, Studholme C. A Supervised Patch-Based Approach for Human Brain Labeling. *IEEE Trans Med Imag*. 2011; 30(10):1852–1862.
84. Manjón JV, Carbonell-Caballero J, Lull JJ, Garcia-Marti G, Marti-Bonmati L, Robles M. MRI denoising using non-local means. *Medical Image Analysis*. 2008; 12(4):514–523. [PubMed: 18381247]
85. Wiest-Daessle N, Prima S, Coupé P, Morrissey SP, Barillot C. Rician noise removal by non-Local Means filtering for low signal-to-noise ratio MRI: applications to DT-MRI. *Med Image Comp and Comp Asst Intervention (MICCAI)*. 2008; 11:171–179.
86. Roy S, Carass A, Shiee N, Pham DL, Prince JL. MR Contrast Synthesis for Lesion Segmentation. *Intl Sym on Biomedical Imaging*. 2010:932–935.
87. Wang L, Lai H, Barker GJ, Miller DH, Tofts PS. Correction for variations in MRI scanner sensitivity in brain studies with histogram matching. *Mag Reson Med*. Feb; 1998 39(2):322–327.

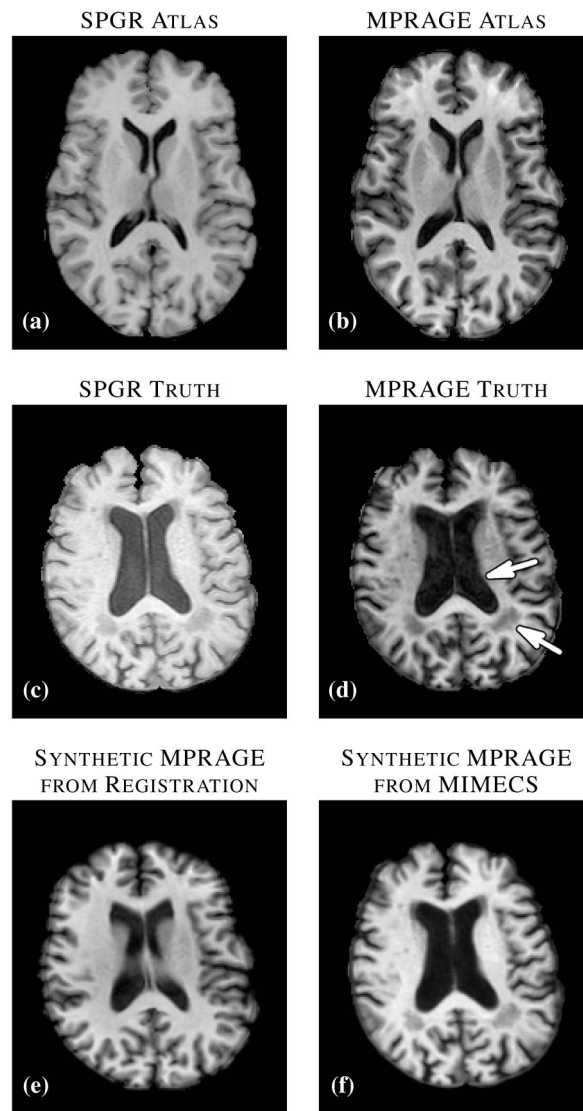


Fig. 1.

(a) Atlas T1-weighted SPGR and (b) its corresponding T1-weighted MPRAGE. (c) A subject T1-weighted SPGR scan and (d) its T1-weighted MPRAGE image. The atlas SPGR is deformably registered (using SyN [7]) to the subject SPGR. This deformation is applied to the atlas MPRAGE to obtain (e) a synthetic subject MPRAGE. (f) The synthetic MPRAGE image generated by our algorithm, MIMeCS.

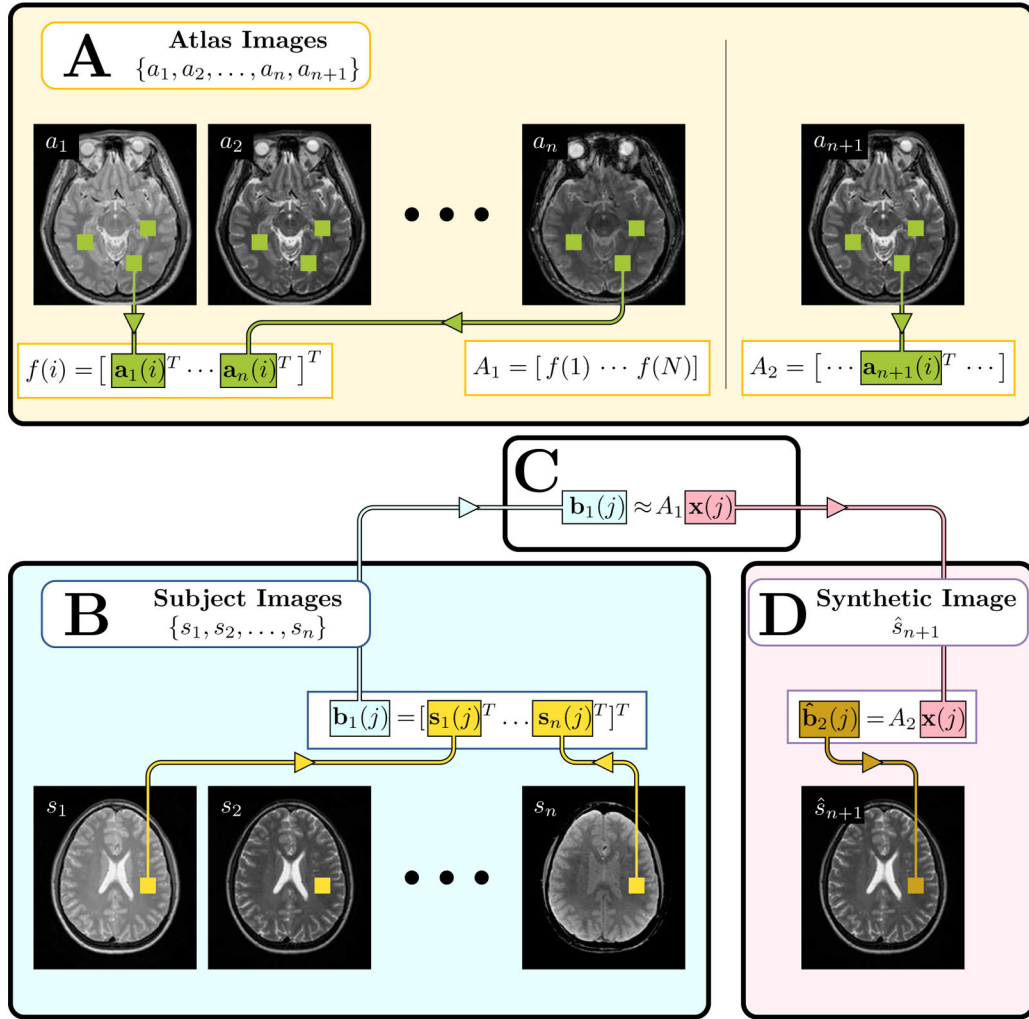


Fig. 2. An illustration of the MIMACS algorithm. The region of the image labeled A, shows the construction of the atlases A_1 and A_2 , see Section III-A for details. Region B shows the input subject images converted into patches $\mathbf{b}_1(j)$. Region C denotes the estimation of the coefficients $\mathbf{x}(j)$ which relate $\mathbf{b}_1(j)$ to the patches of the atlas A_1 . Finally, region D shows the computation of the synthetic image \hat{s}_{n+1} by using the learned coefficients $\mathbf{x}(j)$ to compute the patch $\hat{\mathbf{b}}_2(j)$ based on the atlas A_2 . Section III-B describes regions B, C, and D.

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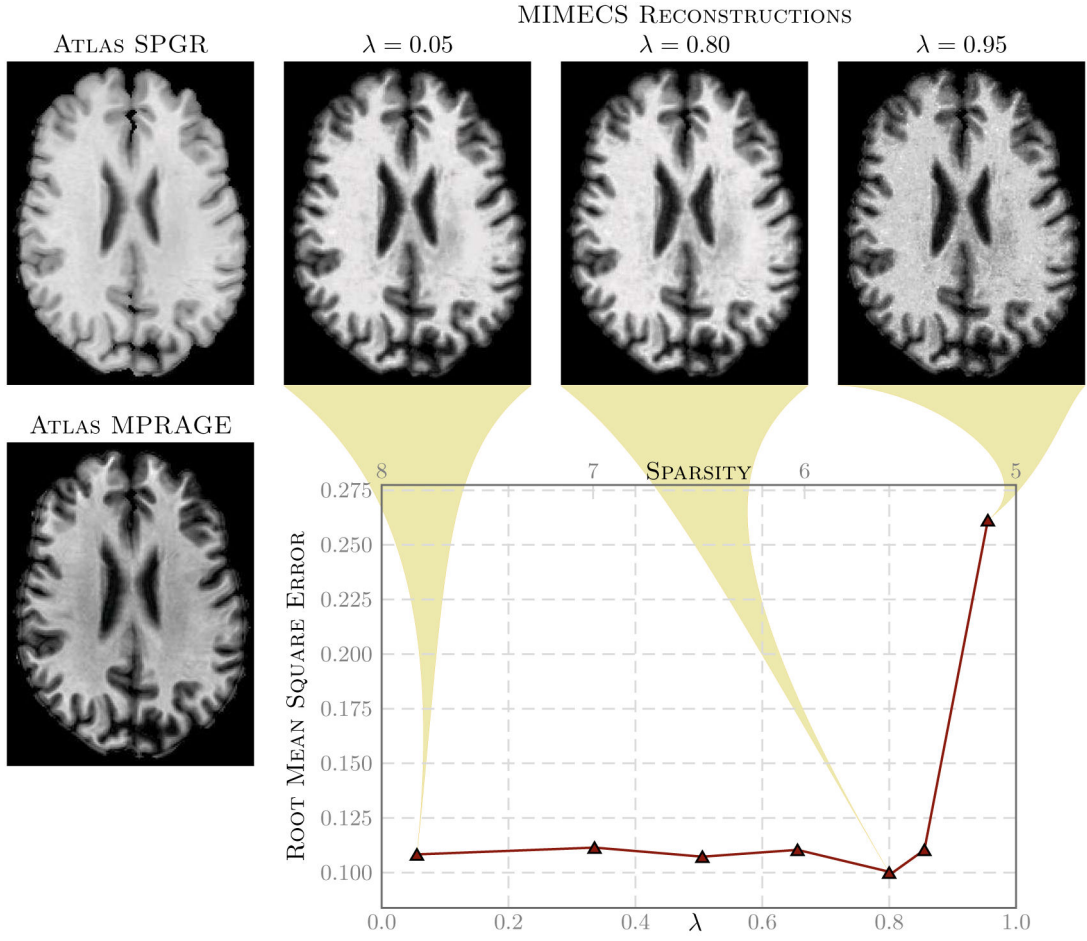


Fig. 3. Effect of sparsity of $x(j)$ on the c_2 contrast
 The left most column shows a subject’s SPGR (top) and MPAGE (bottom) acquisitions. The second, third, and fourth columns of the top row show synthetic MPAGEs generated using another portion of the subject’s MPAGE as the atlas. The synthetic MPAGEs were generated using λ values of 0.05, 0.80, and 0.95, respectively. The plot shows the MPAGE synthesis error vs. the average sparsity of all $x(j)$ ’s, averaged over all non-zero voxels. The average sparsity scale is on the top of the plot while the sparsity regularization parameter, λ , is plotted on the bottom axis.

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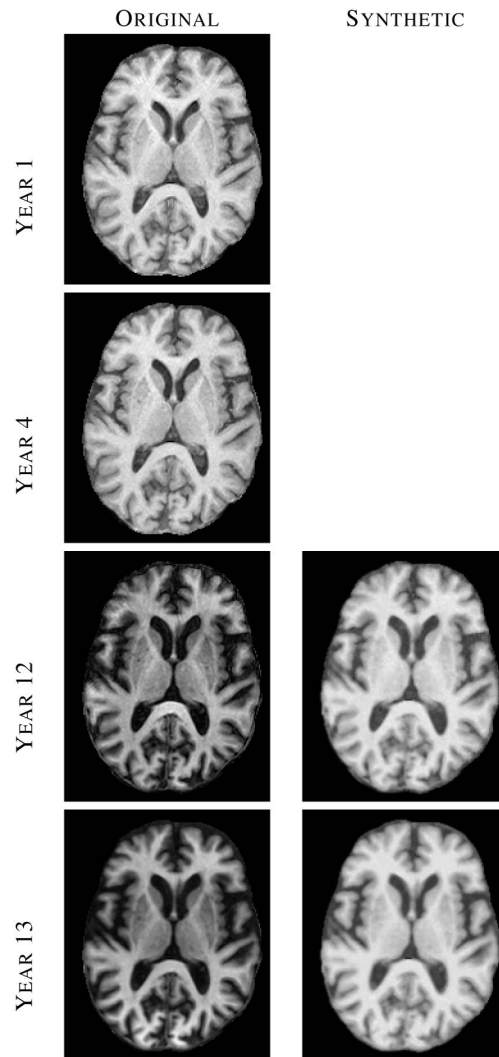


Fig. 4. The left column shows images from four (out of fourteen) time-points of a normal BLSA subject, where each image was acquired approximately one year apart. The right column shows synthetic SPGR images, synthesized using MIMICS applied to the corresponding MPRAGE images in the left column.

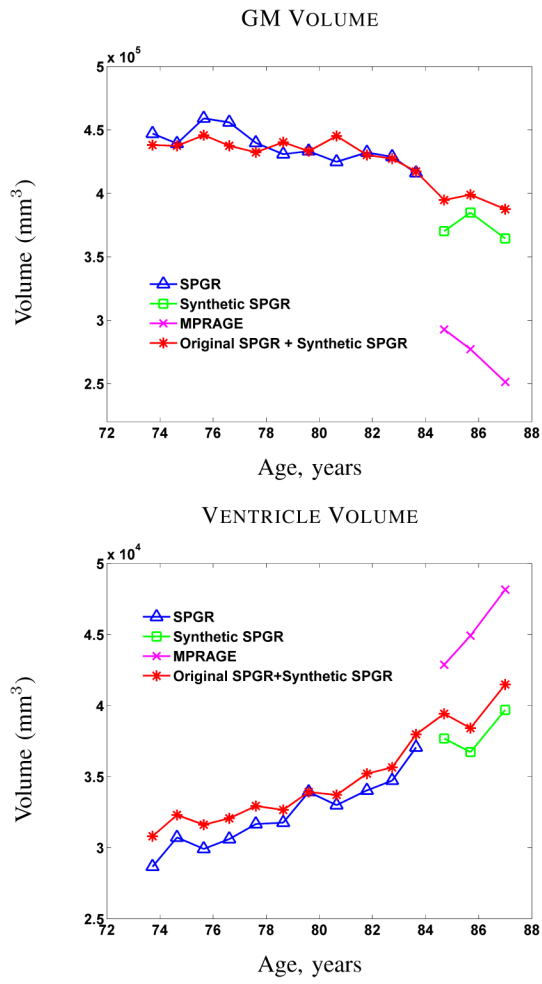


Fig. 5. GM and ventricle volumes from longitudinal FreeSurfer [71] of a normal subject with 14 scans (first 11 are SPGR, last 3 are MPRAGE).

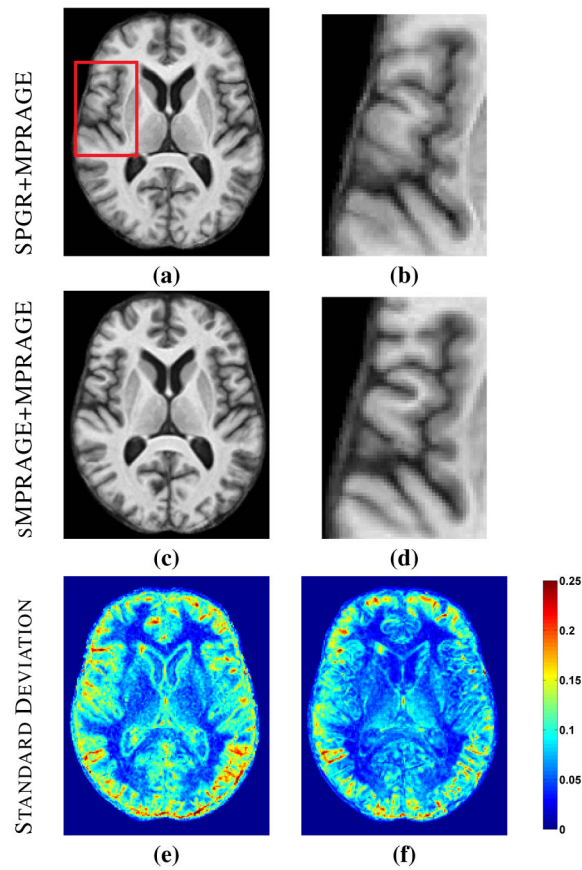


Fig. 6.

(a) Average atlas created using SyN from five BLSA (SPGR) and five OASIS (MPRAGE) images and (b) a zoomed region. (c) Average atlas from five synthetic MPRAGEs from BLSA and five OASIS images (MPRAGE) and (d) a zoomed region. (e) Intensity standard deviation image from SPGR+MPRAGE atlas and (f) intensity standard deviation image from sMPRAGE+MPRAGE atlas.

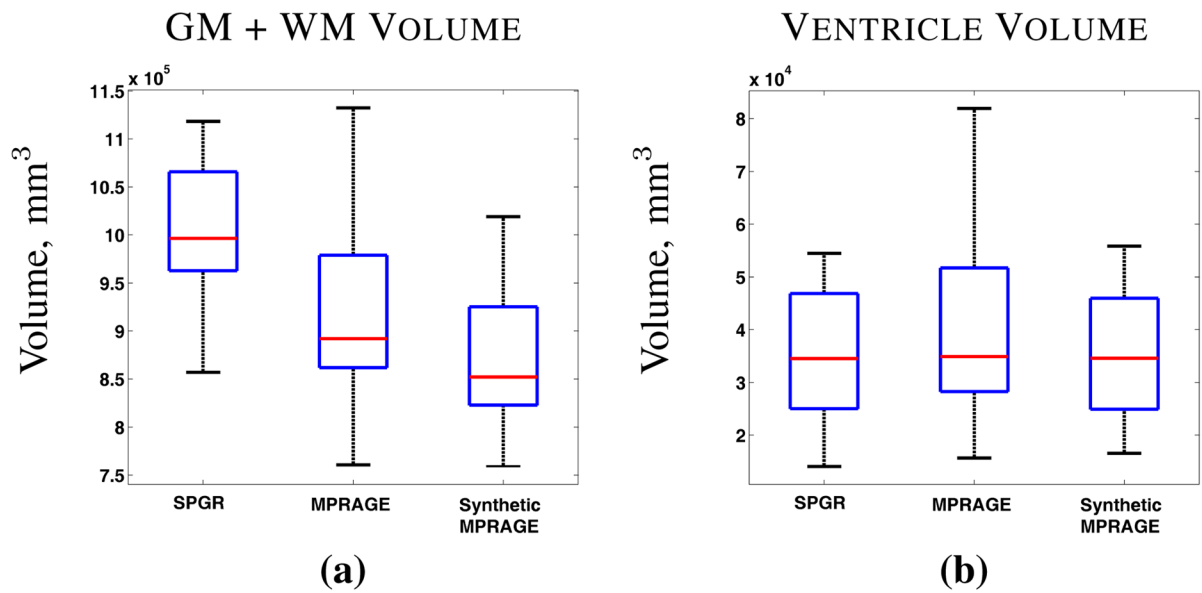


Fig. 7.

(a) Brain volume (GM+WM) and (b) ventricle volume computed using FreeSurfer [74] on 21 normal subjects from the BLSA (SPGR) database and 21 normal subjects from the OASIS (MPRAGE) database, as well as on the 21 synthetic MPRAGEs generated from the BLSA SPGR images.

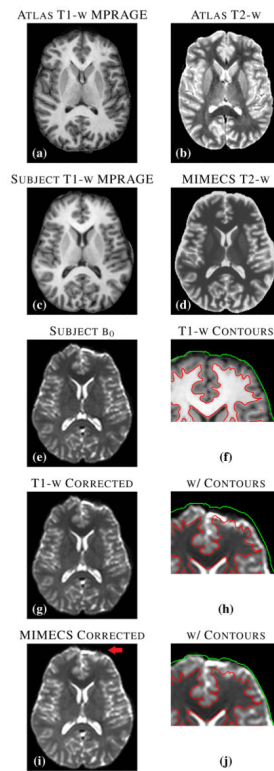


Fig. 8.

(a) and (b) show an atlas pair consisting of MPRAGE and T2-w scans of a normal subject from the Kirby-21 data set. A subject MPRAGE scan (c) is used with MIMECS to synthesize a T2-w image of the subject (d). The acquired subject b_0 image (e) is deformably registered to the subject T1-w image using SyN (with the MI criterion) yielding the corrected image (g). Contours generated from FreeSurfer on the T1-w image (f) are shown on the geometry corrected b_0 image (h). The acquired b_0 image is deformably registered to the synthetic MIMECS image using SyN (CC criterion) to create a MIMECS corrected image (i). Overlaid contours on this image (j) reveal much better geometry correction.

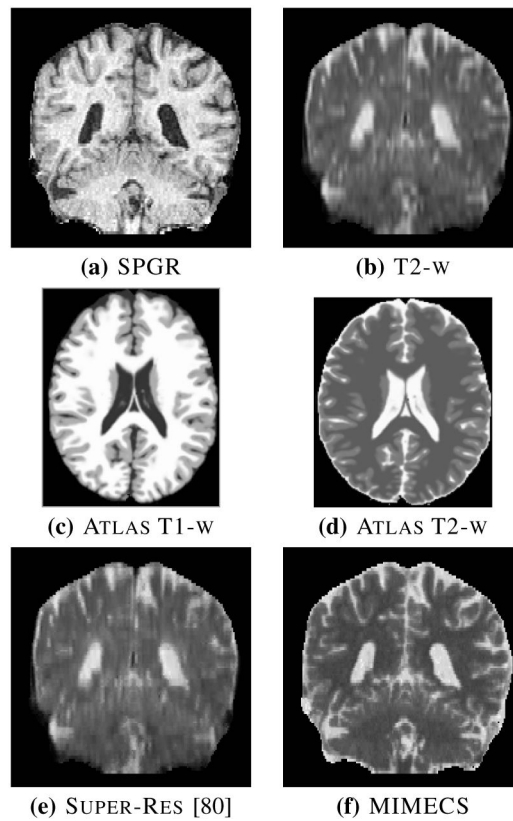


Fig. 9.

Coronal views of (a) SPGR (native resolution $0.94 \times 0.94 \times 1.5\text{mm}$) and (b) T2-w (native resolution $0.94 \times 0.94 \times 5\text{mm}$) scans of a subject. (The T2-w image has been upsampled to the SPGR resolution using trilinear interpolation.) Brainweb (c) T1-w and (d) T2-w phantoms used as an atlas in MIMECS. (e) A hi-res T2-w image upsampled using a non-local super-resolution method. (f) MIMECS synthesized hi-res T2-w image.

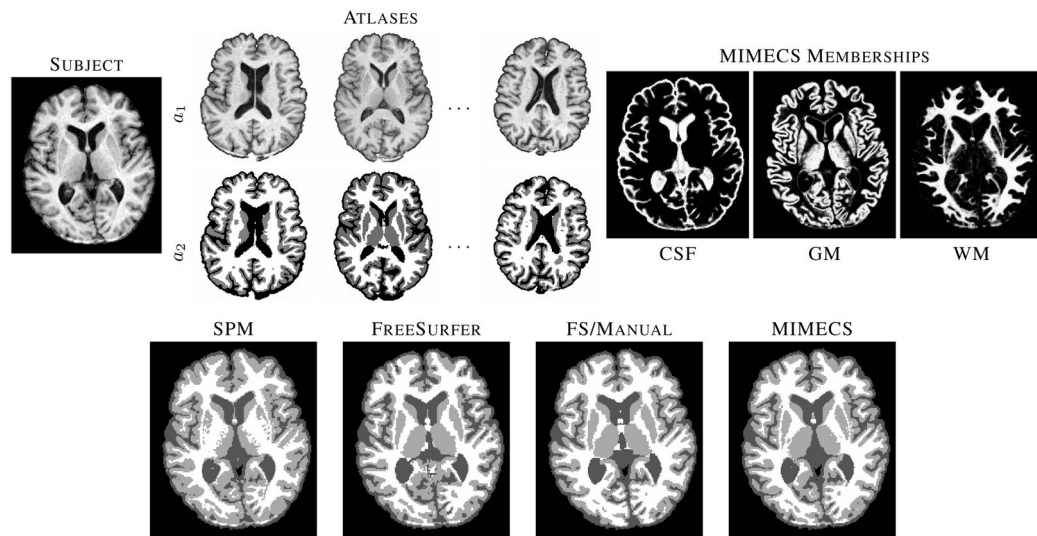


Fig. 10.

The top row shows a subject T1-w SPGR image, three of the five classification atlases, and fuzzy memberships produced by MIMECS-based tissue classification. Hard segmentations from two leading automatic methods and a manually-corrected method are compared to the hard segmentation of MIMECS.

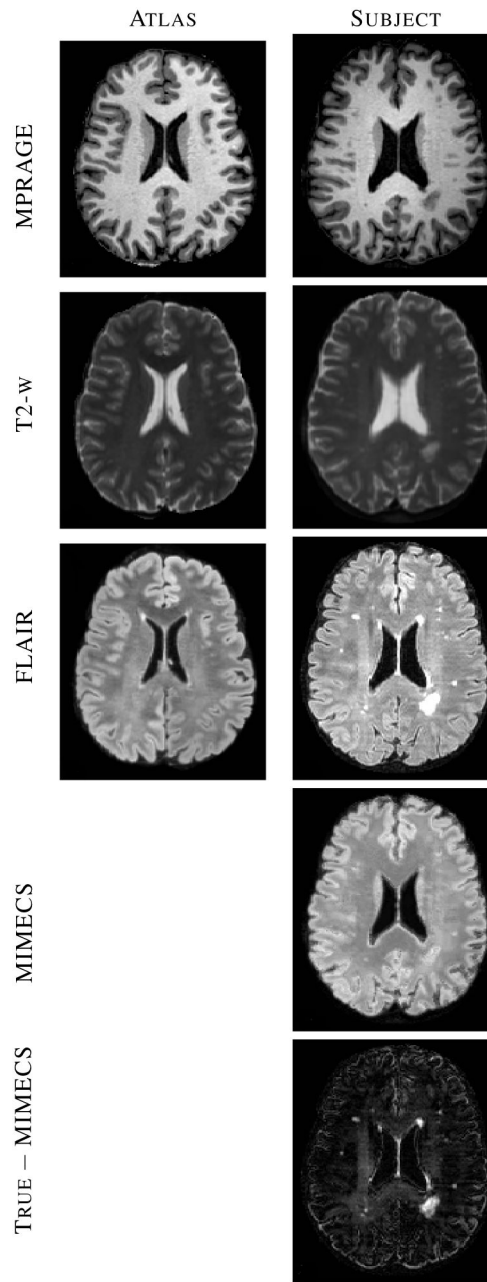


Fig. 11.

The lefthand column contains an atlas for FLAIR synthesis. The righthand column shows the subject's true images, a synthetic MIMECS FLAIR image, and a subtraction image (true FLAIR minus synthetic MIMECS FLAIR).

Fraction of atlas patches used, averaged over all subject patches from a $256 \times 256 \times 199$ image, from each of cerebrospinal fluid (CSF), gray matter (GM), white matter (WM), and the ventricles (VEN).

TABLE I

	Atlas			
	CSF	VEN	GM	WM
CSF	0.71	0.15	0.14	0.00
VEN	0.16	0.78	0.06	0.00
GM	0.10	0.03	0.77	0.10
WM	0.00	0.00	0.23	0.77